Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



RSCPublishing

COMMUNICATION

Synthesis of double-clickable functionalised graphene oxide for biological applications

Cite this: DOI: 10.1039/x0xx00000x

Kuo-Ching Mei, ^a Noelia Rubio, ^a Pedro M. Costa, ^a Houmam Kafa, ^a Vincenzo Abbate, ^a Frederic Festy, ^b Sukhvinder S. Bansal, ^a Robert C. Hider, ^a and Khuloud T. Al-Jamal ^{a*}

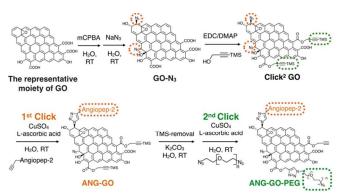
Received ooth Jun 2015, Accepted ooth Aug 2015

DOI: 10.1039/x0xx00000x

www.rsc.org/chemcomm

Azide- and alkyne- double functionalised graphene oxide (Click² GO) was synthesised and characterised with attenuated total reflectance Fourier transform infrared (ATR-FTIR), thermogravimetric analysis spectroscopy (TGA) and Raman spectroscopy. Fourteen-percentage increase in azide content was found, after pre-treatment of meta-chloroperoxybenzoic acid (mCPBA), determined with elemental analysis. No effect on A549 cell viability was found, up to 100 µg/mL and 72 hr of incubation, determined with the modified lactate dehydrogenase (mLDH) assay. Two sequential copper(I) catalysed azide-alkyne cycloaddition (CuAAC) reactions were performed to conjugate the propargyl-modified blood-brain barrier targeting peptide Angiopep-2, and a bis-azide polyethylene glycol (MW = 3500), to the Click² GO. The final conjugate was characterised with ATR-FTIR and TGA.

Graphene and graphene oxide (GO) are single atom thick, 2-D carbon structures that have recently been widely used in many fields including drug delivery, 1,2 gas separation (as membrane material), 3 biosensing, 4,5 sensing, electrochemical bio-imaging photothermal therapy.⁶ One of the very popular methods to functionalise nanomaterials for drug delivery is via click chemistry, in particular, the copper(I) catalysed azide-alkyne cycloaddition (CuAAC) and strained-promoted azide-alkyne cycloaddition (SPAAC). CuAAC click chemistry has also been used to functionalise carbon based nano-materials e.g. fullerene,8 carbon nanotubes, and graphene. Alkyne-modified GO (GO-alkyne) has been used to prepare functionalised GO for different applications e.g. for $\rm CO_2$ absorption, as photo-active sheets, and as efficient absorbents for dye removal. The introduced alkyne, however, also increased GO's hydrophobicity, making it less dispersible in aqueous media. Only one study reported alkyne-modified GO for drug delivery application after conjugation with water-dispersible polymers, used to restore water dispersibility. 18 Double functionalised GO contains both azide and protected alkyne has not been reported. Herein, we propose to synthesise GO containing double clicking sites i.e. azide and TMS-protected alkyne groups, refereed to here as double-clickable or Click² GO, for biological applications. This approach allows sequential CuAAC reactions, on GO, in aqueous media. Sequential CuAAC has been demonstrated on fullerene, ⁸ but not on graphene based material. Compared to fullerene, GO provides a significantly larger surface area and superior water dispersibility thus offering more potential for biomedical applications. As shown in **Scheme 1**, simple quick GO functional groups interconversion followed by two sequential CuAAC clicks for double-GO-functionalisation is presented.



Scheme 1. Sequential functionalisation of GO using two CuAAC click reactions.

In previous reports, GO-N₃ was generated by introducing azide groups onto the carboxylic groups of GO through amidation with 3azidopropan-1-amine $(NH_2(CH_2)_3N_3)^{19}$ or via nucleophilic substitution using 2-chloroethyl isocyanate (Cl(CH₂)₂NCO).²Eigler et al. reported azide functionalisation of GO using NaN₃²¹ The reaction was achieved by the substitution of organosulfate and ring opening of epoxy groups on GO with azide when reacted with NaN₃. GO used in the same study was prepared using the modified Hummer's method. Azide functionlisation of epoxide group can occur via 1,2-epoxide ring opening using NaN3 generating 1,2azidoalcohols.^{22,23} In the current study, azide introduction was achieved using epoxide groups on GO leaving the carboxylic groups available for subsequent functionalisation, i.e. introduction of alkyne groups. Using this approach, it is also expected that the additional hydroxyl groups generated from azidolysis can improve the water ChemComm Page 2 of 6

COMMUNICATION ChemComm

dispersibility of GO-N₃. Azide content was enriched by further epoxidising GO using meta-chloroperoxybenzoic acid (mCPBA), which has been reported to epoxidise fullerenes²⁴ and CNTs²⁵⁻²⁷ but not GO. After azide groups were introduced on GO, alkyne groups were introduced *via* Steglich esterification of the preserved carboxylic groups ²⁸ with trimethylsilyl (TMS)-protected propargyl alcohol, yielding double functionalised GO or Click² GO.

One of the commonly used methods to prepare GO is the chemical exfoliation of graphite by "Hummer's method" firstly reported by Hummers and Offeman, in 1958.²⁹ Many modifications have been proposed since then. This method involves several oxidation and hydration steps carried out at different temperatures. An improved Hummer's method was published by Kovtyukhova et al. 30 In their method, graphite powder was pre-oxidised before proceeding with the modified Hummer's method. In addition to that, the addition of NaNO₃ as an oxidising agent, was omitted. A modified Kovtyukhova-Hummer's method was used to synthesis GO in our recent work and in this study.³¹ Steps carried out in our study are described in SI methods (Figure S1) and schematically summarised in Scheme S1. In brief, Kovtyukhova-Hummer's method was adapted with further modification including: (i) NaNO3 was kept in this reaction, (ii) the water was added in portions at controlled temperature, and (iii) a heating step was introduced at H₂O₂ addition step prior to the washing and purification step. Major steps involved pre-oxidation of graphite, low-, medium-, high-temperature treatment stages followed by washing and purification. At the end, a water-dispersible dark-brown GO suspension was obtained. The final concentration of GO in the dispersion was 6.84 mg/mL, as determined by thermogravimetric analysis (TGA) (eq. S1, SI). The stock GO dispersion was then diluted to the appropriate concentration when desired. The modified Hummer's reported by Ali-Boucetta was used as a control method, ³² yielding ~ 2 mg GO from 300 mg graphite. In contrast, modified Kovtvukhova-Hummer's method generated ~ 5470 mg GO from 4000 mg graphite (Table S1). Elemental analysis (C, H, N, O) was performed to determine the oxidation status of the pre-oxidised graphite and GO (Table S2). Based on final carbon content, % GO yield was calculated to be $\sim 55.63\%$ (eq. S2 and S3, SI).

Thermogravimetric analysis (TGA) was performed to analyse the thermal-decomposition profile of graphite, pre-oxidised graphite and GO (100°C - 978°C). As shown in Figure S2A, the thermal deoxygenation of graphene derivatives was reflected by the reduced residual weight upon heating (10 °C /min in nitrogen). The final residual weight at 978 °C was 99.36%, 94.15% and 38.16% for graphite, pre-oxidised graphite and GO, respectively. Raman spectroscopy confirmed the generation of GO (Figure S2B). D, G and 2D peaks were identified for both graphite and GO. The spectra were normalised to G peak (intensity equals 1). G peak at 1580 cm⁻¹ (graphite) or 1598 cm⁻¹ (GO) represent the bond stretching of all pairs of sp² atoms (double bonds) in both rings and chains. It is used to represent the integrity of the organised graphitic structure.³³ The increased wavenumber of the G peak in GO (by 18 cm⁻¹) indicated an increased sp³ content (single bonds i.e. defects) when compared to graphite.³⁴ The D peak at 1324 cm⁻¹ (graphite) or 1326 cm⁻¹ (GO) is attributed to the breathing modes of sp² atoms in the ring and is usually activated by defects present on the graphitic surface.³⁵ The respective I_D/I_G (peak height) values of graphite and GO were 0.21 \pm 0.01 and 1.22 \pm 0.01, respectively (n = 3). Higher I_D/I_G value indicated an increased disorder of the graphitic structure, thus being more defective.³⁴ The relative thickness (number of layers) of the sheet was reflected by the lower intensity of 2D peaks at 2613 cm⁻¹ for GO (few layers) and higher 2D intensity at 2649 cm⁻¹ for graphite (> 10 layers).³³ Morphological examination of GO was further studied by TEM and AFM as shown in **Figure S3** and **S4**. GO suspension appeared to consist mainly of large and multi-layered GO sheets

GO-N₃ was then prepared using NaN₃ and 1,2-epoxide ring opening reaction as described in SI methods (Scheme S2). GO-N₃ was characterised by Raman spectroscopy and attenuated total reflectance Fourier transform infrared (ATR-FTIR) using the methods described in SI. A unique azide peak at 2121 cm⁻¹ was found when azide groups were introduced on GO. Free azide peak was detected at 2053 cm⁻¹ when the purification was not properly performed (Figure S5). All the oxygen functionalities found in GO were also observed in GO-N₃ except that the intensity of hydroxyl related peaks was higher (3200, 1227, and 1036 cm⁻¹) than that in GO (Figure S6A). The presence of a single azide peak at 2121 cm⁻¹ confirmed the elimination of free azides. Raman spectra were normalised to the G peak (intensity equals 1) (Figure S6B). Higher I_D/I_G (peak height) ratio was obtained for GO-N₃ (1.30 ± 0.05, n = 3) than GO (1.25 \pm 0.01, n = 3). The difference, however, was not significant with t(2) = -1.62, p-value = 0.246. The presence of azide group was further confirmed by Staudinger-Ninhydrin assay (SI methods). The reaction mechanism is described in Scheme S3. Staudinger reduction is a classical reaction reducing azide groups into amino groups with triphenylphosphine (PPh3).36 Cegielska and Kacprzak reported a simple method to visualise organic azides by combining Staudinger reduction with the traditional Ninhydrin assay.37 The azide was firstly reduced to amines then reacted with Ninhydrin forming Ruhemann's purple causing a colour change to either dark-violet to blue or orange-red. In this study, Ninhydrin assay showed negative results (colourless supernatants) for NaN3, GO and GO-N₃. Expectedly, Staudinger-Ninhydrin assay showed positive results (orange-red coloured supernatant) only for NaN₂ (positive control) and GO-N₃ samples (Figure S6C). Studinger-Ninhydrin assay and FT-IR confirmed the introduction of azide group on GO-N₃. In order to increase the azide group content on GO-N₃, m-chloroperoxybenzoic acid (mCPBA), a strong oxidising reagent, commonly used to epoxidise alkenes was used to introduce more epoxide groups on GO. Epoxidation of GO is generally not required as epoxides are known to be naturally present when graphite is oxidised to GO. ^{38,39} Direct epoxidation of GO with H₂O₂, however, has been reported to generate ultra-small graphene quantum dots (GQD) via epoxide ring opening. 40-42 To the best of our knowledge, direct epoxidation of GO with mCPBA has not been reported in the literature. In this work, mCPBA was therefore used to enrich epoxide content on GO surface to form epo-GO. Methods of epo-GO preparation is summarised in SI methods and presented in Scheme S4. Elemental analysis (C, H, N, O) was performed to determine N% in GO and GO-N₃ derivatives (Table S3). The N% for GO, GO-N₃ (- mCPBA), and GO-N₃ (+ mCPBA) were <0.10, 0.71, and 0.81%, respectively. The mCPBA pre-treatment increased the azide content by 14%. The IR spectra of GO and GO-N₃ (± mCPBA) are shown in Figure 1A. The azide peak at 2120 and 2122 cm⁻¹ was found for GO-N₃ (- mCPBA) and GO-N₃ (+ mCPBA), respectively.

After confirming the introduction of azide groups on GO, Click² GO was prepared *via* Steglich esterification as described in SI methods and Scheme 1. Samples were prepared with or without mCPBA, to mimic GO-N₃ preparation condition. The IR spectra of Click² GO are shown in Figure 1B. The azide peak at 2117 or 2120 cm⁻¹ was found in Click² GO (– mCPBA) or Click² GO (+ mCPBA), respectively. Similar to GO-N₃, the epoxidation treatment slightly increased the wavenumber of azide peak. The alkyne signal of Click²

Page 3 of 6 ChemComm

ChemComm COMMUNICATION

GO could not be picked up by Raman spectroscopy (Figure S7). This was expected as the -C=C- stretching vibration on marcomolecules generally has a weak IR transmittance. Furthermore, the peak location overlaps with the azide groups. The Raman spectra I_D/I_G (peak height) ratio was 1.32 ± 0.03 and $1.28 \pm$ 0.01 (n = 3) for Click² GO, without and with mCPBA, respectively. TGA was used to characterise GO, GO-N₃, and Click² GO. As shown in **Figure 2**, GO exhibited 4 stages of thermal-decomposition, represented by the 4 disconnected linear weight-loss intervals with curvy turning points. GO-N₃ also showed 4 linear weight-loss intervals with relatively sharp turning points. A higher and a sharper weight-loss slope was found in the linear range Ib than in Ia. These changes could be due to the decomposition of azide groups when the temperature approaches 200°C, as previously reported in the literature. 43 Click GO also showed a deep and steep drop in weight within the linear range Ic, however; only two linear ranges were found in Click² GO; the stage 4 decomposition observed in GO (IVa) and GO-N₃ (IVb) could not be seen in Click² GO. The introduction of TMS-protected propargyl alcohol on the carboxylic groups seems to have stabilised the GO at 800°C or higher. Detailed information on slopes analysis is summarised in Table S4 – S6.

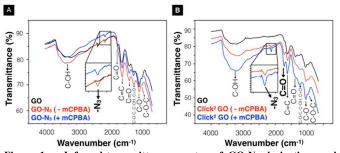


Figure 1. Infrared-transmittance spectra of GO-N₃ derivatives and Click² GO derivatives. (A) GO-N₃ derivatives. The introduction of azide groups was confirmed at 2120 cm⁻¹ (GO-N₃ – mPCBA), 2122 cm⁻¹ (GO-N₃ + mCPBA). (B) Click² GO derivatives. (Click² GO + mCPBA). Enhanced C-OH peaks (3200 cm⁻¹, 1227 cm⁻¹ and 1036 cm⁻¹) were observed. Full description of the peak locations can be found in SI.

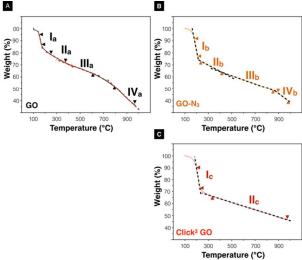


Figure 2. Thermogravimetric analysis. (A) GO and (B) GO-N₃ exhibit four linear decomposition ranges when heated from 100 - 978 °C. A steeper curve was found in linear range Ib than Ia. (C) Click² GO. Only two linear thermal-decomposition ranges were found throughout the heating range. It was noticed that the introduction of alkyne groups eliminated stage IV decomposition event observed in IVa and IVb.

In preparation of introducing the synthesised GO derivatives as potential drug carriers, a size reduction step was introduced by bath sonication up to 4-6 hr. Transmission electron microscopy (TEM) and Atomic Force Microscopy (AFM) were used to analyse the flake surface area (**Figure S3 and S4**). Maximum/medium size reduction from 13.19 μ m²/186 nm² (prior sonication, n = 370) to 2.15 μ m²/62 nm² (after 4 hr bath sonication, n = 621) was confirmed by AFM.

The Propargyl-modified blood-brain barrier targeting peptide angiopep-2 (TFFYGGSRGKRNNFKTEEYG)⁴⁴ and bis-azide polyethylene glycol (N₃-PEG₃₅₀₀-N₃) were synthesised using Fmoc solid phase peptide synthesis and reaction with imidazole-1-sulfonyl azide hydrochloride⁴⁵, respectively (SI method, Figure S8-9). The as-synthesised Click² GO was double-functionalised using two sequential CuAAC clicks. In brief, propargyl- modified angiopep-2 was firstly clicked to the azide groups of the GO yielding ANG-GO. The TMS-alkyne was then de-protected and clicked to N₃-PEG₃₅₀₀-N₃ yielding ANG-GO-PEG. Control click reactions were run in parallel. The first and second control click reactions were run lacking angiopep-2 and CuSO₄/L-ascorbic acid, respectively. The latter was performed to evaluate the physical absorption of PEG on the GO. Conjugation of angiopep-2 was confirmed by the reduction in the azide IR intensity (2120 cm⁻¹) and the appearance of amide I (1647 cm⁻¹, C=O stretching vibration) and amide II (1515 cm⁻¹, N-H bending vibration) signals on IR spectra of ANG-GO (Figure 3A). The conjugation of N₃-PEG₃₅₀₀-N₃ to GO in ANG-GO-PEG was confirmed by the appearance of the two C-H stretching vibration signals at 2869 and 2920 cm⁻¹ (Figure 3A). TGA showed evidence of thermal decomposition between 200-300°C, corresponding to angiopep-2, in both ANG-GO and ANG-GO-PEG. PEG decomposition on the other hand was observed at a temperature above 480°C (Figure 3B). The IR and TGA results of the control experiments are shown in Figure S10 further confirming that sequential click reactions took place. The TEM images of ANG-GO and ANG-GO-PEG are shown in Figure S11 and S12, respectively.

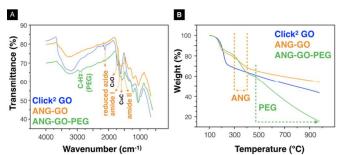


Figure 3. Characterisation of ANG-GO and ANG-GO-PEG. (A) Infrared-transmittance spectra and (B) thermogravimetric analysis of aniopep-2 functionalised Click² GO (ANG-GO) and angiopep-2/PEG double-functionalised Click² GO (ANG-GO-PEG).

Optical microscopy images of A549 incubated with GO, GO-N₃, Click² GO, ANG-GO, and ANG-GO-PEG were taken at 24 and 72 hr time points. Cells incubated with functionalised GO showed normal morphology at all concentrations tested, while cells treated with DMSO appeared unhealthy and detached from the plate (Figures S13). Click² GO then GO-N₃ appeared to interact better with cells compared to plain GO sheets, as shown by the dark signals localised to cells. The extent of GO uptake was concentration and time-dependent (Figures S14-15). This interaction was further enhanced in both ANG-GO and ANG-GO-PEG (Figure S16-17). The modified lactate dehydrogenase (LDH) assay was used to assess the cytotoxicity of all GO derivatives on A549 cells (adenocarcinoma human alveolar basal epithelial cells) using the method described in SI.46 Cells were incubated with GO, GO-N₃, Click²GO, ANG-GO, and ANG-GO-PEG at 10, 50, and 100 µg/mL for 24 and 72 hr. Dimethyl sulfoxide (DMSO) was used as a positive

control. The cytotoxicity result from the mLDH assay is shown in Figure 4. No significant effect on cell viability was observed in GO, CO-N₃, and Click² GO. Lower viability was observed in ANG-GO at 50 and 100 μ g/mL at 72 hr (p < 0.001). The viability of GO-ANG was restored when PEGylated (ANG-GO-PEG).

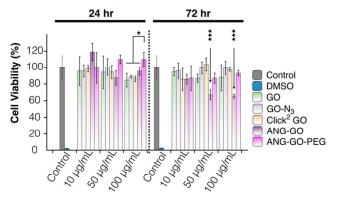


Figure 4. Modified LDH assay of A549 cells incubated with GO, GO-N₃, Click GO, ANG-GO, and ANG-GO-PEG. The toxicity was assessed using the modified LDH assay. No significant effect on cell viability was observed in GO, CO-N₃, and Click² GO. Lower viability was observed in ANG-GO at 50 and 100 µg/mL at 72 hr (p < 0.001). (* p< 0.05; ***p < 0.001). The viability was recovered in ANG-GO-PEG.

The double-clickable GO is reported for the first time, offering an eco-friendly method to perform CuAAC at room temperature in aqueous media, i.e., organic solvent-free. This is particularly important for GO functionalization as the GO sheets do not disperse well in organic solvents with layers tend to fold-up. Water-based reaction is expected to offer maximum exposure of GO surface to the reactants.

Conclusions

In this study, GO was synthesised using a modified Kovtyukhova-Hummer's method. 31 This method greatly improved the yield of GO (55.63%) when compared to the traditional modified Hummer's method (< 1%). The azidefunctionalisation of GO was achieved using NaN3. The azide content was boosted by 14% upon pre-treatment with mCPBA. Double functionalisation of Angiopep-2 and N₃-PEG₃₅₀₀-N₃ on Click² GO using sequential CuAAC clicks was demonstrated. No major effect on cell viability was found for Click² GO. Lower cell viability was found in ANG-GO at 50/100 µg/mL at 72 hr, which was reversed in ANG-GO-PEG. Click² GO is proposed here as an alternative graphene-based platform for applications in drug delivery.

Acknowledgements

K.-C.M. would like to thank King's College London for the Graduate School International Research Award (GSIRA) scholarship. Funding from Biotechnology and Biological Sciences Research Council (BB/J008656/1) and Wellcome Trust (WT103913MF) is acknowledged. H.K. was sponsored by the Atomic Energy Commission of Syria. P.M.C. is a Sir Henry Wellcome postdoctoral fellow. The author would also like to thank Dr. K.L. Andrew Chan at King's College London for constructive discussions.

Notes and references

- ^a Institute of Pharmaceutical Science, King's College London, Franklin-Wilkins Building, 150 Stamford Street, London SE1 9NH, United Kingdom
- ^b Biomaterials and Biomimetics Department, King's College London Dental Institute, London SE1 9RT, United Kingdom.

† Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here] DOI: 10.1039/c000000x/

- L. Feng, L. Wu and X. Qu, Adv. Mater., 2012, 25, 168-186.
- J. Liu, L. Cui and D. Losic, Acta Biomater., 2013, 9, 1-15.
- H. Li, Z. Song, X. Zhang, Y. Huang, S. Li, Y. Mao, H. J. Ploehn, Y. Bao and 3 M. Yu, Science, 2013, 342, 95-98.
- Y. Shao, J. Wang, H. Wu, J. Liu, I. A. Aksay and Y. Lin, Electroanalysis, 2010,
- Y. Song, K. Qu, C. Zhao, J. Ren and X. Qu, Adv. Mater., 2010, 22, 2206-2210.
- J.-L. Li, B. Tang, B. Yuan, L. Sun and X.-G. Wang, Biomaterials, 2013, 34, 6 9519-9534.
- 7 E. Lallana, A. Sousa-Herves, F. Fernandez-Trillo, R. Riguera and E. Fernandez-Megia, Pharm. Res., 2011, 29, 1-34.
- 8 J. Iehl and J.-F. Nierengarten, Chem. Commun., 2010, 46, 4160-4162.
 - G. Clavé and S. Campidelli, Chem. Sci., 2011, 2, 1887-1896.
- 10 M. Castelaín, G. Martínez, C. Marco, G. Ellis and H. J. Salavagione, Macromolecules, 2013, 46, 8980-8987.
- 11 S. Sun, Y. Cao, J. Feng and P. Wu, J. Mater. Chem., 2010, 20, 5605-5607.
- 12 H.-X. Wang, K.-G. Zhou, Y.-L. Xie, J. Zeng, N.-N. Chai, J. Li and H.-L. Zhang, Chem. Commun., 2011, 47, 5747-5749.
- 13 Z. Jin, T. P. McNicholas, C.-J. Shih, Q. H. Wang, G. L. C. Paulus, A. J. Hilmer, S. Shimizu and M. S. Strano, Chem. Mater., 2011, 23, 3362-3370.
- Y. Jing, H. Tang, G. Yu and P. Wu, Polym. Chem., 2013, 4, 2598-2607
- D. Zhou, Q.-Y. Cheng, Y. Cui, T. Wang, X. Li and B.-H. Han, Carbon, 2014, 66. 592-598.
- C. Deetuam, C. Samthong, S. Thongyai, P. Praserthdam and A. Somwangthanaroj, Compos. Sci. and Technol., 2014, 93, 1-8.
- M. Namvari and H. Namazi, Int. J. Environ. Sci. Technol., 2014, 11, 1527-
- Y. Pan, H. Bao, N. G. Sahoo, T. Wu and L. Li, Adv. Funct. Mater., 2011, 21, 18 2754-2763
- 19 L. Kou, H. He and C. Gao, Nano-Micro Lett., 2010, 2. 20
 - Z. Wang, Z. Ge, X. Zheng, N. Chen, C. Peng, C. Fan and Q. Huang, Nanoscale, 2012 4 394-399
- 21 S. Eigler, Y. Hu, Y. Ishii and A. Hirsch, Nanoscale, 2013, 5, 12136-12139.
- R. Salvio, S. Krabbenborg, W. J. M. Naber, A. H. Velders, D. N. Reinhoudt and 22
- W. G. van der Wiel, Chem. Eur. J., 2009, 15, 8235-8240. 23
- M. Namvari and H. Namazi, CARBOHYDRATE RESEARCH, 2014, 396, 1-8. 24 W.-C. Oh, A.-R. Jung and W.-B. Ko, J. Ind. Eng. Chem., 2007, 13, 1208-1214.
- M.-L. Chen and W.-C. Oh, Nanoscale Res. Lett., 2011, 6, 398.
- A. Karmakar, S. M. Bratton, E. Dervishi, A. Ghosh, M. Mahmood, Y. Xu, L. M. Saeed, T. Mustafa, D. Casciano, A. Radominska-Pandya, A. Biris and A. S. Biris, Int. J. Nanomedicine, 2011, 6, 1045-1055.
- 27 M.-L. Chen, Z.-D. Meng, L. Zhu, C.-Y. Park, J.-G. Choi, T. Ghosh, K.-Y. Cho and W.-C. Oh, J. Nanomater., 2012, 2012, 1-7.
- W. Steglich and B. Neises, Angew. Chem. Int. Ed., 1978, 17, 522-524.
- W. S. Hummers and R. E. Offeman, J. Am. Chem. Soc., 1958, 80, 1339-1339.
- 30 N. I. Kovtyukhova, P. J. Ollivier, B. R. Martin, T. E. Mallouk, S. A. Chizhik, E. V. Buzaneva and A. D. Gorchinskiy, Chem. Mater., 1999, 11, 771-778.
- K.-C. Mei, Y. Guo, J. Bai, P. Costa, H. Kafa, A. Protti, R. C. Hider and K. T. Al-Jamal, ACS Appl. Mater. Interfaces, 2015, 7, 14176–14181. 31
- H. Ali-Boucetta, D. Bitounis, R. Raveendran-Nair, A. Servant, J. Van den Bossche and K. Kostarelos, Adv. Healthc. Mater., 2013, 2, 433-441.
- 33
- A. C. Ferrari, *Solid State Commun.*, 2007, **143**, 47–57. A. Ferrari and J. Robertson, *Phys. Rev. B*, 2000, **61**, 14095–14107. 34
- 35 L. G. Cançado, A. Jorio, E. H. M. Ferreira, F. Stavale, C. A. Achete, R. B. Capaz, M. V. O. Moutinho, A. Lombardo, T. S. Kulmala and A. C. Ferrari, Nano Lett., 2011, 11, 3190-3196.
- H. Staudinger and J. Meyer, Helv. Chim. Acta., 1919, 2, 635-646.
- 37 B. Cegielska and K. M. Kacprzak, Chem. Anal., 2009, 54, 807-812.
- 38 J.-A. Yan and M. Y. Chou, Phys. Rev. B, 2010, 82, 125403.
- 39 J. R. Rani, J. Lim, J. Oh, J.-W. Kim, H. S. Shin, J. H. Kim, S. Lee and S. C. Jun, J. Phys. Chem. C, 2012, 116, 19010-19017.
- 40 F. Jiang, D. Chen, R. Li, Y. Wang, G. Zhang, S. Li, J. Zheng, N. Huang, Y. Gu, C. Wang and C. Shu, Nanoscale, 2013, 5, 1137-1142.
- 41 L. Li, G. Wu, G. Yang, J. Peng, J. Zhao and J.-J. Zhu, Nanoscale, 2013, 5,
- 42 T. Palaniselvam, M. O. Valappil, R. Illathvalappil and S. Kurungot, Energy Environ. Sci., 2014, 7, 1059-1067.
- 43 S. Eigler, Y. Hu, Y. Ishii and A. Hirsch, Nanoscale, 2013, 5, 12136-12139.
- R. Kurzrock, N. Gabrail, C. Chandhasin, S. Moulder, C. Smith, A. Brenner, K. 44 Sankhala, A. Mita, K. Elian, D. Bouchard and J. Sarantopoulos, Mol. Cancer Ther., 2012, 11, 308-316,
- E. D. Goddard-Borger and R. V. Stick, Org. Lett., 2007, 9, 3797-3800 45
- H. Ali-Boucetta, K. T. Al-Jamal and K. Kostarelos, in Biomedical Nanotechnology, ed. S. J. Hurst, Humana Press, Totowa, NJ, 2011, vol. 726, pp. 299-312

RSCPublishing

COMMUNICATION