ChemComm

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemcomm

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxx

ARTICLE TYPE

Simulating the Inhibition Reaction of *Mycobacterium tuberculosis* L,D-Transpeptidase 2 by Carbapenems

José Rogério A. Silva,^{*a,c*} Thavendran Govender,^{*b*} Glenn E. M. Maguire,^{*b*} Hendrik G. Kruger,^{*b*} Jerônimo Lameira,^{*a*} Adrian E. Roitberg, ^{**,c*} and Cláudio Nahum Alves^{**,a,c*}

s Received (in XXX, XXX) Xth XXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

A theoretical free energy study describes the inactivation of a new Tuberculosis target, the L,D-transpeptidase 2 enzyme. A new reaction mechanism of two carbapenem inhibitors is 10 proposed and molecular features are determined from QM/MM and PMF approaches. The theoretical findings with

the new proposed mechanism agree in principle with experimental data.

The *Mycobacterium tuberculosis* (*Mtb*) peptidoglycan layer is ¹⁵ maintained mainly by cross-link bonds built by transpeptidase enzymes. They cross-link the glycan chain by forming (D,D) $4\rightarrow 3$ and (L,D) $3\rightarrow 3$ peptide linkages, which are formed by D,D and L,D-transpeptidases, respectively.¹ Since 80% of the peptidoglycan cross-links are of the $3\rightarrow 3$ linkages, the L,D-²⁰ Transpeptidase (LDT) enzyme may have a major role in

peptidoglycan biosynthesis.² Among the five LDTs paralogues encoded by the chromosome Mtb strain, the LDT_{Mt2} show to be essential for virulence of acute infection in a mouse model.³

The LDTs enzymes are efficiently inhibited by carbapenems, ²⁵ such as meropenem, imipenem, doripenem and ertapenem.⁴ Carbapenems form an extremely important class of antibiotic β lactam compounds. Among hundreds of different β -lactams, carbapenems exhibit a wide spectrum of activity with the strongest potency against Gram-negative and Gram-positive

³⁰ bacteria.⁵ The development of carbapenems in treating of tuberculosis disease has raised wide interest since these compounds, in association with clavulanic acid as a β-lactamase inhibitor, are evenly active against extensively drug-resistant *Mtb* (XDR-TB).⁶ These drugs have been shown to kill both dormant ³⁵ and growing forms of the bacilli.^{6b}

The inhibition reaction of LDTs by carbapenems is similar to the first step of the LDT catalytic mechanism, which leads to enzyme acylation by the donor stem peptide⁷, where a covalent adduct is formed by the LDT active-site residue (cysteine) and

- ⁴⁰ the carbapenem β -lactam ring.⁸ Triboulet and co-workers have proposed that LDT inactivation by carbapanems is carried out in a two-step reaction.⁹ In the first part, a (reversible) tetrahedral intermediate is formed (EI^{ox}), the oxyanion. Whereas, the second step (irreversible) leads to formation of the acyl enzyme (EI*),
- ⁴⁵ the adduct form. However, their proposal from experimental observations does not take into account the probable protonation states for the catalytic residues.

Our recent theoretical study described that the acylation step is

preceded by thiol-imidazole pair formation (Cys and His catalytic ⁵⁰ residues) in its zwitterionic form,¹⁰ which will potentially be involved in the mechanism of LDT enzymes reacting with carbapenems.



Fig. 1 Carbapenem inhibitors of LDT employed in the present report.

In this report, we have applied hybrid quantum 55 $(QM/MM)^{11}$ mechanics molecular mechanics/molecular dynamics (MD) simulations and umbrella sampling¹² approaches to investigate the inactivation mechanism of LDT_{Mt2} by meropenem and imipenem, respectively. The calculations were 60 carried out with the self-consistent charge density functional tight binding (SCC-DFTB) semi-empirical method¹³ in the QM part of the system, using ff99SB classical force field14 for the protein and the TIP3P water model¹⁵ for the explicit solvent. The atom link method¹⁶ was used to satisfy the QM-MM boundary conditions. 65 The system building, theory level and energy minimization protocols are described in the ESI file. They are similar to the protocols applied previously on the LDT_{Mt2} catalytic mechanism.¹⁰ The SCC-DFTB method was successfully applied to β-lactam systems.¹⁷ Besides, to provide an independent vision 70 on the QM/MM systems described above, a model for the reaction in water was also simulated using same conditions (see ESI file for details).

The QM/MM umbrella sampling approach was performed in two steps (Scheme 1). (a) In the first one, two reaction ⁷⁵ coordinates were selected for the sampling: $RC_X = d(S_\gamma - C_7)$ (nucleophilic attack from S_γ of Cys354 side chain to C_7 of carbonyl group in the β -lactam ring of the inhibitor, sampled from 1.60 to 3.40 Å) and $RC_Y = d(C_7 - N_4)$ (breaking of $C_7 - N_4$ bond in the β -lactam ring of the inhibitor, sampled from 1.30 to $\epsilon_0 2.80$ Å), to simulate the acylation step. (b) In the second, just, one reaction coordinate was used for the sampling: $RC = d(N_\epsilon - H_\epsilon) - d(N_4 - H_\epsilon)$ (proton transfer from N_ϵ of His336 side chain to N_4 atom of carbonyl group in the β -lactam ring of the inhibitor, sampled from -2.50 to 1.50 Å). For each simulation window, 20 55

ps of MD production were preceded by 5 ps of equilibration with a time step of 1.0 fs. All the scans were executed in steps of 0.20 Å. This procedure follows the protocol applied previously for the catalytic mechanism.¹⁰ The QM/MM umbrella sampling simulations were performed within the Amber12 MD package.¹⁸ The 2D and 1D free energy surface (FES) for the acylation and proton transfer reaction steps, respectively, were calculated using the variational free energy profile (VFEP)¹⁹ (see ESI for details).¹⁰ Finally, the PMFs were corrected by means of M06-¹⁰ 2X-D3/MM single-point energies for the minimum free energy

 2X-D3/MM single-point energies for the minimum free energy path (MFEP) (see ESI for details).



Scheme 1 The inactivation mechanism proposal for the LDT_{Mt2} by carbapenem taking into account previous theoretical findings for the 1s catalytic mechanism.¹⁰ The blue atoms are in QM part of simulation models. The details about cutted bond on the QM/MM frontier are included on the ESI.

The 2D FES for the first step of the inactivation mechanism of LDT_{Mt2} by meropenem inhibitor is presented in Fig. 2. According ²⁰ to the results, the tetrahedral intermediate (EI^{ox}) proposed by Triboulet and co-workers,⁹ will not be formed during the reaction studied, since this step would be thermodynamically unfavourable ($\Delta G^{o}_{calc} > 20.00$ kcal mol⁻¹). Moreover, the ΔG^{o}_{calc} for reaction involving this inhibitor and LDT_{Mt2} is –13.08 kcal ²⁵ mol⁻¹ making the nucleophilic attack of thiol group of Cys354

- catalytic residue to carbonyl group in the β -lactam ring of the meropenem inhibitor thermodynamically favourable. These results are taking into account that the Cys-thiolate/His-imidazolium pair is in its zwitterionic form, as described
- ³⁰ previously.¹⁰ In addition, the transition state (TS₁) and anionic INT found on the MFEP (Fig. 2) are similar to structures proposed for the mechanism of carbapenem hydrolysis by CphA,^{17c} and subclass B2 metallo-β-lactamase.²⁰ Similar results were found for the imipenem (see ESI, Fig. S1). Graphical ³⁵ presentations for TS₁ and TS₂ are also presented in the ESI (Fig. S4).

Instead, the MFEP on this reaction shows a concerted mechanism to form an anionic intermediate state (INT, see Fig S4 in ESI for a graphical presentation) for the whole reaction, which ⁴⁰ is in accordance with experimental kinetics data.²¹ In energetic terms, considering this MFEP (Fig 2), the calculated activation

⁴⁰ Is in accordance with experimental kinetics data.²⁷ In energetic terms, considering this MFEP (Fig 2), the calculated activation free energy ($\Delta G_{calc}^{\ddagger} = 7.27$ kcal/mol) is underestimating the experimental value of activation free energy ($\Delta G_{exp}^{\ddagger} = 15.74$ kcal

mol⁻¹).²¹ In order to fix inaccuracies in estimating the 45 experimental absolute value by DFTB/MM potential, we have corrected the activation barrier using single-point at M06-2X-D3/MM level, where we have obtained $\Delta G_{calc}^{\dagger} = 17.41$ kcal mol⁻¹ at this potential, which is in remarkable agreement with the experimental value. Despite the DFTB/MM potential have failed 50 in describe the absolute value of $\Delta G_{exp}^{\ddagger}$, it is important to point out that the theoretical energetics obtained for imipenem and meropenem have the same trend of reported experimental data²⁰, which support the mechanism described by our DFTB/MM free energy surface (see Table S2 and S3).



Fig. 2 2D Free energy surface (FES) obtained by DFTB/MM for the first step of meropenem in LDT_{Mt2}. R: reactant; TS₁: transition state; INT: intermediate; EI^{ox}: oxyanion state. The energy values are report in the ⁶⁰ kcal mol⁻¹. The red dashed line present the minimum energy path (MFEP), while the black dashed line present the experimental proposal.

For the second step, the value for $\Delta G_{calc}^{\ddagger}$ at M06-2X-D3/MM level is 20.00 kcal mol⁻¹ (~2.60 kcal mol⁻¹ higher than the preceding step) and shows a concerted mechanism similar to the ⁶⁵ first step (see ESI, Fig. S2). This result agrees with the experimental results.²¹ A crystal structure of the LDT_{M12}meropenem complex adopts two different adduct forms for the drug side chain after covalent bond linkage,²² referred as state I and II. Our computational study shows that the P state found on ⁷⁰ the second step is in good accordance to the experimental state I (see ESI, Fig. S3), which is not the ground state for the meropenem inhibitor covalently linked to Cys catalytic residue.⁸, ²²

More thorough investigation of this reaction mechanism ⁷⁵ (covalent bond formation between the inhibitor and the catalytic site) is necessary to better understand the main features that contribute to the inactivation of LDTs by carbapanems and for others β -lactam compounds. The relevant calculated parameters (bond distances) found for the chemical species upon inactivation ⁸⁰ of the active site by these carbapenem inhibitors are summarized in the ESI file (Table S1).

As highlighted previously, the experimental proposal of inactivation of LDTs by carbapenems does not consider the zwitterionic form of the catalytic residues as the most stable sprotonation state in the enzyme active site. In this report, the theoretical results provided from the MFEP about the inhibition

75

95

100

105

mechanism of LDT_{Mt2} agree in principle with experimental values. The complete reaction mechanisms including the experimental and theoretical values are summarized in the ESI (Fig. S4 and Table S2, respectively). The $\Delta G_{calc}^{\ddagger}$ values obtained

⁵ for the whole reaction at M06-2X-D3/MM level are 17.41 and 20.00 kcal mol⁻¹ for the first and second step, respectively, which is suitable to experimental evidences. Finally, Our QM/MM simulations may be useful for the design of more potent carbapenem LDT inhibitors, leading to the development of new ¹⁰ antibiotics against tuberculosis and other bacilli infections.

We thank to University of Florida Research Computing for the allotment of computational resources. The authors J.R.A.S and C.N.A. gratefully acknowledge the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and

¹⁵ Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Brazilian agencies, UKZN and NRF for financial support. Finally, this work was conducted during a scholarship supported by Program PNPD/CAPES at the Federal University of Pará.

20 Notes and references

^aLaboratório de Planejamento e Desenvolvimento de Fármacos, Instituto de Ciências Exatas e Naturais, Universidade Federal do Pará, Belém, Pará, Brazil. Fax: 55 91-3201 7363; Tel: 55 91-3201 8235; E-mail: nahum@ufpa.br

- ^bCatalysis and Peptide Research Unit, School of Health Sciences, University of Kwazulu-Natal, Durban 4001, South Africa.
 ^cDepartment of Chemistry, University of Florida, Gainesville, Florida, United States. Fax: 352 392 8722; Tel: 352 392 6972; E-mail: roitberg@ufl.edu
- ³⁰ † Electronic Supplementary Information (ESI) available: [major details of materials and methods, including the PDB code for the crystal structure used. Free energy surfaces, relevant parameters and theoretical findings for the all reactions calculated]. See DOI: 10.1039/b000000x/
- ³⁵ 1(a) Wietzerb.J, B. C. Das, J. F. Petit, E. Lederer, Leyhboui.M and J. M. Ghuysen, *Biochemistry-Us*, 1974, 13, 3471; (b) G. L. Moraes, G. C. Gomes, P. R. M. Sousa, C. N. Alves, T. Govender, H. G. Kruger, G. Lamichhane and J. Lameira, ¹ *Tuberculosis*, 2015, 95, 95.
- 40 2 M. Lavollay, M. Arthur, M. Fourgeaud, L. Dubost, A. Marie, N. Veziris, D. Blanot, L. Gutmann and J. L. Mainardi, J Bacteriol, 2008, 190, 4360.
 B. Gunta, M. Lavollay, L. L. Mainardi, M. Arthur, W. R. ¹¹⁵
- 3 R. Gupta, M. Lavollay, J. L. Mainardi, M. Arthur, W. R. Bishai and G. Lamichhane, *Nat Med*, 2010, **16**, 466.
- 45 4 J. L. Mainardi, J. E. Hugonnet, F. Rusconi, M. Fourgeaud, L. Dubosth, A. N. Moumi, V. Delfosse, C. Mayer, L. Gutmann, L. B. Rice and M. Arthur, *J Biol Chem*, 2007, **282**, 30414.
- 5 K. M. Papp-Wallace, A. Endimiani, M. A. Taracila and R. A. Bonomo, *Antimicrob Agents Ch*, 2011, 55, 4943.
- 50 6(a) J. E. Hugonnet and J. S. Blanchard, *Biochemistry-Us*, 2007, 46, 11998; (b) J. E. Hugonnet, L. W. Tremblay, H. I. Boshoff, C. E. Barry and J. S. Blanchard, *Science*, 2009, 323, 1215.
- 7 D. J. Tipper and Stroming.Jl, *P Natl Acad Sci USA*, 1965, **54**, 1133.
- 8 H. S. Kim, J. Kim, H. N. Im, J. Y. Yoon, D. R. An, H. J. Yoon, J. Y. Kim, H. K. Min, S. J. Kim, J. Y. Lee, B. W. Han and S. W. Suh, *Acta Crystallogr D*, 2013, **69**, 420.
- 9(a) S. Triboulet, M. Arthur, J. L. Mainardi, C. Veckerle, V. Dubee, A. NGuekam-Moumi, L. Gutmann, L. B. Rice and J. E. Hugonnet, *J Biol Chem*, 2011, 286, 22777; (b) S. Triboulet, V. Dubee, L. Lecoq, C. Bougault, J. L. Mainardi, L. B. Rice, M. Etheve-Quelquejeu, L. Gutmann, A. Marie, L. Dubost, J. E. Hugonnet, J. P. Simorre and M. Arthur, *Plos One*, 2013, 8.

- 10 J. R. A. Silva, A. E. Roitberg and C. N. Alves, J Chem Inf Model, 2014, 54, 2402.
- A. Warshel and M. Levitt, J Mol Biol, 1976, 103, 227; (b)
 H. M. Senn and W. Thiel, Angew Chem Int Edit, 2009, 48, 1198.
- B. Honarparvar, T. Govender, G. E. M. Maguire, M. E. S. Soliman and H. G. Kruger, *Chem Rev*, 2014, 114, 493; (b) J. Kastner, *Wires Comput Mol Sci*, 2011, 1, 932.
- M. Elstner, D. Porezag, G. Jungnickel, J. Elsner, M. Haugk, T. Frauenheim, S. Suhai and G. Seifert, *Phys Rev B*, 1998, 58, 7260; (b) M. Elstner, D. Porezag, G. Jungnickel, T. Frauenheim, S. Suhai and G. Seifert, *Mater Res Soc Symp P*, 1998, 491, 131.
- 14 V. Hornak, R. Abel, A. Okur, B. Strockbine, A. Roitberg and C. Simmerling, *Proteins*, 2006, 65, 712.
- 15 W. L. Jorgensen, J. Chandrasekhar, J. D. Madura, R. W. Impey and M. L. Klein, *J Chem Phys*, 1983, **79**, 926.
- 16 M. J. Field, P. A. Bash and M. Karplus, *J Comput Chem*, 1990, **11**, 700.
- ⁸⁵ 17(a) D. Xu, H. Guo and Q. Cui, J Am Chem Soc, 2007, **129**, 10814;
 (b) D. G. Xu, H. Guo and Q. Cui, J Phys Chem A, 2007, **111**, 5630;
 (c) S. S. Wu, D. G. Xu and H. Guo, J Am Chem Soc, 2010, **132**, 17986;
 (d) E. I. Chudyk, M. A. L. Limb, C. Jones, J. Spencer, M. W. van der Kamp and A. J. Mulholland, Chem Commun, 2014, **50**, 14736;
 (e) M. Zheng and D. G. Xu, J Phys Chem B, 2013, **117**, 11596.
 - D. A. Case, T. A. Darden, T. E. Cheatham, C. L. Simmerling, J. Wang, R. E. Duke, R. Luo, R. C. Walker, W. Zhang, K. M. Merz, B. Roberts, S. Hayik, A. Roitberg, G. Seabra, J. Swails, A. W. Goetz, I. Kolossváry, K. F. Wong, F. Paesani, J. Vanicek, R. M. Wolf, J. Liu, X. Wu, S. R. Brozell, T. Steinbrecher, H. Gohlke, Q. Cai, X. Ye, J. Wang, M. J. Hsieh, G. Cui, D. R. Roe, D. H. Mathews, M. G. Seetin, R. Salomon-Ferrer, C. Sagui, V. Babin, T. Luchko, S. Gusarov, A. Kovalenko and P. A. Kollman, University of California, San Francisco, 2012; (b) G. D. Seabra, R. C. Walker, M. Elstner, D. A. Case and A. E. Roitberg, J Phys Chem A, 2007, 111, 5655.
 - T. S. Lee, B. K. Radak, A. Pabis and D. M. York, J Chem Theory Comput, 2013, 9, 153; (b)
 T. S. Lee, B. K. Radak, M. Huang, K. Y. Wong and D. M. York, J Chem Theory Comput, 2014, 10, 24.

20(a) D. L. Gatti, *Plos One*, 2012, **7**; (b) S. H. Ackerman and D. L. Gatti, *Plos One*, 2013, **8**.

- 110 21 M. Cordillot, V. Dubee, S. Triboulet, L. Dubost, A. Marie, J. E. Hugonnet, M. Arthur and J. L. Mainardi, *Antimicrob Agents Ch*, 2013, 57, 5940.
 - 22 W. J. Li, D. F. Li, Y. L. Hu, X. E. Zhang, L. J. Bi and D. C. Wang, *Cell Res*, 2013, **23**, 728.

This journal is © The Royal Society of Chemistry [year]

