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Dioxygenation of cysteamine to hypotaurine at a tris(pyrazolyl)borate iron(II) unit – cysteamine dioxygenase mimicking?†‡

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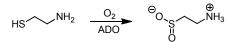
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The novel complex $Tp^{Me,Ph}Fe(SCH_2CH_2NH_2)$ has been synthesized as a speculative model for the ADO. Indeed its reaction with O₂ led to the dioxygenation of the S atom and thus to hypotaurine. This finding may allow to draw conclusions on the constitution of the ADO active site, whose structure is still unknown.

Beside the cysteine dioxygenase (CDO) the only other thiol dioxygenase active in mammals is the cysteamine (2-aminoethanethiol) dioxygenase (ADO).¹ It catalyses the dioxygenation of the thiol group in 2-aminoethanethiol released during co-enzyme A degradation with dioxygen to produce hypotaurine (Scheme 1).



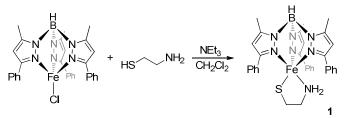
Scheme 1 Oxidation of protonated cysteamine to hypotaurine catalyzed by the ADO. $^{\rm 1}$

While the CDO, its mechanism and its mimicking has been intensely investigated within the last 10 years,²⁻⁸ the ADO has received comparatively little attention. It is known to be based on iron and to belong to the cupine family but apart from that many questions remain open with respect to its active site and functioning.^{1a} The fact that CDO and ADO mediate a similar reaction setting out from a thiol substrate suggests that there is a phylogenetic connection between them.⁹

The CDO active site contains an iron centre surrounded by three facially arranged histidine ligands, and to this unit the S-deprotonated cysteine substrate then binds in a bidentate fashion via the amine and the thiolate functions, leaving one coordination site open for the coordination and activation of O_2 .³ Through a complex mechanism the two O atoms are finally transferred to the sulfur atom yielding a cysteine sulfinic acid.

Modelling this reactivity with molecular compound is complicated by the tendency of iron(II) thiolates to form Fe^{III}-O-Fe^{III} units and disulfanes upon addition of O_2 .⁵ Nevertheless research aimed at mimicking the CDO has met with some success in recent years.⁵⁻⁸ After first reports by Goldberg and co-workers on the trioxygenation of thiolates coordinated to iron,⁵ we were able to report the successful coordination and dioxygenation of a protected cysteinato ligand at a (His)₃Fe^{II} analogue, namely a Tp^{Me,Ph}Fe⁺ complex metal fragment.⁷ To test the hypothesis that the ADO active site is related to the one of CDO an investigation on the oxidation of cysteamine at the Tp^{Me,Ph}Fe⁺ scaffold thus suggested itself.

To synthesise a $Tp^{Me,Ph}Fe$ -cysteamine complex a solution of 2aminoethane thiol in dichloromethane was treated with triethylamine and subsequently a solution of $Tp^{Me,Ph}FeCl^{10}$ in the same solvent was added. After stirring for 12 h work-up yielded a light yellow solid of pure $Tp^{Me,Ph}Fe(SCH_2CH_2NH_2)$, 1, in 43 % yield (Scheme 2).



Scheme 2 Synthesis of [Tp^{Me,Ph}Fe(SCH₂CH₂NH₂)], **1**.

Through slow evaporation of the solvent from a solution of **1** in dichloromethane crystals could be grown that were investigated via X-ray diffraction, and the result is shown in Figure 1.

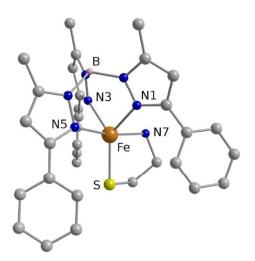
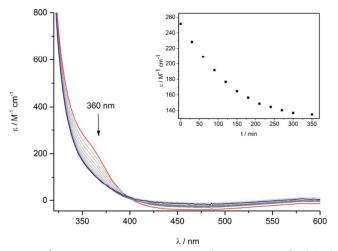


Fig. 1 Molecular structure of **1** (all hydrogen atoms have been omitted for clarity). Selected bond lengths (Å) and angles (°): Fe–S 2.3174(6), Fe–N7 2.2520(18), Fe–N1 2.1531(16), Fe–N3 2.1102(16), Fe–N5 2.2489(16), S–Fe–N7 84.96(5).

As expected S-deprotonated cysteamine coordinates as a chelating ligand, so that a distorted trigonal bipyramidal coordination sphere results for the iron(II) centre ($\tau = 0.57$). The Fe–S and Fe–N bond lengths of 2.3174(6) Å and 2.2520(18) Å differ only slightly from those found in the cysteine-based analogue.⁷ As there the iron centre is prepared for O₂ binding and activation, since sufficient space remains open within its coordination sphere for O₂ to bind and it exists in its high-spin state: The effective magnetic moment of **1** as determined for a C₆D₆ solution by the Evans method at room temperature amounted to 5.36 µB, which is close to the spin-only value expected for four unpaired electrons (4.90 µB).

To investigate the reactivity of $[Tp^{Me,Ph}Fe(SCH_2CH_2NH_2)]$ towards O₂ a solution in dichloromethane was exposed to O₂ under anhydrous conditions. Within 4 h a colour change from light yellow to dark green occurred that was monitored by UV/Vis spectroscopy (Figure 2).



Scheme 3 The reaction of **1** with O_2 and conceivable products resulting from work-up: cysteamine (a), hypotaurine (b), 2-aminoethanesulfenic acid (c), taurine (d) and cystamine (e).

Fig. 2 UV/Vis spectral changes in course of the reaction of **1** (blue) dissolved in dichloromethane with O_2 within 6 h (red); the inset shows the behaviour of the 360 nm absorbance with time.

Work-up of the reaction mixture was performed by addition of 3 M hydrochloric acid as well as chelex in MeOH to separate

In the course of the reaction a shoulder found in the spectrum of $[Tp^{Me,Ph}Fe(SCH_2CH_2NH_2)]$, 1 at 360 nm ($\epsilon = 231 \text{ M}^{-1} \text{ cm}^{-1}$), which may be assigned to a S \rightarrow Fe charge-transfer loses in intensity and after 6 h it is vanished almost completely. A similar observation had been made in case of the cysteinate complex, and it may indicate the cleavage of the Fe–S bond.

The final reaction mixture was also investigated by ESI-MS. A dichloromethane solution of 1 gives rise to a major signal at 615.18 m/z corresponding to $[1]^+$ as well as to a signal at 638.18 m/z assigned to the Na⁺ adduct of 1 (see Supporting information). After O2 treatment two new signals were observed at 670.17 m/z, matching the spectrum calculated for $[(1+O_2)+Na]^+$, as well as a signal at 685.16 m/z belonging to the corresponding K⁺ analogue (as sodium ions also potassium ions from the environments are sometimes observed to form adducts⁷). To confirm these assignments, also experiments with ¹⁸O-enriched dioxygen (99 %) were performed. In the ESI-TOF spectrum recorded subsequently from the reaction solution, a shift of the $[(1+O_2)+Na]^+$ peak by four mass units to 674.18 m/z could be observed, indicating that in the course of the reaction with O2 two O atoms are incorporated, that is, a dioxygenation takes place (see Supporting information). Under the more dilute conditions of the ¹⁸O₂ experiment additionally a signal at 697.25 m/z appeared, which could not be assigned. It sometimes was also observed in the ¹⁶O₂ experiments and thus does not belong to an oxygenation product.

Recording an EPR spectrum of the final product mixture in the perpendicular mode did not reveal any signal, that is, the sample is EPR silent, as one would expect for a product complex with iron in the oxidation state +II.

To demonstrate that the dioxygenation occurs selectively at the cysteamine ligand, a work-up procedure was developed for the reaction mixture, which finally allowed for the isolation of the cysteamine part of the reaction product. First of all three compounds were conceivable and had to be distinguished: (a) cysteamine in case of selective oxygenation of the Tp ligand, (b) the sulfinic acid hypotaurine after selective dioxygenation of the cysteamine ligand and (c) the corresponding sulfenic acid as a product of monooxygenation. Beyond that, two other products had to be considered, although these are not in agreement with the ESI-TOF results: An SS-bond formation, which often occurs on coordination of thiolates to highly oxidized metal centres, leads to the disulphide (e), a tripleoxygenation, as observed in the case of some models for the CDO reported in the literature,^{5,6} to taurine (d), as shown in Scheme 3.

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all iron ions, and washing with toluene removed the decomposition product of Tp. The investigation of all residual organic components was performed by ¹H NMR spectroscopy, and the spectrum obtained was then compared with the spectra of cysteamine, hypotaurine, taurine and cystamine, all of which feature two triplets at characteristic chemical shifts (see Figure 3).

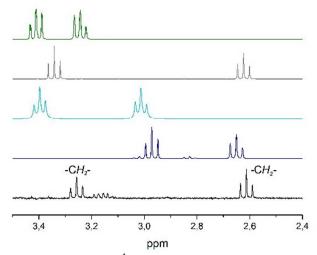


Fig. 3 Comparison of the ¹H-NMR spectra of the isolated oxidation product (black), cysteamine (dark blue), cystamine (light blue), hypotaurine (grey) and taurine (green) dissolved in D_2O .

Clearly, the spectrum of hypotaurine fitted best to the one recorded after work-up, although the low-field triplet occurred shifted by 0.09 ppm to higher field. To test whether impurities like residual iron ions are responsible for this shift commercially obtained hypotaurine was added to the sample of the oxygenation product, and as still only one signal set was observed afterwards, it thus became clear that hypotaurine is indeed the dioxygenation product. Consequently, the reaction of 1 with O_2 leads to the product $[Tp^{Me,Ph}Fe(O_2SCH_2CH_2NH_2)]$, 2. Due to the instability of 2 all attempts to grow single crystals that might have allowed a characterization via X-ray diffraction have failed so far.

Altogether the results suggest the following logic operation: The active site structure of the ADO is still unknown but it may resemble the one of the CDO, which shows a rather similar reactivity. The active centre of CDO contains an iron(II) ion coordinated by three histidine residues, which binds the cysteine in a chelating fashion. In previous work a TpFe^{II} complex with a cysteinate ethyl ester ligand proved capable of mimicking the CDO both structurally and functionally.⁷ Hence, in this work a complex Tp^{Me,Ph}Fe(SCH₂CH₂NH₂), **1**, has been synthesized as a speculative model for the ADO. Indeed its reaction with O₂ led to the dioxygenation of the S atom and thus to hypotaurine. This result lends further support to the hypothesis that the active sites of CDO and ADO are quite similar.

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Notes and references

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‡ Electronic Supplementary Information (ESI) available: Additional information pertaining to the method, synthesis and analysis as well as crystallographic data are presented. See DOI: 10.1039/c000000x/

- a) J. E. Dominy, Jr., C. R. Simmons, L. L. Hirschberger, J. Hwang, R. M. Coloso, M. H. Stipanuk, J. Biol. Chem., 2007, 282, 25189-25198; b) beyond that two bacterial homologues of CDO were identified, which catalyse the oxidation of 3-mercaptopropionate: J. E. Dominy Jr., C. R. Simmons, L. L. Hirschberger, J. Hwang, R. M. Coloso, M. H. Stipanuk, J. Biol. Chem. 2007, 282, 25189.
- 2 J. G. McCoy, L. J. Bailey, E. Bitto, C. A. Bingman, D. J. Aceti, B. G. Fox, G. N. Phillips, Jr., *Proc. Natl. Acad. Sci. USA*, 2006, **103**, 3084-3089.
- 3 S. Ye, X. Wu, L. Wei, D. Tang, P. Sun, M. Bartlam, Z. Rao, J. Biol. Chem., 2007, 282, 3391-3402.
- 4 S. Aluri, S. P. de Visser, J. Am. Chem. Soc., 2007, 129, 14846-14847.
- 5 Y. Jiang, L. R. Widger, G. D. Kasper, M. A. Siegler, D. P. Goldberg, J. Am. Chem. Soc., 2010, 132, 12214-12215.
- 6 Y. M. Badiei, M. A. Siegler, D. P. Goldberg, J. Am. Chem. Soc., 2011, 133, 1274-1277.
- 7 M. Sallmann, I. Siewert, L. Fohlmeister, C. Limberg, C. Knispel, Angew. Chem., 2012, 124, 2277-2280; Angew. Chem. Int. Ed., 2012, 51, 2234-2237.
- 8 A. C. McQuilken, Y. Jiang, M. A. Siegler, D. P. Goldberg, J. Am. Chem. Soc., 2012, 134, 8758-8761.
- J. E. Dominy, Jr., C. R. Simmons, L. L. Hirschberger, J. Hwang, R. M. Coloso, M. H. Stipanuk, *J. Biol. Chem.*, 2007, 282, 25189-25198;
 D. Buongiorno, G. D. Straganz, *Coord. Chem. Rev* 2013, 257, 541–563.
- 10 T. Tietz, C. Limberg, R. Stößer, B. Ziemer, *Chem. Eur. J.*, 2011, 17, 10010–10020.