

# Analytical Methods

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4 **1 Highly sensitive and selective colorimetric detection of Hg<sup>2+</sup> based on**  
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6 **2 separation of Hg<sup>2+</sup> and formation of catalytic DNA-gold**  
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8 **3 nanoparticles**  
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13 4 Chi-Fang Peng\*, Na Pan, Zheng-Jun Xie and Liang-Liang Wu  
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21 **7 Abstract:** Hg<sup>2+</sup> ions can be absorbed onto DNA-AuNPs complex and separated from  
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23 8 water samples while the catalytic activity of DNA-AuNPs can be promoted. Based on  
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25 9 the above principle, a highly sensitive and selective colorimetric assay for the  
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27 10 detection of Hg<sup>2+</sup> was developed. The proposed method for Hg<sup>2+</sup> has a detection limit  
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29 11 of 1.5 nM with a linear range from 5.0 nM to 500 nM. Moreover, this detection  
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31 12 method for Hg<sup>2+</sup> demonstrated more than 1000-fold selectivity toward most of the  
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33 13 possible interfering ions and can be used for tap water detection. The results indicate  
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35 14 the high potential of AuNPs as enzyme mimic for sensing heavy metal ions by  
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37 15 metallophilic interactions.  
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43 **16 Introduction**  
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46 17 Mercuric ions (Hg<sup>2+</sup>) are widely distributed in the environment and have  
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48 18 deleterious effects on the environment and human health. Monitoring Hg<sup>2+</sup> level in  
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50 19 water is a very important task in terms of water safety and water quality. The U.S.  
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52 20 Environmental Protection Agency (EPA) has set the maximum allowable level of Hg  
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54 21 in drinking water at 10 nM (2.0 ppb).<sup>1</sup> At present, there are various classical methods  
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56 22 for detecting Hg<sup>2+</sup>, including atomic fluorescence spectrometry,<sup>2</sup> atomic absorption  
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3 23 spectrometry,<sup>3</sup> inductively coupled plasma mass spectrometry,<sup>4</sup> These methods are  
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5 24 selective and sensitive, but require expensive instruments and complicated sample  
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7 25 pretreatments.<sup>5</sup>  
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10 26 In order to overcome these problems, many metallic nanoparticle-based sensors  
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12 27 have been developed.<sup>6-15</sup> In these nanosensors, various specific ligands of Hg<sup>2+</sup> were  
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14 28 carefully selected to achieve selective respond to Hg<sup>2+</sup>. For example, DNA-gold  
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16 29 nanoparticle (AuNP) assays have demonstrated excellent selectivity for Hg<sup>2+</sup> that can  
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18 30 interact with T-T mismatches to form T-Hg<sup>2+</sup>-T complexes.<sup>8-10, 15</sup> Many small ligands,  
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20 31 including (11-mercapto-undecyl)-trimethyl-ammonium, mercaptopropionic acid,<sup>13</sup>  
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22 32 homocystine, lysine,<sup>14</sup> diethyldithiocarbamate,<sup>6</sup> 5-methyl-2-thiouracil,<sup>12</sup>  
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24 33 dithiocarbamate derivative of calixarene,<sup>7</sup> and 1-dodecanethiol,<sup>11</sup> modified on the  
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26 34 surface of AuNPs also have demonstrated specific interaction with Hg<sup>2+</sup>. This  
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28 35 ligand-mediated interaction can be measured by fluorescent, colorimetric,  
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30 36 chemiluminescent, electrochemical and electrochemiluminescent methods, etc.<sup>6, 11, 14,</sup>  
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32 37 <sup>16-18</sup> Interestingly, Hg<sup>2+</sup> also can interact strongly and selectively with gold  
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34 38 nanoparticles,<sup>19</sup> gold nanorods<sup>5</sup> and silver nanoparticles,<sup>20</sup> which results in forming  
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36 39 Au/Hg and Ag/Hg alloys (amalgam). This specific metallophilic interaction<sup>21</sup> has  
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38 40 been used to develop fluorescent and colorimetric sensors of Hg<sup>2+</sup>.<sup>5, 19, 20, 22, 23</sup>  
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40 41 Although most of these methods have demonstrated high sensitivity to Hg<sup>2+</sup> and good  
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42 42 selectivity to other common metal ions, it is still difficult to eliminate the interference  
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44 43 from the high concentration of various metal ions in real samples without proper  
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46 44 sample purification. In order to solve this problem, some specific materials, such as  
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48 45 YPA4 resin microcolumn and Hg<sup>2+</sup>-imprinted polymers,<sup>24, 25</sup> were prepared to  
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50 46 separate or preconcentrate Hg<sup>2+</sup> from real samples.  
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3 47 Recently, some methods for  $\text{Hg}^{2+}$  detection were developed based on the  
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5 48 formation of Hg-Au alloy NPs which possess higher peroxidase-like activity than  
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7 49 naked AuNPs. For example, Long *et al.*<sup>23</sup> utilized the above Hg-Au alloy NPs to  
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9 50 catalyze TMB- $\text{H}_2\text{O}_2$  redox reaction and developed a colorimetric assay for  $\text{Hg}^{2+}$   
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11 51 detection. Yan *et al.*<sup>22</sup> and Wang *et al.*<sup>26</sup> developed fluorescent detections of  $\text{Hg}^{2+}$  on  
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13 52 the basis of the oxidation of o-phenylenediamine and Amplex UltraRed, catalyzed by  
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15 53 the above Au-Hg peroxidase-mimic. Although these methods have demonstrated  
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17 54 excellent analytical performance for  $\text{Hg}^{2+}$  detection, the high concentration of other  
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19 55 metal ions in samples may produce serious matrix interference. Improving the  
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21 56 selectivity and sensitivity of  $\text{Hg}^{2+}$  detection is an effective strategy to avoid matrix  
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23 57 effects.

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27 58 In this work, we improved Long's method for  $\text{Hg}^{2+}$  detection by using a  
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29 59 DNA-AuNPs complex. In our strategy,  $\text{Hg}^{2+}$  ions were deposited onto DNA-AuNPs  
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31 60 complex and separated from water samples, and then quantified by measuring the  
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33 61 catalytic activity of the formed Au/Hg amalgam. This new method has several  
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35 62 advantages: (i) The selectivity of this detection for  $\text{Hg}^{2+}$  was outstanding due to the  
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37 63 combination of the metallophilic interaction of Au-Hg and further separation of  
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39 64 Au-Hg amalgam through centrifugation. (ii) This proposed method was highly  
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41 65 sensitive with a detection limit of 1.5 nM and a linear range from 5.0 nM to 500 nM.  
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43 66 The sensitivity of this method is satisfactory for environmental water samples. (iii)  
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45 67 The preparation of DNA- AuNPs complex is simple and the detection is low-cost.  
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## 51 **Experimental Section**

### 52 53 54 69 **Reagents and chemicals**

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3 70 Hydrogen tetrachloroaurate (III) tetrahydrate ( $\text{HAuCl}_4 \cdot 4\text{H}_2\text{O}$ ) were purchased from  
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5 71 Aldrich (Milwaukee, WI). Trisodium citrate ( $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7 \cdot 2\text{H}_2\text{O}$ ) and citric acid  
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7 72 monohydrate ( $\text{C}_6\text{H}_8\text{O}_7 \cdot \text{H}_2\text{O}$ ) were obtained from Sigma (St. Louis, MO). The  
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9 73 thiolated single strand oligonucleotides, 5'-SH-TTTTTTTTTT-3' ( $\text{T}_{10}$ ) and  
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11 74 5'-SH-GCGACATGGTAATGG-3' (a random sequence, rDNA), were purchased  
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13 75 from Sangon Biotech (Shanghai) Co., Ltd.  $\text{Hg}(\text{NO}_3)_2$  and all the other metal salts  
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15 76 were purchased from the national institute of metrology P. R. China standard. 3, 3, 5,  
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17 77 5-tetramethylbenzidine (TMB) was supplied by Sinopharm Group Chemical Regent  
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19 78 Co., Ltd (Shanghai, China). All of the reagents used were of analytical grade.  
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21 79 Ultra-pure water prepared with a Milli-Q Pure system was used throughout the  
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23 80 experiments.  
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### 27 81 **Synthesis of AuNPs**

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29 82 AuNPs were prepared through citrate-mediated reduction of  $\text{HAuCl}_4$ .<sup>27</sup> Firstly,  
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31 83 0.01% chloroauric acid solution (100 mL) was added to a flask which had been  
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33 84 soaked with aqua regia solution and cleaned with ultra-pure water. Then the flask was  
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35 85 stirred and heated until boiling for 10 min. 1.0% sodium citrate (2.0 mL) was  
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37 86 subsequently added quickly to the boiling solution. An obvious color change of the  
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39 87 reaction mixture was observed from transparent to dark blue and finally wine red. The  
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41 88 mixture was further boiled for another 5 min and cooled to room temperature.  
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### 45 89 **Preparation of DNA-AuNP complex**

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47 90 The DNA-functionalized Au nanoparticles were prepared by directly incubating  
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49 91 the thiolated DNA strands with AuNP solution. Firstly, the prepared AuNP solution  
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51 92 (3 nM) was concentrated to 6 nM by centrifugation. Then the concentrated AuNPs (6  
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53 93 nM, 990  $\mu\text{L}$ ) were mixed with thiolated DNA (100  $\mu\text{M}$ , 10  $\mu\text{L}$ ). After incubated at  
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55 94 25°C for 24 h, the mixtures were centrifuged for 15 min at 10000 rpm and the excess  
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3 95 DNA was removed. After repeating the centrifugation for twice the obtained  
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5 96 DNA-AuNP complex was resuspended in ultra-pure water and stored at 4°C.  
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### 7 **Analytical procedure for the colorimetric detection of Hg<sup>2+</sup>**

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10 98 Different concentrations of Hg<sup>2+</sup> (500 μL) were mixed with DNA-AuNPs solution  
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12 99 (0.6 nM, 25 μL) and trisodium citrate (8 mM, 175 μL), and then incubated for 30 min  
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14 100 at room temperature. Then the mixtures were centrifuged at 10000 rpm for 15 min  
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16 101 and the supernatants were removed, by which 50 μL of concentrated solution was  
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18 102 obtained. Afterward, 90 μL of citrate buffer (100 mM, pH 4.5), 100 μL of TMB (1.5  
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20 103 mM) and 60 μL of H<sub>2</sub>O<sub>2</sub> (1.5 M) were pipetted into the above concentrated solution.  
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22 104 The absorbance value of the mixture was then measured at 650 nm by a microplate  
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24 105 reader (PowerWave XS2, Bio-Teck, USA).  
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## 29 **Results and discussion**

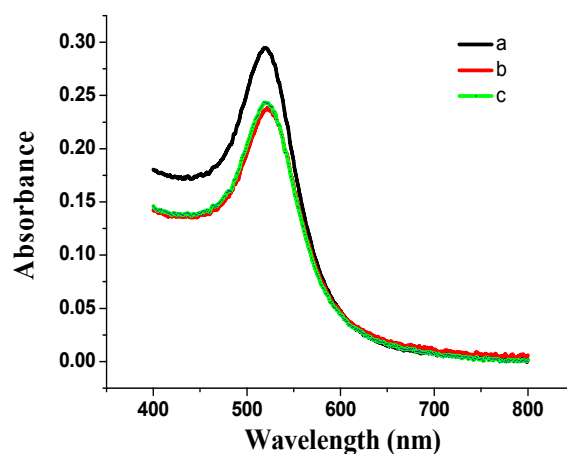
### 30 **Characterization of AuNPs and DNA-AuNPs complex**

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33 107  
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35 108 The dimension of the AuNPs was measured with a transmission electron  
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37 109 microscope (TEM, H7100, Hitachi High-Technologies Corporation, Tokyo, Japan).  
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39 110 The AuNPs were monodisperse with an average size of 16 nm (Figure S1). The  
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41 111 concentration of AuNPs was estimated to be 3.0 nM. Hg<sup>2+</sup> can be absorbed by AuNPs  
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43 112 through strong Au-Hg metallophilic interaction, but the formed Au/Hg alloy is  
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45 113 unstable.<sup>19</sup> In order to separate Hg<sup>2+</sup> from solution, AuNPs should be protected and  
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47 114 stabilized by some molecules.<sup>19</sup> Considering that AuNPs can be modified controllably  
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49 115 with thiolated DNA, a DNA-AuNP complex was prepared and used to separate Hg<sup>2+</sup>  
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51 116 ions.  
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56 117 Figure 1 shows the UV-vis absorption spectra of the AuNPs with a maximum  
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58 118 absorption peak (λ<sub>max</sub>) at 519 nm. After AuNPs modified with T<sub>10</sub>, the λ<sub>max</sub>  
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3 119 red-shifted from 519 nm to 522 nm. These results confirmed the successful  
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5 120 conjugation of AuNPs and DNA strands. After T<sub>10</sub>-AuNPs were incubated with a high  
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7 121 concentration of Hg<sup>2+</sup> (500 nM), the  $\lambda_{\max}$  blue-shifted from 522 nm to 520 nm, which  
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9 122 should attribute to that Hg<sup>2+</sup> ions were reduced to Hg<sup>0</sup> and then adsorbed onto the  
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11 123 surface of the DNA-AuNPs.<sup>28</sup> It should be noted that the spectrum of DNA-AuNPs  
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13 124 kept almost unchanged in terms of intensity and shape after incubated with high  
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15 125 concentration of Hg<sup>2+</sup>. The TEM images of AuNPs, T<sub>10</sub>-AuNPs and T<sub>10</sub>-AuNPs  
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17 126 treated with Hg<sup>2+</sup> (500 nM) also showed that all three kinds of Au NPs are  
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19 127 mono-dispersed (Figure S1).

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23 128 The above result confirmed the high stability of DNA-AuNPs/Hg<sup>2+</sup> complex that  
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25 129 will be beneficial to the separation of Hg<sup>2+</sup> from water samples.  
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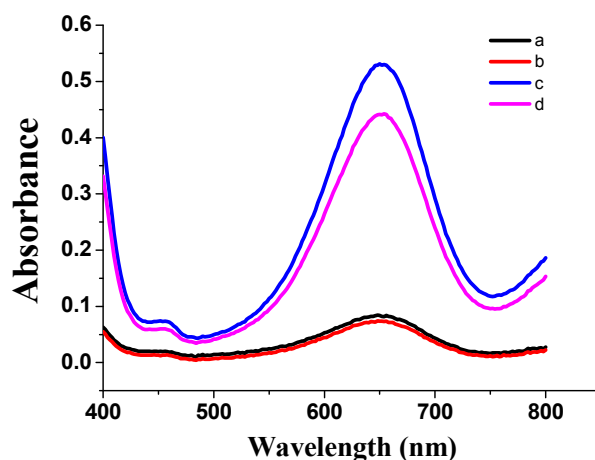
44 131  
45 132 Figure 1. UV-Vis spectra of (a) AuNPs, (b) T<sub>10</sub>-AuNPs and (c) T<sub>10</sub>-AuNPs treated with Hg<sup>2+</sup> (500  
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47 133 nM).  
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#### 50 134 **Effects of DNA on the detection of Hg<sup>2+</sup>**

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52 135 Long *et al.*<sup>23</sup> found that the forming of Hg-Au alloy NPs stimulated the catalytic  
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54 136 ability of Au NPs to TMB-H<sub>2</sub>O<sub>2</sub> redox reaction by accelerating the decomposition of  
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3 137 H<sub>2</sub>O<sub>2</sub>. Here this mechanism can be used to detect Hg<sup>2+</sup> further after Hg<sup>2+</sup> was  
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5 138 separated from water sample by DNA-AuNPs.

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7 139 To confirm that the redox reaction between TMB and H<sub>2</sub>O<sub>2</sub> was accelerated by the  
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10 140 formed Hg–Au alloys, the UV-Vis spectra were recorded separately in the presence of  
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12 141 AuNPs, T<sub>10</sub>-AuNPs, AuNPs/Hg<sup>2+</sup> and T<sub>10</sub>-AuNPs/Hg<sup>2+</sup>. As shown in Figure 2, the  
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14 142 absorbance value at 650 nm (A<sub>650</sub>) in the presence of AuNPs or T<sub>10</sub>-AuNPs is very  
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16 143 low, indicating their very weak catalysis toward the redox reaction of TMB-H<sub>2</sub>O<sub>2</sub>  
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18 144 (curve a and curve b in Figure 2 ). However, this redox reaction can be catalyzed by  
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20 145 AuNPs/Hg<sup>2+</sup> system obviously (curve c in Figure 2), indicating the catalytic capacity  
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22 146 promotion of AuNPs by Hg<sup>2+</sup>. Moreover, the catalytic capacity of the T<sub>10</sub>-AuNPs can  
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24 147 be promoted almost the same as that of the AuNPs (curve d in Figure 2).  
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45 150 Figure 2. UV-vis spectra of TMB and H<sub>2</sub>O<sub>2</sub> after incubation with different catalytic system: (a)  
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47 151 AuNPs; (b) T<sub>10</sub>-AuNPs; (c) AuNPs + Hg<sup>2+</sup> (500 nM); (d) T<sub>10</sub>-AuNPs + Hg<sup>2+</sup> (500 nM).  
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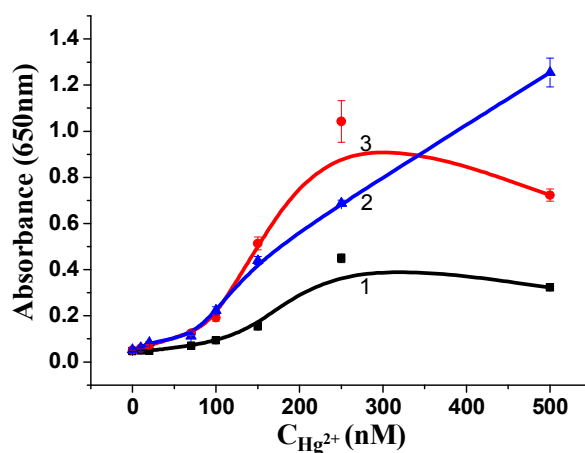
51  
52 153 The AuNPs modified with T<sub>10</sub> (T<sub>10</sub>-AuNPs) or a random ssDNA (rDNA-AuNPs)  
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54 154 were utilized to separate and detect Hg<sup>2+</sup> ions. As expected, the absorbance values at  
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56 155 650 nm (A<sub>650</sub>) increased rapidly as the increasing of Hg<sup>2+</sup> concentrations in the range  
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3 156 of 50-500 nM when the T<sub>10</sub>-AuNPs complex was utilized to separate and detect Hg<sup>2+</sup>  
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5 157 (Figure 3). In contrast, the A<sub>650</sub> increased slowly when AuNPs without modification  
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7 158 of DNA were used. In the case of the rDNA-AuNPs complex, the A<sub>650</sub> increased more  
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9 159 rapidly over the range 10-250 nM than that of T<sub>10</sub>-AuNPs. However, the A<sub>650</sub> started  
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11 160 to decline as the Hg<sup>2+</sup> concentration increased above 250 nM.

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14 161 It was known that T<sub>10</sub> and Hg<sup>2+</sup> can interact and form T-Hg<sup>2+</sup>-T complex, which  
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16 162 probably reduced the absorption of Hg<sup>2+</sup> on the surface of AuNPs. Meanwhile, it was  
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18 163 reported that T<sub>n</sub> sequences possess more extended and upright conformations on the  
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20 164 surface of AuNPs than rDNA.<sup>29</sup> The different performance between rDNA-AuNPs  
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22 165 and T<sub>10</sub>-AuNPs in the sensing of Hg<sup>2+</sup> should be due to the different interaction of  
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24 166 DNA strands and Hg<sup>2+</sup> as well as the different conformations of T<sub>10</sub> and rDNA  
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26 167 sequences on the surface of AuNPs. Considering the sensitivity and a wide linear  
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28 168 respond to Hg<sup>2+</sup> concentration, T<sub>10</sub>-AuNPs complex was selected for the next  
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30 169 experiment.

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51 172 Figure 3. Effect of DNA on the colorimetric detection for Hg<sup>2+</sup>. The TMB-H<sub>2</sub>O<sub>2</sub> redox reaction  
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53 173 catalyzed by citrate-capped AuNPs (1), T<sub>10</sub>-AuNPs (2) and rDNA-AuNPs (3) after the addition of  
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55 174 various concentrations of Hg<sup>2+</sup> ions (0, 10, 20, 70, 100, 150, 250 and 500 nM). The concentration  
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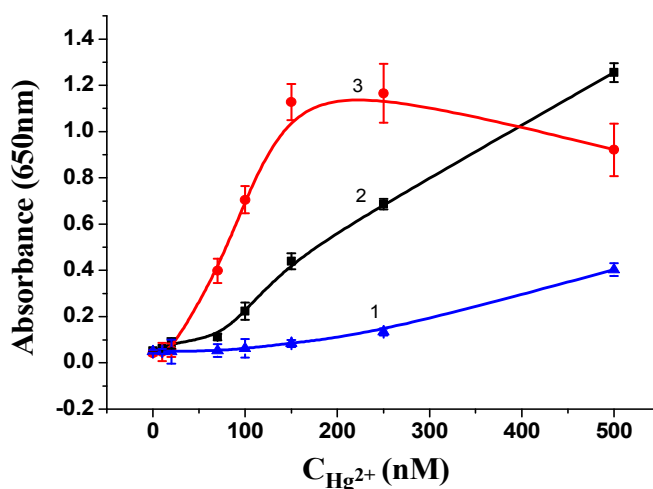
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3 of AuNPs, T<sub>10</sub>-AuNPs and rDNA-AuNPs, 0.6 nM; TMB, 1 mM; H<sub>2</sub>O<sub>2</sub>, 1.5 M; pH, 4.5; reaction  
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5 176 time, 25 min.

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8 177 Since macromolecules on the metallic nanoparticles can affect their catalytic  
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10 178 activity,<sup>30</sup> the concentrations of T<sub>10</sub> used to modify the AuNPs were investigated. It  
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12 179 was found that the T<sub>10</sub>-AuNPs showed highest catalytic activity when 1.0 μM of T<sub>10</sub>  
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14 180 was utilized (Figure S2).

### 181 **Effect of Hg<sup>2+</sup> volume**

182 Overall, the catalytic activity of T<sub>10</sub>-AuNPs increased with increasing the Hg<sup>2+</sup>  
183 volume from 100 μL to 1000 μL. As a result, sharper respond to Hg<sup>2+</sup> concentration  
184 occurred when bigger volume of Hg<sup>2+</sup> solution was utilized to detect Hg<sup>2+</sup> (Figure 4).  
185 However, a shoulder peak appeared when using 1000 μL of Hg<sup>2+</sup> solution for Hg<sup>2+</sup>  
186 detection. This phenomenon should be due to the excessive deposition of Hg<sup>2+</sup> onto  
187 the surface of the T<sub>10</sub>-AuNPs complex, which led to destabilization of the T<sub>10</sub>-AuNPs  
188 complex. A wide linear range of 10-500 nM can be obtained whether 100 μL or 500  
189 μL of Hg<sup>2+</sup> was mixed with the T<sub>10</sub>-AuNPs complex, but much higher sensitive  
190 respond was found with 500 μL of Hg<sup>2+</sup> used than 100 μL of Hg<sup>2+</sup>.

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3 193 Figure 4. The colorimetric detection for  $\text{Hg}^{2+}$  by  $\text{T}_{10}$ -AuNPs after incubated with different  
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5 194 volume of  $\text{Hg}^{2+}$  (From 1 to 3, the volumes of  $\text{Hg}^{2+}$  were: 100  $\mu\text{L}$ , 500  $\mu\text{L}$  and 1000  $\mu\text{L}$ ,  
6  
7 195 respectively. The concentrations of  $\text{Hg}^{2+}$  were 0, 10, 20, 70, 100, 150, 250 and 500 nM,  
8  
9 196 respectively).  $\text{T}_{10}$ -AuNPs, 0.6 nM; TMB, 1.0 mM;  $\text{H}_2\text{O}_2$ , 1.5 M; pH, 4.5; reaction time, 25 min.

### 197 **Sensitivity and selectivity of the detection of $\text{Hg}^{2+}$**

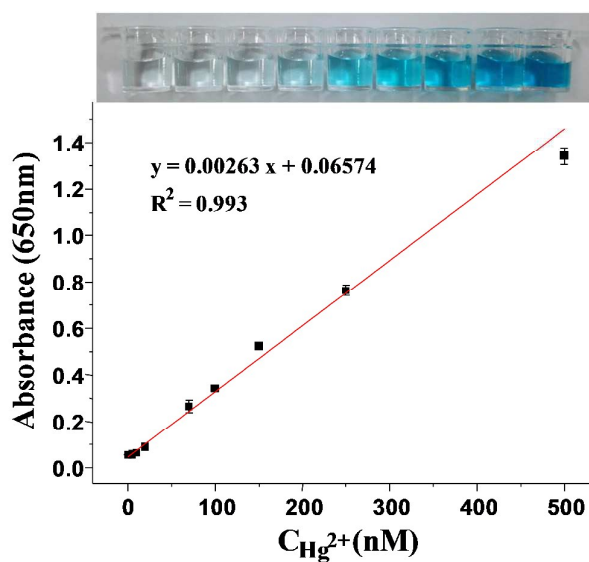
198 Several parameters including the concentrations of TMB and  $\text{H}_2\text{O}_2$  and reaction  
199 time were investigated to optimize the conditions for the colorimetric detection of  
200  $\text{Hg}^{2+}$  ions. As shown in Figure S3, the  $A_{650}$  increased as reaction time increasing and  
201 reached a plateau after 20 min. In Figure S4, it can be seen that the highest  $A_{650}$  was  
202 found at 1.5 M over the range of 0.9-1.5 M  $\text{H}_2\text{O}_2$ . The maximal difference of  
203 absorbance values in the presence and absence of  $\text{Hg}^{2+}$  was achieved at 1.5 M of  
204  $\text{H}_2\text{O}_2$ .

205 In the presence of 200 nM  $\text{Hg}^{2+}$ , the  $A_{650}$  increased sharply with increasing the  
206 concentration of TMB in the range of 0.1-1.0 mM and then reached a plateau at 1.5  
207 mM TMB (Figure S5). The highest  $A_{650}$  was found at 0.6 nM of  $\text{T}_{10}$ -AuNPs over the  
208 range 0.2-3.0 nM (Figure S6). In addition, the background signal increased with the  
209 increasing concentration of  $\text{T}_{10}$ -AuNPs.

210 Under the optimal conditions,  $\text{Hg}^{2+}$  level as low as 20 nM can be clearly detected  
211 by the naked eye and a calibration curve for  $\text{Hg}^{2+}$  detection was obtained with a linear  
212 range from 5 nM to 500 nM ( $R^2 = 0.993$ ) (Figure 5). The limit of detection was  
213 estimated to be 1.5 nM at a signal-to-noise ratio of 3, which was more sensitive than  
214 most of previously reported colorimetric and some of the fluorescent methods (some  
215 representative methods were listed in Table S1).<sup>5-9, 13, 14, 17, 22, 26, 31, 33, 34</sup>

216 To investigate the selectivity of the  $\text{T}_{10}$ -AuNPs probe (0.6 nM) toward  $\text{Hg}^{2+}$ , 10  $\mu\text{M}$   
217  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Sr}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Al}^{3+}$ ,  $\text{Co}^{2+}$ ,  $\text{Bi}^{3+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Pb}^{2+}$  or

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3 218  $\text{Fe}^{3+}$  was added into the  $\text{T}_{10}$ -AuNPs, separately. As a control, we added 20.0 nM and  
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5 219 200 nM  $\text{Hg}^{2+}$  ions into the probe solutions. Figure 6 reveals that only mercury can  
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7 220 stimulate the peroxidase-like activity of the AuNPs. As expected, the tolerance of the  
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9 221  $\text{T}_{10}$ -AuNPs probe for  $\text{Hg}^{2+}$  ions toward most of the possible interfering ions was more  
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11 222 than 1000-fold except that it was more than 500-fold toward  $\text{Ca}^{2+}$ . The selectivity of  
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13 223 our method for  $\text{Hg}^{2+}$  detection was higher than most of the reported colorimetric  
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15 224 methods.<sup>5, 7, 19, 23, 31, 32</sup> This high selectivity should be due to the selective separation of  
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17 225  $\text{Hg}^{2+}$  from water as well as the consequent  $\text{Hg}^{2+}$ -stimulated the peroxidase-like  
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19 226 activity of AuNPs.  
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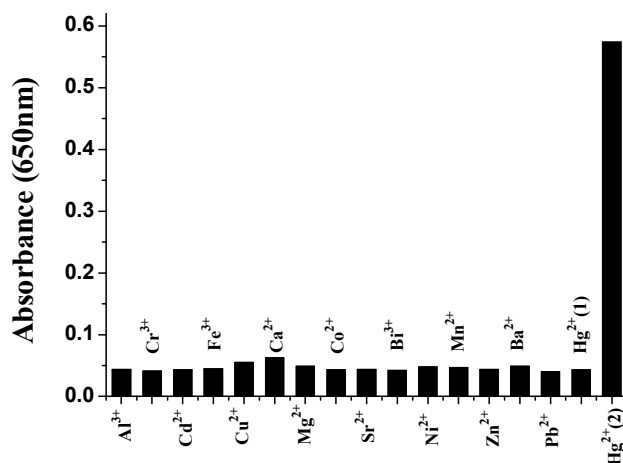


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229 Figure 5. Photographic images of the colors and calibration curve for the detection of  $\text{Hg}^{2+}$  after  
230 the addition of various concentrations of  $\text{Hg}^{2+}$  ions (0, 5, 10, 20, 70, 100, 150, 250 and 500 nM).

231  $\text{T}_{10}$ -AuNPs, 0.6 nM; TMB, 1.5 mM;  $\text{H}_2\text{O}_2$ , 1.5 M; pH, 4.5; reaction time, 20 min.

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234 Figure 6. Catalytic activity of T<sub>10</sub>-AuNPs stimulated by various metal ions. All metal ions are 10235  $\mu\text{M}$  except for Hg<sup>2+</sup> (1) (20 nM) and Hg<sup>2+</sup> (2) (200 nM). T<sub>10</sub>-AuNPs, 0.6 nM; TMB, 1.5 mM;236 H<sub>2</sub>O<sub>2</sub>, 1.5 M; pH, 4.5; reaction time, 20 min.237 **Detection of Hg<sup>2+</sup> ions in tap water samples**238 To test the practicality of the proposed method, the T<sub>10</sub>-AuNPs were used to detect239 Hg<sup>2+</sup> ions in tap water. The tap water samples collected from our laboratory were240 detected by atomic fluorescence spectrometry and the concentration of Hg<sup>2+</sup> was

241 lower than 2.5 nM. After aliquots of tap water were spiked with 10, 50 and 200 nM

242 Hg<sup>2+</sup> ions, respectively, the samples were filtered through microfiltration membranes

243 and measured by our method. The recoveries of 90.9-112.0% with relative standard

244 deviation (RSD) of 2.0-8.9% were shown in Table 1. The results demonstrated that

245 the proposed colorimetric method can accurately and reliably determine Hg<sup>2+</sup> in real

246 samples.

247 Table 1. Determination of Hg<sup>2+</sup> in tap water samples (n = 3)

Added Hg <sup>2+</sup> (nM)	Found Hg <sup>2+</sup> (nM)	Recovery (%)	RSD (%)
10	11.2	112.0	8.9
50	53.1	106.2	2.0

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## 248 Conclusions

249 A highly sensitive and selective colorimetric assay for the detection of  $\text{Hg}^{2+}$  ions  
250 was developed based on the simple separation of  $\text{Hg}^{2+}$  by DNA-AuNPs and the  
251  $\text{Hg}^{2+}$ -stimulated the peroxidase-like activity of AuNPs. The DNA strands modified on  
252 the surface of AuNPs can stabilize the Au-Hg alloy NPs and keep their  
253 peroxidase-like activity. The proposed method has a detection limit of 1.5 nM  $\text{Hg}^{2+}$   
254 with a linear range from 5.0 nM to 500 nM. Moreover, the above method of  $\text{Hg}^{2+}$   
255 detection showed ultra-high selectivity toward other common metal ions. The  
256 developed colorimetric method opens up a new possibility for monitoring trace level  
257 of heavy metal ions in water samples.

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261 and the Natural Science Foundation of Jiangsu Province (BK20141108).

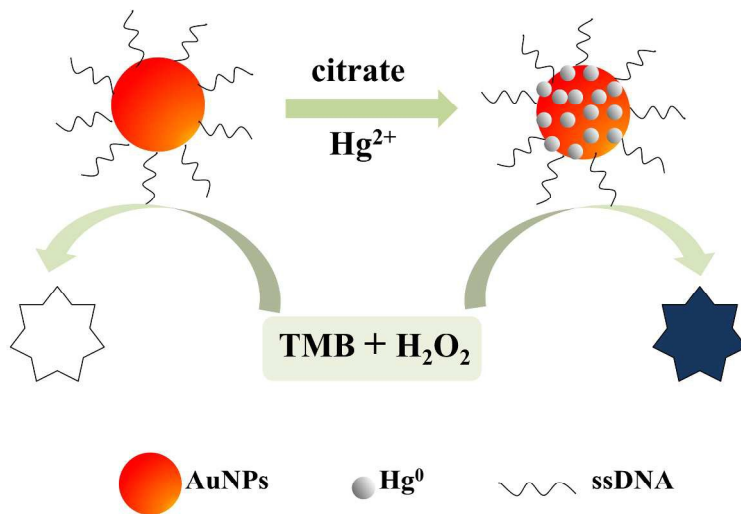
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