

Analytical Methods

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6 2 **Ionic liquid-based microextraction method for the determination of Cu and Ni in**
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9 3 **high-salinity produced waters from offshore petroleum exploration by GF AAS**
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Abstract

This work reports the development of a novel method for the ionic-liquid microextraction and determination of Cu and Ni in high-salinity produced waters by Graphite Furnace Atomic Absorption Spectrometry (GF AAS). Studies were performed to set a suitable temperature program and to optimize the extraction conditions. Due to the presence of the ionic liquid in the extracts, a drying temperature of 300 °C was chosen for the temperature program. The pyrolysis and atomization curves indicated that the pyrolysis temperature for Cu and Ni should be set at 900 °C, and that the best atomization temperatures were 2100 and 2500 °C for Cu and Ni, respectively. The ionic-liquid microextraction was carried out by mixing 10 mL of sample with 1 mL of a 1% m/v ammonium pyrrolidine dithiocarbamate (APDC) solution and 1 mL of a Britton-Robinson buffer solution (total concentration 0.1 mol L⁻¹) with pH = 4.0. Then, 100 µL of the ionic liquid (1-hexyl-2-methylimidazolium-hexafluorophosphate, C₆MImPF₆) were added and the mixture was shaken manually for 2 min. The obtained suspension was centrifuged for 15 min at 4500 rpm in order to promote the sedimentation of the ionic liquid. The ionic-liquid was separated from the upper aqueous phase using a micropipette and then, absolute ethanol enough to complete 1 mL was added. This final solution was introduced into the graphite tube for the determination of Cu and Ni. The method presented limits of quantification of 0.37 and 0.61 µg L⁻¹ for Cu and Ni, respectively. The calibration of the method was possible using a matrix-matching approach using synthetic saline samples. Recovery tests were performed with the addition of aqueous standard solutions of the metal cations and resulted in the recovery percentages in the range of 76 - 126%.

Keywords: produced water, ionic liquid microextraction, GF AAS, trace elements

1. Introduction

Nowadays, the exploration/exploitation of petroleum is one of the most important industrial activities worldwide. Several countries, including Brazil, has spent many efforts to find new reservoirs in order to ensure an adequate supply of crude oil for fuel production, industrial uses, heating, electric energy generation and others. In Brazil, recent discoveries of large reservoirs of petroleum in the pre-salt layer is increasing the investments in this area and demanding the development of new technologies.^{1,2}

Under the environmental point of view, the petroleum exploitation can cause many problems. Certainly, one of them is the production of large amounts of water along with petroleum. In some cases, the amount of water produced by a reservoir is five times the amount of oil. In the beginning of the operation of a well, the inner pressure is high, which allow the extraction of the crude oil without using any fluid to enhance the oil recovery. With time, the inner pressure decreases and water must be injected into the well to maintain the inner pressure and enhance the secondary recovery of crude oil. The water injected into the well is called of "injection water". The injection water is mixed with the water already present inside the reservoir (called formation water) and the mixture is taken out along with the crude oil. This water is called "produced water" and can contain a number of toxic organic substances such as benzene, toluene, xylenes, polycyclic aromatic hydrocarbons (PAH), phenols, alkylated phenols and organic acids. Additionally, inorganic contaminants such as radionuclides (NORM, from *normally occurring radioactive materials*) and toxic elements (As, Cd, Cr, Cu, Ni, Pb, V, among others) can be found in produced waters. The main characteristic of the produced water

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3 70 extracted from offshore platforms is the high salinity, which can reach values as high as
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6 71 270‰.^{3,4}
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8 72 The extracted produced water has, in general, two destinations. It can be re-injected
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10 73 into the well or discarded in the ocean.⁵ For discharging in the ocean, the production water
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12 74 must be treated to eliminate possible toxic contaminants and achieve the (good) quality
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14 75 imposed by the legislation. One of the most important parts of the treatment process is the
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16 76 determination of the toxic substances present in the produced water. This task is especially
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18 77 difficult due to the complex nature of this kind of sample, which includes its very high salinity.
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21 78 In the specific case of metals, some papers report their presence in produced waters from
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24 79 different origins.⁶⁻⁹
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28 80 Despite the fact that the development of methods for the determination of metals in
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31 81 saline samples by atomic spectrometric techniques is frequently reported in the literature,¹⁰⁻¹⁵
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33 82 including the use of ionic liquids for separation/preconcentration purposes,¹⁶⁻¹⁸ the
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35 83 development of analytical strategies for the determination of metals in produced waters is a
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37 84 subject relatively new. In this field, two different strategies have been followed: (i) direct
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39 85 determination of the analytes, which implies in the direct injection of the samples into the
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42 86 instruments¹⁹⁻²² and; (ii) determination of the analytes after their separation/preconcentration
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45 87 from the samples.²³⁻²⁴
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48 88 Oliveira et al.¹⁹⁻²⁰ proposed the use of combined chemical modifiers for the direct
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51 89 determination of Pb and Cr by ET AAS in produced waters from offshore petroleum exploration.
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53 90 In the first work, the use of HF combined with Ir-W permanent modifier provided efficient
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56 91 matrix elimination and suitable stabilization of Pb, allowing its quantification in samples with
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3 92 salinities up to 82%.¹⁹ Similar approach was employed in the determination of Cr in produced
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6 93 waters, but using HF combined with Pd modifier.²⁰
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9 94 Cassella et al.²¹ employed $\text{NH}_4\text{H}_2\text{PO}_4$ as chemical modifier for the direct determination
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11 95 of vanadium in produced waters by ET AAS. The use of this modifier allowed the use of a
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13 96 pyrolysis temperature of 1500 °C, which was sufficient to set the background signals close to
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16 97 zero without significant loss of the analyte. Also, Cassella et al.²² tested the Ir-W permanent
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18 98 modifier for the direct determination of Mn in produced waters using the same analytical
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21 99 technique. Samples with salinities that varied between 74 and 84‰ were analyzed.
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23 100 Nevertheless, the standard addition calibration had to be employed to overcome non-spectral
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26 101 interferences derived from the complex composition of the waters.
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28 102 The separation of the analytes from the matrix is another option for the determination
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31 103 of metals in produced waters. In this approach, the analytes are transferred to another phase,
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33 104 which is simpler than the original sample. This process helps one to control possible
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36 105 interferences on the measurements and, in some cases, allows the preconcentration of the
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38 106 analytes, improving the limits of detection and quantification.
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41 107 Oliveira et al.²³ determined ten elements (Cd, Pb, Ni, Zn, U, Mo, Fe, Co, V e Mn) in
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43 108 produced waters by ICP-MS after their separation from the saline matrix using the chelating
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46 109 resin Toyopearl AF-Chelate-650M. The Chelex-100 resin was also successfully employed for this
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48 110 purpose. In this work, the authors separated five trace metals (Co, Cu, Mn, Ni and Pb) from the
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51 111 samples and quantified the analytes in the extracts by high resolution continuum source
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53 112 graphite furnace atomic absorption spectrometry.²⁴
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3 113 The goal of this work was to introduce, for the first time, the use of a hydrophobic ionic
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6 114 liquid (1-hexyl-3-methylimidazolium hexafluorophosphate, C_6MImPF_6) to promote the
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9 115 microextraction of Cu(II) and Ni(II) ions from high-salinity produced waters extracted along with
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11 116 petroleum in offshore operations. The quantification of the analytes in the extracts was
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13 117 performed by GF AAS. Also, a detailed study on the thermal behavior of the analytes in the
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16 118 presence of the ionic liquid was performed in order to set the best temperature program for
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19 119 the determination of the analytes by GF AAS.
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23 121 **2. Experimental**

28 123 **2.1. Apparatus**

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33 125 The absorbance measurements were performed with a Varian (Mulgrave, Australia),
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36 126 model AA240Z, atomic absorption spectrometer with a Varian GTA 120 graphite furnace
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39 127 atomizer unit and a Varian PSD 120 autosampler. The instrument was equipped with a Zeeman
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41 128 effect background corrector system (constant magnetic field of 0.8 T), which was utilized in all
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44 129 measurements. Graphite tubes with integrated L'vov platform were employed for the
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46 130 atomization of the analytes. They were also supplied by Varian. Argon (99.99% purity), supplied
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49 131 by Linde Gases (Macaé, Brazil), was employed as protective gas. Copper was measured at 324.8
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51 132 nm with a slit width of 0.5 nm and using an individual hollow cathode lamp operated at 4.0 mA.
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54 133 Nickel was measured at 232.0 nm with a 0.2 nm slit width and using a hollow cathode lamp also
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56 134 operated at 4.0 mA.
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3 135 The thermogravimetric analysis of the liquid ionic was performed with a SDT Q600
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6 136 analyzer from TA Instruments (New Castle, DE, USA). Approximately 10 mg of the liquid ionic
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9 137 was directly inserted into the instrument and the temperature was varied from laboratory
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11 138 ambient temperature to 1000 °C under N₂ atmosphere. A heating rate of 20 °C min⁻¹ was
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14 139 employed in the analysis.

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16 140 An Eppendorff centrifuge (Hamburg, Germany), model 5804, was utilized to promote
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18 141 the separation of the phases. It was equipped with a variable angle rotor and an adaptor for
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21 142 polyethylene capped tubes of 15 mL capacity.

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23 143 A Digimed (São Paulo, Brazil) pHmeter, model DM-22, was employed to determine the
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26 144 pH of the solutions. The pHmeter was equipped with a combined glass electrode, also furnished
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28 145 by Digimed. The salinity of the samples was measured with a hand-held refractometer, model
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31 146 RTS-101ATC, from Instrutherm (São Paulo, Brazil).

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35 36 148 **2.2. Reagents and solutions**

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41 150 Deionized water was obtained with a Direct Q-3 system, supplied by Millipore (Milford,
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43 151 USA). The deionized water employed throughout the experimental work always had resistivity
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46 152 of 18.2 MΩ cm, at least.

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48 153 Standard solutions of Cu and Ni were prepared from the dilution of the stock standard
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51 154 solutions of 1000 mg L⁻¹, purchased from Tedia (Fairfield, OH, USA). The dilution was always
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54 155 carried out with a solution compatible with the solutions utilized in the extractions.

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4 156 Ammonium pyrrolydine dithiocarbamate (APDC) was employed as ligand. A 1% m/v
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6 157 APDC solution was prepared by dissolving 0.1 g of the reagent, supplied by Sigma-Aldrich (St.
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8 158 Louis, MO, USA), in approximately 5 mL of deionized water. Afterwards, the obtained solution
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10 159 was diluted to 100 mL with deionized water.

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13 160 The ionic liquid employed in the present work, 1-Hexyl-3-Methylimidazolium
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15 161 Hexafluorophosphate (C_6MImPF_6), was supplied by Sigma-Aldrich (St. Louis, MO, USA). Absolute
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17 162 ethanol employed in the dilution of the extracts was furnished by Tedia (Fairfield, OH, USA).

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20 163 The Britton-Robinson buffer solution was prepared by dissolving convenient masses of
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22 164 sodium acetate (Sigma-Aldrich, Rio de Janeiro, Brazil), disodium hydrogen phosphate (Sigma-
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24 165 Aldrich, Rio de Janeiro, Brazil) and boric acid (Sigma-Aldrich, Rio de Janeiro, Brazil) in water
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26 166 sufficient to complete a final concentration of 0.1 mol L^{-1} of each component.

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29 167 The samples of produced waters evaluated in this work were supplied by Petrobras.
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31 168 They were acidified to pH 1.8 with trace metal grade nitric acid (Seastar Chemicals, Sidney,
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33 169 Canada) and maintained in the refrigerator (4°C) until their analysis. No additional treatment
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35 170 was dispensed to the water samples.

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42 43 172 **2.3. Extraction procedure**

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49 174 In a 15 mL polyethylene tube, 10 mL of sample (or diluted sample), 1 mL of buffer
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51 175 solution (Britton Robinson 0.1 mol L^{-1} , pH = 4.0) and 1 mL of a 1% m/v APDC solution were
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53 176 added and a clear solution was obtained. Then, 100 μL of the ionic liquid were introduced and
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55 177 the system was manually agitated for 2 min in order to ensure uniform dispersion of the ionic
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3 178 liquid in the medium. At this point, the formation of a turbid solution was observed due to the
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6 179 dispersion of the hydrophobic ionic liquid throughout the aqueous solution. Afterwards, the
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9 180 polyethylene tube was centrifuged for 15 min at 4500 rpm to promote the separation of the
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11 181 phases. After centrifugation, the ionic liquid phase containing the extracted metals was
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13 182 deposited in the bottom of the polyethylene tube. Then, the aqueous upper phase was taken
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16 183 out with the aid of a micropipette and absolute ethanol sufficient to complete 1 mL was added
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19 184 to the ionic liquid. This final extract was employed to determine Cu and Ni by GF AAS.

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21 185 The optimization of the temperature program was performed with a $25 \mu\text{g L}^{-1}$ Cu and Ni
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23 186 ethanol solution containing 20% v/v (200 μL in each 1 mL of solution) of ionic liquid.
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27 28 188 **2.4. GF AAS measurements**

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33 190 A volume of 20 μL of extract was introduced into the graphite tube for the
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36 191 determination of Cu and Ni in the extracts. In the case of Cu, 10 μL of a $500 \mu\text{g mL}^{-1}$ Pd modifier
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39 192 solution was introduced immediately after the extract. Then, the temperature program was run
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41 193 and the measurements were carried out in integrated atomic absorbance mode. The
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44 194 temperature programs used for Cu and Ni determination in the extracts are listed in Table 2
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46 195 and were obtained after a proper optimization, which will be described in the section 3.1.
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3. Results and discussion

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3.1. Optimization of the temperature program

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204 The first part of this work was to set a convenient temperature program for the
205 determination of the analytes by GF AAS in the ethanolic medium containing the ionic liquid. It
206 is important to remember that the ionic liquids present very low vapor pressure and tend to
207 volatilize at high temperatures. In this sense, initially, the drying step of the temperature
208 program was optimized.

209 Before starting the experiments with the GF AAS, a thermogravimetric analysis of the
210 ionic liquid was performed to identify its thermal characteristics and orientate us in the
211 optimization of the drying step of the temperature program. As displayed in Fig. 1, the loss of
212 mass of the ionic liquid started at approximately 275 °C, which indicated that a regular
213 temperature used for drying aqueous samples (120 °C) could not be adequate to promote a
214 convenient drying of the extracts inside the graphite tube. Therefore, the first parameter
215 optimized in the drying step was the final temperature of this step.

216 The drying step was composed of different stages. In the first stage, the temperature
217 was elevated up to 95 °C and then to 120 °C, in order to eliminate all ethanol present in the
218 extracts and any residue of water. Then, the temperature was increased to a final temperature,
219 which was evaluated in this study. Additionally, the influence of the heating ramp and the
220 holding time of this stage on the signals of Cu and Ni was evaluated.

221 Firstly, the final temperature of the drying step was varied from 150 to 600 °C. The
222 absorbance signals for both Cu and Ni did not vary significantly, being only observed a little
223 decrease of them at temperatures higher than 550 °C. It is important to remark that, at the
224 lowest temperatures employed in the study, typically between 150 and 250 °C, an incomplete
225 drying of the extracts was observed. So, we decided to set the final temperature of the drying
226 step at 300 °C, which was compatible with the data obtained in the thermogravimetric analysis.

227 After optimizing the final temperature of the drying step, the ramp (18 - 180 s) and
228 holding (3 - 7 s) times were evaluated. Both parameters did not influence the absorbance
229 signals of Cu and Ni. In order to avoid that the temperature program was excessively long, the
230 ramp and holding times were then set at the lowest values, i.e., 18 s and 3 s, respectively. A
231 ramp of 18 s represented a heating rate of 10 °C s⁻¹.

232 Optimum pyrolysis and atomization temperatures were determined according to the
233 classical procedure, which was based on the construction of the pyrolysis and atomization
234 curves. In this experiment, the curves were constructed using two solutions: (i) a 25 µg L⁻¹ of Cu
235 and Ni in water and (ii) a 25 µg L⁻¹ of Cu and Ni in ethanol containing the ionic liquid (see section
236 2.4). The obtained results are shown in Fig. 2A and 2B.

237 As it can be seen in Fig. 2, the difference observed between the curves obtained with
238 the solutions prepared in water and in the ethanol/ionic liquid medium was small, evidencing
239 that the adopted drying step was efficient for the elimination of the solvents (and ionic liquid)
240 without volatilizing the analytes. Even though, the pyrolysis and atomization temperatures were
241 chosen taking into account the values obtained with the solution prepared in the medium
242 containing ethanol and ionic liquid, since it tends to be very similar to the composition of the

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3 243 extracts. Under these conditions, the pyrolysis temperature set for Cu and Ni was the same,
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6 244 900 °C; and the atomization temperatures were 2100 and 2500 °C, for Cu and Ni, respectively.
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10 11 246 **3.2. Optimization of the extraction conditions** 12 13 14 247

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16 248 Once established the operational conditions for the determination of Cu and Ni in the
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18 249 extracts by GF AAS, the next step of the work was the optimization of the experimental
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21 250 conditions for the microextraction of the analytes using the ionic liquid.
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25 26 252 **3.2.1. Effect of the pH on the extraction** 27 28 29 253

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31 254 The first parameter evaluated, related to the microextraction process, was the pH. It
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33 255 was expected that pH presented a remarkable effect on the complexing capacity of APDC,
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36 256 which behaves as an weak acid in solution. Also, pH affects the formation of hydroxy complexes
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39 257 of Ni(II) and Cu(II), which could impair the formation of the respective metal-APDC complexes.
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41 258 The experiment was carried out using the same conditions expressed in the section 2.3,
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44 259 but using 200 µL of ionic liquid. The pH of the solution was varied by changing the pH of the
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47 260 Britton-Robinson buffer in the range of 4 - 10. The results obtained are shown in Fig. 3. This
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50 261 variable had no noticeable influence on the extraction in the range of pH studied, being
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53 262 observed a practically constant signal along the whole range. This result indicated that the
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56 263 complexing capacity of APDC was not affected by the pH in this range, which was already
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59 264 expected, since the pKa of the APDC is 3.29.²⁵ Studies with pH lower than 4 were not performed
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3 265 due to the difficult to maintain the pH without increasing the buffer concentration and to avoid
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6 266 the degradation of the reagent, a typical phenomenon observed for dithiocarbamate-
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9 267 containing reagents. In front of these results, the pH 4.0 was employed in all further
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11 268 experiments.

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16 270 **3.2.2. Effect of the APDC concentration**17
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21 272 Other parameter that could influence the microextraction of the metal cations from
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23 273 solution was the reagent concentration. The study of this parameter is important because the
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26 274 amount of reagent must be enough to chelate the metal cations and provide their quantitative
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28 275 extraction to the hydrophobic ionic liquid phase

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31 276 The concentration of APDC was tested in the range of 0.25 - 1.5% m/v. All other
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33 277 conditions were those described in section 2.3, except the volume of ionic liquid, which was
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36 278 200 μL . The signals of both Cu and Ni presented the same behavior with the variation of the
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38 279 APDC concentration (Fig. 4). A soft increase of the signals was observed when the APDC
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41 280 concentration was increased from 0.25% m/v to 1.0% m/v. Beyond this point, the signals
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43 281 remained practically constant, indicating that no more ions could be chelated by the reagent.
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46 282 Therefore, in order to use a lower amount of reagent as possible, an APDC concentration of
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48 283 1.0% m/v was set for the method.

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3.2.3. Influence of the volume of ionic liquid

Liquid-liquid microextraction is characterized by the large difference between the volumes of extractant solution and sample employed in the process. The use of a very small volume of extractant phase (in relation to the sample volume), in general, results in the observation of a large preconcentration factor. However, the use of an excessively small volume of extractant phase can result in incomplete or inefficient extractions. In the case of the use of ionic liquids as extractant, their high-cost must also be considered.

In order to achieve the best condition, in terms of volume of ionic liquid, an experiment was performed to evaluate the influence of this variable on the method. In the experiments, the volume of sample was kept constant (10 mL) and the volume of ionic liquid employed in the extraction was varied between 25 and 200 μL . The other conditions were those already optimized.

The increase of the volume of ionic liquid caused a remarkable increase of the analytical signals for both Cu and Ni, to volumes up to 100 μL (Fig. 5). Beyond this volume, for Cu, the signals remained practically constant, indicating that maximum extraction of this analyte could be achieved even with 100 μL of ionic liquid were used. In the case of Ni, only a little increase in the signals was observed when 200 μL of ionic liquid was used. Therefore, we decided to set a volume of 100 μL of ionic liquid for the method in order to save this high-cost reagent without losing extraction efficiency.

3.2.4. Influence of salinity

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311 The method presented in this work was conceived to be employed in the separation and

312 determination of the analytes in highly saline waters obtained from offshore petroleum

313 exploration. In order to evaluate the performance of the extraction procedure under these

314 conditions, an experiment was performed to investigate the influence of salinity on the

315 extraction process. Also, the results obtained in this experiment could indicate the efficiency

316 observed in the separation of the analytes from the rest of the sample.

317 The influence of salinity was studied by carrying out the extraction of Cu and Ni from

318 solutions containing NaCl concentrations in the range of 0 (no addition of salt) - 2 mol L⁻¹. Blank

319 experiments were always run by applying the same procedure in the analysis of NaCl solutions

320 without adding the analytes. Figure 6 shows the results obtained in this experiment. It is

321 important to highlight that such results show the net integrated absorbances, i.e., the

322 integrated absorbance obtained in the experiment with addition of the analytes less the

323 integrated absorbance obtained in the experiment without adding them.

324 The increase of NaCl concentration in the solutions caused a decrease of the analytical

325 signals for both Cu and Ni. This decrease was more pronounced for Ni than for Cu, being

326 observed analytical signals approximately 60 and 40%, respectively, smaller than those

327 observed when no NaCl was present in the samples. These results can be credit to the influence

328 of NaCl on the extraction process, since no significant disturbance of the peak profiles and the

329 increase of background absorbance were verified in the determination of the metals by GF AAS.

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3.3. Evaluation of the calibration strategy

The results obtained in the evaluation of the effect of the salinity indicated that this parameter could cause significant influence on the accuracy of the method, being responsible for important interference on the quantification strategy. In order to find a suitable a calibration approach, taking into account the salinity of the samples, two curves were prepared for each metal in the range of 1 - 10 $\mu\text{g L}^{-1}$. The first curve was constructed using standard solutions of the metal cations prepared in deionized water, whereas the second one was constructed by spiking the sample A (160‰ salinity) with Cu(II) and Ni(II). The sample was 10-fold diluted for application of the procedure. The results are shown in Table 3.

Remarkable differences were noted when the calibration was performed with the saline sample, for both Cu and Ni. The curves prepared in saline medium presented smaller slopes than the curves prepared in water, which indicated the occurrence of a matrix interference on the extraction due to the ionic strength (salinity). These differences evidenced that the calibration could not be performed with aqueous standard solutions of the analytes and that a matrix-matching approach should be tested. Therefore, in order to evaluate this hypothesis, a new calibration curve was prepared with standard solutions prepared in a synthetic saline sample with 150‰ salinity (also 10-fold diluted). The composition of the synthetic saline sample is shown in Table 4.

As expected, the slopes were not significantly different of those observed when the calibration curves were constructed with the sample A, which had similar salinity. This result reinforced the idea that the salinity was the parameter responsible for the matrix interference

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3 353 on the extraction process and that a matrix matching approach must be used as calibration
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6 354 strategy in the quantitative analysis of the samples.
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9 355 Once the experimental conditions were established for the application of the proposed
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11 356 method, the analytical features were estimated. The limits of detection and quantification were
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13 357 estimated using a calibration curve prepared in water. The estimated limits of detection (3σ
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16 358 criterion) for Cu and Ni were 0.1 and 0.2 $\mu\text{g L}^{-1}$, respectively, whereas the limits of
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18 359 quantification (10σ criterion) were 0.4 and 0.6 $\mu\text{g L}^{-1}$, respectively. These limits were derived for
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21 360 guidance purposes only, since they can be affected by the salinity of the medium employed for
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23 361 calibration. Linear calibration curves were always constructed in the range of 2.5 - 10 $\mu\text{g L}^{-1}$ for
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26 362 both analytes. The typical equations were: $A = 0.0346 [\text{Cu } (\mu\text{g L}^{-1})] - 0.0086$ ($r^2 = 0.999$) and $A =$
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28 363 $0.0353 [\text{Ni } (\mu\text{g L}^{-1})] + 0.0059$ ($r^2 = 0.999$). Relative standard deviations of 4.0 and 7.5% were
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31 364 verified for Cu and Ni, respectively, in five independent measurements of an extract obtained
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33 365 from the extraction of a solution containing 1.0 $\mu\text{g L}^{-1}$ of each analyte. The intermediary
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36 366 precision was evaluated by the analysis of the sample D in three different days and it was 11
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38 367 and 13% for Cu and Ni, respectively.
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42 43 369 **3.4. Application of the proposed methodology**

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48 371 The developed method was applied in the determination of Cu and Ni in six samples of
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51 372 produced water, using two synthetic saline samples (55 and 150‰) for the preparation of
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53 373 calibration curves. The synthetic samples (Table 4) were diluted to provide a medium with a
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56 374 salinity as close as possible of the samples, which were 10-fold diluted.
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3 375 Recovery tests were also performed to evaluate the accuracy of the method. For this
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6 376 purpose, the samples were spiked with 2.5, 5.0 and 10 $\mu\text{g L}^{-1}$ of each analyte and analyzed
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8 377 employing the optimized conditions. The results obtained in the recovery tests are presented in
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10 378 Table 5 and the results obtained in the analysis of the samples are displayed in Table 6.

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12 379 As it can be seen, the proposed extraction method showed to be suitable for the
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14 380 separation of both Cu and Ni from the saline matrices in order to allow their determination by
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16 381 GF AAS. The proposed method presented suitable analytical capacity for the quantification of
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18 382 the analytes in all samples, which were present in concentrations always higher than the limit
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20 383 of quantification derived for the methods.

21 384 Recovery percentages in the range of 76 - 126% were observed, indicating that the
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23 385 method has suitable accuracy for the determination of low concentrations of Cu and Ni in this
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25 386 kind of sample.

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27 388 **4. Conclusions**

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29 390 The separation method developed in this work was efficient for the isolation of the
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31 391 targeted analytes from the very saline samples, since no influence of the matrix was observed
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33 392 in the determination of both Cu and Ni in the extracts by GF AAS. The results obtained also
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35 393 indicated that the microextraction using ionic liquids can be considered an excellent option for
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37 394 the separation of metal cations from high-salinity produced waters originated from petroleum
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39 395 exploration in offshore conditions. Maybe, the use of other analytical techniques such as ICP-

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3 396 MS for the determination of the metals in the extracts can increase the sensitivity of the
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6 397 method.

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8 398 The results obtained in the study of the thermal behavior of the analytes in the medium
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11 399 of the ionic liquid showed that elevated drying temperatures should be used in the
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13 400 temperature program to eliminate adequately the ionic liquid, which was in agreement with
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16 401 data obtained in the thermogravimetric analysis of the ionic liquid. Also, it is important to
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18 402 remark that the choice of the calibration strategy was of fundamental importance for ensuring
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21 403 the accuracy of the proposed method. The saline matrix caused significant non-spectral
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23 404 interferences, which could be conveniently corrected by using the matrix-matching approach.
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27 28 406 **Acknowledgments**

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40
41 412 providing the samples analyzed in this work.
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485 **Table 1.** Operational conditions employed in the determination of Cu and Ni by GF AAS.

Parameter	Cu	Ni
Wavelength (nm)	324.8	232.0
Lamp current (mA)	4.0	4.0
Slit width (nm)	0.5	0.2
Measurement mode	Integrated absorbance	Integrated absorbance

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6 502 **Table 2.** Temperature program employed in the determination of Cu and Ni in the ionic liquid-
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8 503 rich extracts by GF AAS.
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Step	Temperature (°C)	Ramp (s)	Hold (s)	Ar flow rate (mL min ⁻¹)
Drying I	95	5	-	300
Drying II	120	10	-	300
Drying III	300	18	3	300
Pyrolysis	900	1	5	300
Atomization	2100 (Cu) 2500 (Ni)	1	4	0
Cleaning	2200 (Cu) 2600 (Ni)	2	-	300

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6 516 **Table 3.** Slopes of the calibration curves prepared in different media, expressed as slope value \pm
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8 517 standard error.
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Medium	Cu	Ni
Water	$0.035 \pm 0.002 \text{ L } \mu\text{g}^{-1} \text{ s}^{-1}$	$0.035 \pm 0.003 \text{ L } \mu\text{g}^{-1} \text{ s}^{-1}$
Sample A (160%)*	$0.018 \pm 0.004 \text{ L } \mu\text{g}^{-1} \text{ s}^{-1}$	$0.023 \pm 0.003 \text{ L } \mu\text{g}^{-1} \text{ s}^{-1}$
Synthetic saline sample (150%)*	$0.020 \pm 0.003 \text{ L } \mu\text{g}^{-1} \text{ s}^{-1}$	$0.026 \pm 0.003 \text{ L } \mu\text{g}^{-1} \text{ s}^{-1}$

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21 518 * Both samples were 10-fold diluted before application of the extraction procedure.
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534 **Table 4.** Composition of the synthetic saline samples utilized in this work.

Substance	Concentration (g L ⁻¹)	
	Synthetic sample (150‰)	Synthetic sample (55‰)
KCl	0.50	0.10
MgCl ₂ ·6H ₂ O	0.66	0.32
CaCl ₂ ·2H ₂ O	5.60	0.84
BaCl ₂ ·2H ₂ O	0.19	0.070
SrCl ₂ ·6H ₂ O	0.40	0.20
FeCl ₃ ·6H ₂ O	0.030	-
NaHCO ₃	-	0.44
Na ₂ SO ₄	0.070	-
NaCl	91	30
NaBr	0.15	0.080
Sodium acetate	0.029	-

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546 **Table 5.** Results obtained in the recovery test applied to the analysis of the produced waters by
547 the proposed procedure. Results are expressed as mean \pm standard deviation ($n = 3$). The
548 salinities of the samples are indicated between parenthesis.

Sample	Concentration added ($\mu\text{g L}^{-1}$) ^a	Cu recovery (%)	Ni recovery (%)
A (160‰)	2.5	88 \pm 2	80 \pm 12
	5.0	98 \pm 1	76 \pm 15
	10	89 \pm 8	78 \pm 9
B (62‰)	2.5	88 \pm 5	80 \pm 3
	5.0	97 \pm 2	95 \pm 9
	10	100 \pm 6	98 \pm 9
C (33‰)	2.5	88 \pm 14	76 \pm 4
	5.0	84 \pm 7	89 \pm 12
	10	76 \pm 4	88 \pm 2
D (234‰)	2.5	84 \pm 13	120 \pm 5
	5.0	85 \pm 2	110 \pm 11
	10	81 \pm 13	118 \pm 3
E (182‰)	2.5	88 \pm 7	80 \pm 7
	5.0	88 \pm 9	95 \pm 5
	10	97 \pm 8	98 \pm 5
F (270‰)	2.5	92 \pm 4	126 \pm 6
	5.0	86 \pm 8	125 \pm 10
	10	89 \pm 16	118 \pm 10

549 ^a The analytes were added directly to the diluted sample employed in the procedure.

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56 553 **Table 6.** Concentrations of Cu and Ni found in the samples of produced waters employing the7
8 554 matrix matching approach. Results expressed as mean \pm standard deviation (n = 3).
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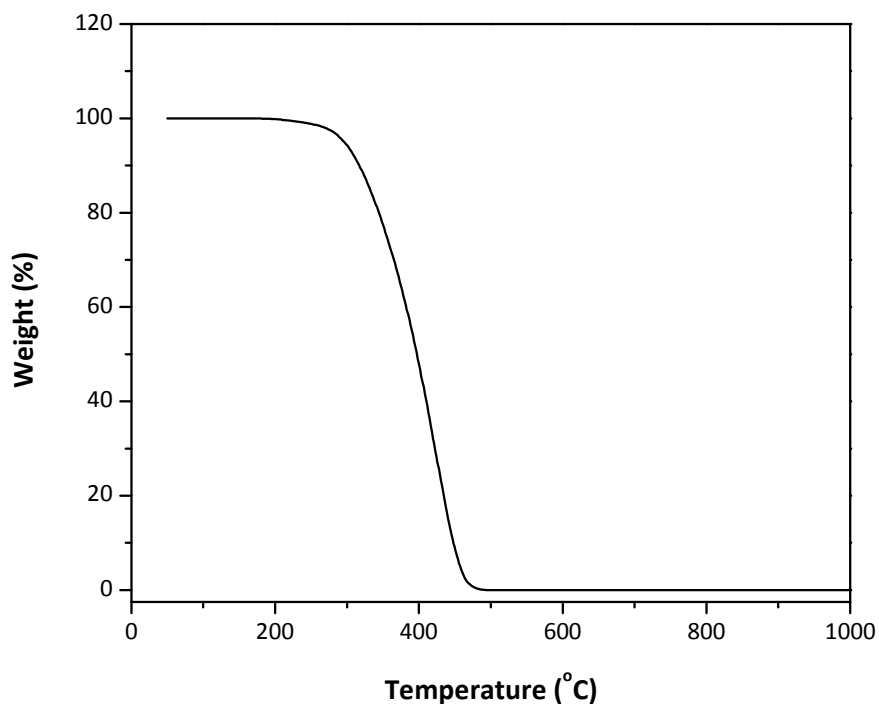
Sample	Cu ($\mu\text{g L}^{-1}$)	Ni ($\mu\text{g L}^{-1}$)
A	29.9 \pm 1.4	75.4 \pm 13.4
B	< LOQ	< LOQ
C	17.9 \pm 0.6	2.64 \pm 0.13
D	20.1 \pm 2.7	29.9 \pm 3.3
E	13.1 \pm 1.4	7.13 \pm 1.42
F	22.1 \pm 1.7	11.9 \pm 1.3

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567 **Figure 1**

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570 **Figure 1.** Thermogravimetric curve of the ionic liquid (C_6MImPF_6) employed in this study. The
571 thermogram was obtained under nitrogen atmosphere and using a heating rate of $20\text{ }^\circ\text{C min}^{-1}$.

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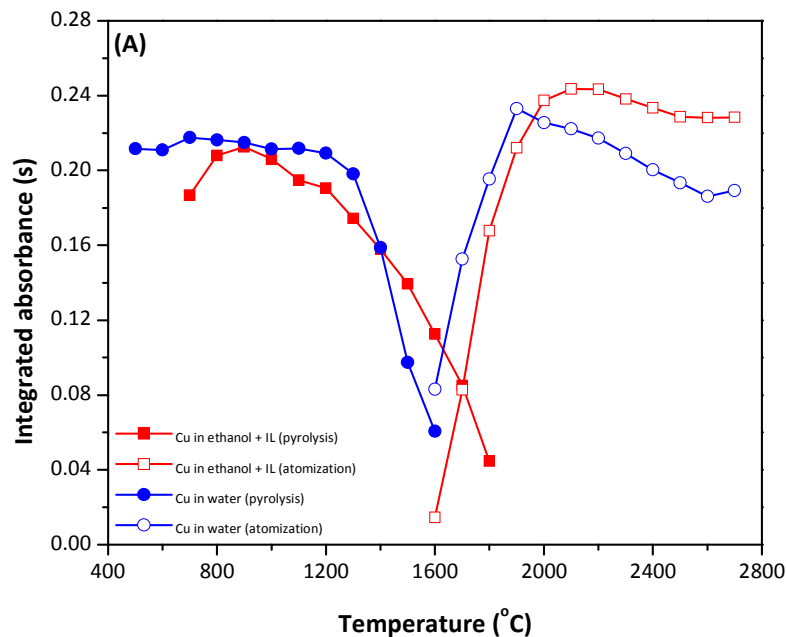
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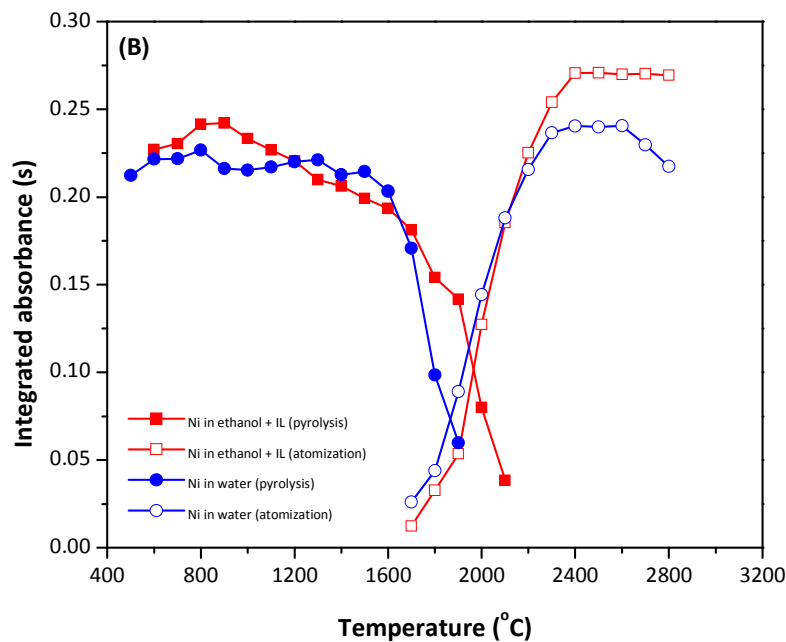
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578 **Figure 2**

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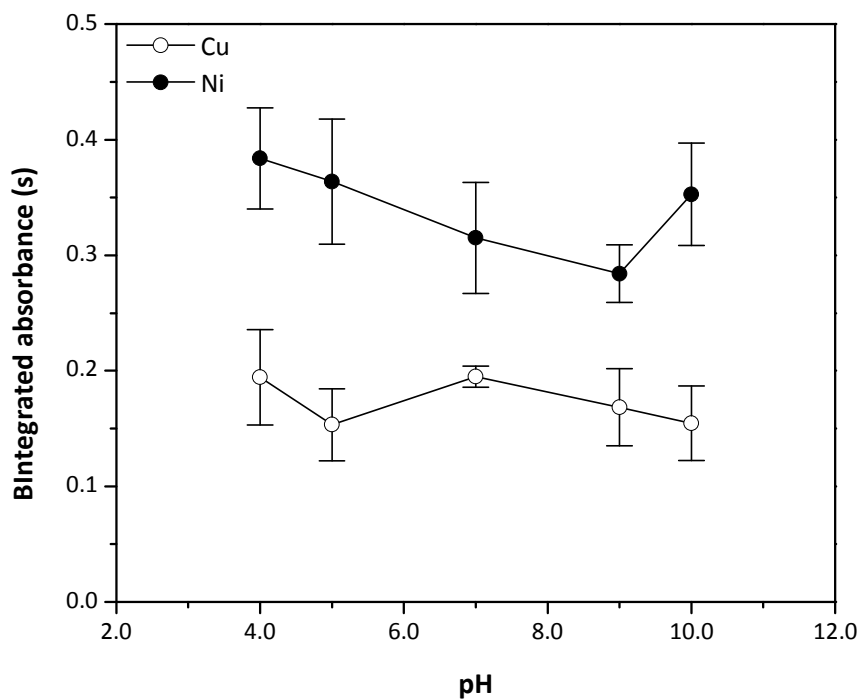
581 **Figure 2.** Pyrolysis and atomization curves for Cu (A) and Ni (B) in water and ionic liquid-rich

582 extract media.

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584 **Figure 3**

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587 **Figure 3.** Influence of pH on the extraction of Cu(II) and Ni(II) using the ionic liquid-based588 microextraction. Volume of ionic liquid = 200 μ L and APDC concentration = 1% m/v.

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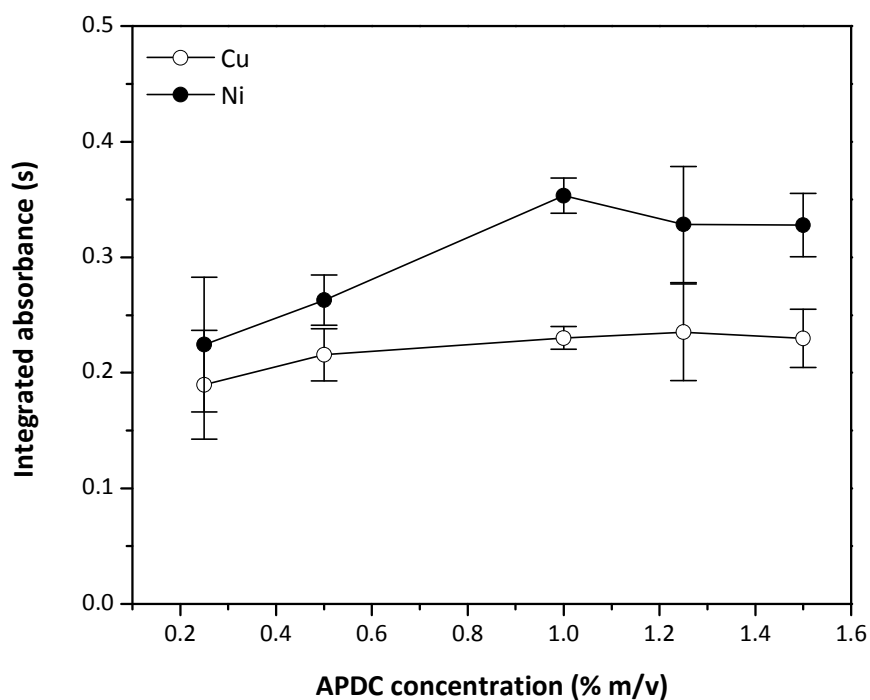
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596 **Figure 4**

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599 **Figure 4.** Influence of APDC concentration on the extraction of Cu and Ni using the ionic liquid-600 based microextraction. Volume of ionic liquid = 200 μ L and pH = 4.0

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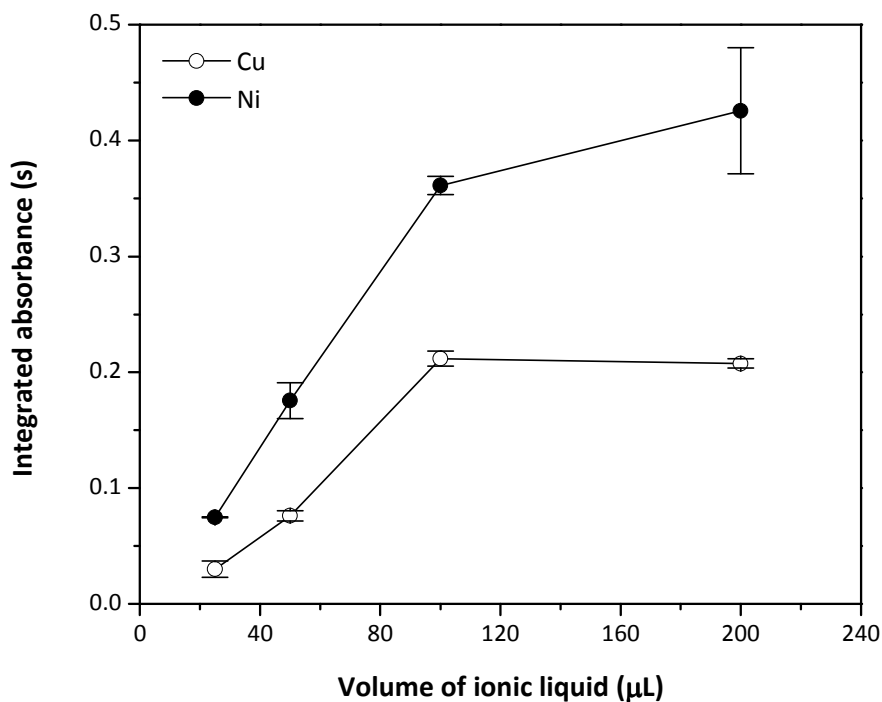
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608 **Figure 5**

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611 **Figure 5.** Influence of the volume of ionic liquid on the extraction of Cu and Ni using the ionic

612 liquid-based microextraction. APDC concentration = 1% m/v and pH = 4.0.

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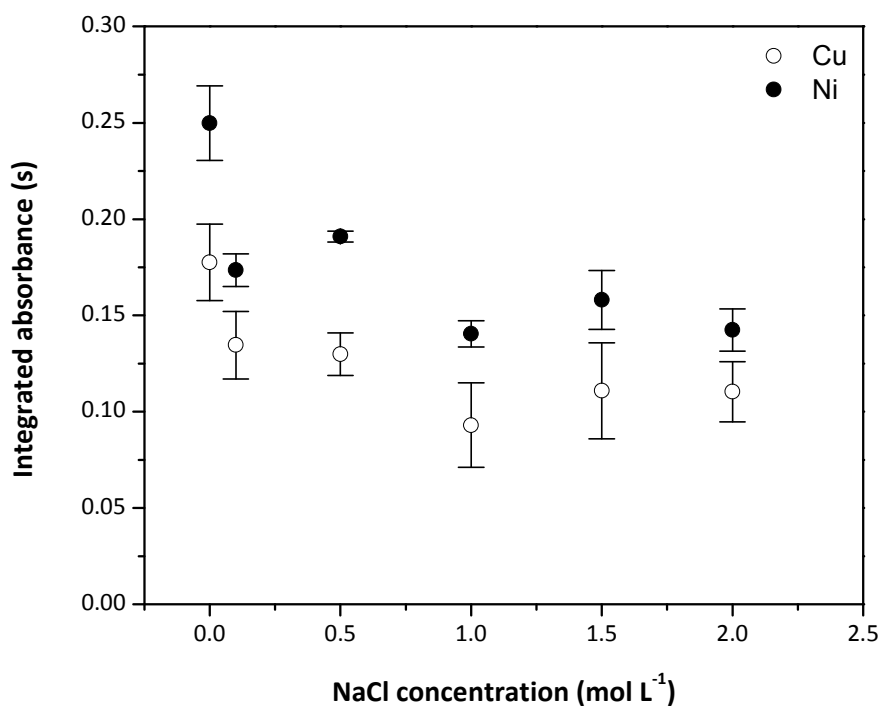
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620 **Figure 6**

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623 **Figure 6.** Effect of salinity on the ionic-liquid based microextraction of Cu and Ni by the

624 proposed method.

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