

Analytical Methods

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4 **1 Application of SPE followed by large-volume injection GC/MS for analysis of Geosmin**
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6 **2 and 2-Methylisoborneol in water**
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26 **14 ABSTRACT**
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29 15 A new method comprising of solid phase extraction (SPE) and subsequent large volume
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31 16 injection-gas chromatograph/mass spectrometer (LVI-GC/MS) was developed to analyze 2-
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33 17 methylisoborneol (2-MIB) and geosmin in water. The method with the injection volume of 25
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35 18 μL showed a good linearity (i.e., $R^2 > 0.999$) over the concentration range of 0.5-20 ng L^{-1}
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37 19 and good repeatability and recovery. The MDLs of the method for 2-MIB and geosmin were
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39 20 determined 0.87 and 0.62 ng L^{-1} , respectively, which are lower than one tenth of the
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41 21 compounds' published odor thresholds (i.e., 5-10 ng L^{-1}). If the injection volume was further
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43 22 increased, even lower MDLs could be obtained. In short, considering its ease of use, and high
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45 23 accuracy and sensitivity, the proposed SPE-LVI-GC/MS method can be easily applied for
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47 24 routine analysis of the target compounds in water.
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53 26 **Keywords:** 2-MIB, geosmin, SPE, large volume injection, SPME, GC/MS,
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1. INTRODUCTION

Taste and odor of drinking water have been a concern for both consumers and water supply authorities. As the national economy grows around the world, more public complaints on tastes and odors from drinking water are received. The public reluctance to directly consume tap water is mainly due to its musty or earthy odors and tastes. For example, most citizens in Korea do not drink water right from their taps; as of 2008, only 1.4% of the people consume tap water without further treatment^[1]. The earthy and musty odors from tap water are characteristics of 2-methylisoborneol (2-MIB) and geosmin produced by actinomycetes, cyanobacteria, and fungi in source water^[2-6]. They can cause odor sensation to the public even at an extremely low concentration; the odor threshold for 2-MIB or geosmin is 5-10 ng L⁻¹, which is 10⁻⁴–10⁻³ times lower than that of methyl tertiary-butyl ether^[7-9]. Due to low odor thresholds of 2-MIB and geosmin, it is critical that drinking water suppliers to have an easy analytical method to accurately determine concentrations of odorous compounds on a routine basis.

The quantitative analysis of 2-MIB and geosmin in water has been carried out with capillary gas chromatograph/mass spectrometer (GC/MS) methods because of their high separation power and sensitivity^[10]. Since the target analytes usually exist at extremely low concentration (often at ng L⁻¹ levels), however, the sensitivity of the GC-MS method remains challenging. In addition, an extensive sample preparation step is often required before injecting the sample into the GC/MS system. A few sample pretreatment techniques have been suggested and applied to analyze 2-MIB and geosmin in water^[11-13]. Examples include closed loop stripping analysis (CLSA)^[14], purge and trap^[13, 15], open stripping analysis^[16], simultaneous distillation extraction (SDE)^[17], liquid- liquid extraction (LLE)^[5, 18], liquid-liquid microextraction (LLME)^[19], continuous liquid-liquid extraction (CLLE)^[19, 20], solid-phase extraction (SPE)^[4, 18, 21], solid-phase microextraction (SPME)^[13, 22, 23] and stir bar

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4 54 sorptive extraction (SBSE) ^[24-26].
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7 55 Although the specific procedure of each technique for sample pretreatment is different,
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9 56 the principles are basically similar. Target analytes are first extracted from aqueous samples,
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11 57 such as drinking water, surface water, ground water, and wastewater, and subsequently
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13 58 trapped on or in a sorbent. Then, the target analytes are concentrated before injected into an
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15 59 instrument for quantitation. In fact, some of them (e.g., SDE, LLE, CLLE) are very labor
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17 60 intensive, and require large sample volume. In addition, LLE and CLLE require specially
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19 61 designed apparatus and the use of potentially harmful solvents ^[26, 27]. Albeit, these methods
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21 62 often suffer from deteriorated sensitivity and repeatability and result in to some extent large
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23 63 errors; it is simply because the sample volume is required to be reduced to 1-2 μL before GC
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25 64 analysis ^[28].
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28 65 Currently, the methods requiring less or no solvents, e.g., SPE, SPME, and SBSE, are
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30 66 more prevalent nowadays. In practice, SPME coupled with GC/MS is more commonly
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32 67 applied to quantitate 2-MIB and geosmin in water samples, since it allows extraction and pre-
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34 68 concentration of the analytes to be carried out in one step. Although the method has been
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36 69 successfully applied for extracting and pre-concentrating 2-MIB and geosmin, its use for
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38 70 accurate quantitative analysis is still limited. Two different phase equilibriums (one between
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40 71 liquid to headspace and the other between headspace and adsorbant of a fiber), extraction
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42 72 (such as temperature), variations in sample matrixes, etc. result in large variations in results;
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44 73 the reported relative standard deviations (RSDs) were as large as 40% for pond water ^[29].
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48 74 An easy way to overcome the above-mentioned limitation is injecting a sample of larger
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50 75 volume than typical injection, e.g., 1 μL . Recently, a programmable temperature vaporizing
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52 76 (PTV) technique was developed to facilitate large-volume injection (LVI) ^[30-32], resulting in
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54 77 significantly improved sensitivity of GC analysis. For example, sub-ppt levels of 2-MIB and
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56 78 geosmin in water could be determined using CLLE coupled with GC/MS equipped with a
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4 79 PTV device ^[20].
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6 80 This paper aims to develop a new analytical method to quantify 2-MIB and geosmin in
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8 81 raw and tap waters by coupling SPE with LVI-GC/MS technique. The developed method
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10 82 utilized a GC/MS system with a specially designed LVI liner (so called Stomach Insert) and a
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12 83 PTV to quantitate the target analytes that were pre-concentrated with SPE. Using this method,
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14 84 improvement in the sensitivity of GC/MS measurements to determine 2-MIB and geosmin
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16 85 could be achieved with the minimum use of environmentally-harmful solvents. The
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18 86 developed method was validated and compared with the headspace SPME/GC/MS method.
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20 87 Finally, the method of the present study was applied to analyze 2-MIB and geosmin in
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22 88 samples of source waters.
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29 90 **2. MATERIALS AND METHODS**

30 91 **2.1. Standards and reagents**

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33 92 Both 2-MIB and geosmin standards were purchased from Sigma-Aldrich (St. Louise,
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35 93 MO, USA). The stock solution of 1 $\mu\text{g mL}^{-1}$ for method development was prepared by
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37 94 diluting 100 $\mu\text{g mL}^{-1}$, when needed. However, the working solutions of 0.5, 1, 5, 10, and 20
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39 95 ng mL^{-1} were daily prepared by diluting the stock solution. The diluting water of 18.2 $\text{M}\Omega$
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41 96 cm^{-1} was produced using the AquariusTM purification system (Advantec, Kashiwa-shi, Japan).
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43 97 Acetone and hexane as extracting solvents of target analytes were purchased from TEDIA
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45 98 (Fairfield, OH, USA). Phenanthrene- d_{10} and polyethylene glycol 200 were acquired from
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47 99 Sigma-Aldrich (St. Louise, MO, USA).
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52 101 **2.2. Instrument**

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55 102 In this study, a GC/MS system (GCMS-QP2010Plus, Shimadzu, Kyoto, Japan),
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57 103 equipped with a PTV for facilitating LVI developed by AiSTI (Fig. 1a; LVI-S200, AiSTI,
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4 104 Wakayama, Japan), was used to quantify 2-MIB and geosmin in water; particularly, a spiral-
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6 105 shaped liner, named Stomach Insert made by AiSTI (Fig. 1b; Wakayama, Japan), was used
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8 106 for LVI. By twisting a conventional liner, the liner could hold inside a large volume of a
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10 107 liquid sample. Once a sample was injected in the Stomach Insert, hot air was supplied into the
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12 108 PTV unit to increase inside temperature according to a pre-set temperature program. Then,
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14 109 solvent was vaporized and target analytes were introduced to a capillary column to achieve
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16 110 separation.
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22 **[Figure 1 here]**
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27 114 The PTV was connected to an auto-sampler (AOC-20i, Shimadzu, Kyoto, Japan) with a
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29 115 50 μL syringe (SGE, Victoria, Australia). The operating condition of the PTV along with the
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31 116 temperature program is provided in Table 1. The procedure consisted of the following steps;
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33 117 injection, solvent vaporization, split transfer, and cleaning. During injection and vaporization
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35 118 of solvent, the split purge line was open. In these steps, the PTV temperature was raised from
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37 119 70 to 210 $^{\circ}\text{C}$ for about 1.2 min and was still below the boiling point of 2-MIB or of geosmin;
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39 120 boiling points of 2-MIB and geosmin were 207 and 210 $^{\circ}\text{C}$, respectively. Using this approach,
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41 121 the target analytes could be concentrated in the Stomach Insert liner while the solvent was
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43 122 removed through the split purge. After solvent was completely vaporized, the analytes
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45 123 remaining in the liner were transferred to the capillary column as the PTV temperature was
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47 124 rapidly raised to 270 $^{\circ}\text{C}$, at which the temperature was held for 20 min to clean up the insides
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49 125 of the liner.
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53 126 For separation of 2-MIB and geosmin, a Shim-5 MS column (30 m \times 0.25 mm i.d., 0.25
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55 127 μm film thickness; Shimadzu, Kyoto, Japan) was used with helium as a carrier gas. The
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57 128 GC/MS analysis was performed in both SCAN and SIM modes. The detailed operating
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4 129 condition for the GC/MS along with the oven temperature program is also provided in Table
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11 **[Table 1 here]**
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16 134 For comparison, SPME followed by GC/MS analysis was also applied to the water
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18 135 samples. The procedure to extract the target analytes from the headspace of water samples
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20 136 using SPME fibers along with GC condition is provided as Supporting Information (Table
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22 137 SM-1).
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26 27 139 **2.3. Sample pretreatment procedure**

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29 140 The SPE method with a PBX cartridge (20 mg; AiSTI, Wakayama, Japan) was applied
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31 141 to selectively extract 2-MIB and geosmin from a water sample of 100 mL (Fig. 2). The
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33 142 extraction procedure is briefly described below.
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38 144 **[Figure 2 here]**
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43 146 Prior to extracting target chemicals, the SPE cartridge was washed twice with 2 mL pure
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45 147 water, and dried for 5 min. Then, a sample of 100 mL underwent the loading process under
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47 148 approximately 3 kPa in order to allow target analytes to be adsorbed on to the SPE cartridge.
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49 149 The SPE cartridge was then mounted on a vacuum manifold, washed twice with 2 mL water,
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51 150 and dried for 5 min. After drying, the SPE cartridge was eluted with a 1-mL mixture of
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53 151 acetate and hexane (3:7). A 2- μ L acetone mixture containing 0.2% polyethylene glycol 200
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55 152 as an analyte protectant and 10-ppm phenanthrene- d_{10} as an internal standard was added to
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57 153 the eluted sample; in particular, the analyte protectant was added to prevent matrix-induced
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4 154 chromatographic response enhancements caused by the undesirable interactions of matrix
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6 155 compoenets with active sites in the PTV inlet and the GC column. The sample volume was
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8 156 subsequently adjusted to 1 mL by adding a mixture of acetate and hexane (3:7). Finally, the
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10 157 sample was injected into the LVI-GC/MS for quantitation of target compounds.
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159 **2.4. Validation of proposed analytical method**

160 The performance of the sample extraction and analytical method for quantitation of 2-
161 MIB and geosmin developed in this study was evaluated by determining linearity of
162 calibration curves, repeatability and recovery efficiencies, RSDs, instrumental limit of
163 quantifications (LOQs), and method detection limits (MDLs).

164 For developing the calibration curves for 2-MIB and geosmin, standards of each
165 compound at five different concentrations (0.5-20 ng L⁻¹) were prepared by diluting
166 appropriate amounts of the stock solution. All standards were prepared in duplicate. For the
167 repeatability test, 7 standard samples of 5 ng L⁻¹ and 3 standard samples of 50 ng L⁻¹ were
168 prepared by adding each of the two compounds to water, and were analyzed using the
169 proposed method. Recovery tests were performed with source water and produced drinking
170 water. Test samples were prepared by spiking 2-MIB and geosmin in raw or tap water at
171 three different levels (i.e., 5, 10, and 50 ng L⁻¹); five samples were prepared for each level.

172 For repeatability and recovery tests, raw water samples were collected from a water reservoir.

173 The instrumental LOQ was determined by calculating the signal-to-noise (S/N) ratio for
174 the standard solution of the lowest concentration in the calibration curve; 10 times of a signal
175 to noise (S/N) ratio was considered as the LOQ for each target compound. Since each
176 compound has different instrumental responses, determining the MDL of each compound was
177 separately carried out. For MDL determination, solutions containing the target compounds of
178 1 ng L⁻¹ each were prepared in target compounds-free raw water samples (total organic

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4 179 carbon of about $2 \pm 0.1 \text{ mg L}^{-1}$). The MDL with a 99% confidence level that the
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6 180 concentration of a target analyte was considered greater than zero was calculated.
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10 182 **3. RESULTS AND DISCUSSION**

11 183 **3.1. Chromatogram from analysis of 2-MIB and Geosmin**

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15 184 Figure 3 shows the chromatogram for 2-MIB and geosmin, obtained by 25 μL injection
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17 185 (Table 1). Sharp peaks for both target compounds were observed; even at the concentration of
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19 186 0.5 ng L^{-1} , discernable peaks could be observed for both target compounds.
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28 190 In Fig. 4, target ions for 2-MIB and geosmin are shown; the concentration of each
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30 191 analyte was 100 ng L^{-1} . For each of the target compounds, the most abundant fragment ion
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32 192 was used as the quantifier and two additional mass ions were selected as qualifiers (Table 1).
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34 193 Mass ions used for quantification, i.e., 112 and 95 m/z for 2-MIB and geosmin, respectively,
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36 194 are clearly shown in Fig. 4. For the confirmation of target compounds, the tolerance intervals
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38 195 for mass ion ratios between quantifier and qualifiers were set $\pm 30\%$. Considering the
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40 196 injection volume size used in this study (i.e., 25 μL), the sensitivity of the proposed method
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42 197 was promising; using the current PTV system, the injection volume could be increased up to
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55 202 **3.2. Validation of the proposed method**

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57 203 The linearity of the calibration curves, the repeatability, the recovery, LOQ, and the
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4 204 MDL for each target compound were evaluated to validate the suitability of the developed
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6 205 method. Calibration standards were prepared at five different concentrations (0.5-20 ng L⁻¹)
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9 206 to cover the respective dynamic range for both analytes. The linearities of the calibration
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11 207 curves developed for 2-MIB and geosmin were excellent; R²s of both calibration curves were
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13 208 all > 0.999 (See Fig. SM-1).

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15 209 The repeatability test was carried out with standard solutions of two different
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17 210 concentrations, i.e., 5, and 50 ng L⁻¹ (Table 2). The proposed method showed good
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19 211 repeatability for both target compounds. In the case of 2-MIB, RSD values were 6.9% and
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21 212 6.2% for 5 and 50 ng L⁻¹, respectively, while those for geosmin were 4.5% and 3.7%. The
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23 213 repeatability of the proposed method was also tested with raw water samples spiked with the
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25 214 target analytes. Larger RSDs, especially at lower level were observed possibly due to
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27 215 presence of other constituents of the water samples. Noticeably, RSDs were still within 15%
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29 216 (Table 2). At the level of 5 ng L⁻¹, the error value calculated from the repeatability test with
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31 217 raw water was more than two times larger than that with pure water.
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37 219 **[Table 2 here]**

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41 221 The recovery test was performed with both raw water and tap water samples, which
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43 222 were prepared by spiking 2-MIB and geosmin. Before the water samples were used, their 2-
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45 223 MIB and geosmin concentrations were analyzed. 2-MIB and geosmin concentrations of the
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47 224 raw water were 44.6 ± 0.3 (n = 5) and 2.6 ± 0.2 (n = 5) ng L⁻¹, respectively. However, the two
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49 225 target analytes were not detected from the tap water. After background concentrations of the
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51 226 target compounds in the water samples were determined, each water sample was spiked with
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53 227 2-MIB and geosmin. The expected concentration increases of the samples were 5, 10, and 50
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55 228 ng L⁻¹. In general, the method showed to some degree better recovery ratio for geosmin than
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4 229 that for 2-MIB (Table 3). In the test performed with raw water, the proposed method showed
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6 230 72-104% recovery ratio for 2-MIB, and 93-105% for geosmin. On the other hand, it showed
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8 231 61-67% for 2-MIB and 74-85% for geosmin, when tests were performed with tap water. The
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10 232 relatively lower recovery for tap water samples was attributed to the interaction between
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12 233 target compounds or adsorption sites on SPE material and residual chlorine. Lin et al. also
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14 234 reported that residual chlorine causes enlarged errors in 2-MIB and geosmin analysis ^[33]. If
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16 235 residual chlorine was removed from water samples, much better recovery efficiency could be
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18 236 obtained.
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24 238 **[Table 3 here]**
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28 240 The instrumental LOQs were determined by considering the peak area corresponding 10
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30 241 times the S/N ratios for 2-MIB and geosmin. The calculated LOQs for 2-MIB and geosmin
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32 242 were 7.7 and 6.7 ng L⁻¹, respectively. Similar levels of LOQs were also reported with a
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34 243 method based on headspace-SPME coupled to GC/MS ^[22].
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37 244 For the determination of MDLs of the developed method for 2-MIB and geosmin, a total
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39 245 of 7 raw water samples were prepared. The obtained MDLs for 2-MIB and geosmin were
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41 246 0.87, and 0.62 ng L⁻¹, and were compared well with those reported by others. Especially, for
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43 247 the SPME followed by GC/MS analysis, which is the most commonly applied approach, the
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45 248 reported MDLs for 2-MIB and geosmin are 0.5-5 ng L⁻¹ and 0.5-3.3 ng L⁻¹, respectively
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47 249 (Table 4). In this study, the method using SPME-GC/MS was also applied, and its MDLs for
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49 250 2-MIB and geosmin were calculated as 1.5 and 0.6 ng L⁻¹, respectively, which are comparable
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51 251 with those reported by others. However, the main disadvantage of SPME is reduced limited
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53 252 concentration capability due to the small amount of polymer coating on the fiber. In addition,
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55 253 the SPME fiber coating is easily breakable and hence has limited lifetime.
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[Table 4 here]

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3.3. Application of proposed method for analysis of 2-MIB and geosmin in real water.

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The proposed method was applied to quantify the target compounds in raw water samples collected from upstream sites of Han River; it is located approximately 30 km away from the eastern boundary of the Seoul Metropolitan. The sample temperature was about 23

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4 279 °C and dissolved oxygen and total organic carbon concentrations were $8.6 \pm 0.2 \text{ mg L}^{-1}$ and
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6 280 $2.1 \pm 0.2 \text{ mg L}^{-1}$, respectively. The concentrations of 2-MIB and geosmin determined using
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8 281 the developed method were 59.3-65.6 and 8.9-9.4 $\mu\text{g L}^{-1}$, respectively. This result is
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10 282 comparable with those reported by a study previously conducted for Han River [39]. As stated
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12 283 in Section 3.1, the tolerance interval for the ion ratios between quantifier and each of two
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14 284 qualifiers was set at 30% for both 2-MIB and geosmin in raw water samples. The ion ratios
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16 285 calculated for 2-MIB were within the ranges of 19-27% and 10-14% whereas those for
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18 286 geosmin were 15.0-25.7% and 13.4-21.7%, indicating the preset tolerances of absolute ion
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20 287 abundances ratios set for both odorants satisfied.
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25 26 289 **4. CONCLUSION**

27
28 290 A new method consisting of SPE and subsequent LVI-GC/MS was developed to analyze
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30 291 2-MIB and geosmin in water. The proposed method showed a good linearity over the wide
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32 292 calibration range for 2-MIB and geosmin, and good repeatability and recovery. Large volume
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34 293 injection practiced in the proposed method resulted in a good sensitivity for the target
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36 294 analytes. The MDLs of the method for 2-MIB and geosmin are lower than one tenth of the
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38 295 compounds' published odor thresholds, and are comparable with or lower than those obtained
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40 296 using SPME-GC/MS. However, the proposed method is free from the concern about breaking
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42 297 of adsorbent on a fiber frequently witnessed in the practice with SPME. Overall, due to its
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44 298 relative ease of practice and high accuracy and sensitivity, the proposed method can be easily
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46 299 applied to a water quality monitoring program for the odorants.
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51 300 52 53 301 **ACKNOWLEDGEMENT**

54
55 302 This work was supported by the R&D program of MOTIE/KEIT (R&D program number:
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57 303 10037331, Development of Core Water Treatment Technologies based on Intelligent BT-NT-
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366 **Table 1.** Operating conditions for PTV and GC/MS used in this study

| Operating Parameters for PTV | |
|---|---|
| Injector Temp. Program | 70 to 240 °C 120 °C min ⁻¹ , to 270 °C 50 °C min ⁻¹ , and held for 20 min |
| Injector Solvent Purge Time, sec | 7 |
| Injection Volume, µL | 25 |
| Operating Parameters for GC/MS | |
| Oven Temp. Program | held at 50 °C for 3 min, to 180 °C at 10 °C min ⁻¹ , and to 310 °C at 25 °C min ⁻¹ , and held for 5 min |
| Column Flow, mL min ⁻¹ | 1 |
| Carrier Gas Saver at Split Ratio of 20, min | 4 |
| Split Ratio | 150 |
| Ion Source Temp., °C | 280 |
| Interface Temp., °C | 290 |
| Scan Range, <i>m/z</i> | 50-250 |
| SIM, <i>m/z</i> | 97, 112 (quantifier), 125 for geosmin 95 (quantifier), 108, 135 for 2-MIB |

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369 **Table 2.** Results of repeatability tests for 2-MIB and gesmin (n = 5)

| | | 2-MIB | | | | Geosmin | | | |
|--------------------|--------------------|--------------------|--------------------|-----------|--------------------|------------|--------------------|-----------|--|
| | | Pure water | | Raw water | | Pure water | | Raw water | |
| Level, | SD ^a , | RSD ^b , | SD, | RSD, | SD, | RSD, | SD, | RSD, | |
| ng L ⁻¹ | ng L ⁻¹ | % | ng L ⁻¹ | % | ng L ⁻¹ | % | ng L ⁻¹ | % | |
| SPE-LVI-GC/MS | | | | | | | | | |
| 5 | 0.29 | 6.9 | 0.7 | 14.6 | 0.2 | 4.5 | 0.67 | 14.2 | |
| 50 | 3.2 | 6.2 | 3.2 | 6.5 | 1.9 | 3.7 | 3.8 | 8.0 | |
| SPME-GC/MS | | | | | | | | | |
| 5 | 0.48 | 9.7 | | | 0.14 | 3.0 | | | |
| 50 | 1.9 | 3.8 | | | 4.0 | 8.0 | | | |

370 a: standard deviation

371 b: relative standard deviation

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374 **Table 3.** Results of recovery test for 2-MIB and gesmin (n = 5)

| Levels, ng L ⁻¹ | 2-MIB | | | Geosmin | | |
|----------------------------|-------|------|------|---------|------|------|
| | 5 | 10 | 50 | 5 | 10 | 50 |
| Raw water, % | 104.4 | 72.1 | 72.9 | 104.7 | 93.0 | 95.3 |
| Tap water, % | 61.3 | 66.5 | 62.2 | 85.3 | 74.1 | 79.8 |

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377 **Table 4.** Method detection limit values for 2-MIB and geosmin reported in literature and
 378 obtained in this study

| 2-MIB, ng L ⁻¹ | Geosmin, ng L ⁻¹ | Method | Matrix | Reference |
|---------------------------|-----------------------------|----------------------|------------------|-------------------|
| 1.0 | 5.0 | LLE-GC/MS | Pure water | [18] |
| 1.0 | 1.0 | LLE-GC/ITMS | Pure water | [34] |
| 5.0 | 5.0 | CLSA-GC/MS | Pure water | [14] |
| 1.0 | 1.1 | Headspace LLME-GC/MS | Raw water | [35] |
| 0.5 | 0.5 | SPME-GC/MS | Raw water | [36] |
| 0.9 | 0.6 | Headspace SPME-GC/MS | Pure water | [22] |
| 9.7 | 0.94 | SPME-GC/MS | Pure water | [36] |
| 1.52 | 0.6 | SPME-GC/MS | Raw water | This study |
| 9 | 2 | USADLLME-GC/MS* | Pure water | [38] |
| 0.25 | 0.1 | SBSE-GC/MS | Pure water | [24] |
| 0.18 | 0.09 | SBSE-GC/MS | Pure Water | [36] |
| 0.04-0.16 | 0.03-0.11 | SBSE-GC/MS | Mineral water | [37] |
| 0.3 | 0.05 | CLLE-LVI-GC/MS | Pure water | [20] |
| 0.91 | 0.63 | SPE-LVI-GC/MS | Raw water | This study |

379 * USADLLME-GC/MS; ultrasound-assisted dispersive liquid–liquid microextraction coupled to GC/MS.

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3 **Figure legend**
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6 **Fig. 1.** Schematic diagram for PTV unit (a) and picture of Stomach Insert (b) used in
7 this study
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10 **Fig. 2.** Extraction procedure for 2-MIB and geosmin in water
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12 **Fig. 3.** Chromatogram of 2-MIB and geosmin analyzed by GC/MS
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14 **Fig. 4.** Target mass ions for 2-MIB and geosmin analyzed by GC/MS
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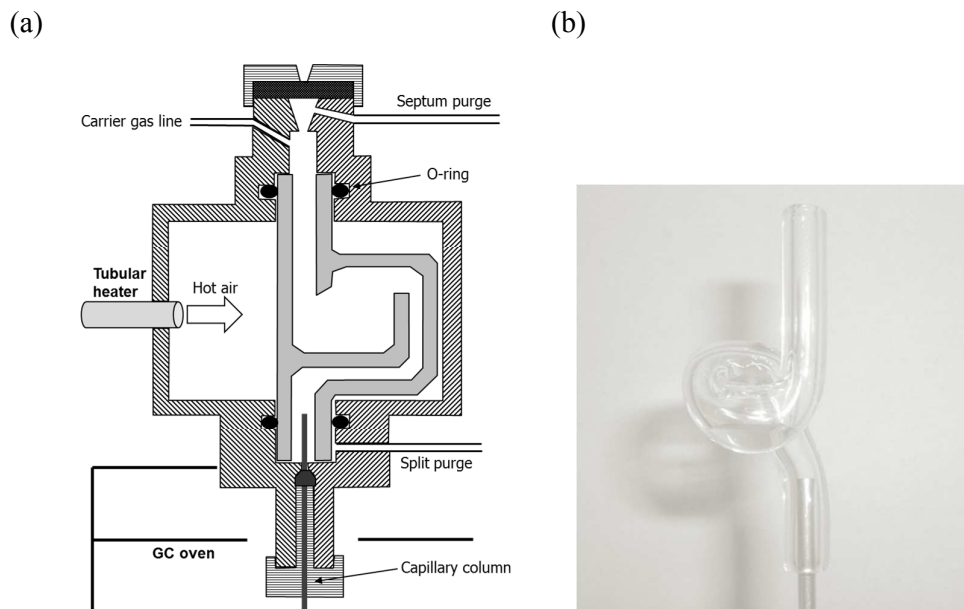


Fig. 1. Schematic diagram for PTV unit (a) and picture of Stomach Insert (b) used in this study

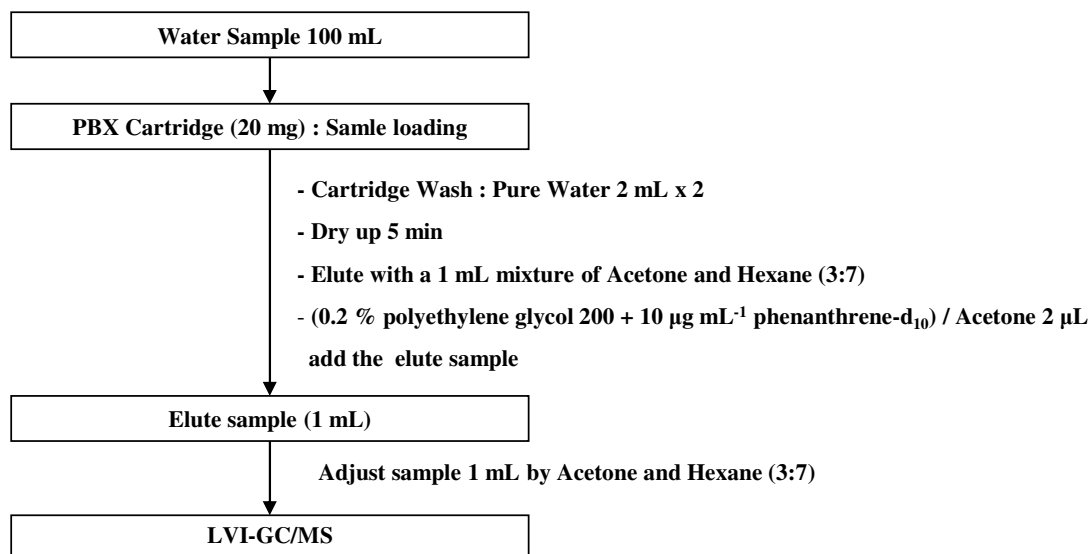


Fig. 2. Extraction procedure for 2-MIB and geosmin in water

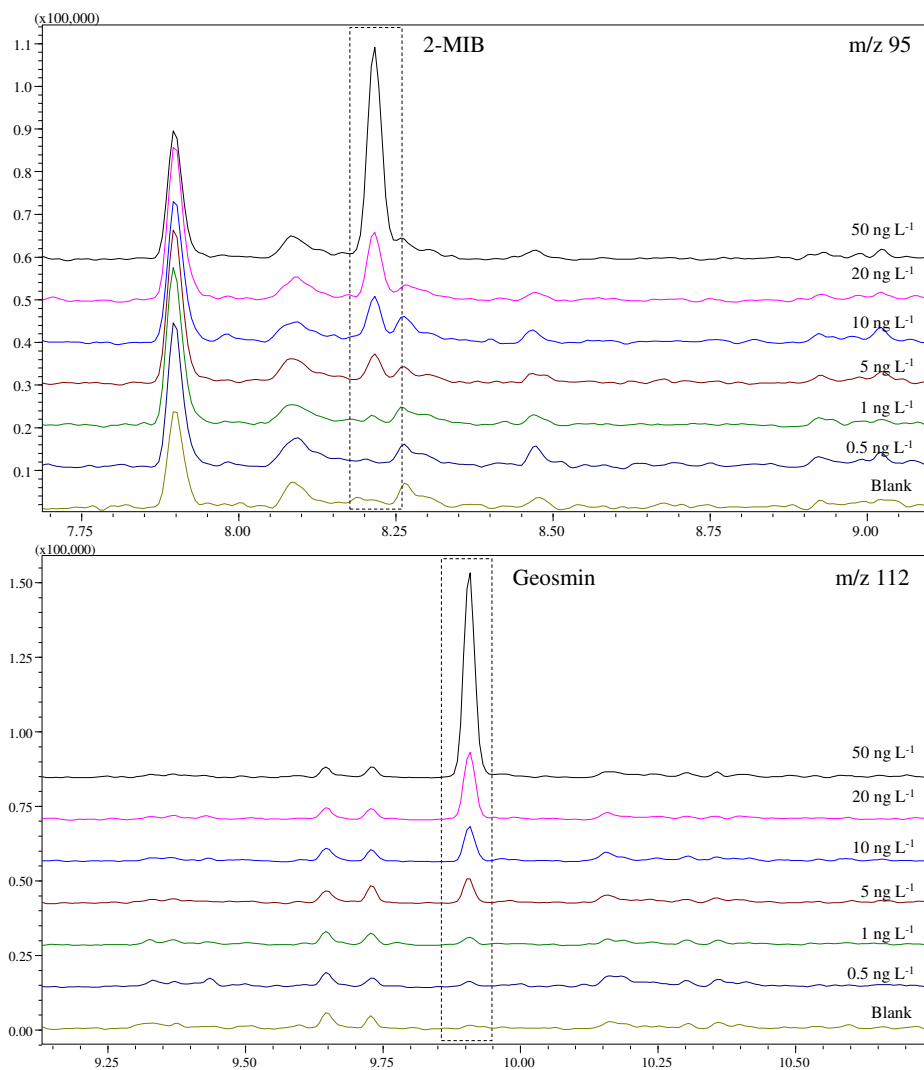


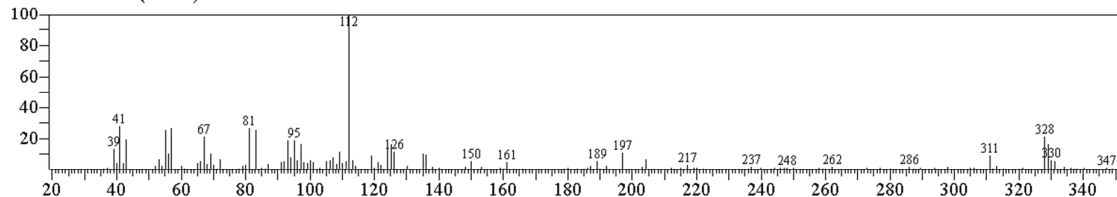
Fig. 3. Chromatogram of 2-MIB and geosmin analyzed by GC/MS

(a) Target mass ions for Geosmin

<< Target >>

R.Time:9.908(Scan#:1341) Retention Index:2612

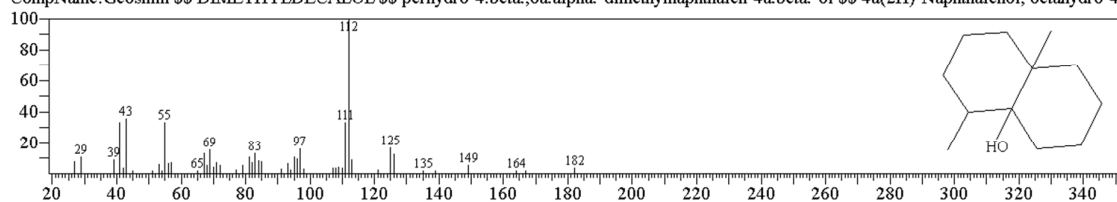
BasePeak:112.10(10000)



Hit#:2 Entry:106651 Library:Wiley9.lib

SI:72 Formula:C12H22O CAS:19700-21-1 MolWeight:182 RefIndex:0

CompName:Geosmin \$\$ DIMETHYLDECALOL \$\$ perhydro-4.beta.,8a.alpha.-dimethylnaphthalen-4a.beta.-ol \$\$ 4a(2H)-Naphthalenol, octahydro-4

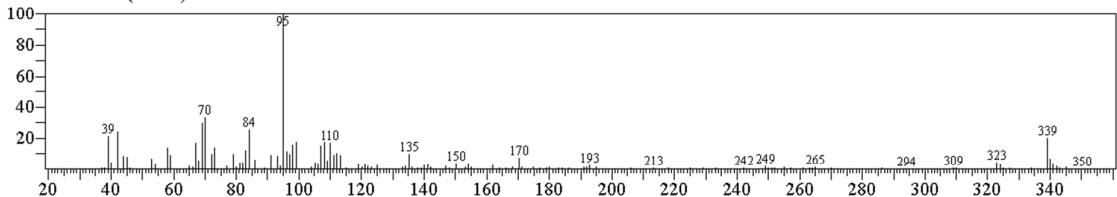


(b) Target mass ions for 2-MIB

<< Target >>

R.Time:8.217(Scan#:761) Retention Index:2134

BasePeak:95.10(10000)



Hit#:1 Entry:81118 Library:Wiley9.lib

SI:67 Formula:C11H20O CAS:91278-70-5 MolWeight:168 RefIndex:0

CompName:2-METHYLISOBORNEOL \$\$ 2-Bornanol, 2-methyl- \$\$ 1,2,7,7-Tetramethylbicyclo[2.2.1]heptan-2-ol \$\$ 1,2,7,7-TETRAMETHYLBIC

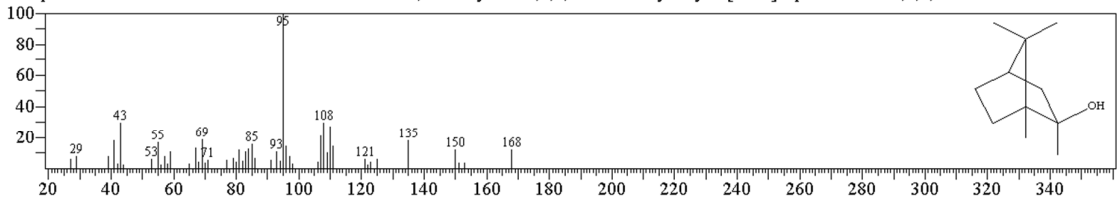


Fig. 4. Target mass ions for 2-MIB and geosmin analyzed by GC/MS