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Fluorescent Mimics of Cholesterol that Rapidly Bind Surfaces of Living Mammalian Cells

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Mammalian cells acquire cholesterol, a critical membrane constituent, through multiple mechanisms. We synthesized mimics of cholesterol, fluorescent N-alkyl-3β-cholesterylamine-glutamic acids, that are rapidly incorporated into cellular plasma membranes compared with analogous cholesteryl amides, ethers, esters, carbamates, and a sitosterol analogue. This process was inhibited by ezetimibe, indicating a receptor-mediated uptake pathway.

Cholesterol (1) is a critical constituent of membranes of mammalian cells. Cells acquire exogenous forms of this sterol through multiple mechanisms involving structurally distinct cell surface receptors. Lipoprotein particles such as low-density lipoprotein (LDL) and high-density lipoprotein (HDL) carry cholesteryl esters (2) and associated protein and lipid components throughout the bloodstream. Cells expressing LDL and HDL receptors actively internalize these natural nanoparticles via receptor-mediated endocytosis. In contrast, Niemann-Pick C1 Like 1 protein (NPC1L1) plays key roles in the cellular uptake of dietary (unesterified) cholesterol (1), as found in mixed micelles. This receptor was identified in 2004 as a target of ezetimibe (3), a drug used to treat hypercholesterolemia. More recent studies suggest that although NPC1L1 is a primary target of ezetimibe and its active glucuronide metabolite, other proteins, such as the HDL receptor SR-BI, can also be inhibited by this drug. Recent proteomics experiments have identified over 250 cholesterol-binding proteins, including receptors, channels, and enzymes.

Derivatives of cholesterol have numerous biological applications. These compounds have been used to facilitate the delivery of small inhibitory RNA (siRNA), enhance DNA transfection, probe cellular membrane subdomains, and assay cholesterol transport processes. Cholesteryl carbamates have been extensively investigated, and cellular uptake of cholesteryl carbamate-conjugated siRNA has been reported to be similar to uptake of cholesteryl esters, requiring...
binding to HDL or LDL, followed by internalization via HDL or LDL receptors. This initial lipoprotein-binding step can slow cellular uptake, and the presence of high concentrations of serum (e.g. 10%) in media typically reduces the activity of cellular uptake, and the presence of high concentrations of LDL receptors. This initial lipoprotein-binding step can slow inhibition upon coaddition with excess (200 µM) ezetimibe (3), ester (5) or carbamates (10) or cholesterol (9, 11−13) linked to the hydrophobic Pennsylvania Green fluorophore through amino acid subunits. We hypothesized that the carbonyl linked to the steroid in amide 10, ester 11 or carbamates 12 and 13 would be similar to natural cholesteryl esters, and this structural modification might correspondingly affect their ability to bind cell surfaces.

As shown in Figure 1, confocal laser scanning microscopy was employed to compare living human Jurkat lymphocyte cells after treatment with 4−13. These experiments demonstrated that a brief (5 minute) treatment of cells with 4 or 5 (2 µM) at 37 °C results in robust fluorescent staining of plasma membranes. Examination of these cells after 1 hour showed enhanced cellular binding, uptake of the probe, and localization in transferrin-positive early/recycling endosomes (Figure 1 and Figure S1 of the supporting information). Binding of 4 (and 5) to cell surfaces was predominantly receptor-mediated as evidenced by up to 80% inhibition upon coaddition with excess (200 µM) ezetimibe (3). Compare Figure 1A and 1C, and see Figure S3 of the supporting information). This inhibition indicates that proteins on cell surfaces may recognize these compounds as mimics of free cholesterol or related cholesterol metabolites. These metabolites might include structurally related cholesterol sulfate and cholesterol glucuronide, which are present in micromolar concentrations in the bloodstream of animals. Comparison of 4 with 6 lacking the glutamic acid residue in the linker region revealed that the anionic moiety of 4 is critical for rapid high affinity/efficacy binding to cells (Figure 1, compare panels B and G). Moreover, despite the presence of a derived intermediates, were prepared as previously reported. The novel building block 3β-sitosterylamine was prepared from sitosterol using methodology described for the synthesis of 3β-cholesterylamine. The 4-carboxy Pennsylvania Green fluorophore was prepared as previously described. Full synthetic details are provided in Scheme S1 and Scheme S2 of the supporting information. These probes were designed to systematically compare membrane anchors derived from N-alkyl-3β-cholesterylamines (4−7), a sitosteryl analogue (8), a N-acyl-3β-cholesterylamine (10), or cholesterol (9, 11−13) linked to the hydrophobic Pennsylvania Green fluorophore through amino acid subunits. We hypothesized that the carbonyl linked to the steroid in amide 10, ester 11 or carbamates 12 and 13 would be similar to natural cholesteryl esters, and this structural modification might correspondingly affect their ability to bind cell surfaces.

As shown in Figure 1, confocal laser scanning microscopy was employed to compare living human Jurkat lymphocyte cells after treatment with 4−13. These experiments demonstrated that a brief (5 minute) treatment of cells with 4 or 5 (2 µM) at 37 °C results in robust fluorescent staining of plasma membranes. Examination of these cells after 1 hour showed enhanced cellular binding, uptake of the probe, and localization in transferrin-positive early/recycling endosomes (Figure 1 and Figure S1 of the supporting information). Binding of 4 (and 5) to cell surfaces was predominantly receptor-mediated as evidenced by up to 80% inhibition upon coaddition with excess (200 µM) ezetimibe (3). Compare Figure 1A and 1C, and see Figure S3 of the supporting information). This inhibition indicates that proteins on cell surfaces may recognize these compounds as mimics of free cholesterol or related cholesterol metabolites. These metabolites might include structurally related cholesterol sulfate and cholesterol glucuronide, which are present in micromolar concentrations in the bloodstream of animals. Comparison of 4 with 6 lacking the glutamic acid residue in the linker region revealed that the anionic moiety of 4 is critical for rapid high affinity/efficacy binding to cells (Figure 1, compare panels B and G). Moreover, despite the presence of a
structurally analogous glutamic acid, the cholesteryl carbamate 12 showed low cellular binding and cellular uptake compared with 4, supporting the hypothesis that carbamates engage a mechanistically distinct cellular uptake pathway. However, this loss of activity of 12 could be at least partially rescued by addition of a second glutamic acid, as found in 13.

Using ezetimibe (3, 200 µM, 0.2% DMSO) as a specific competitor, we quantified the relative affinities (Kd) of rapid binding of 4-13 to Jurkat lymphocytes in media containing 10% serum. Data from saturation binding experiments after treatment for five minutes, designed to limit cellular uptake by endocytosis, are shown in Figure 2, Table 1, and the supporting information. The high affinity and efficacy of binding of 4 and 5 to cell surfaces compared to 6-12 revealed that N-alkyl cholesterylamines bearing a anionic functional group and a spacer residue such as β-alanine most efficiently bind cell surfaces, a critical initial step for initiation of endocytosis and delivery of linked agents. The absence of substantial cellular binding of the sitosterol analogue (8), differing from 4 by the presence of an additional ethyl group in the tail of the sterol, further supports a specific receptor-mediated uptake mechanism. This result is consistent with limited receptor-mediated uptake of phytosterols such as sitosterol by mammalian cells.

To investigate whether other cell types show differential effects, we examined rapid (5 min) binding of 4, 5, 8, and 13 to three different human cell lines. Human Jurkat cells, grown in suspension were compared with the human cell lines HeLa and HEK-293, which were suspended in media prior to treatment.

Table 1. Left columns: Apparent affinity (Kd, app) and efficacy (Bmax) of binding of 4–13 to plasma membranes of living Jurkat cells. Cells were treated with compounds at 37 °C for 5 min in media containing 10% FBS (± SEM). Non-specific binding was quantified with ezetimibe (200 µM) as a competitor; vehicle = 0.2% DMSO; NC: Not calculated due to low efficacy. Right columns: Values of Kd and Vmax calculated from Michaelis-Menten analysis of rapid time-dependent cellular uptake at 22 °C (Figure 3). ND: Not determined. MEF: molecules of equivalent fluorescein. Data, based on measurements in triplicate, is reported as mean ± SEM.
For these assays, cellular fluorescence was converted to molecular equivalents of fluorescein (MEFL) using fluorescent bead standards. For the Jurkat cell line, this analysis indicated that treatment with **4** or **5** at a concentration of 2 µM for 5 minutes loads 0.5–1.5 × 10⁶ molecules into the plasma membrane of individual cells. This rapid and massive increase in cellular fluorescence suggested that an enzyme may be actively inserting these compounds into the cellular plasma membrane. Further analysis of time-dependent fluorescence resulting from treatment with **4**, **5**, **8**, and **13** using the Michaelis-Menten model of enzyme kinetics is shown in Figure 4. These studies revealed that the cellular uptake of **4**, **5**, and to a lesser extent **13**, is highly efficient, with 1.8–5.0 × 10⁷ molecules incorporated per minute per cell, consistent with a catalytic process. Kinetic values of **K**ₘ and **V**ₘₐₓ from this analysis are shown in Table 1.

In conclusion, we identified novel structure-activity relationships that govern binding of fluorescent cholesterol mimics to the surface of living mammalian cells. New cholesterol-mimetic membrane anchor motifs of **4**, **5**, and **13** were identified that engage a rapid cellular uptake pathway, consistent with a receptor-mediated process, that catalytically inserts these compounds into the plasma membrane. Although the receptor or enzyme targeted by these compounds has not yet been identified, it is unlikely to be NPC1L1, the classical pharmacological target of ezetimibe, because this protein is not highly expressed outside of the liver and intestine, and the active metabolite ezetimibe-glucuronide, prepared as previously reported, does not inhibit binding of **4** to Jurkat cell surfaces (data shown in Figure S2 of the supporting information). Given that cholesterol trafficking and distribution involves dynamic receptor-mediated and vesicular processes that are not completely understood, these compounds have potential as novel probes and tools for the delivery of impermeable molecules into mammalian cells.

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