

# ChemComm

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

## Robust silver-mediated imidazolo-dC base pairs in metal DNA: dinuclear silver bridges with exceptional stability in double helices with parallel and antiparallel strand orientation†

Received 00th January 20xx,  
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

Sunit Kumar Jana,<sup>a</sup> Xiurong Guo,<sup>a,c</sup> Hui Mei,<sup>a</sup> and Frank Seela<sup>\*a,b</sup>

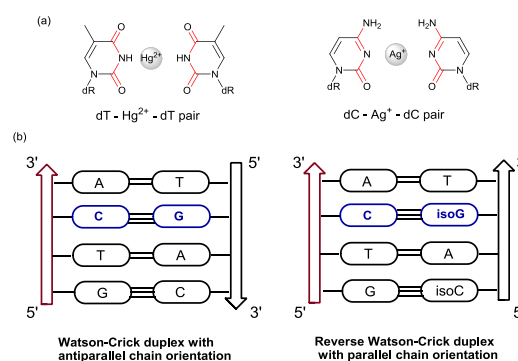
www.rsc.org/

A new unprecedented metal-mediated base pair was designed that stabilizes reverse Watson-Crick DNA (parallel strand orientation, ps) as well as canonical Watson-Crick DNA (antiparallel strand orientation, aps). This base pair contains two imidazolo-dC units decorated with furan residues.  $T_m$  measurements and spectroscopic studies reveal that each silver-mediated furano-imidazolo-dC forms exceptionally stable duplexes with ps and aps chain orientation. This stability increase by a silver-mediated base pair is the highest reported so far for ps and aps DNA helices.

Metal-mediated base pairs have generated significant attention as substitutes of Watson-Crick pairs.<sup>1</sup> These base pairs not only expand the existing genetic coding system but also allow the construction of extremely stable DNA structures. Thus, metal-mediated base pairs are considered as a high-tech alternative to canonical base pairs. Nevertheless, transition metals have been associated with the oxidative damage of DNA.<sup>2</sup> Metal base pairs between canonical nucleobases have been developed in which  $Hg^{2+}$  or  $Ag^+$  take over the function of hydrogen bonds (Fig. 1a).<sup>3</sup> Generally, metal-mediated pairing systems bind only one metal ion per base pair. Depending on the type of ligand, a metal-mediated base pair can even bind two transition metal ions.<sup>4</sup>

DNA is known to adopt a wide range of structures. Among those parallel-stranded (ps) DNA represents an unique DNA assembly characterized by a sugar-phosphate backbone pointing in the same direction while canonical DNA forms duplexes with antiparallel chains (aps DNA) (Fig. 1b).<sup>5</sup> Although, ps DNA is rather stable, it is less stable than aps DNA.<sup>6</sup> As chemical modification of ps DNA is

more restricted, its stabilization remains a challenge.<sup>7</sup>



**Fig. 1** (a) Proposed structure for metal-mediated base pairs.<sup>3</sup> (b) Schematic presentation of ps and aps duplex DNA. isoG = 2'-deoxyisoguanosine, isoC = 2'-deoxy-5-methylisocytidine, dR = 2'-deoxyribofuranosyl. A, T, G and C refer to 2'-deoxyribonucleosides.

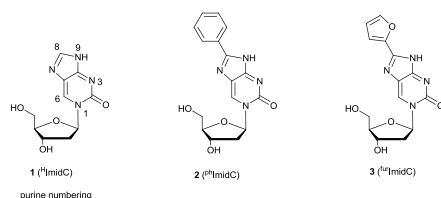
Recently, it was reported that the pyrrolo-dC-dC base pair binds one silver ion.<sup>3c</sup> Our laboratory discovered that pyrrolo-dC pyrrolo-dC base pairs can bind two silver ions.<sup>4c,8</sup> Then, for aps DNA it was observed that the imidazolo-dC (ImidC) system functionalized with a phenyl residue at the 8-position (<sup>ph</sup>ImidC, **2**, Fig. 2) is even a better silver ion binder.<sup>9</sup> This positive effect has to be caused by the purine skeleton replacing the pyrrolo[2,3-*d*]pyrimidine system. Thus, we envisaged that a furyl moiety decorating the imidazolo-dC system might add extra coordination forces for silver ions to the system and as a side effect, it might improve the fluorescence properties.<sup>10</sup> We expected that the modified purine nucleobases provide flexibility to the system and act as silver ion binder on both ps as well as on aps DNA structures. Herein we describe the synthesis of imidazolo-dC (<sup>h</sup>ImidC, **1**) and 8-furyl imidazolo-dC (<sup>fur</sup>ImidC, **3**) and disclose their effect on ps and aps duplexes in the presence of silver ions. Pleasingly, we observed that <sup>fur</sup>ImidC forms exceptional strong silver-mediated imidazolo-dC base pairs that stabilize both parallel and antiparallel DNA far beyond the stability of canonical base pairs in natural DNA.

<sup>a</sup> Laboratory of Bioorganic Chemistry and Chemical Biology, Center for Nanotechnology, Heisenbergstraße 11, 48149 Münster, Germany  
Fax: (+49) 251-53406857; E-mail: frank.seela@uni-osnabrueck.de

<sup>b</sup> Laboratorium für Organische und Bioorganische Chemie, Institut für Chemie neuer Materialien, Universität Osnabrück, BarbarasträÙe 7, 49069 Osnabrück, Germany

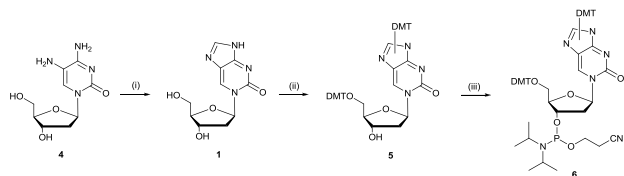
<sup>c</sup> Institute for Nanobiomedical Technology and Membrane Biology, Sichuan University, No. 1 Keyuan 4<sup>th</sup> Road, Gaopengdadao, Chengdu 610041, P.R. China

† Electronic Supplementary Information (ESI) available: Experimental details; supplementary figures. See DOI: 10.1039/x0xx00000x

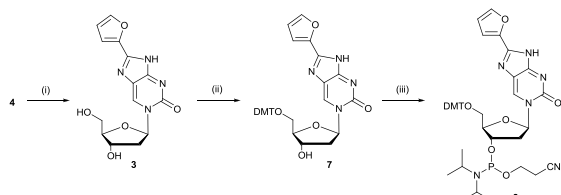


**Fig. 2** The structures of ImidC nucleosides **1–3**.

<sup>H</sup>ImidC nucleoside **1** was synthesized following a procedure described for the corresponding ribonucleoside.<sup>11</sup> To this end, 5-amino-dC (**4**) was treated with diethoxymethyl acetate to give **1** (Scheme 1). Treatment of **1** with excess of 4,4'-dimethoxytriphenylmethyl chloride (DMT-Cl) (2.2 equiv) leads to the bis-DMT derivative **5** with one protecting group at the 5'-position and the other at the base. Phosphitylation under standard conditions afforded phosphoramidite **6**.



**Scheme 1** Synthesis of phosphoramidite **6**: (i) (EtO)<sub>2</sub>CHOAc, 100 °C, 3 h, 29%; (ii) DMT-Cl, pyridine, Et<sub>3</sub>N, rt, 16 h, 44%; (iii) NC(CH<sub>2</sub>)<sub>2</sub>OP(Cl)N(*i*-Pr)<sub>2</sub>, (*i*-Pr)<sub>2</sub>EtN, rt, 20 min, 29%.



**Scheme 2** Synthesis of phosphoramidite **8**: (i) furfural, CAN, DMF, 55 °C, 24 h, 57%; (ii) DMT-Cl, pyridine, rt, 16 h, 47%; (iii) NC(CH<sub>2</sub>)<sub>2</sub>OP(Cl)N(*i*-Pr)<sub>2</sub>, (*i*-Pr)<sub>2</sub>EtN, rt, 10 min, 79%.

For the synthesis of <sup>fur</sup>ImidC **3** initially a Traube condensation was employed which was used earlier for the synthesis of **2**.<sup>9,12</sup> However, condensation between 5-amino-dC (**4**) with furfural in the presence of para-toluenesulfonic acid led to the formation of only 4% of desired **3**. Thus, we examined several oxidants including PhI(OAc)<sub>2</sub>, Cu(OH)<sub>2</sub>, Cu(OAc)<sub>2</sub> and ceric ammonium nitrate (CAN) at different conditions. The desired <sup>fur</sup>ImidC **3** was obtained in 57% yield using CAN and DMF as solvent at 55 °C. Protection of 5'-OH with DMT-Cl followed by phosphitylation yielded phosphoramidite **8** (Scheme 2). Next, the imidazolo-dC derivatives **1–3** were incorporated near the center of the two parallel stranded duplexes ODN-1 • ODN-2 and ODN-9 • ODN-13 (Table 1) thereby replacing strongly bonded central dC-iG<sub>d</sub> base pairs. For comparison aps duplexes were designed in which one strand is identical to the ps duplexes and the central dG-dC pair was replaced by compounds **1–**

**3**. Oligonucleotide synthesis was performed on solid phase using the phosphoramidites of canonical nucleosides, iG<sub>d</sub> (2'-deoxyisoguanosine),<sup>5b,13</sup> and <sup>5me</sup>iC<sub>d</sub> (2'-deoxy-5-methyl-isocytidine)<sup>13</sup>, the phosphoramidite of **2**, as well as phosphoramidites **6** and **8**.

The most obvious feature of metal-mediated base pairs is their significant contribution to the stabilization of DNA duplexes, since the strength of the coordinative bond is much stronger than that of hydrogen bonds. Table 1 summarizes the  $\Delta T_m$  values in the presence and absence of silver ions with regard to duplexes with a dC-dC pair, ImidC pairs as well as relative to the unmodified reference duplexes. Addition of silver ions to the reference duplexes (ODN-1 • ODN-2 and ODN-1 • ODN-9) caused no significant  $T_m$  changes showing that silver ions are not involved in base pairing. Only one silver ion was captured by a dC-dC pair. This base pair caused a notable increase in aps duplex stability ( $\Delta T_m = +8.5$  °C), but still shows lower strength than the canonical one.

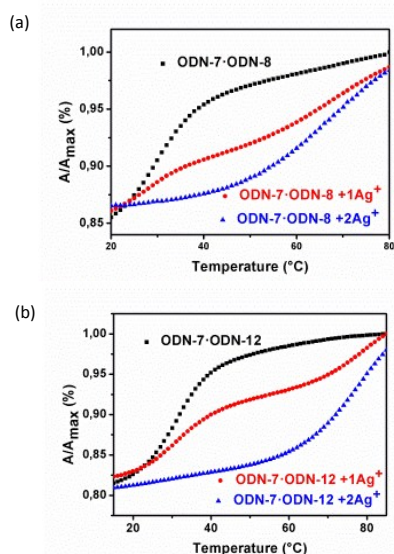
However, the situation is entirely different when ImidC residues are present and facing each other. Now, the  $T_m$  increased tremendously for ps and aps DNA by addition of 2 equiv. of silver ions (Table 1). The  $\Delta T_m$  values relative to mismatches amount to 27.0–38.0 °C for ps DNA and to 38.5–48.0 °C for aps DNA. The  $T_m$  increase is unique for compound **3** bearing a furan residue; the phenyl residue of **2** did not stabilize the silver-mediated base pair over that of the non-functionalized nucleoside **1** (Table 1). For ps DNA two different sequence motifs (series I and II) were studied. This positioned the silver-mediated base pairs in different environments and changed the interaction with neighboring base pairs. However, the  $T_m$  increase was almost the same for both series. For aps DNA, the  $T_m$  values were 10 °C higher than for ps DNA. Also in this case, the stabilization of the furan residue of **1** is exceptional high while the phenyl residue of **2** does not add stability. The silver-mediated base pairs of **1–3** are much stronger than the dG-dC or iG<sub>d</sub>-dC pair. The enhanced stability probably results from additional interactions of the furan ligands with silver ions. To the best of our knowledge, such a strong stabilization induced by only one silver-mediated base pair in ps and aps DNA has never been observed before.

Typical  $T_m$  curves of ImidC modified ps and aps duplexes in the absence and presence of silver ions are shown in Fig. 3. Biphasic  $T_m$  curves with a low and a high  $T_m$  value were observed by adding only one equivalent of silver ions. This indicates the existence of two species; duplexes without silver ions (low  $T_m$ ) and duplexes with two silver ions (high  $T_m$ ).<sup>8,9</sup> Addition of two equiv. of silver ions leads to monophasic melting curves showing only the high  $T_m$  value. From that it is obvious that the ImidC-ImidC base pair captures two silver ions instantaneously in a cooperative way.<sup>9</sup> To verify the stoichiometry of silver ion binding, fluorescence and UV titration experiments were performed (see ESI<sup>†</sup>). ImidC derivatives show strong fluorescence around 390 nm ( $\Phi = 0.03$  for <sup>H</sup>ImidC (**1**),  $\Phi = 0.33$  for <sup>ph</sup>ImidC (**2**), and  $\Phi = 0.39$  for <sup>fur</sup>ImidC (**3**); for details see ESI<sup>†</sup>). These experiments confirm that each duplex with an ImidC-ImidC pair captures two silver ions. Coordination forces between the furan oxygen and silver ions might be the central reason for the exceptional stability of this silver ion mediated base pair. Nonetheless,  $pK_a$  changes induced by the substituents have to be considered as it is documented that the ease of base deprotonation plays a key role in the formation of silver-mediated base pairs.<sup>14</sup>

**Table 1**  $T_m$  Values of DNA duplexes containing ImidC nucleosides **1-3** in the presence or absence of silver ions<sup>af</sup>

Duplexes	$T_m$ [°C] with $n$ equiv of $\text{AgNO}_3$			$\Delta T_m$ [°C] relative to match <sup>b</sup>	$\Delta T_m$ [°C] relative to mismatch <sup>c</sup>
	$n = 0$	$n = 1$	$n = 2$		
ps duplexes series I					
5'-d(TAG GTC AAT ACT) ODN-1 5'-d(ATiCiCAiG TTATiGA) ODN-2	34.5	n.d.	33.5	-	-
5'-d(TAG GT1 AAT ACT) ODN-3 5'-d(ATiCiCA1 TTATiGA) ODN-4	28.0	n.d./55.0 <sup>d</sup>	55.0	+20.5	+27.0
5'-d(TAG GT2 AAT ACT) <sup>g</sup> ODN-5 5'-d(ATiCiCA2 TTATiGA) ODN-6	31.0	n.d./58.0 <sup>d</sup>	58.0	+23.5	+27.0
5'-d(TAG GT3 AAT ACT) ODN-7 5'-d(ATiCiCA3 TTATiGA) ODN-8	29.0	27.0/67.0 <sup>d</sup>	67.0	+32.5	+38.0
ps duplexes series II					
5'-d(AGTATTGAC CTA) ODN-9 5'-d(TiCATAAiC TiGiGAT) ODN-13	42.0	42.0	42.0	0	-
5'-d(AGTATT1AC CTA) ODN-10 5'-d(TiCATAA1 TiGiGAT) ODN-14	28.0	n.d./57.0 <sup>d</sup>	57.0	+15.0	+29.0
5'-d(AGT ATT 2AC CTA) ODN-11 5'-d(TiCATAA 2TiGiGAT) ODN-15	32.5	n.d.	61.0	+19.0	+28.5
5'-d(AGTATT3AC CTA) ODN-12 5'-d(TiCATAA3 TiGiGAT) ODN-16	28.0	27.0/66.0 <sup>d</sup>	66.0	+24.0	+38.0
aps duplexes					
5'-d(TAGGTCAACT) ODN-1 3'-d(ATCCAGTTATGA) ODN-9	46.5	47.0	46.5	-	-
5'-d(TAGGTCAACT) ODN-1 3'-d(ATCCA1TTATGA) <sup>hb</sup> ODN-1a	26.0	32.5 <sup>e</sup>	34.5	+12.0	+8.5
5'-d(TAGGT1AATACT) ODN-3 3'-d(ATCCA1TTATGA) ODN-10	32.0	32.0/71.0 <sup>d</sup>	71.0	+24.5	+39.0
5'-d(TAGGT2AATACT) <sup>g</sup> ODN-5 3'-d(ATCCA2TTATGA) ODN-11	35.5	32.0/72.0 <sup>d</sup>	74.0	+27.5	+38.5
5'-d(TAGGT3AATACT) ODN-7 3'-d(ATCCA3TTATGA) ODN-12	30.0	25.0/75.0 <sup>d</sup>	78.0	+31.5	+48.0

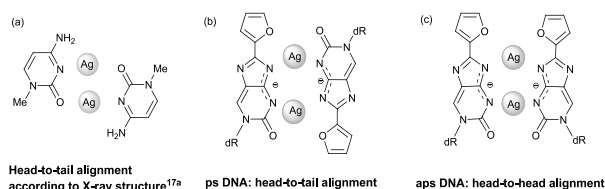
<sup>a</sup>Measured at 260 nm with 5  $\mu\text{M}$  + 5  $\mu\text{M}$  single-strand concentration in 100 mM NaOAc, 10 mM  $\text{Mg}(\text{OAc})_2$ , pH 7.5 in the presence of  $\text{AgNO}_3$  (0-2 equivalents). <sup>b</sup> $\Delta T_m = T_m$  after the addition of 2.0 equivalents  $\text{AgNO}_3 - T_m$  of reference duplex. <sup>c</sup> $\Delta T_m = T_m$  after the addition of 2.0 equivalents  $\text{AgNO}_3 - T_m$  before the addition of  $\text{AgNO}_3$ . <sup>d</sup>Biphasic melting. <sup>e</sup>Binds only one silver ion (ESI<sup>†</sup>, Fig. S11). <sup>f</sup>For thermodynamic data see ESI<sup>†</sup>, Table S6. <sup>g</sup>iC<sub>d</sub> corresponds to <sup>5me</sup>iC<sub>d</sub>. n.d. not determined.

**Fig. 3** Thermal melting ( $T_m$ ) curves measured at 260 nm in 100 mM NaOAc, 10 mM  $\text{Mg}(\text{OAc})_2$  at pH 7.5 in presence of 0-2 equivalents of  $\text{Ag}^+$ . (a) ps ODN-7 • ODN-8, (b) aps ODN-7 • ODN-12.

Indeed, the  $\text{pK}_a$  value of compound **3** ( $\text{pK}_a = 7.3$ ) is the lowest compared to **2** ( $\text{pK}_a = 7.9$ ) and **1** ( $\text{pK}_a = 8.8$ ) (see ESI<sup>†</sup>). In accordance with the  $\text{pK}_a$  values, the furano derivative **3** forms the most stable silver-mediated base pair while the non-functionalized derivative **1** shows the lowest stability.

Earlier, we proposed a structure for the silver-mediated <sup>ph</sup>ImidC-<sup>ph</sup>ImidC base pair in aps DNA.<sup>9</sup> Herein, a structure for the ps duplex is suggested which aligns the purine bases head-to-tail instead of head-to-head as suggested for canonical DNA which is consistent with the observation for dC residues in silver-mediated dC-dC base pairs made by Ono<sup>15</sup> and by the computational studies of Marino<sup>16</sup> (Fig. 4). Strong support for the head-to-tail alignment in ps DNA comes from the single crystal X-ray structure of the dimeric 1-methylcytosine silver nitrate

complex by Marzilli in which the two silver ions connect the heterocyclic ligands with bonds between the atoms N-3 and O-2 in that way that the bases are aligned head-to-tail (Fig. 4).<sup>17</sup> Connectivity between silver ions and purine bases should be different in ps and aps DNA. According to the size of the silver ions, the base connectivity is not strictly linear but bent by 132°. A similar spacing of the Ag-Ag repeat length between columnar stacks (3.642 Å) and the base stacking distance in duplex DNA (about 3.5 Å) suggests that this dimeric unit provides a model for the cross-linking of DNA.



**Fig. 4** Proposed silver-mediated base pairs. (a) Dimeric 1-methylcytosine silver complex. <sup>fur</sup>ImidC-<sup>fur</sup>ImidC pair with two silver ions in (b) ps DNA and (c) in aps DNA. dR = 2'-deoxyribofuranosyl. Ag corresponds to Ag<sup>+</sup>.

In summary, we designed an exceptional strong silver-mediated imidazolo-dC base pair decorated with furan residues. The purine nucleobases act as silver ion binder and provide flexibility to form base pairs in ps and aps DNA. Two silver ions are bound to a single imidazolo-dC pair. The incorporation of only one silver-mediated furano-imidazolo-dC pair increased the  $T_m$  value of a 12-mer duplex with parallel strand orientation by 38.0 °C relative to the silver-free duplex. An even higher increment was observed for canonical DNA with antiparallel chains ( $\Delta T_m = 48.0$  °C). Almost no sequence dependent stability increase was observed when the silver-mediated base pairs were located in different helical environment as shown for the ps duplexes (series I and II; Table 1). The extremely high stability of these base pairs probably superimposes other possible changes in the double helix structure. For silver-mediated base pairs of lower stability, sequence dependent stability changes were detected.<sup>9</sup>

In recent years, various new base pairs have been developed to increase duplex stability. In this context, the silver-mediated base pair of **1** is a unique example. Incorporation of multiple base pairs with two silver ions per base pair is feasible.<sup>8a</sup> Strong and robust functional materials can be constructed with possible applications in metal-ion based nanomechanical devices<sup>18</sup> and biomedical science. To the best of our knowledge, the furan functionalized imidazolo-dC (**3**) pair is the most stable Ag<sup>+</sup>-mediated base pair in ps and aps DNA reported so far.

We would like to thank Dr. S. Budow-Busse and Dr. P. Leonard for helpful discussions, Mr. Nhat Quang Tran for oligonucleotide syntheses and Dr. M. Letzel, Universität Münster, Germany, for the MALDI spectra. Financial support by ChemBiotech, Münster, Germany, is highly appreciated.

## Notes and references

- (a) G. H. Clever, C. Kaul and T. Carell, *Angew. Chem. Int. Ed.*, 2007, **46**, 6226-6236; (b) A. Ono, H. Torigoe, Y. Tanaka and I. Okamoto, *Chem. Soc. Rev.*, 2011, **40**, 5855-5866; (c) Y. Takezawa and M. Shionoya, *Acc. Chem. Res.*, 2012, **45**, 2066-2076; (d) P. Scharf and J. Müller, *ChemPlusChem*, 2013, **78**, 20-34; (e) S. Taherpour, O. Golubev and T. Lönnberg, *J. Org. Chem.*, 2014, **79**, 8990-8999.
- P. Ghude, M. A. Schallenberger, A. M. Fleming, J. G. Muller and C. J. Burrows, *Inorg. Chim. Acta*, 2011, **369**, 240-246.
- (a) Y. Miyake, H. Togashi, M. Tashiro, H. Yamaguchi, S. Oda, M. Kudo, Y. Tanaka, Y. Kondo, R. Sawa, T. Fujimoto, T. Machinami and A. Ono, *J. Am. Chem. Soc.*, 2006, **128**, 2172-2173; (b) A. Ono, S. Cao, H. Togashi, M. Tashiro, T. Fujimoto, T. Machinami, S. Oda, Y. Miyake, I. Okamoto and Y. Tanaka, *Chem. Commun.*, 2008, 4825-4827; (c) K. S. Park, J. Y. Lee and H. G. Park, *Chem. Commun.*, 2012, **48**, 4549-4551.
- (a) D. A. Megger, C. F. Guerra, J. Hoffmann, B. Brutschy, F. M. Bickelhaupt and J. Müller, *Chem. Eur. J.*, 2011, **17**, 6533-6544; (b) I. Okamoto, T. Ono, R. Sameshima and A. Ono, *Chem. Commun.*, 2012, **48**, 4347-4349; (c) H. Mei, I. Röhl and F. Seela, *J. Org. Chem.*, 2013, **78**, 9457-9463.
- (a) J. H. van de Sande, N. B. Ramsing, M. W. Germann, W. Elhorst, B. W. Kalisch, E. V. Kitzing, R. T. Pon, R. C. Clegg and T. M. Jovin, *Science*, 1988, **241**, 551-557; (b) F. Seela, C. Wei and Z. Kazimierzczuk, *Helv. Chim. Acta*, 1995, **78**, 1843-1854; (c) F. Seela, Y. He and C. Wei, *Tetrahedron*, 1999, **55**, 9481-9500.
- (a) P. S. Pallan, P. Lubini, M. Bolli and M. Egli, *Nucleic Acids Res.*, 2007, **35**, 6611-6624; (b) S. A. Ingale, P. Leonard, Q. N. Tran and F. Seela, *J. Org. Chem.*, 2015, **80**, 3124-3138.
- (a) F. Seela, X. Peng and H. Li, *J. Am. Chem. Soc.*, 2005, **127**, 7739-7751; (b) I. Sinha, C. F. Guerra and J. Müller, *Angew. Chem. Int. Ed.*, 2015, **54**, 3603-3606.
- (a) H. Mei, H. Yang, I. Röhl and F. Seela, *ChemPlusChem*, 2014, **79**, 914-918; (b) H. Yang, H. Mei and F. Seela, *Chem. Eur. J.*, 2015, **21**, 10207-10219.
- H. Mei, S. A. Ingale and F. Seela, *Chem. Eur. J.*, 2014, **20**, 16248-16257.
- N. J. Greco and Y. Tor, *Tetrahedron*, 2007, **63**, 3515-3527.
- J. J. Fox and D. Van Praag, *J. Org. Chem.*, 1961, **26**, 526-532.
- M. Kovaliov, M. Weitman, D. T. Major and B. Fischer, *J. Org. Chem.*, 2014, **79**, 7051-7062.
- Y. Tor and P. B. Dervan, *J. Am. Chem. Soc.*, 1993, **115**, 4461-4467.
- (a) I. Okamoto, K. Iwamoto, Y. Watanabe, Y. Miyake and A. Ono, *Angew. Chem. Int. Ed.*, 2009, **48**, 1648-1651; (b) T. Matsui, H. Miyachi, T. Baba and Y. Shigeta, *J. Phys. Chem. A*, 2011, **115**, 8504-8510; (c) C. F. Guerra, P. J. S. Miguel, A. Cebollada, F. M. Bickelhaupt and B. Lippert, *Chem. Eur. J.*, 2014, **20**, 9494-9499.
- T. Ono, K. Yoshida, Y. Saotome, R. Sakabe, I. Okamoto and A. Ono, *Chem. Commun.*, 2011, **47**, 1542-1544.
- M. Fortino, T. Marino and N. Russo, *J. Phys. Chem. A*, 2015, **119**, 5153-5157.
- (a) L. G. Marzilli, T. J. Kistenmacher and M. Rossi, *J. Am. Chem. Soc.*, 1977, **99**, 2797-2798; (b) T. J. Kistenmacher, M. Rossi and L. G. Marzilli, *Inorg. Chem.*, 1979, **18**, 240-244.
- N. C. Seeman, *Annu. Rev. Biochem.*, 2010, **79**, 65-87.

Journal Name

COMMUNICATION

ChemComm Accepted Manuscript