

Analytical Methods

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1 ASSESSMENT OF PHOTO-OXIDATIVE ALTERATIONS TO NATURAL ORGANIC MATTER IN
2 WATER USING FLUORESCENCE EXCITATION EMISSION MATRICES AND THE LIQUID
3 CHROMATOGRAPHY-ORGANIC CARBON DETECTION TECHNIQUES

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9 **Abstract**

10 The fluorescence excitation emission matrices (FEEM) and the liquid chromatography-
11 organic carbon detection (LC-OCD) are relatively novel techniques currently used in
12 analysing natural organic matter (NOM) in water. Regardless of the complexity of the
13 natural water matrices, the LC-OCD is able to characterize and quantify NOM as five
14 fractions; biopolymers, humic substances, building blocks, low molecular weight neutrals,
15 low molecular weight acids. The FEEM on the other hand characterises chromophoric
16 dissolved organic matter and maps out their emission and excitation spectral contours. In
17 the current study, these techniques were used to characterize and quantify NOM fractions
18 in raw water used for cooling at Eskom power stations, as well as monitor the changes in
19 their composition when irradiated with different intensities of sunlight using a sun
20 simulator. The effect of hydrogen peroxide (H₂O₂) (an oxidant) together with exposition to
21 light on NOM was also investigated. Firstly, results showed that humic substances decreased
22 with increased sunlight intensities. Secondly, a similar trend was observed with the
23 introduction of an oxidant such as H₂O₂. The dissolved organic carbon (DOC) in irradiated
24 samples decreased from 9.100 ppm (in raw water) to 8.832, 8.838 and 8.175 ppm for 0.6-
25 sun, 1.2-sun and 2.0-sun, respectively. At constant sunlight intensity (of 1.2-sun), addition of
26 increasing amounts of 30% H₂O₂ showed that the distribution of NOM fractions changed
27 significantly. Generally, a decreasing trend in molecular weight of NOM was observed with
28 increased irradiation intensity as well as increased oxidant amounts. The fluorescent
29 signature peaks showed that the humic substances (HS) contours changed but remained
30 relatively stable with an increase in light intensity. The results of the peroxide effect on
31 humic substances showed disappearing spectral signature peaks. No spectral contours in
32 the HS region were formed for samples illuminated by near natural sunlight intensity in the
33 presence of H₂O₂. These results are particularly important in understanding NOM in water
34 used for cooling in power generation stations.

35 **Keywords:** Fluorescence excitation emission matrices; Humic substances; Natural organic
36 matter, Liquid chromatography-organic carbon detection; Sunlight

1. INTRODUCTION

Natural organic matter (NOM) research has witnessed ground breaking studies over the last 10 years. This decadal research has generated valuable information on NOM characterisation and fractionation in soils and natural waters (1-4). Over these years, NOM is reported to play an important role in chemical interactions in water (5, 6). However, NOM being a highly complex matrix, only studies on its fractions will give useful information on how it chemically interacts with other dissolved components in water. It is therefore necessary, to characterise and discuss individual fractions of NOM and their chemical behaviour. For example, humic acid (which dissolves in alkaline conditions) and fulvic acid can control a number of chemical interactions in natural water. These include dissolution, mineralization, precipitation, adsorption, chemical transport and complexation (7-9). Therefore, the NOM fractions ought to be well characterized and quantified. In addition, their alterations due to chemical changes need to be addressed.

It is noteworthy that NOM serves as microbial food and the resulting by products of organic nature termed as autochthonous (microbial) NOM [10]. Furthermore, NOM is important in many other processes including UV-visible light attenuation, buffering pH and binding metals, among others (10-16). The NOM metal binding processes control the transport and bio-cycling of carbon and nitrogen compounds (17). These could be in the form of organometallic and coordination compounds. Compared to allochthonous (terrestrial) NOM (25–30% aromatic carbon), autochthonous NOM has a lower aromaticity degree (12–17% aromatic carbon). In addition, autochthonous NOM is more labile and has a lower capability to block UV-radiation (18, 19).

However, the phenomenon of UV-penetration into surface waters has an impact on NOM fractions. Changes in UV light penetration and NOM lability needs to be investigated in our natural surface water that serve domestic and industrial purposes. Power generating companies in South Africa (Eskom), for example, use water for conventional cooling of turbines. Their cooling chambers are supplied by raw water from the rivers with NOM inputs and other chemical constituents. When metals such as Ca and Mg combine with inorganic anions in the raw water to form salts at specific physicochemical conditions, the salts precipitate on pipe walls forming scale deposits, which are very difficult to remove. This forces the power generation companies to replace their water pipes from time to time (20).

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3 70 It is important to note that the presence of certain NOM fractions in a higher proportion,
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5 71 may be a solution to problems such as scaling in pipes. This is because, some NOM fractions
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7 72 are large organic molecules (such as humic acid (HA) and fulvic acid (FA)) and could act as
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9 73 excellent ligands that coordinate with metals to form relatively stable complexes. Other
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11 74 literature expresses this in terms of chelate effect (21). However, the complexation capacity
12
13 75 will depend on the physico-chemical conditions (such as pH, temperature, light penetration,
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15 76 presence of oxidants and their composition. This study seeks to show the changes in
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17 77 composition of raw water as a function of changing sunlight intensities. Furthermore, the
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19 78 changes to NOM in the presence of an oxidant were investigated. This 'photo-oxidation' of
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21 79 natural organic matter was determined using raw water obtained from Vaal River, which
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23 80 serves Lethabo and Kriel power stations (of Eskom, South Africa) with cooling water.

23 81 **2. EXPERIMENTAL**

24 82 *2.1. Sampling and sample pre-treatment*

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28 83 Raw water samples from Vaal River were collected at the points feeding Lethabo and Kriel
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30 84 power stations and immediately filtered through a 0.45 μm PES membrane and stored at 4
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32 85 °C. Cooling water was collected at Lethabo and Kriel and similar treatment to that of raw
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34 86 water applied and stored for analysis.

35 36 87 *2.2 Instrumental analysis*

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39 88 The FEEM measurements were conducted using the fluorescence AquaLog Spectrometer
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41 89 (Horiba, New Jersey, USA). The NOM fractions were characterized and quantified by the LC-
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43 90 OCD (DOC-Labor, Karlsruhe, Germany). Prior to LC-OCD chromatographic separation,
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45 91 samples were passed through 0.45 μm filters and all the soluble organics were identified by
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47 92 the LC-OCD system and the FEEM.

48 49 93 *2.2.1 Light simulation studies*

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51 94 The UV-visible light used in this study was obtained from a light simulator (**Fig. 1**) that
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53 95 producing light that was measured with the equivalent of natural sunlight. Accordingly, an
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55 96 equivalent of natural sunlight was termed *1-sun*; twice the natural sunlight termed *2-sun*;
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57 97 thrice termed as *3-sun*; and so on. These intensities are obtained by adjusting the sample
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59 98 distance between **a** and **b** at a power of 400 to 500W (**Fig. 1**).

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3 99**Fig. 1**4
5 100 2.2.2 LC-OCD analysis

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7 101 The LC-OCD technique, developed to identify fractions natural organic matter in water, gives
8 102 quantitative information and qualitative results regarding molecular size distribution of
9 103 organic matter in water (22). This technique separates components on the basis of their
10 104 molecular size. The separated fractions include humic substances, biopolymers, building
11 105 blocks, low molecular weight organic acids, neutrals and hydrophobic organic carbon. In
12 106 addition, it generates an improved diagram for the characterisation of aquatic humic
13 107 substances (HS-diagram) (7). The HS-diagram is a plot of the spectral absorption coefficient
14 108 (SAC) to the organic carbon (OC) ratio (aromaticity) of HS fractions against their nominal
15 109 molecular weights. The spectral absorption coefficient (SAC) is obtained with the ultra-violet
16 110 detection. The SAC/OC ratio describes the specific UV absorption of the HS peak, and a
17 111 measure of HS aromaticity, since the response in UVD reflects aromatic and unsaturated
18 112 structures. As shown, a close correlation exists between HS aromaticity and HS-molecularity
19 113 (7).

20 114 Water samples were injected into a column filled with a chromatographic gel material
21 115 where large molecules are eluted first followed by the smaller compounds (22). To achieve
22 116 quantification of organic carbon, organic nitrogen and specific UV absorbance at λ -254, their
23 117 signal responses in the samples at different retention times were measured with an online
24 118 organic carbon detector, organic nitrogen detector and UV detectors, respectively (**Fig. 2**)
25 119 (7). The LC-OCD analysis was carried out following the procedure described by Huber *et al.*
26 120 (7).

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45 122 2.1 Fluorescence excitation emission matrices (FEEM)

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47 123 The FEEM spectrometer used a xenon excitation flash lamp with excitation and emission
48 124 slits set to a 10 nm band pass. To obtain excitation emission maps, excitation wavelengths
49 125 were varied from 200 nm to 600 nm at 3 nm band pass. For each excitation wavelengths,
50 126 the emission was detected at longer wavelengths (0.3 nm steps). A 1 cm quartz fluorescence
51 127 cuvette held the water samples during analysis. To account for Rayleigh scattering, a blank
52 128 solution (Type 1 water) response was subtracted from the fluorescence spectra of the

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3 129 sample to be analysed. All Spectra were then corrected by Rayleigh 1st and 2nd (10 band
4 130 slits) order masking and inner filter effect corrections were made. Spectral correction of
5 131 fluorescence EEMs was done to reduce instrumental and sample biases as a result of
6 132 wavelength-dependent variability. Processed FEEM spectra were then plotted using Origin
7 133 Lab. FEEM distinguishes chromophoric dissolved organic compounds by assessing the
8 134 absorption/emission wavelength spectral information for different groups of molecules.
9 135 These are classified into spectral groups based on excitation and emission wavelengths
10 136 where the molecules fluoresce (**Fig. 3**) (23). The simultaneous working setup of the FEEM
11 137 technique incorporates a light source (usually a xenon lamp and a detector. Overall, a FEEM
12 138 measurement is a rapid, quantitative evaluation (15 seconds to 2 minutes) that determines
13 139 all the species independent of fluorescent properties.

140 **Fig. 3**

141 **3.0 RESULTS AND DISCUSSION**

142 The LC-OCD results (in which all the fractions identified in water were quantified in terms of
143 organic carbon concentration) are presented in tabular, chromatogram and HS-diagram
144 forms (**Table 1, Fig. 4-6**). The FEEM spectral signatures for raw, cooling, irradiated and
145 oxidized raw water samples are shown in **Fig. 7-8**.

146 *3.1 LC-OCD results*

147 The dissolved organic carbon (DOC) concentrations in raw water was found to be 9.106
148 ppm. Results showed that light irradiation of a 50 mL sample of raw water decreased the
149 DOC after the 4 hours of measurement. The DOC in irradiated samples decreased from
150 9.100 ppm to 8.832, 8.838 and 8.175 ppm for *0.6-sun*, *1.2-sun* and *2.0-sun*, respectively. On
151 the other hand, the oxidation experiment results (at constant sunlight intensity of *1.2-sun*)
152 showed that the distribution of NOM fractions changed on addition of hydrogen peroxide
153 (H₂O₂), from the original distribution in raw water. This is expected as oxidation leads to
154 increased dissolved organic compounds, due to the decomposition of organic particles. On
155 the other hand, sunlight has an effect on NOM as it results in its breakdown. Combining light
156 and an oxidant, as done in this study shows that NOM will change and this could be
157 attributed to the breakdown of NOM and or aggregation of molecules which r-arrange to
158 form new organic compounds. At *1.2-sun*, the corresponding DOC values for 250, 500 and

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3 159 750 μL of H_2O_2 were 16.81, 9.946 and 8.832 ppm, respectively. Generally, exposition to light
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5 160 conditions showed decreased DOC values from those of raw water, whereas combined
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7 161 exposition to light and H_2O_2 showed an increase in DOC values. Apparently, the dissolved
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9 162 organic nitrogen distribution in the various samples did not show any particular trend.
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11 163 However, the dissolved organic nitrogen values were relatively reduced in the samples with
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13 164 hydrogen peroxide.

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15 165 The average molecular weights of HS in all the analysed water samples are shown in **Table 1**.
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17 166 These nominal molecular weights were plotted in the humic substances-diagram (HS-
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19 167 diagram) alongside the aromaticities of the HS, to give a picture of how
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21 168 hydrophobic/hydrophilic NOM was in the various water samples. The highest molecular
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23 169 weight of HS was observed in raw water samples (523 g mol L^{-1}). A decreasing trend in
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25 170 molecular weights was observed with increased exposition to light and increased H_2O_2
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27 171 amounts. The range of molecular weight was found to be $204\text{-}510 \text{ g mol L}^{-1}$. The observed
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29 172 decrease in molecular weights when samples were exposed to light in the presence of H_2O_2 ,
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31 173 was significantly different from the values of photo-irradiation in absence of H_2O_2 ($P < 0.05$,
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33 174 one-way ANOVA, generated by Origin Lab V8 at 0.05 confidence limits).

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Table 1

176 The level of unsaturation in NOM fractions could be expressed by aromaticity levels. In the
177 current study, the NOM aromatic character as shown in the HS-diagrams follows the order
178 $\text{C} > \text{B} > \text{A}$ (**Fig. 4a**) for the photo-irradiated water samples. **Fig. 4b** shows only one photo-
179 oxidised sample data point, D, measured for $250 \mu\text{L H}_2\text{O}_2$. The corresponding molecularities
180 of water samples follow the order $\text{A} > \text{B} > \text{C}$. Generally, the aromaticity and molecularity
181 decreased with increased radiation which implied decreased hydrophobicity. The HS-
182 diagram showed that irradiated samples were relatively on the same position on the plot.
183 Only one sample exposed to light and hydrogen peroxide was mapped on the HS-diagram
184 (i.e. the $250 \mu\text{L H}_2\text{O}_2$ irradiated sample).

185 The aromaticity relationship is critical in describing the NOM composition. In essence, it
186 allows discrimination of relatively higher molecular weight humic acids and fulvic acids from
187 water. However, aromaticity of compounds with the same molecular weight could be
188 different depending on the source. For example, Huber *et al.* (2011) reported that
189 aromaticity of fulvic acid was higher in brown water lakes (7). Their reports are attribute the

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3 190 disparity to sunlight induced bleaching of substances, which is related to the current study.
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5 191 The photo-bleaching however depends on the depth of water since deeper layers are not
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7 192 relatively accessible and hence lower impact of solar radiation.

8
9 **Fig. 4**

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11 194 The chromatograms based on the organic carbon detector, the organic nitrogen detector
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13 195 and ultraviolet detector responses of samples exposed to light and those exposed to light
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15 196 and H₂O₂ are presented in **Fig. 6**. The NOM of raw water from Vaal River was mainly
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17 197 comprised of humic substances of fulvic acid character (**Fig. 4**). As a function of the intensity
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19 198 of radiation, the humic substances decreased in percentage from 61.5% in raw water to 57.8
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21 199 and 61.4 down, for *0.6-sun*, and *2.0-sun*, respectively. At *1.2-sun*, however a slight increase
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23 200 in humic substances was observed (i.e from 61.5 to 61.7%). Furthermore the molecular
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25 201 masses decreased from 523 g mol L⁻¹ to 510, 469 and 408 g mol L⁻¹ for 0.6-sun, 1.2-sun and
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27 202 2.0-sun, respectively. Regardless of the concentration of humic substances, the specific
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29 203 absorbance at λ₂₅₄ decreased with an increase in the light intensity. The corresponding SUVA
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31 204 values were, 4.24, 3.77 and 3.74, for 0.6-sun, 1.2-sun and 2.0-sun, respectively. These values
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33 205 which deviate from the raw water SUVA (4.11), show a decreasing trend as the intensity of
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35 206 light was increased. The same observation was made in biopolymers with a decrease from
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37 207 3.9% in raw water to 3.4%, 3.4% and 3.6% for samples exposed to sunlight of *0.6-sun*, *1.2-*
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39 208 *sun* and *2.0-sun* intensities, respectively. The decreasing SUVA values observed showed
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41 209 increased hydrophilicity of NOM present in the water samples (24).

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43 210 In the presence of H₂O₂ and exposition to light conditions, the SUVA values were found to
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45 211 reduce even further to 0.81, 0.57 and 0.27 for 250, 500 and 750 μL aliquots of H₂O₂. The
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47 212 molecular masses decreased from 523 g mol L⁻¹ to 275, 278 and 204 g mol L⁻¹ for 250, 500
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49 213 and 750 μL, respectively. The concentration of humic substances decreased even further. An
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51 214 interesting observation was made on the behaviour of the building block peaks (42-50 min),
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53 215 that is, the concentration increased with the amount of radiation and concentration of the
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55 216 oxidant.

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57 217 The biopolymer fraction and the corresponding concentrations of dissolved organic nitrogen
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59 218 increased with irradiation but decreased with oxidation (all oxidative processes being
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219 carried out at *1.2 sun*) (**Fig. 5**), as expected. Literature indicates that the physico-chemical
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treatments such as oxidation or adsorption have degradative effects on NOM. These studies

221 have also indicated that oxidation processes such as ozonation reduce the biopolymer
222 fraction of NOM (8, 24). The biopolymer fraction mainly consists of polysaccharides,
223 proteins and some humic-like substances (25). These compounds are generally large
224 molecules and henceforth their decrease suggests that advanced oxidation processes have a
225 degradative of NOM.

226 Besides the humic substances, biopolymers and building blocks, the low molecular weight
227 acids and low molecular weight neutrals (weakly and uncharged organic compounds)
228 identified are shown in **Fig. 5**. The low molecular weight organic responses of samples
229 exposed to light and samples exposed to both light and H₂O₂ showed a high percentage of
230 low molecular weight neutrals and a non-significant percentage of low molecular weight
231 acids. In most water samples exposed to light, the low molecular weight acids were not
232 produced. The values for the low molecular weight neutrals decreased from 1.38 ppm in
233 raw water to 1.09, 0.918, 0.708 ppm in *0.6*, *1.2* and *12.0 sun*, respectively. In the presence
234 of H₂O₂ and exposition to light conditions, the low molecular weight neutrals showed a
235 decreasing trend i.e. 2.32, 2.31 and 1.32 ppm for 250, 500 and 750 µL aliquots of H₂O₂
236 added. In the presence of an oxidant, low molecular weight acids were formed which
237 decreased drastically with increased H₂O₂ aliquots. These acids may have been as a result of
238 oxidation of NOM in the raw water.

239 **Fig. 6** shows chromatograms for the various NOM fractions. The ultra-violet detector
240 response in the HS peak was observed to reduce with intensity showing decreased levels of
241 unsaturation and aromaticity of humic substances. We conclude that exposition to light
242 decreases aromaticity. Combined H₂O₂ and exposition to light conditions minimise the HS
243 peak, an indication of changing the form and structure of HS.

244 **Fig. 5**

245 **Fig. 6**

246 *3.2 FEEM signatures of raw and cooling water*

247 Complete NOM characterization of the raw, samples exposed to light and those exposed to
248 combined H₂O₂ and exposition to light conditions was achieved using combined LC-OCD and
249 FEEM techniques. In addition, the fluorescent contours of cooling water samples of two
250 stations are discussed. The FEEM spectra of fluorescent NOM for samples of raw and cooling

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3 251 water collected from Lethabo and Kriel power stations are shown in **Fig. 7**. The raw water
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5 252 used for cooling in these stations is sourced from the same river, the Vaal. It is for this
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7 253 reason that the FEEM spectra of raw and cooling water used in these two stations are
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9 254 compared. Subsequently, the discussions on FEEM spectral signatures were based on raw
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11 255 water that was sampled at Lethabo power station. The results in this study show that the
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13 256 FEEM signatures were independent of the sampling power station but depended on the
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15 257 stage at which sampling took place (i.e raw and cooling water sampling stages). The cooling
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17 258 water showed more intense spectral contours, all of which were obtained at the primary
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19 259 humic-like region (excitation wavelength 280-350 nm/ emission wavelength 350-500 nm).
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21 260 The increase in the intensity of fluorescent signatures can be explained by the water
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23 261 processes that lead to concentration of fluorescing compounds in water.

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25 262 The spectral analysis was performed at 0 to 1146 spectral units for raw water and 0 to 1650
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27 263 units for cooling water. The effect of exposition to light conditions and combined H₂O₂ and
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29 264 light conditions on fluorescent organic matter are discussed in **Section 3.2.1** and **3.2.2**. The
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31 265 only fluorescing organic material found in this study corresponds to signatures obtained for
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33 266 the humic-like substances region. The FEEM signatures observed in this study, showed the
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35 267 changes occurring to humic substances as a result of increasing exposure to light intensities
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37 268 and oxidizing substances in water. The main essence of our study, therefore, is to describe
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39 269 the processes that bring about primary changes in raw water before use in the cooling
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41 270 water chambers, where recycling takes place.

42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 271 **Fig. 7**

272 *3.2.1 The effect of light intensity*

273 The intensity of spectral contours (indicating humic-like substances) was dependent on the
274 intensity of light (**Fig. 8**). The identified EEM spectra of humic substances obtained by using
275 different light intensities showed that the spectral contours reduced with increased UV-
276 visible radiation. In all the samples exposed to light, the spectral signatures (predominantly
277 appearing in the humic-like region), showed a significant decrease in fluorescence. The
278 results in this study showed that spectral response was lowest at *1.2-sun*. However, the
279 decrease in spectral contour intensities with increasing sunlight was not linear. We noted
280 similar signatures for *0.6-sun* and *2.0-sun*. We thus concluded that near natural or natural

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3 281 sunlight (that is, 1.2 *sun* ~1.0 *sun*) could be the most appropriate condition for attenuating
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5 282 the humic substances in water. The EEM spectra of raw water before exposition to light was
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7 283 in close agreement with what was reported in other studies. The effect of surface waters on
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9 284 the reduction of the HS peak due to exposure to sunlight, has also been reported elsewhere
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11 285 (2, 26). Moreover, these previous studies have shown that natural light has a significant
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13 286 effect on the alterations to humic-like materials, which means the total fluorescence
14
15 287 intensity of humic-like materials could be much higher in closed water systems, such as
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17 288 those used in the cooling water chambers.

17 289 **Fig. 8**

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20 290 *3.2.2 Effect of H₂O₂ to NOM in surface water*

21 291 The results of the peroxide effect on humic substances showed disappearing spectral
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23 292 signatures (**Fig. 9**). No spectral contours in the HS region were formed for samples exposed
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25 293 to light with near natural sunlight intensity in the presence of hydrogen peroxide. The
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27 294 oxidative reactions between NOM and oxidizing compounds such as peroxides, dictate the
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29 295 destiny of humic substances in natural water. While recycle systems produce HS-
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31 296 concentrates, oxidizing agents affect their composition and probably, the physico-chemical
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33 297 characteristics of NOM. Overall, the FEEM spectra supported the results of LC-OCD analyses
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35 298 regarding the breakdown and oxidation of humic substances in NOM in the presence of UV-
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37 299 visible light. Oxidation processes, including chlorination, ozonation and advanced oxidation
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39 300 processes application of both UV alone and UV/hydrogen peroxide are reported to
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41 301 breakdown humic substances, hence reduced humic like peaks/spectra (9, 26). Decreases in
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43 302 fluorescent intensity of humic materials when advanced oxidation processes are used have
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45 303 been attributed to the decrease in molecular weight since the hydroxyl radicals are
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47 304 postulated to attack high electron density regions within double bonds (9).

47 305 **Fig. 9**

49 306 *3.3 Overall implications of advanced analytical assessments of NOM attenuation*

51 307 The techniques, SUN simulation, LC-OCD and FEEM used for the assessment process of NOM
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53 308 in river water (for Eskom cooling in condenser tubes in power stations), gave important
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55 309 information on NOM fractions and how they change due to the influence of sunlight and the
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57 310 presence of an oxidant. Advanced treatment processes which apply to both solar radiation

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3 311 and solar radiation combined with hydrogen peroxide, showed reduced humic like
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5 312 peaks/spectra, especially for the latter. However, the changing intensities of solar radiation
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7 313 did not impact greatly on the humic substances. A similar study by Legrini et al in 1993,
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9 314 reported this observation: “that the humic substances in water remain relatively stable
10
11 315 when irradiated with UV light” (27). Since humic acids and fulvic acids are structurally large
12
13 316 molecules with many carboxylic and hydroxyl groups, it is therefore expected that they
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15 317 remain relatively stable in solution. In terms of denticity, Mg and Ca ions will prefer multi-
16
17 318 dentate binding with the large molecules due to their chelate effect. Once Ca and Mg form
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19 319 stable complexes with these large molecules, precipitation of their carbonate salts is
20
21 320 inhibited. This is because the complexation of the Ca and Mg by the organic matter reduces
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23 321 the total concentration of the metals and hence inhibits the formation of their carbonate
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25 322 salts that cause scaling. That way these cations (whose salts normally form scale in pipes
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27 323 carrying cooling water) are kept in solution, with reduced potential for scale formation.

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29 324 The impact of solar radiation on the distribution of NOM fractions (including HS) must
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31 325 therefore be measured appropriately. With the advanced LC-OCD, it is possible to quantify
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33 326 various NOM fractions (discussed in **Section 3.1**). The FEEM technique on the other hand,
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35 327 offers capacity to explain the spectral maps and signatures formed by fluorescing groups
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37 328 shown in **Fig. 3**.

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330 **4.0 CONCLUSIONS**

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41 331 The LC-OCD technique was used to quantify the distribution of NOM fractions in water
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43 332 samples after irradiation with simulated sunlight alone as well as sunlight in the presence of
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45 333 an oxidizing agent such as H₂O₂. The DOC values and the NOM molecular weights were
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47 334 altered by light and more significantly by a combination of light and H₂O₂. Except under the
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49 335 influence of H₂O₂, the HS peak remained, relatively stable.

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51 336 All irradiated sample aromaticity versus molecularity relationships could be explained on the
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53 337 HS-diagram. On the contrary only one ‘photo-oxidised’ sample data point (measured for 250
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55 338 µL H₂O₂) could be shown on the HS-diagram. This implied that a combination of light and an
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57 339 oxidant (H₂O₂ in this study) is very effective in breaking down aromatic and unsaturated
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59 340 structures in the NOM matrix. This was confirmed by the decrease SUVA values.

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3 341 Low molecular weight acids formed were found to increase with increase in hydrogen
4 342 peroxide amounts. In this case, the low molecular weight neutrals showed a decreasing
5 343 trend i.e. 2.32, 2.31 and 1.32 ppm for 250, 500 and 750 μL aliquots of H_2O_2 added.

8
9 344 The cooling water at Eskom undergoes continuous recycling processes which in turn
10 345 concentrate the organic constituents. The type of NOM classified as the “humic-like”
11 346 fraction was successfully identified and quantified using the FEEM spectral contours for raw
12 347 and cooling water.

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17 348 A decrease in spectral contour peak intensities was observed for the light-attenuated
18 349 samples. No spectral contours in the HS region were formed for the samples illuminated by
19 350 near natural sunlight intensity in the presence of hydrogen peroxide.

22 23 351 **5.0 ACKNOWLEDGEMENTS**

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427 **Table 1:**

428 Distribution of various NOM fractions in various raw, in samples exposed to light and samples exposed to both light and H₂O₂ (DOC = dissolved
 429 organic carbon and HS = humic substances)

Sample	Concentration in ppb								
	DOC	Biopolymers	HS	Dissolved organic nitrogen	SUVA	Molecular weight (gmol ⁻¹)	Building blocks	Low molecular weight neutrals	Low molecular weight acids
Raw water	9106.0	354	5597	321	4.11	523	1109	1382	nq
0.6 Sun	8832.0	304	5104	188	4.24	510	1013	1085	24.0
1.2 Sun	8838.0	287	5146	325	3.77	469	1095	918.0	Nq
2.0 Sun	8175.0	297	5021	300	3.74	408	1054	708.0	Nq
1.2 Sun + 250 μL H ₂ O ₂	16813	139	5932	321	0.81	275	2266	2324	475
1.2 Sun + 500 μL H ₂ O ₂	9946.0	145	4434	147	0.57	278	2253	2315	299
1.2 Sun + 750 μL H ₂ O ₂	8832.0	55.0	3924	84	0.27	204	1653	1316	6.00

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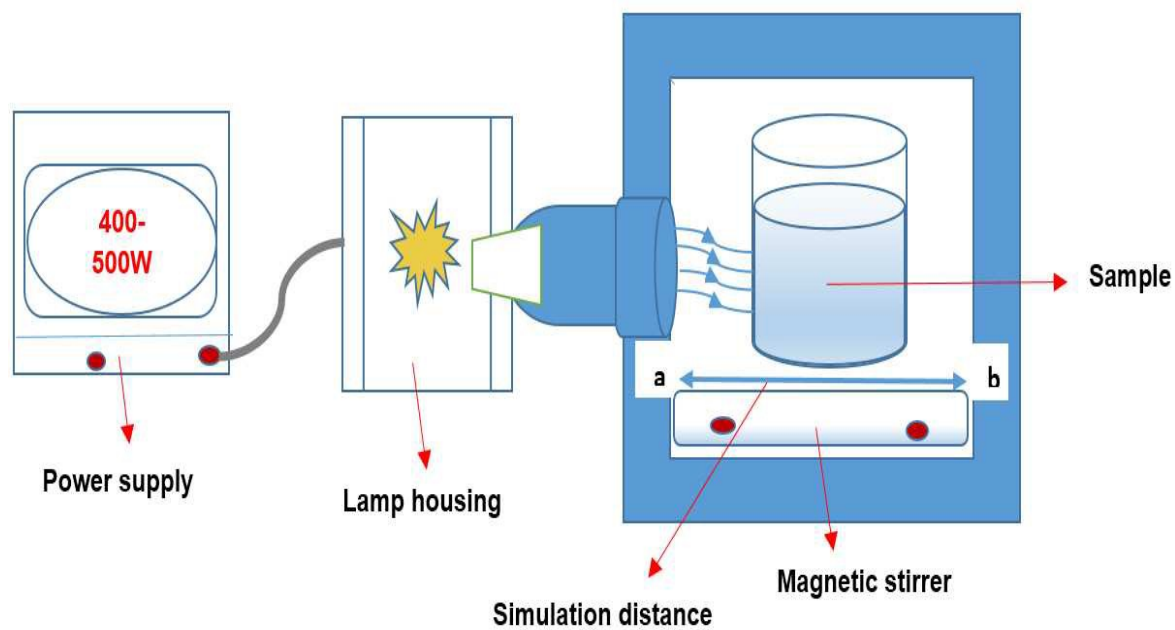
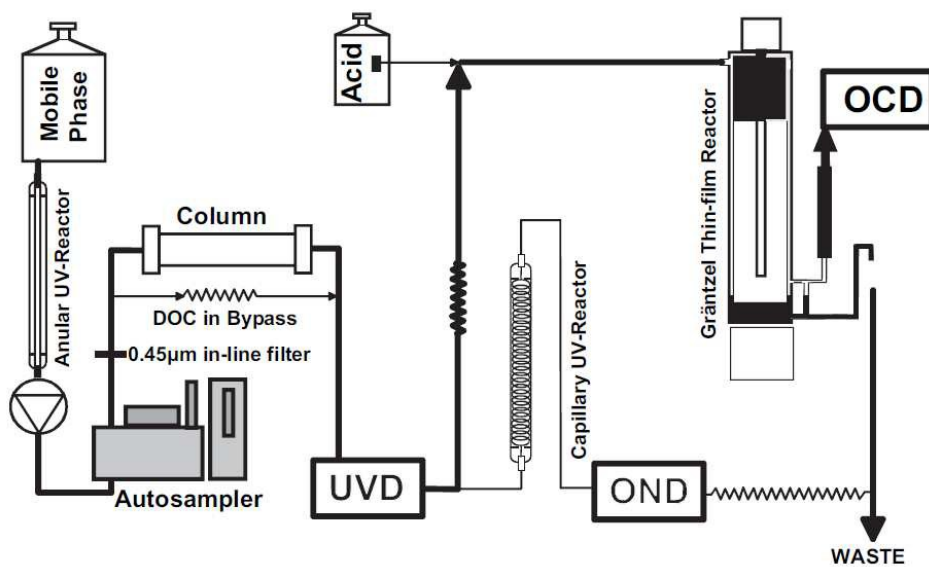


Fig. 1: A scheme showing the sun simulator

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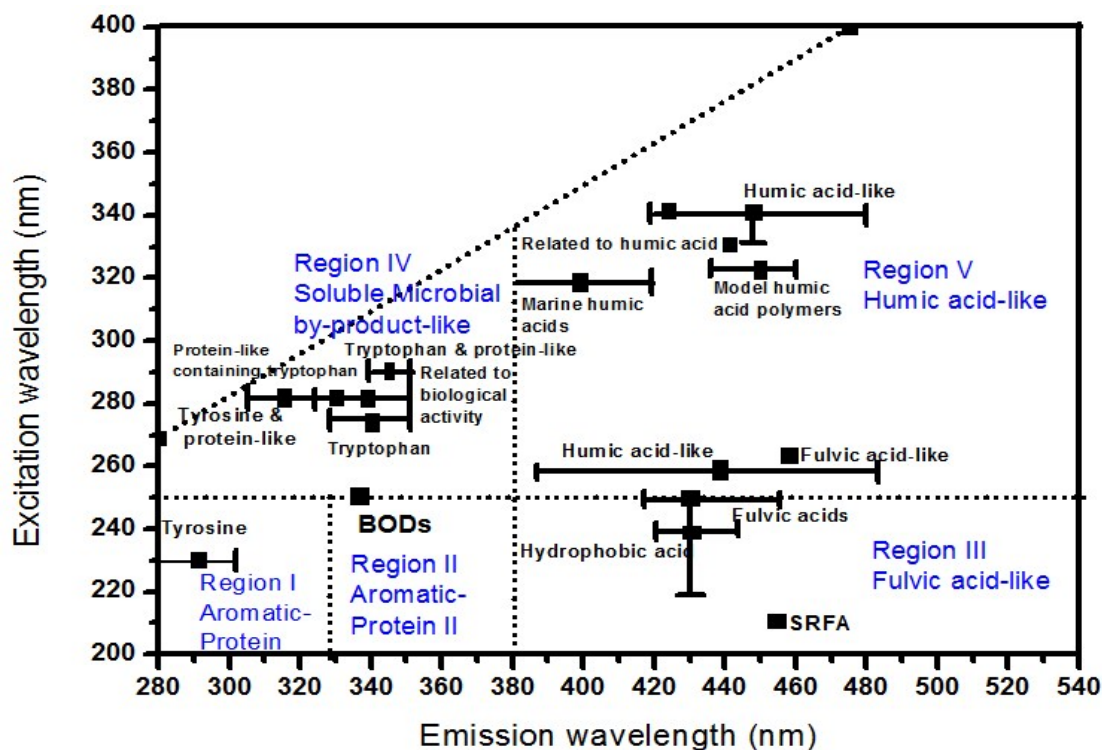


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436 **Fig. 2:** Scheme of the liquid chromatographic LC-OCD system. The UVD detector for specific
437 UV absorbance at λ -254 responses is the first detector (non-destructive) (Adopted from
438 Huber et al., 2011)

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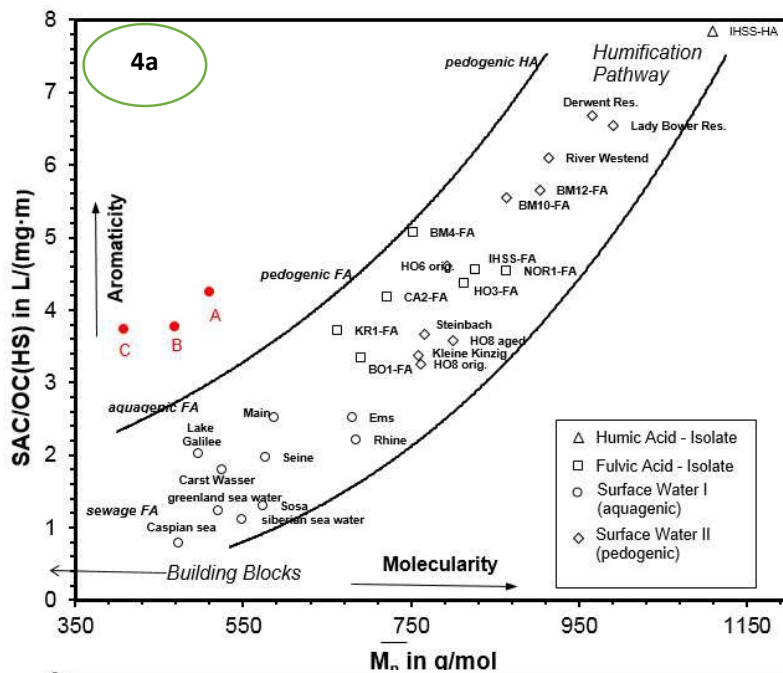


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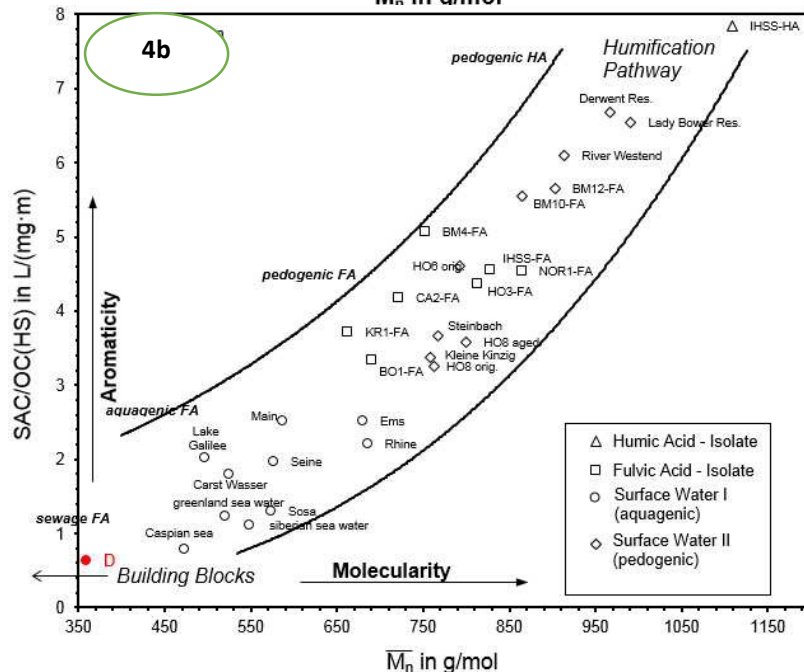
442 **Fig. 3:** FEEM spectral regions based on reported definite excitation (Ex) and emission (Em)
 443 wavelengths (I. Ex 250 nm, Em 350 nm; II. Ex 280 nm, Em 380 nm (soluble microbial by
 444 products); III. Ex 250 nm, Em 280 nm (humic acid-like organics); IV. Ex 300 nm, Em 400 nm
 445 and V. Ex 214 nm, Em 400 nm (fulvic acid-like molecules) (Nkambule et al., 2012).

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450 **Fig. 4:** The HS diagrams showing the aromatic character (x-axis) of HS fractions as a function of
 451 their molecularity (y-axis) as a plot of the Spectral Absorption Coefficient (SAC) to the organic
 452 carbon (OC) ratio (aromaticity) of each HS-fraction against its nominal molecular weight (\bar{M}_n). **4a**
 453 shows three components: A raw water irradiated by 0.6-sun, B water irradiated by 1.2-sun and C
 454 irradiated by 2.0-sun. **4b** only shows element D, the raw water irradiated by 1.2-sun in the
 455 presence of 750 μL of H_2O_2 .

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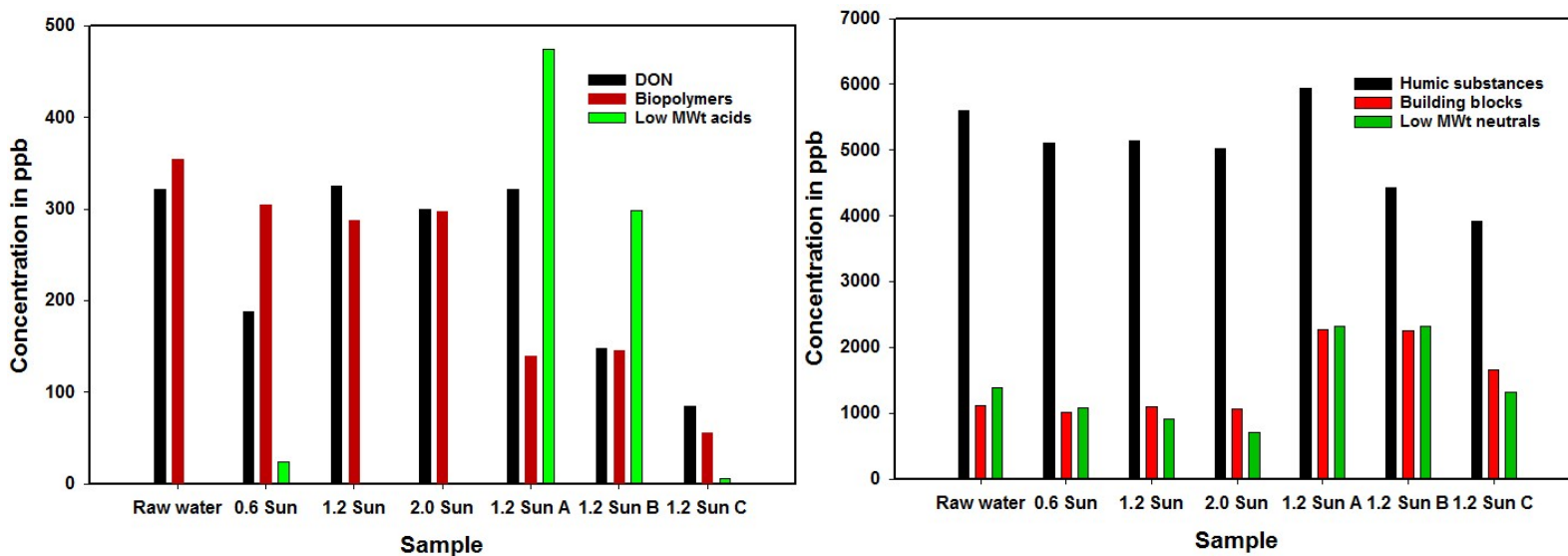


Fig. 5: The NOM fractions distribution after of exposition to sunlight. A, B and C represent increasing amounts of hydrogen peroxide i.e. 250, 500 and 750 μL , respectively.

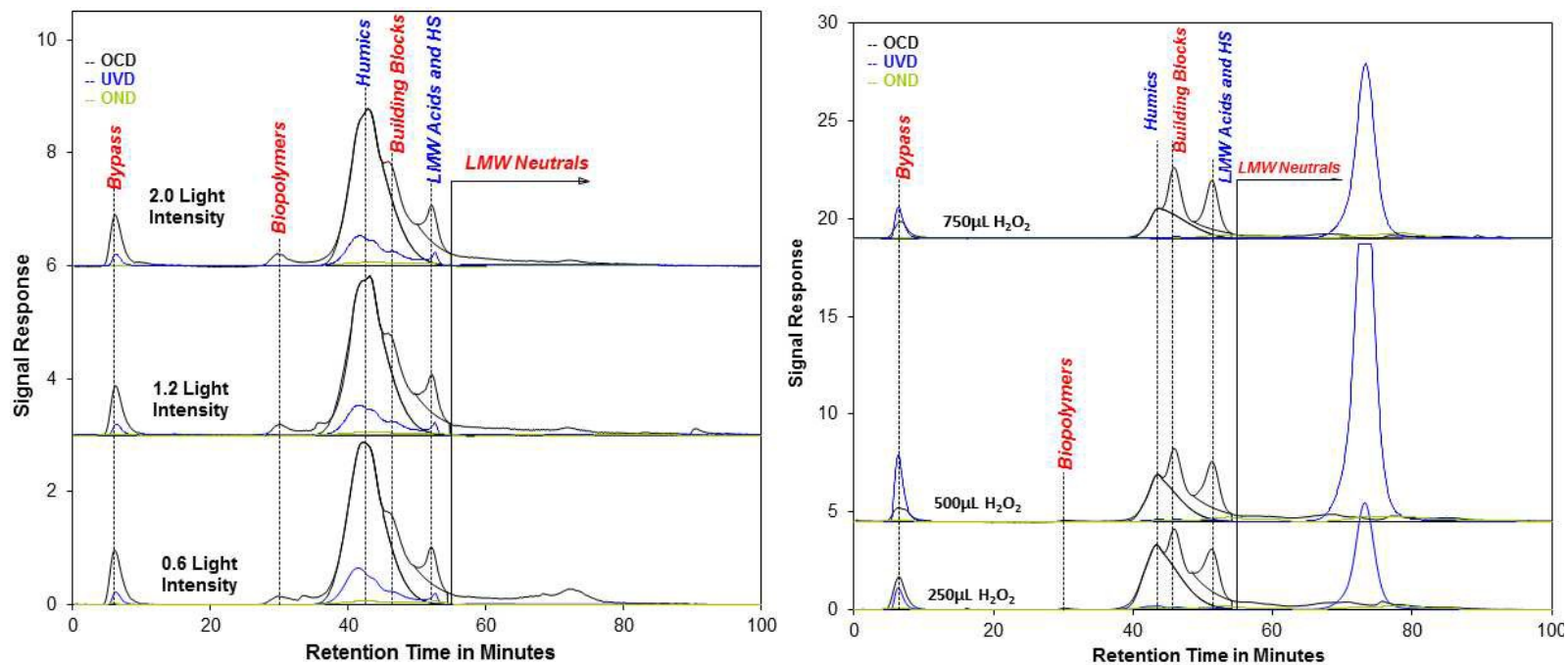
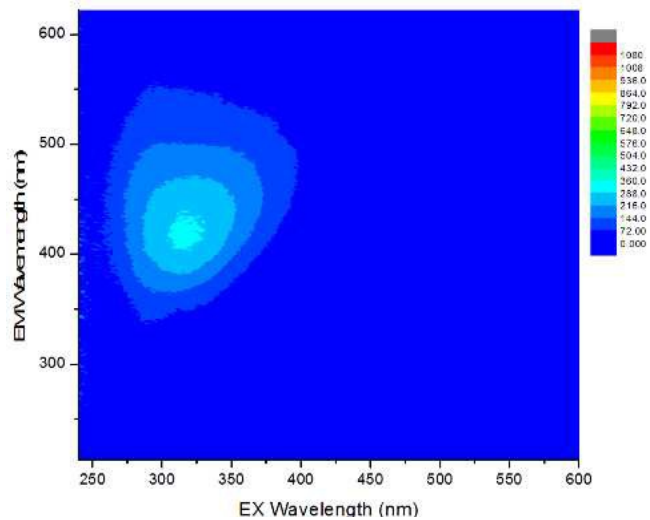


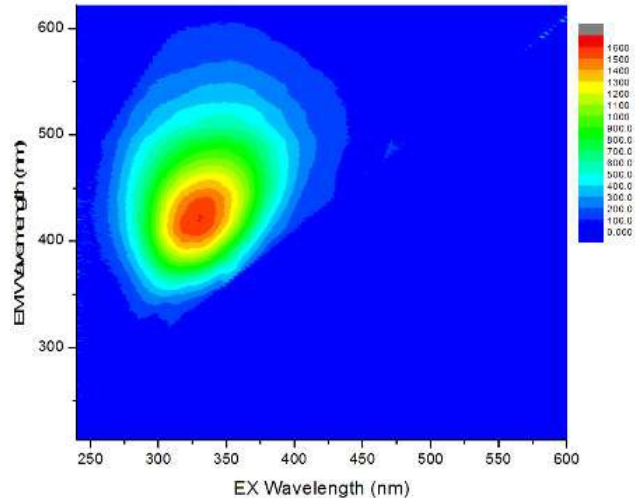
Fig. 6: Chromatograms water samples of exposed to light only and those exposed to light and hydrogen peroxide conditions

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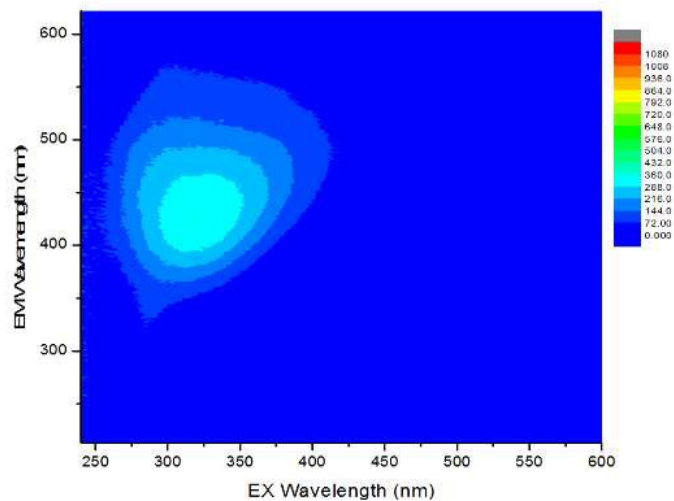
Lethabo raw water



Lethabo cooling water



Kriel raw water



Kriel cooling water

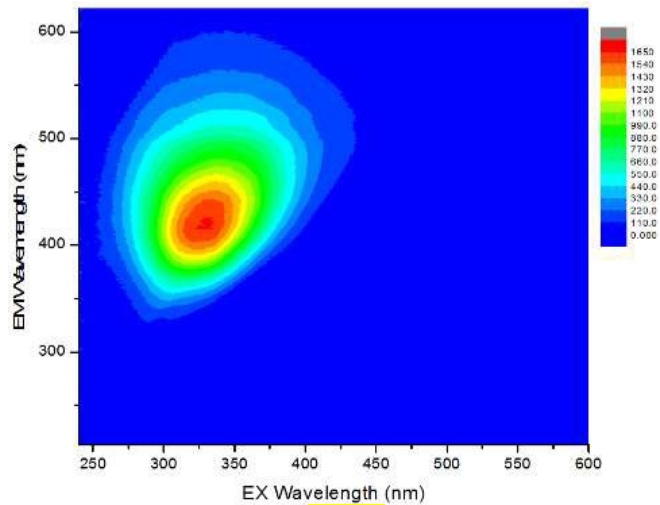


Fig. 7: FEEM spectral signatures obtained from raw and cooling water from Kriel and Lethabo Eskom power stations.

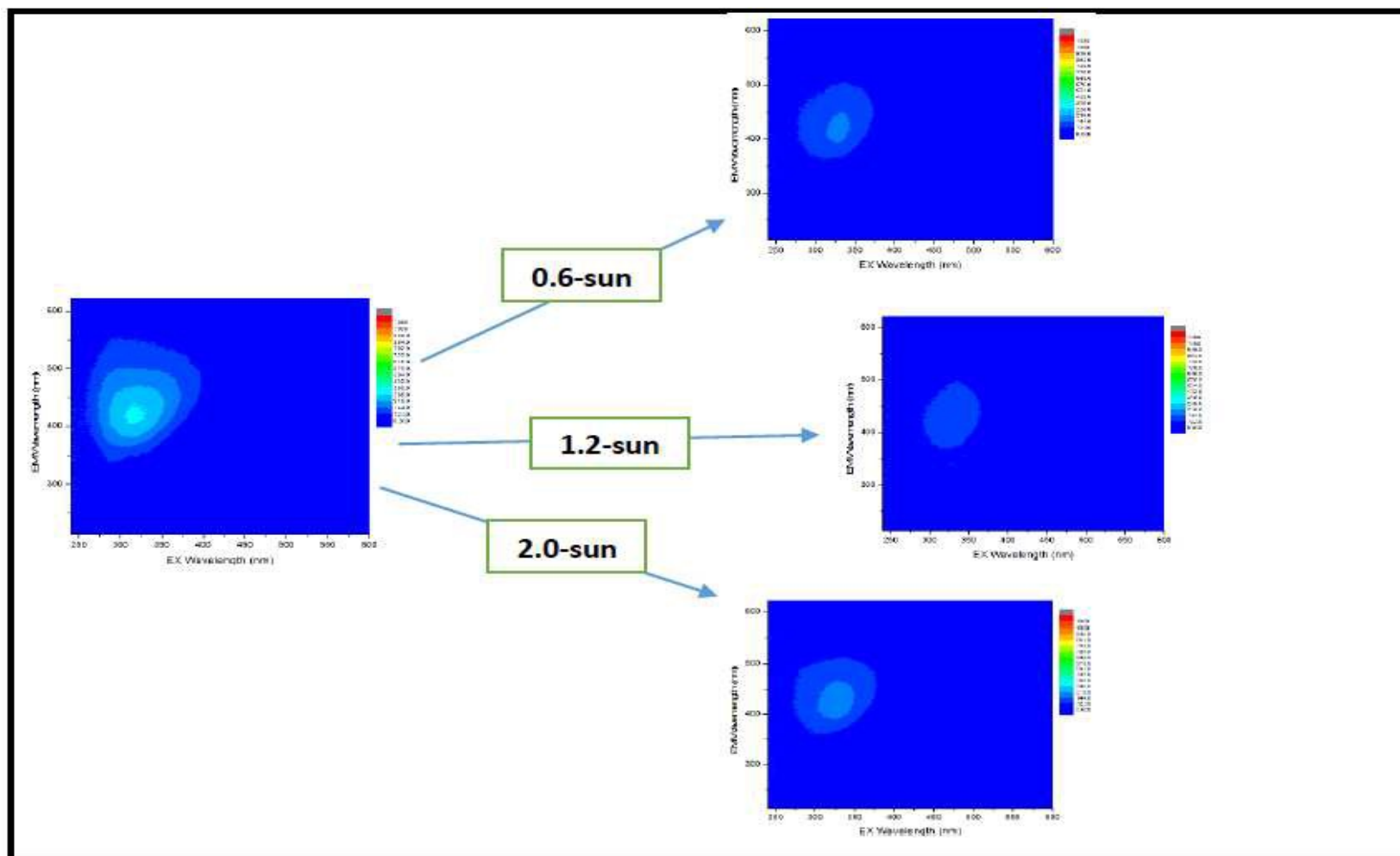


Fig. 8: Typical fluorescent EEM spectral signatures obtained of raw at Lethabo power station and after exposition to various intensities of sunlight for 4 hours i.e. 0.6-sun, 1.2-sun and 2.0-sun, respectively

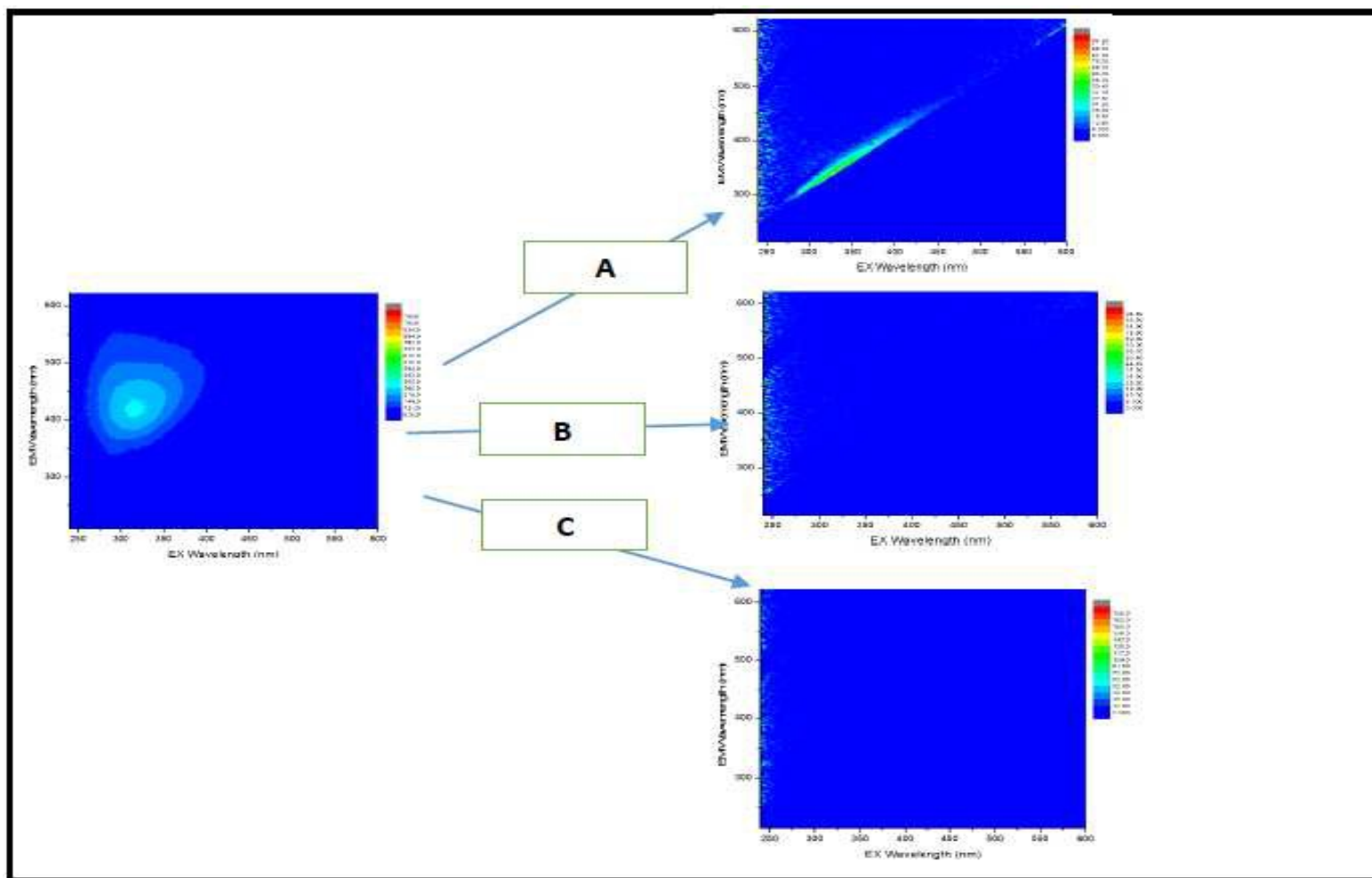


Fig. 9: Fluorescent EEM spectral signatures obtained from raw at Lethabo power station, after photo-irradiating with it at 1.2-sun with varying amounts for 4 hours of H₂O₂ (250, 500 and 750 μ L)

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