# ChemComm

# Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](http://www.rsc.org/Publishing/Journals/guidelines/AuthorGuidelines/JournalPolicy/accepted_manuscripts.asp).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](http://www.rsc.org/help/termsconditions.asp) and the Ethical quidelines still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemcomm

## **Journal Name**

# **COMMUNICATION**



Accepted 00th January 20xx DOI: 10.1039/x0xx00000x

Received 00th January 20xx,

### Dewey A. Sutton,<sup>a</sup> Seok-Ho Yu,<sup>b</sup> Richard Steet,<sup>b</sup> and Vladimir V. Popik<sup>a\*</sup>

**Cyclopropenone-caged Sondheimer Diyne (Dibenzo[a,e]cy-**

**clooctadiyne): A Photoactivatable Linchpin for Efficient SPAAC** 

**www.rsc.org/**

**The first fully conjugated bis-cyclopropenone (photo-DIBOD), a derivative of dibenzo[***a,e***][8]annulene, has been synthesized. 350-420 nm irradiation of this robust compounds results in the efficient formation of dibenzo [***a, e***] cyclooctadiyne, an unstable, but useful SPAAC cross-linking reagent. Since photo-DIBO doesn't react with organic azides, this method allows for the spatiotemporal control of the ligation of two azide-tagged substrates.**

**Crosslinking** 

In recent years the copper (I) - catalyzed 1,3-dipolar cycloaddition of azides to terminal acetylenes  $(CuAAC)$ , as well as the strainpromoted, catalyst – free version of this reaction  $(SPAAC)$ , have become very popular tools for the functionalization, cross-linking, and immobilization of various substrates.<sup>3</sup> These quintessential "click-chemistries" have found numerous applications in molecular biology,<sup>[3b](#page-4-0)-c,4</sup> drug development,<sup>5</sup> biotechnology,<sup>6</sup> and materials science.<sup>3a,7</sup> The SPAAC ligation is better suited for applications in living systems as it avoids the use of cytotoxic copper catalysts.<sup>8</sup> In addition, terminal acetylenes were found to inhibit cysteine proteases by forming thioethers with the catalytically active thiol.<sup>9</sup> The utility of the SPAAC technique, on the other hand, is somewhat restricted by the requirement for the derivatization of the coupling partners with two different functionalities. While introduction of an azide group is well developed for a large variety of substrates, the incorporation of cyclooctyne moieties often requires multiple synthetic steps. A limited number of prefunctionalized SPAAC reagents are available commercially, but are rather expensive. Alternatively, two azide-derivatized substrates can be ligated using a bifunctional linker. $10,11$ 

Dibenzo[*a,e*]cyclooctadiyne (DIBOD **2a**, a.k.a. Sondheimer diyne), represents a unique cross-linking platform in this sense, as it contains two azide-reactive strained triple bonds within the same cyclic structure and allows for double-SPAAC click ligation of azide-tagged substrates.<sup>12</sup> Diyne **2a** has been employed for the cross-coupling of various azide-functionalized substrates from biomolecules to metal – organic frameworks.<sup>12,13</sup> The utility of this method, however, is limited by the low stability of the Sondheimer diyne. In the neat form **2a** completely decomposes within two days and exhibits half lifetime of only 10 min in a neutral aqueous solution (1mM in PBS, pH 7.4, r.t.).<sup>14</sup>

To address this deficiency of the DIBOD cross - linker, we envisaged protecting (or "caging") triple bonds in the molecul with a photolabile functionality. Photochemical decarbonylation cyclopropenones is one of the most efficient methods for the generation of reactive acetylenes, as it is characterized by very fast rate, quantitative chemical and high quantum yields.<sup>15</sup> This reaction can be induced by NIR light under two-photon excitation conditions<sup>16</sup> and proceeds smoothly in the solid state.<sup>17</sup> We have already employed this reaction for the generation of reactive triple bonds in ynols, $18$  enediynes, $19$  and dibenzocyclooctynes. Irradiation of bis-cyclopropenone **1** (photo-DIBOD) results in the formation of the Sondheimer diyne 2, which, in turn reacts wi<sup>1</sup> two azide-functionalized substrates to create a stable covalent cross-link (Scheme 1). It is important to note that the addition second equivalent of azide to the diyne 2a proceeds at much high-rate than the first cycloaddition.<sup>[12a](#page-4-1)</sup>



**Scheme 1.**

Photochemical generation of cross-linker 2 from azide-vert precursor **1** is not only reagent-free, but also allows for the spatiotemporal control of the ligation process. This reaction expands  $t_{\parallel}$ . family of recently developed "photo-click" ligation methods, $21$  including cycloaddition of alkenes to photochemically generated nitrile imines, photo-initiated thiol-ene, $^{23}$  and thiol-yne<sup>24</sup> reactions, light-induced Diels Alder,<sup>25</sup> as well as photo-Michael reaction.<sup>26</sup> Additionally, phot DIBOD (**1**) is an interesting molecule in its own right, representing the first known fully conjugated bis-cyclopropenone. Despite the presence of the cyclooctatetraene core and the obvious angle strain, this

*a.Department of Chemistry, University of Georgia, Athens, GA 30602*

*b. Complex Carbohydrate Research Center, University of Georgia, Athens, GA 30602* Electronic Supplementary Information (ESI) available: synthetic procedures and spectral data for the new compounds. See DOI: 10.1039/x0xx00000x

compound is surprisingly stable in the solid state and in solutions (*vide infra*).

We envisioned the preparation of dibenzo $[a,e]$ dicyclopropa $[c,g]$ [*8*]annulene-1,6-dione (**1a**) and its derivatives by the double addition of dihalocarbene across the triple bonds in dibenzocyclooctyne **2** followed by the controlled hydrolysis. To address the limited solubility of the parent diyne **2a**, we have also synthesized 2,8-dibutoxy- (**2b**) and 2,8 bis-[tetra(ethylene glycol)]- (**2c**) substituted analogs. Diynes **2a-c** were prepared using the modified method of Otera and coworkers.<sup>[14,2](#page-4-2)7</sup> Standard protocols for the generation of difluorocarbene require elevated temperature and harsh conditions $^{28}$  that are not compatible with the highly sensitive diynes **2a-c**. The milder conditions of the recently developed Hu-Prakash reaction<sup>29</sup> allow for the efficient formation of difluorocarbene without significant decomposition of the substrate. Treatment of diynes  $2a,b,d$  with TMSCF<sub>3</sub> resulted in the formation of the bis-difluorocyclopropenes **4a**,**b**,**d**, which were subsequently hydrolyzed on wet silica gel to afford the target bis-cyclopropenones **1ac** (Scheme 2). The tetra(ethylene glycol) substituents in **2c** were capped with acetyl groups prior to cyclopropenation. The ester protection was later removed by base-promoted hydrolysis (Scheme 2).<sup>1</sup>



Scheme 2. Synthesis of photo-DIBOD *Reagents and conditions: i*) TMSCF<sub>3</sub>, NaI, 110°C, 4a: 60%, 4b: 80%, 4c: 25%; *ii*) wet silica gel, 1a: 81%, 1b: 73%; *iii*) K<sub>2</sub>CO<sub>3</sub>, MeOH (**1c**: 53% over 2 steps, *ii* and *iii*)

Bis-cyclopropenones **1a-c** are bright yellow high-melting crystalline compounds<sup>[14](#page-4-2)</sup> that have a long shelf life if stored in the dark. The spectrum of photo-DIBOD **1a** in the UVA region (Fig. 1) contains two close-lying bands at 342 (log  $\varepsilon$ =4.0) and 361 nm (log  $\varepsilon$ = 4.1), as well as two intense bands at shorter wavelengths (260 and 268 nm, log  $\varepsilon$  – 4.8). Two weaker bands in the visible region (409 nm, log  $\varepsilon$  = 2.9; 438 nm,  $log \varepsilon = 2.6$ ) are responsible for the color of these compounds (Fig. 1). Alkoxy-substitution in **1b** and **1c** results in the 17 nm red shift of the UVA bands, which is accompanied by a moderate hyperchromic effect (360 nm, log ε ~ 4.5, 378 nm, log ε~ 4.4). Photo-DIBODs **1a-c** do not react with organic azides under ambient conditions. Irradiation of a methanol solution of photo-DIBO **1a** with 350 nm light in the presence of butyl azide results in efficient decomposition of the starting material  $(\Phi=0.05)$ , which could be observed by the bleaching of the 342-361 nm bands. An isomeric mixture of head-to-tail (**5**) and head-to-head (**6**) bistriazole adducts was isolated from the photolysate in 77% preparative yield (Fig. 1).

Interestingly, while the absorbance bands at 409 and 438 nm have relatively low intensity, irradiation of photo-DIBOD **1a** with 420 nm fluorescent tubes in the presences of butyl azide produced the same mixture of triazoles **5** and **6**. Complete conversion, however, requires longer irradiation times than with 350 nm lamps. Sensitivity to the violet light makes the photo-DIBOD cross-linking procedure compatible with the majority of modern confocal fluorescent microscopes.

Since photo-DIBOD derivatives **1a-c** have very low aqueous solubility, we decided to explore the feasibility of conducting photocrosslinking experiments employing nanocrystalline suspension of photo-DIBOD. Garcia-Garibay has previously reported that irradiation of the nanocrystalline suspension of diphenylcyclopropenone results the efficient decarbonylation of the substrate.<sup>[17](#page-4-3)</sup> In fact, the quantum yie<sup>ld</sup> of the process in the solid state is about 4 times higher than in solution due to "quantum chain reaction."<sup>[17,3](#page-4-3)0</sup> The nanocrystalline suspension of photo-DIBOD **1a** was prepared using reprecipitation techniques.<sup>17</sup> methanol solution of the substrate was slowly added to an aqueous PP buffer containing 15 mM of SDS under sonication. Dynamic light scattering (DLS) studies performed on a photo-DIBOD suspension revealed a mean distribution of particles at  $572\pm130$  nm.<sup>[14](#page-4-2)</sup> The aqueo suspension of photo-DIBOD is chemically stable in the dark. While the intensity of the absorbance bands of **1a** slowly decreases with time, this observation can be attributed to the precipitation of the substrate. Even after 48 h, full absorbance value was restored by sonication of  $t \bar{t}$ . mixture for 15 min. In accordance with previously reported observations,<sup>[17,](#page-4-3)[30](#page-4-4)</sup> the efficiency of photodecarbonylation of 1a show dramatic enhancement in crystalline state ( $\Phi = 0.41 \pm 0.8$ ).



**Figure 1.** UV spectra of ca. 80  $\mu$ M solutions of *photo-DIBOD* (1a, solid line) and a mixture of bis-triazoles (5 and 6, dashed line), as well as  $0.78$  mM solution  $p^2$ DIBOD (1a, dash-dotted line) in MeOH-CH<sub>2</sub>Cl<sub>2</sub> (9:1).

To demonstrate the utility of the photo-DIBOD crosslinker for biochemical ligations, we have derivatized a model protein, azide-tagged bovine serum albumin (azido-BSA), with a variety of function... moieties. The latter was prepared by treating the commercially availab' BSA with 1-azido-3-iodopropane in TRIS buffer.<sup>[14](#page-4-2)</sup> Fluorescent labelin of azido-BSA was achieved by irradiation of 0.4  $\mu$ M PBS solution of the protein containing  $4 \mu M$  of the nanocrystalline suspension of **1a** and  $4 \mu M$  $\mu$ M of Rhodamine B azide for 8 min using 350 nm fluorescent tub (Scheme 3).



**Scheme 3.** Photo-labeling of azido-BSA

#### **Journal Name COMMUNICATION**

The SDS-PAGE analysis of the photolysate, conducted after overnight incubation in the dark, clearly shows the efficient crossconjugation of azido-BSA with fluorescent dye (Fig. 2). Incubation of the same mixture in the dark overnight, as well as irradiation of azido-BSA – Rhodamine-azide mixture, does not produce labelled BSA **7a**. As a positive control experiment, we have incubated azido-BSA – Rhodamine-azide mixture with freshly prepared Sondheimer diyne **2a**. As the gel image illustrates, the photo-DIBOD conjugation is at least as efficient as the use of dibenzocyclooctadiyne **2a** (Fig. 2).



**Figure 2.** SDS-PAGE analysis of BSA - Rhodamine B conjugation. Lane 1: photo-DIBOD **1a**, azido-BSA and Rhodamine B - azide irradiated; Lane 2: **2a**, azido-BSA, and Rhodamine B - azide incubated overnight; Lane 3: photo-DIBOD **1a**, azido-BSA, and Rhodamine B - azide incubated overnight in the dark. Lane 4: azido-BSA and Rhodamine B - azide incubated overnight.

Photo-biotinylation of azido-BSA has been conducted following a similar procedure. A PBS solution of azido-BSA  $(0.4 \mu M)$ , biotin-azide (4  $\mu$ M), and nanocrystalline suspension of **1a** (4  $\mu$ M) was irradiated with 350 nm light for 5 min and incubated overnight. The protein was separated from low-molecular weight compounds by spin-filtration and further purified by size-exclusion chromatography. Western blot - antibiotin-HRP analysis demonstrates the efficient photo-biotinylation of azido-BSA (Fig. 3). $^{14}$  $^{14}$  $^{14}$  As in the previous experiment, azido-BSA – biotin-azide cross-linking using Sondheimer diyne **2a** in the dark has been employed as a positive control.

The labeling of BSA with Rhodamine B and biotin demonstrated the utility of photo-DIBOD for protein functionalization with relatively small moieties. To test the capability of this platform for the conjugation of larger substrates, we conducted the cross-linking of azido-BSA with PEG<sub>5000</sub>-azide<sup>31</sup> (Scheme 3). A PBS solution of azido-BSA (0.4  $\mu$ M), PEG<sub>5000</sub>-azide (4  $\mu$ M), and nanocrystalline suspension of **1a** (4  $\mu$ M) was irradiated with 350 nm light for 8 min and incubated overnight. The functionalized protein (**7c**) was concentrated via spin filtration (MWCO 10,000), purified on a Sephadex column, and lyophilized. Surprisingly, the MALDI-TOF mass spectrum revealed a mixture of BSA derivatives with various degrees of functionalization (Fig.  $S1$ ).<sup>[14](#page-4-2)</sup> To rule out the possibility of non-specific reaction of photo-DIBOD **1a** or of photogenerated **2a** with the protein, we have conducted copper-catalyzed

"click" conjugation of azido-BSA with  $PEG<sub>5000</sub>$ -acetylene. Gratifyingly, the MALDI-TOF spectrum of the resulting product show the same distribution of PEGylated BSA derivatives as in the phot-conjugation experiment (Fig. S2).<sup>[14](#page-4-2)</sup> These experiments demonstrate that the photo-derivatization of proteins with **1a** occurs only at azide moieties and is as efficient as CuAAC ligations. Incubation of azido-BSA, photo-DIBOD, and  $PEG<sub>5000</sub>$ -azide in the dark does not produce any BS/ PEG<sub>5000</sub> conjugates.<sup>[14](#page-4-2)</sup>



Figure 3. Western blot analysis of azido-BSA – Biotin-azide conjugation using different protein loading: lanes 1-5, 2  $\mu$ g per lane and lanes 6-7, 2  $\mu$ g per lane. Lanes 2,7: photo-DIBOD **1a**, azido-BSA and Biotin-azide irradiated; Lanes 3.8: **2a**, azido-BSA, and Biotin-azide incubated overnight; Lanes 4,9: photo-DIBOD 1a, azido-BSA, and Bioti azide incubated overnight in the dark; Lane 5,10: azido-BSA and Biotin-azide incubated overnight.

Finally, photo-DIBOD can be useful for the patterned immobilization of various substrates. To illustrate this capability of the new photo-crosslinking platform, we have immobilized fluorescent dye in selected wells of 96-well plates.<sup>[14](#page-4-2)</sup> Amine binding plates we functionalized with azide moieties by incubation with 3-azidopropan-1 amine. Methanol solution of Rhodamine B-azide and photo- $DIP^{\frown}$ were added to each well and the loaded plate was irradiated with a  $35<sub>o</sub>$ nm handheld fluorescent lamp via a shadow mask (Scheme 4). The plate was thoroughly washed with methanol and fluorescent intensity of individual wells was recorded using a plate reader (Table S1).<sup>[14](#page-4-2)</sup> Inten. fluorescence of the irradiated wells demonstrates covalent immobilization of the dye.



**Scheme 4.** Light-directed Immobilization of Rhodamine B on a 96-well plate

In conclusion, we have reported the first synthesis of a conjugated bis-cyclopropenone-fused dibenzo[a,e]cyclo-octatetraene (phot DIBOD **1a**) and its butoxy- (**1b**) and tetra-ethylene glycol  $(1c)$ derivatives. Irradiation of photo-DIBODs gives the corresponding bi alkyne with a quantum yield of  $\Phi = 0.05$ , which acts as a linchpin in

#### **COMMUNICATION Journal Name**

conjugating two azides. In aqueous solutions, nanocrystalline suspension of photo-DIBOD **1a** serves as an efficient photo-crosslinker. The use of the nanocrystalline suspension has an additional benefit by enhancing the quantum yield of the process by almost an order of magnitude. We have demonstrated the general applicability of this platform by functionalizing a model protein, BSA, with a variety of functional fragments. In addition, photo-DIBOD was employed for the lightdirected immobilization of azide-derivatized substrates.

The authors gratefully acknowledge Dr. Y. G. Zheng and Mr. Z. Han's help in the analysis of dye-labeled BSA. This project was supported by grants from the National Science Foundation (VP, CHEM-1213789) and the National Institutes of Health (RS, GM107012).

#### **Notes and references**

- 1 H. C. Kolb, M. G. Finn, and K. B. Sharpless, *Angew. Chem., Int. Ed*. 2001, **40**, 2004; C. W. Tornøe, C. Christensen, and M. J. Meldal, *Org. Chem*. 2002, **67**, 3057.
- 2 J. Baskin, J. Prescher, S. Laughlin, N. Agard, P. Chang, I. Miller, A. Lo, J. Codelli, and C. R. Bertozzi*, Proc. Natl. Acad. Sci. U.S.A.,* 2007, **104**, 16793.
- <span id="page-4-0"></span>3 Some recent reviews: (a) G. Delaittre, N. K. Guimard, and C. Barner-Kowollik, *Acc. Chem. Res*. 2015, **48**, 1296; (b) P. Shieh, and C. R. Bertozzi, *Org. Biomol. Chem*. 2014, **12**, 9307; (c) W. Tang, and M. L. Becker, *Chem. Soc. Rev*. 2014, **43**, 7013.
- 4 N. J. Agard, J. M. Baskin, J. A. Prescher, A. Lo, and C. R. Bertozzi, *ACS Chem. Biol.* 2006, **1**, 644; I. C. Tanrikulu, E. Schmitt, Y. Mechulam, W. A. Goddard, and D. A. Tirrell, *Proc. Natl. Acad. Sci. U.S.A*. 2009, **106***,* 15285.
- 5 K. B. Sharpless and R. Manetsch, *Exp. Opin. Drug Discov*. 2006, **1**, 525.
- 6 M. T. Smith, A. K. Hawes, and B. C. Bundy, *Curr. Op. Biotech*. 2013, **24**, 620; K. E. Sapsford, W. R. Algar, L. Berti, K. B. Gemmill, B. J. Casey, E. Oh, M. H. Stewart, and I. L. Medintz, *Chem. Rev*. 2013, **113**, 1904.
- 7 J. Lahann, Click Chemistry for Biotechnology and Materials Science, Wiley, West Sussex, 2009; J. E. Moses and A. D. Moorhouse, *Chem. Soc. Rev*. 2007, **36**, 1242; R. M. Arnold, D. L. Patton, V. V. Popik, and J. Locklin, J. *Acc. Chem. Res*. 2014, **47**, 2999.
- 8 L. M. Gaetke and C. K. Chow, *Toxicology* 2003, **189**, 147; C. J. Burrows and J. G. Muller, *Chem. Rev.* 1998, **98**, 1109.
- 9 C. Arkona and J. Rademann, *Angew. Chem. Int. Ed*. 2013, 52, 8210; S. Sommer, N. D. Weikart, U. Linne, and H. D. Mootz, *Bioorg. Med. Chem*. 2013, **21**, 2511.
- 10 D. M. Beal, V. E. Albrow, G. Burslem, L. Hitchen, C. Fernandes, C. Lapthorn, L. R. Roberts, M. D. Selby, and L. H. Jones, *Org. Biomol. Chem.* 2012, **10**, 548; H. Elamari, F. Meganem, J. Herscovici, C. Girard, *Tetrahedron Lett*. 2011, **52**, 658; H. Zong, S. N. Goonewardena, H. N. Chang, J. B. Otis, and J. R. Baker, *Bioorg. Med. Chem*. 2014, **22***,* 6288; C.-H. Lai, T. C. Chang, Y. J. Chuang, D. L. Tzou, and C. C. Lin, *Bioconjugate Chem*. 2013, **24**, 1698.
- 11 S. Arumugam and V. V. Popik, *J. Org. Chem.* 2014, **79**, 2702.
- <span id="page-4-1"></span>12 (a) L. Kii, A. Shiraishi, T. Hiramatsu, T. Matsushita, H. Uekusa, S. Yoshida, M. Yammamoto, A. Kudo, M. Hagiwara, and T. Hosoya, *Org. Biomol. Chem*. 2010. **8**. 405; (b) F. Xu, L. Peng, K. Shinohara, T. Morita, S. Yoshida, T. Hosoya, A. Orita, and J. J. Otera, *Org. Chem*. 2014, **79**, 11592.
- 13 S. Yoshida, A. Shiraishi, K. Kanno, T. Matsushita, K. Johmoto, H. Uekusa, and T. Hosoya, *Sci. Rep*. 2011, **1**, 8; A. Horner, D. Volz, T. Hagendorn, D. Furniss, L. Greb, F. Ronicke, M. Nieger, U. Schepers, and S. Brase, *RSC Adv*. 2014, **4**, 11528.
- <span id="page-4-2"></span>14 Supporting Information
- 15 (a) A. Poloukhtine and V. V. Popik, *J. Org. Chem*. 2003, *68*, 7833; (b) A. Poloukhtine and V. V. Popik, *J. Phys. Chem. A* 2006, **110**, 1749.
- 16 N. K. Urdabaev, A. Poloukhtine, and V. V. Popik, *Chem. Comm.* 2006, 454.
- <span id="page-4-3"></span>17 G. Kuzmanich, M. N. Gard, and M. A. Garcia-Garibay, *J. Am. Chem. Soc*. *2009*, **131**, 11606.
- 18 Y. Chiang, A. J. Kresge, S. W. Paine, and V. V. Popik, *J. Phys. Org. Chem*. 1996, **9**, 361.
- 19 D. R. Pandithavidana, A. Poloukhtine, and V.V. Popik, *J. Am. Chem. Soc*. 2009, **131**, 351; A. Poloukhtine and V. V. Popik, *J. Org. Chem.* 2006, **71**, 7417.
- 20 C. D. McNitt and V. V. Popik, *Org. Biomol. Chem*. 2012, **10**, 8200.
- 21 (a) M. A. Tasdelen and Y. Yagci, *Angew. Chem. Int. Ed*. 2013, **52**, 5930. (b) G. Delaittre, A. S. Goldmann, J. O. Mueller, C. Barner-Kowollik, *Angew. Chem. Int. Ed*. 2015, **54**, 11388.
- 22 (a) R. K. Lim and Q. Lin, *Acc. Chem. Res.* 2011**, 44**, 828. (b) Z. Yu, T. Y. Ohulchanskyy, P. An, P. N. Prasad, and Q. Lin, *J. Am. Chem. Soc.* 2013**, 135**, 16766.
- 23 C. A. De Forest, B. D. Polizzotti, and K. S. Anseth, *Nature Materials* 2009, **8**, 659; M. Fiore, A. Marra, and A. Dondoni, *J. Org. Chem.* 2009, **74**, 4422; K. L. Killops, L. M. Campos, and J. Hawker, *J. Am. Chem. Soc*. 2008 **130**, 5062; J. W. Chan, B. Yu, C. E. Hoyle, and A. B. Lowe, *Chem. Commun*. 2008, 4959.
- 24 R. M. Hensarling, V. A. Doughty, J. W. Chan, and D. L. Patton, *J. Am. Chem. Soc*. 2009, **131**, 14673.
- 25 (a) W, Dillmore, M. Yousaf, M. Mrksich, *Langmuir* 2004, **20**, 7223; (b) F. Rusmini, Z. Zhong, J. Feijen, *Biomacromolecules* 2007, **3**, 1775; (c) S. Arumugam, S. Orski, J. Locklin, V. V. Popik, *J. Am. Chem. Soc*. 2012, **134**, 179; (d) D. M. Bauer, A. Rogge, L. Stolzer, C. Barner-Kowollik, L. Fruk, *Chem. Commun*. 2013, **49**, 8626.
- 26 (a) S. Arumugam and V. V. Popik, *J. Am. Chem. Soc*. 2012, **134**, 8408; (b) C. N. Bowman, W. Xi, C. Kloxin, and M. Kriege *Chem. Commun*. 2013, **49**, 4504.
- 27 A. Orita, D. Hasegawa, T. Nakano, and J. Otera, *Chem. Eur. J*. 2002, **8**, 2000.
- 28 D. Brahms and W. Dailey, *Chem Rev*. 1996, **96**, 1585; F. Tien, V. Kruger, O. Bautista, J.-X. Duan, A-R. Li, W. Dolbier, and Q. Chen, *Org. Lett.* 2000, **2**, 563.
- 29 F. Wang, T. Luo, J. Hu, Y. Wang, H. Krishnan, P. Jog, S. Ganesh, G. Prakash, and G. Olah, *Angew. Chem. Int. Ed*. 2011, **50**, 7153; F. Wang, W. Zhang, J. Zhu, H. Li, K.-W. Huang, and J. Hu, *Chem. Comm*. 2011, **47**, 2411.
- <span id="page-4-4"></span>30 A. Nielsen, G. Kuzmanich, and M. A. Garcia-Garibay, *J. Phys. Chem. A* 2014, **118**, 1858.
- 31 M. Gauthier and H. Klok, *Chem. Comm*. 2008, **23**, 2591.
- 32 F. Friscourt, C. Fahrni, and G.-J. Boons, *J. Am. Chem. Soc.* 2012, **134**, 18809.

**Cheman Replacement of the Communication** of the Communication of the Commu mudor