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The Culprit of Gout: Triggering Factors and
Formation of Monosodium Urate Monohydrate

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ABSTRACT

Gout is a crystal-induced inflammatory arthritis by the deposition of monosodium urate monohydrate (MSUM). Hyperuricemia is considered as an essential factor for gout formation. However, in vivo MSUM crystallization mechanism is still uncertain, and only some of the patients with hyperuricemia are able to develop gout. Attempts were made to answer those unsolved questions based on the experimental methods developed by Perrin and Swift including (1) morphological studies of MSUM under various Na^+ ion levels, (2) pH effect and conversion from uric acid dihydrate (UAD) to MSUM, and (3) synergistic effect of

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Na\(^+\), K\(^+\) and Ca\(^{2+}\) ions and hyaluronate chains on MSUM crystallization. Various MSUM morphologies such as “beachball”, “urchin-like aggregate” and “bow-like aggregate” could be prepared at different Na\(^+\) ion concentrations at 37\(^\circ\)C.

Consequently, the pathogenesis of gout might be related to the MSUM morphological transformation from “beachball” to needle. The conversion of UAD to MSUM was studied by adjusting the pH value. The determined UAD-to-MSUM pathway was thought to be followed by the MSUM deposition during phagocytosis when lactic acid was present. In addition, a new type of MSUM “fishtail” morphology was observed in the hyaluronate-, Na\(^+\) ions-, and Ca\(^{2+}\) ions-containing solutions. The synergistic effect of hyaluronate and cations on the inhibition of MSUM crystallization was further verified based on the crystal yields. The higher water solubility of the hyaluronate-Ca-urate complex than the one of the urate ion might explain why only a fraction of hyperuricemic patients developed gout. Although there was no reported evidence about the complex, it was probable for it to minimize the risk of gout formation in the hyperuricemic patients. Finally, the puzzle of gout formation was pieced together and illustrated

**Keywords:** Gout, monosodium urate monohydrate (MSUM), hyaluronate, uric acid dihydrate (UAD).
INTRODUCTION

Gout is a common inflammatory arthritis induced by the deposition of monosodium urate monohydrate (MSUM) and might cause serious joint deformity and disability. MSUM originated from uric acid ($pK_a = 5.75$), which is an end-product of purine metabolism.\(^1\)\(^,\)\(^2\) Although MSUM had been identified over 100 years\(^3\) and its molecular structure and solid-state properties had also been well characterized,\(^4\)\(^-\)\(^11\) the prevalence and incidence of gout have continued to rise in the US,\(^12\) UK\(^13\) and Taiwan R.O.C.\(^14\) However, the *in vivo* crystallization mechanism of MSUM is still uncertain to date.

MSUM crystal has a crystallographically triclinic structure.\(^15\) Its crystal habit usually exhibits a needle shape, and rarely a spherical morphology for which the two medical doctors, Fiechtner and Simkin, had dubbed the sphere “beachball”.\(^7\)\(^,\)\(^8\) However, the “bow-like shaped” MSUM needle-clusters were often appeared *in vivo* under the influence of biological substances.\(^9\) Besides, the MSUM clusters were also reported to be produced in a buffer solution exceeding 100 mM of Na\(^+\) ion.\(^16\) The many interesting morphological appearances of MSUM crystals have prompted us to investigate the influence of Na\(^+\) ions on MSUM crystal morphology. Interestingly, the “beachball-like” MSUM morphology was seldom discussed and was regarded as an amorphous form by others.\(^6\) For the biological strategy of making the
mineralized skeletons, a recent study suggested that stable crystals in living organisms are usually developed by the transformation of the first-formed solids which are usually disordered deposits.\(^{17}\) This crystallization strategy is widely discovered in nature such as chiton teeth, mollusks shell and sea urchin spicules and spines.\(^{18}\)

Therefore, we speculate that MSUM “beachballs” may serve as an essential transient precursor during MSUM morphological evolution. Although the MSUM beachballs had been prepared by quenching, the preparation method was completely deviated from the \textit{in vivo} conditions.\(^{6,7}\) Consequently, the first aim of our study is to reproduce MSUM beachballs and its other morphologies under various Na\(^+\) ion levels and to shed some light on the pathogenesis of gout.

Accordingly, there were several factors, which were considered to facilitate the MSUM crystallization: (1) hyperuricemia,\(^9\) (2) temperature,\(^{19-21}\) (3) ionic strength,\(^{4,19,21,22}\) (4) pH,\(^{21-24}\) and (5) promoter/inhibitor molecules.\(^{22,25-27}\) Among these factors, the temperature effect had been used to explain why MSUM is preferred to crystallize out mainly in human joint and connective tissue. This is because the lower local temperatures in joint and connective tissue than in other positions led to the decrease in MSUM solubility. Some cations, biomolecules and proteins could also affect the MSUM precipitation and be regarded as promoters or inhibitors. For example, the presence of K\(^+\) ions would stabilize urate ions in an aqueous solution to
inhibit the crystallization of uric acid and MSUM.

The pH effect is related to inflammatory response\textsuperscript{28} and acidosis.\textsuperscript{29}

Noticeably, the pH-solubility relationships of uric acid and MSUM in Figure S1 established by Wang and Königsberger\textsuperscript{24} had shown that the solubility of MSUM increases as the pH value decreases.\textsuperscript{22,24} However, this unusual observation is against the fact that the decline in pH would promote the MSUM nucleation \textit{in vitro}.\textsuperscript{22}

Moreover, uric acid is quite sensitive to pH fluctuation as reported in the renal system.\textsuperscript{30} There were three identified phases of uric acid, including uric acid anhydrate (UAA), uric acid monohydrate (UAM), and uric acid dihydrate (UAD).

All of them were observed in the renal stone,\textsuperscript{31} and the relationships among UAA, UAD and MSUM were shown in Scheme S1 of the Supplementary Information.

Therefore, the second aim of our study is to investigate the pH effect on MSUM formation and the conversion from uric acid to MSUM in a simulated body fluid (serum) system.

Hyperuricemia is an abnormally high level of uric acid in the blood when the concentration of urate in serum is greater than 6.8 mg/dL. It is a prerequisite for the MSUM deposition, but intriguingly, only a fraction of hyperuricemic subjects suffer from gout disease.\textsuperscript{9} There seems to be physiological substances capable of inhibiting urate crystallization. Individual synovial fluid component such as
hyaluronic acid had no significant effect on the growth rate constant of MSUM. However, the interactions among hyaluronate chains, and Na\(^+\), K\(^+\) and Ca\(^{2+}\) ions which are abundant in synovial fluid, may synergistically influence *in vivo* MSUM crystallization. Therefore, the third aim is to explore the synergistic effect of hyaluronate chains and various ions on MSUM crystallization.

**EXPERIMENTAL SECTION**

All chemicals were used as received without further purification and listed in Table S1.

**Morphological studies of MSUM under various Na\(^+\) ion levels**

The method developed by Perrin and Swift\(^\text{11}\) was modified and then used in our studies. Solutions A and B were first prepared in a 37 °C water bath. 15 mL of aqueous solution A was composed of (1) 13.3 mM of uric acid and (2) 15 mM of NaOH. 5 mL of aqueous solution B was composed of (1) 20 mM of KCl, (2) 8 mM of CaCl\(_2\)-2H\(_2\)O and (3) 400, 2400 or 6000 mM of NaCl, and the resulting solutions were called B1, B2 and B3, respectively. The pH values of those solutions were adjusted to 7.4 by 1 M HCl (aq) or 1 M NaOH (aq). Upon mixing of both solutions A and B, the supersaturated solutions of MSUM containing of (1) 10 mM of uric acid, (2) 5 mM of KCl, (3) 2 mM of CaCl\(_2\)-2H\(_2\)O and (4) 100, 600 or 1500 mM of NaCl, respectively, were produced and were called C1, C2 and C3, respectively (Table 1).
Mixing the pre-dissolved solutions of A and B together was necessary to avoid the occurrence of heterogeneous nucleation induced by undissolved NaCl, KCl and CaCl$_2$·2H$_2$O solids. In a separate experiment, the concentration of NaCl was fixed to 1500 mM and MSUM crystals would be harvested after 2 months. All MSUM crystals were filtered, rinsed by reversible osmosis water, oven dried at 40°C for 24 hr, and characterized by polarized optical microscopy (POM), scanning electron microscopy (SEM), Fourier-transform infrared spectroscopy (FT-IR), powder X-ray diffraction (PXRD), and thermogravimetric analysis (TGA).

Based on the pH-solubility relationship shown in Figure S1, the saturation of MSUM was around 0.26 mM in 150 mM Na$^+$ aqueous solution under the *in vivo* normal condition of pH = 7.4 and 37°C as a reference.

**Effect of pH and conversion from UAD to MSUM**

Presores and Swift had discussed the conversion and transformation of uric acid in urine-like systems$^{33}$ such as McIlvaine buffer and artificial urine solutions.$^{34}$ In this study, we chose the simulated body fluid (SBF)$^{35}$ to mimic synovial fluid instead. SBF comprising 92.45 mM of NaCl, 8.81 mM of NaHCO$_3$, 19.3 mM of Na$_2$CO$_3$, 3.02 mM of KCl, 1.01 mM of K$_2$HPO$_4$·3H$_2$O, 1.50 mM of MgCl$_2$, 0.5 mM of Na$_2$SO$_4$, and 50 mM of HEPES was prepared accordingly.$^{35}$ The pH value of SBF was adjusted to 7.4 by 1 M NaOH (aq).
The very same procedure in the section of “Morphological studies of MSUM under various Na\(^+\) ion levels” would be followed except for the Na\(^+\) ion concentration which was fixed to 140 mM for simulating the \textit{in vivo} condition.\(^{11}\) Instead of using 1 M HCl (aq), about 10 mM aqueous solution of lactic acid\(^{26}\) was chosen as an acidosis factor\(^{28}\) to lower the pH value to 5.0 to 5.8. Lowering the pH to an extreme value of 5.0 was to accelerate the UAD formation within a reasonable time frame.

UAD solids were isolated after a few minutes, filtered, air dried for 10 min and stored at -4 \(^\circ\)C to prevent the dehydration of UAD to UAA.\(^{37}\) About 30 mg of UAD solids were dispersed in 3 mL of SBF at 37\(^\circ\)C.\(^{35}\) Solids were harvested for every 12-hr in a total period of 36 hr, filtered, rinsed with RO water, air dried and characterized immediately by POM and PXRD.

\textbf{Synergistic effect of Na\(^+\), K\(^+\) and Ca\(^{2+}\) ions, and hyaluronate chains on MSUM crystallization}

Since the concentration of hyaluronate in normal joints ranges from 1.45 to 2.94 mg/mL,\(^{38}\) 5 mL of aqueous solution D1 was composed of (1) 560 mM of NaCl and (2) 40 mg of sodium hyaluronate. 5 mL of aqueous solution D2 was composed of (1) 560 mM of NaCl, (2) 20 mM of KCl, and (3) 40 mg of sodium hyaluronate. 5 mL of aqueous solution D3 was composed of (1) 560 mM of NaCl, (2) 8 mM of CaCl\(_2\)·2H\(_2\)O, and (3) 40 mg of sodium hyaluronate. 5 mL of aqueous solution D4...
was composed of (1) 560 mM of NaCl, (2) 20 mM of KCl, (3) 8 mM of CaCl$_2$·2H$_2$O, and (4) 40 mg of sodium hyaluronate. All aqueous solutions were prepared at 37°C and their pH values were adjusted to 7.4 by 1M HCl (aq) or NaOH (aq). The supersaturated solutions of MSUM, called E1, E2, E3 and E4, were prepared by mixing solutions A and D1, D2, D3 and D4, respectively, and their compositions were provided in Table 2. MSUM crystals would be harvested in 5 days, filtered, rinsed by RO water, oven dried at 40°C for 24 hrs and characterized by POM and PXRD.

The constant composition method$^{39}$ developed by Nancollas et al. could overcome the problems associated with the dynamically changing solution composition during precipitation for many crystallization and mineralization kinetics approaches. The concentrations of lattice ions were maintained constant by simultaneous addition of reagent solutions and controlled by a glass electrode probe. However, in our experiments, only the pH values of all the above-mentioned solutions were checked to be around 7.4 by a pH meter due to its simplicity and robustness.

RESULTS AND DISCUSSION

Morphological studies of MSUM under various Na$^+$ ion levels

POM images in Figure 1 showed that MSUM crystals grown under various Na$^+$ ion levels at 37°C were crystalline because of the strong birefringence. 200-300 µm radiating bow-like aggregates, 20-50 µm urchin-like aggregates, and 20-60 µm
“beachballs” were obtained under 100 mM, 600 mM, and 1500 mM Na\textsuperscript{+} ion levels, respectively. It took two months for “beachballs” produced from 1500 mM solution of Na\textsuperscript{+} ion to gradually evolve into “urchin-like” and “bow-like aggregates” (Figure 1d). The SEM images in Figure 2 showed that the “beachball” was composed of many 1-2 µm-sized platelets filled with primary nanometer-sized needles. All solids prepared at pH of 7.4 could guarantee that neither UAD nor UAA was formed by FTIR spectra (Figure S2). All of MSUM crystals were verified according to the characteristic IR peak assignments given in Table S2. The relatively broad peaks with lower intensity in PXRD pattern (Figure S3) further confirmed that “beachball” was composed of nanometer-sized crystallites with poor crystallinity based on Scherrer’s formula.\textsuperscript{40,41} In TGA scans (Figure S4), the first weight losses for both of “bow-like aggregates” and “beachball” were about 7.2 % approximating to the weight percent of hydrate within the crystal lattice of MSUM. However, the dehydration temperature for “beachball” was around 100°C, which was lower than the one for “bow-like” at 150°C. It might be attributed to the relatively large surface area of MSUM beachball for accelerating the dehydration process. They began to degrade when heated to over 300°C. Our in vitro experiments suggested that “beachball”, “urchin-like aggregate” and “bow-like aggregate” were related to the Na\textsuperscript{+} ion levels (Figure 1) and not just
only to the biological\textsuperscript{9} and/or macromolecular\textsuperscript{42} components in synovial fluid as

claimed by others.\textsuperscript{9,42} All MSUM morphologies were mutually related to one

another because the first-formed metastable “beachballs” could transform to

“urchin-like aggregate” and then to “bow-like shaped” clusters or needle-shaped

crystals in solution if given long enough time. According to SEM images in Figure

2, we proposed that MSUM “beachball” might be a mesocrystal having a hierarchical

structure\textsuperscript{43} via an aggregation-mediated pathway by nano-clusters instead of merely

amplification in crystal growth.\textsuperscript{44} However, more studies for the exact pathway

would still be needed in the future.

Since “beachballs” were only discovered in a hypertensive female patient who

also required weekly peritoneal dialysis,\textsuperscript{8} it was speculated that the local fluctuation

of \( \text{Na}^{+} \) ion level or other conditions raising MSUM supersaturation level in her

synovial fluid might play a dominant role in “beachball” formation. The

pathogenesis of gout may be related to the MSUM morphological transformation

from “beachball” to needle.\textsuperscript{8}

Effect of pH and conversion from UAD to MSUM

Within a few minutes upon the addition of lactic acid, the pH was first reduced

to 5.0 and then increased to 5.8 gradually as UAD plates were being precipitated out.

Lowering the pH value to 5.0 was to accelerate the UAD crystallization, and UAD
crystals would precipitate after the pH value was lowered to 5.5. UAD plates were then isolated and converted in a freshly prepared SBF environment to induce the transformation from UAD to MSUM. The POM images and PXRD patterns for UAA, UAD and MSUM also were provided in Figure S5 for comparison. Based on the crystal habits, the POM images in Figure 3 showed that the integrity of UAD plates was partially maintained during dissolution in the first 12 hr. The UAD plates were bristled and covered with many tiny crystals 24 hr later. UAD plates were disappeared after 36 hr and needle-shaped MSUM crystals were found 48 hr later. PXRD patterns in Figure 4 illustrated the conversion of UAD to MSUM thoroughly. Intense diffraction lines for UAD were at 20 = 10° and 20°, for UAA was at 20 = 13.5°, and for MSUM were at 20 = 11.6°, 18.9° and 28.3°. The PXRD patterns in Figures 3(d) and 4(d) also revealed that UAD took 36 hr to convert to MSUM, and UAA was an intermediate phase coexisting with UAD in 24 hr (Figure 4(c)). Finally, MSUM needles would be grown completely after 48 hr later (Figures 3(e) and 4(e)). TGA scans of UAD and UAA were shown in Figure S6.

Under the normal physiological Na⁺ ion concentration of approximately 140 mM, the dominant MSUM morphology was “bow-like aggregates” (Figure 1) which were strikingly similar to the native crystals formed in cartilage. As for the gout formation mechanism in humans, a fundamental question still remains: How does the
MSUM crystal appear initially? Most likely, the triggering factor is the local pH reduction in connective tissues and joints\textsuperscript{25} brought about by the inflammatory reaction\textsuperscript{28} and acidosis.\textsuperscript{29} The sudden decline of local pH did favor the rapid formation of UAD plates more than MSUM crystals according to their solubility behaviors.\textsuperscript{24,37} However, as the pH was returned to about 7.4,\textsuperscript{47} the conversion from UAD to MSUM would be activated and finished after 36 hr (Figure 4). Therefore, UAD crystals are likely to be the triggering factor for \textit{in vivo} gout formation.

This UAD-to-MSUM conversion could offer a more reasonable time frame in human life for gout deposition by acidosis than the time required for direct MSUM crystallization. The growth rate of MSUM crystal at its favorable pH\textsuperscript{23} of 7.0 and a urate concentration of 0.6 mM (i.e. > 6.8 mg/dL, hyperuricemia\textsuperscript{1}) was calculated to be only $10^{-11} \mu$m/min.\textsuperscript{48} However, UAD crystals could precipitate immediately with the temporary decline of local pH value, and UAD would then convert to MSUM at a normal pH value (pH = 7.4) during a relatively short time. The UAD-to-MSUM pathway offered an explanation to the appearance of MSUM crystals in cartilage, synovial and tendon sheaths\textsuperscript{25} because of the pH gradient between human plasma and tissue.\textsuperscript{28} Furthermore, acute gouty arthritis would develop when MSUM needles were ingested by leucocytes. During phagocytosis, the production of lactic acid would further provoke MSUM deposition according to the UAD-to-MSUM pathway.
This self-sustaining cycle provided a stimulus to a greater inflammatory response.\textsuperscript{28}

**Synergistic effect of Na\textsuperscript{+}, K\textsuperscript{+} and Ca\textsuperscript{2+} ions, and hyaluronate chains on MSUM crystallization**

Solids grown in aqueous solutions with sodium hyaluronate and various cations were identified to be MSUM by PXRD (Figure S7) and the crystal yield for each condition was weighted. Because hyaluronate chain would interact with Na\textsuperscript{+}, K\textsuperscript{+} and Ca\textsuperscript{2+} ions,\textsuperscript{32} we broke down the study of those interactions among hyaluronate and cations into four parts. POM images in Figure 5 showed that the number of MSUM crystals decreased as more types of cations were added into the hyaluronate-containing solutions. A new type of MSUM “fishtail” morphology was observed in the hyaluronate-, Na\textsuperscript{+} ions-, and Ca\textsuperscript{2+} ions-containing solutions (Figure 5).

The synergistic effect of hyaluronate and cations on the inhibition of MSUM crystallization was further verified based on the crystal yields (Table S3). SBF with the coexistence of physiological concentrations of hyaluronate, Na\textsuperscript{+}, K\textsuperscript{+} and Ca\textsuperscript{2+} ions gave the minimum MSUM crystal yield of 7.2 wt % which was at least six times less than the crystal yield of 47.3 % from the SBF having only hyaluronate and Na\textsuperscript{+} ions.

The saturation values of MSUM in solutions of E1 to E4 were determined and summarized in Table S3.

Moreover, Ca\textsuperscript{2+} ion was normally believed to reduce the solubility of MSUM,
and the initial nucleus for gout was thought to be calcium urate.\textsuperscript{22,49} However, our findings of the synergistic effect of hyaluronate chains and Ca\textsuperscript{2+} ions and even with Na\textsuperscript{+} and K\textsuperscript{+} ions have shown the otherwise.\textsuperscript{32} The dominant role of Ca\textsuperscript{2+} ions might have been a crystal inhibitor for MSUM deposition instead (Table S3). Since hyaluronate chains contain hydrophilic hydroxyl (–OH) and carboxylate (–COO\textsuperscript{−}) pendent groups, hyaluronate chains are water soluble. With the presence of Ca\textsuperscript{2+} ions and other cations acting as salt bridges between the –COOH\textsuperscript{−} groups of hyaluronate chains and the N3 (pKa = 5.75) sites\textsuperscript{1,50} on the six-member ring of the urate ion, the slightly soluble urate ion has now become a solubilized complex in water thanks to the high solubility power of hyaluronate. The higher water solubility of the hyaluronate-Ca-urate complex than the one of the urate ion might be one of the reasons for only a fraction of hyperuricemic patients to develop gout. Although there was no reported evidence about the complex, it was probable for it to minimize the risk of gout formation in the hyperuricemic patients. If the complex was disrupted, MSUM deposition would occur.

Furthermore, the interactions among hyaluronate chains, Na\textsuperscript{+} and Ca\textsuperscript{2+} ions produced fishtail MSUM clusters (Figure 5). Interestingly, the fishtail clusters had never been reported in gouty patients. Fishtail clusters should have appeared if hyaluronate, Na\textsuperscript{+} ion and Ca\textsuperscript{2+} ion were all abundant in gouty patient’s synovial fluid.
This implied that the decrease in production, polymerization, molecular size and concentration of hyaluronate chains during inflammation\(^{38}\) had prevented the formation of hyaluronate-Ca-urate complex, and thus, only the usual bow-like MSUM aggregates were grown.

Finally, our hypothesis of gout formation was illustrated in Figure 6. Local pH decline and the disruption of hyaluronate-Ca\(^{2+}\)-urate complex are the triggering factors for gout formation. Local pH decline could induce UAD crystals formation at first, and UAD would convert to MSUM after the return of the pH value. The dissolution of UAD may promote MSUM crystallization due to the increase of local urate level. The rise of the concentration Na\(^+\) ions (or the increase of the urate ions level) could increase the degree of supersaturation, and this driving force may promote the “beachball” formation. In addition, MSUM crystals would induce acute gouty arthritis due to leucocytes, and this could decrease the local pH again. This self-sustaining vicious cycle provided a stimulus to an even greater inflammatory response.

**CONCLUSIONS**

Seven key lessons were learned in this study: (1) Various morphologies of MSUM: “beachball”, urchin-like and bow-like aggregates could be prepared under different Na\(^+\) ion levels at 37\(^\circ\)C. (2) Morphological transformation from “beachball”
to urchin-like aggregate to bow-like aggregate can be achieved. (3) “Beachball” is a metastable first-formed precursor for all MSUM morphologies. (4) A new type of MSUM fishtail morphology was observed in the hyaluronate, Na\(^+\), Ca\(^{2+}\) ion containing solutions. (5) The dilemma of MSUM deposition at thermodynamically unfavorable low pH condition was plausibly solved by the kinetic pathway for the phase transformation of UAD to MSUM in the gout study. (6) The UAD-to-MSUM pathway can also be used to explain the effects brought about by the inflammatory response and acidosis, and why MSUM deposition in cartilage and connective tissue. (7) The question of why only a fraction of hyperuricemic patients has gout might be answered by the solubilization of urate through complex formation with hyaluronate chains and Ca\(^{2+}\) ions.

13 **ACKNOWLEDGMENTS**

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FIGURE LEGENDS

Figure 1. POM images of MSUM grown: (a) in 100 mM Na\(^+\) ion for 2 days, (b) in 600 mM Na\(^+\) ion for 2 days, (c) in 1500 mM Na\(^+\) ion for 2 days, and (d) in 1500 mM Na\(^+\) ion after 2 months (N: needle; BLA: bow-like aggregate; ULA: urchin-like aggregate; BB: beachball).
Figure 2. SEM images of a MSUM beachball grown in 1400 mM Na$^+$ ion for 2 days at various magnifications: (a) a full view of “beachball”, (b) oriented platelets, and (c) primary nanometer-sized needles.
Figure 3. POM images during the conversion of UAD to MSUM: (a) initial UAD plates, (b) UAD plates with partially dissolved fragments at 12 hr (D-UAD: dissolved UAD fragments), (c) UAD plates covered with MSUM tiny crystals at 24 hr, (d) aggregation of MSUM tiny needles at 36 hr, (e) well-grown MSUM needles at 48 hr (N: needle; BLA: bow-like aggregate), (f) view of crystal structure of UAD showing the top view along the b-axis, constructed from fractional coordinates in refcode: ZZZPPI02, (g) view of crystal structure of UAA showing the top view along the c-axis, constructed from fractional coordinates in refcode: URICAC, and (h) view of crystal structure of MSUM showing the top view along the c-axis, constructed from fractional coordinates in refcode: NAURAT.
Figure 4. PXRD patterns of solids harvested from SBF during the conversion of UAD to MSUM at (a) 0 hr (UAD), (b) 12 hr (UAD), (c) 24 hr (UAD and UAA), (d) 36 hr (MSUM), and (e) 48 hr (MSUM).
Figure 5. POM images of MSUM crystals grown in solutions: (a) E1: 2 mg/mL of sodium hyaluronate and 140 mM of Na$^+$ ion, (b) E2: 2 mg/mL of sodium hyaluronate, 140 mM of Na$^+$ ion and 5 mM of K$^+$ ion, (c) E3: 2 mg/mL of sodium hyaluronate, 140 mM of Na$^+$ ion and 2 mM of Ca$^{2+}$ ion, (d) E4: 2 mg/mL of sodium hyaluronate, 140 mM of Na$^+$ ion, 5 mM of K$^+$ ion and 2 mM of Ca$^{2+}$ ion (N: needle; BLA: bow-like aggregate; FT: fishtail).
Figure 6. Triggering factors, proposed mechanism and self-sustaining cycle for the crystallization of MSUM and gout.
TABLE LEGENDS

Table 1. The compositions and conditions of supersaturated MSUM solutions under the various Na$^+$ ion levels for MSUM morphological studies.

<table>
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<th>Solution C1</th>
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Table 2. The compositions and conditions of supersaturated MSUM solutions for synergistic effect of Ca\(^{2+}\), K\(^{+}\) and Na\(^{+}\) ions and hyaluronate chains on MSUM crystallization.

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REFERENCES

1 F. Oliviero, A. Scanu, L. Punzi., Reumatismo, 2011, 63, 221-229.


34 T. C. McIlvaine, *J. Biol. Chem.*, 1921, **49**, 183-186.


The Culprit of Gout: Triggering Factors and Formation of Monosodium Urate Monohydrate

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Triggering factors, proposed mechanism and self-sustaining cycle for the crystallization of MSUM and gout.

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