

Analytical Methods

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4 1 **A novel flow injection chemiluminescence method for the**
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6 2 **determination of indole-3-acetic acid in biological sample by using**
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8 3 **trivalent silver**
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Abstract

In this paper, a novel flow-injection chemiluminescence method was developed for the determination of indole-3-acetic acid (IAA). The chemiluminescence signal from the reaction of Ag(III) complex and sulfuric acid system was enhanced in presence of IAA. The conditions of the CL system were investigated and optimized. Under the optimal conditions, the relative chemiluminescence intensity was linear with the IAA concentration in the range of $1.0 \times 10^{-10} \text{ g} \cdot \text{mL}^{-1}$ - $1 \times 10^{-8} \text{ g} \cdot \text{mL}^{-1}$. The detection limit for IAA was $7.7 \times 10^{-11} \text{ g} \cdot \text{mL}^{-1}$, and the relative standard deviation (n = 11) was 0.5%. The proposed method was applied to the analysis of IAA in human urine, mung bean sprouts and soil samples with the recoveries of 103.5%-117.1%, 87.0%-98.4% and 94.1%-118.8%, respectively, and the relative standard deviations was 0.6-2.7%. The results obtained by the proposed method agreed well with those obtained from HPLC method. The chemiluminescence mechanism was discussed by comparison of fluorescence spectra and the UV-vis absorption spectra.

Keywords: Chemiluminescence; Flow injection, Indole-3-acetic acid; Trivalent silver; biological sample.

1. Introduction

Indole-3-acetic acid is released by the terminal bud of a shoot and acts as a true chemical messenger regulating some important biological processes of plants, e.g., division, elongation and differentiation of cells.¹ And the effects of using IAA in agriculture on environment and human health have aroused widespread concern.² However, it is difficult for determination of IAA because the concentration in plants is at very low level(at the ng/g level).³ Therefore, it is significant to search for a method with good sensitivity and high accuracy for IAA detection.

So far, some sensitive methods have been reported for determination of IAA in biological samples, such as immunoassay,⁴ capillary electrophoresis(CE) ,^{5,6} electrochemical methods^{1,7}, colorimetry⁸, fluorometry,^{9,10} chromatography methods(LC or GC) with different detection strategies ,^{3,11-15} and LC/GC-MS.¹⁶⁻²⁰ However, there are some disadvantages exist, for example, enzyme-linked immunosorbent assay suffers from antibodies' cross-reactivity with other compounds of similar structure present in the same sample;¹¹ chromatography and MS need special instruments and strict experimental conditions. The application of CE and electrochemical methods in analysis of complex real sample is limited for the relatively poor reproducibility.¹⁶

Chemiluminescence method (CL) has received much more attention for the analysis of organic and inorganic species owing to its low detection limit, high sensitivity, wide linear dynamic range, short response time and relatively simple instrumentation.²¹ The flow injection technique (FIA) is based on the injection of a sample into a non-segmented carrier stream targeting to the reaction system. The FIA-CL method is characteristic of well

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4 67 sensitivity and reproducibility. Diperoatoargentate (III) (DPA) is widely used as oxidizing
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6 68 agent²² in inorganic chemistry and may also be used as polymerization initiator.²³ In recent
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8 69 years, there are some reports about using DPA in chemiluminescence,²⁴ however, to our best
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11 70 knowledge, no work in literature have reported using DPA to detect IAA without any
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14 71 luminescence reagent such as luminol involved.

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16 72 The weak chemiluminescence in acid conditoinis is result from the DPA as oxidant. It
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18 73 can be enhanced in presence of IAA. Based on the CL reaction, a novel chemiluminescence
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21 74 method has been established for the determination of IAA by combining with flow-injection
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24 75 technique. The experimental conditions have been optimized. It has been proved to be a
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26 76 simple, fast and precise method for the determination of IAA with low detection limit. The
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28 77 proposed method has been successfully applied for the determination of IAA in human urine,
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30
31 78 mung bean sprouts and soil samples. The chemiluminescence mechanism also has been
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34 79 studied by using of the ultraviolet visible absorption spectra and fluorescence spectra.

35 36 37 80 **2. Experimental**

38 39 81 **2.1. Materials**

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42 82 IAA was purchased from Sigma-Aldrich (St. Louis, MO, USA). The used reagents were
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45 83 listed as follow: Potassium periodate (KIO₄), potassium persulfate (Na₂S₂O₈) and argentum
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47 84 nitricum (AgNO₃) was purchased from Guangzhou Guanghua Chemical Factory Co. Ltd.
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50 85 (Guangzhou, China). Potassium hydroxide (KOH) and concentrated sulfuric acid (H₂SO₄)
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52 86 were purchased from Guangzhou Chemical Reagent Factory (Guangzhou, China). All of the
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55 87 chemicals were of analytical reagent grade, and were used without further purification.
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58 88 Doubly distilled water was used throughout the work.

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4 89 The DPA complex was prepared by Ag(I) compound oxidized in an alkaline medium
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6 90 according the known method.²⁵ In brief, KIO₄ (3.24 g), AgNO₃ (1.36 g), Na₂S₂O₈ (3.0 g) and
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9 91 KOH (8.0 g) were added in 200 mL of deionized water. The mixture was heated reflux to
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11 92 boiling for about 40 min on a hot plate with constant stirring. The boiling mixture turned
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13 93 intensely red and the boiling was continued for another 20 min until the completion of the
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16 94 reaction. The mixture was then cooled and filtrated. The obtained DPA complex was stored
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19 95 under refrigeration in dark place, and was found to be fairly stable for several months. DPA
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21 96 solutions were freshly prepared before use. The complex was characterized by the UV/Visible
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23 97 spectrum, which exhibited a brood band at 361 nm with the molar absorptivity (ϵ) of 1.26×10^4
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26 98 L·mol⁻¹·cm⁻¹. The IAA stock solution was 1.6 mg·mL⁻¹, it was prepared by dissolving
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29 99 appropriate amount of standard substance with a small volume of methanol and then diluted
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31 100 into brown volumetric flask with double-distilled water. The IAA working standards solutions
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34 101 were prepared from the stock solution by appropriate dilutions.

35 36 102 **2.2. Apparatus**

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38 103 The CL-FIA system used in this work was shown in **Fig. 1**. Two peristaltic pumps
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41 104 (BT100-1J, Henan Baoding Lange, China) were used to deliver all chemicals at a flow rate of
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43 105 4.0 mL·min⁻¹. PTFE flow tubes (0.8 mm i.d.) were used to connect all components in the
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46 106 system. Injection was made by using a sixteen-port injection valve (Hanzhou, China)
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48 107 equipped with a loop of 100 μ L. The CL signal was monitored by a BPCL ultra-weak
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51 108 luminescence analyzer (Institute of Biophysics, Chinese Academy of Science, Beijing, China)
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53 109 consisting of a flat coil glass flow cell facing the window of the photomultiplier tube (PMT).
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56 110 Data acquisition and treatment were performed with BPCL software. The UV-absorbance was
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4 111 detected with the Cary 300 spectrophotometer (Varian Ltd, American). The CL spectrum was
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6 112 obtained with the RF-5301PC fluorospectrophotometer (Shimadzu Ltd, Japan). MAS-I
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8 113 Microwave extraction response instrument (Shanghai new instrument microwave chemical
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11 114 technology Co., LTD).

12 13 14 115 **2.3. Procedures**

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16 116 FI-CL method: The flow-injection system was easy to operate. As shown in the **Fig. 1**,
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18 117 the peristaltic pump propelled the DPA solution, H₂SO₄ solution and analyte (a standard IAA
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20 118 solution or a sample containing IAA) through the system at 4.0 mL min⁻¹, respectively. When
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22 119 the injection valve was set to the load position, the H₂SO₄ was merged with DPA solution at
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24 120 “M” point before the DPA ran through the whole system until a stable baseline was recorded.
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26 121 When the injection valve was switched to the inject position, the H₂SO₄ carried the sample
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28 122 solution in the reagent loop (100 μL IAA solution), and ran directly through the flow cell,
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30 123 producing CL emission. Also the CL signal was then recorded simultaneity. The reagent IAA
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32 124 injection mode was chosen in the FIA system for the lower baseline was recorder in the
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34 125 absence of IAA solution. The PMT operated at -1200 V. The relative CL intensity, ΔI (defined
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36 126 as the difference of CL intensity between in present and in absence of IAA solution,
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38 127 respectively), was proportional to the corresponding concentration of IAA.
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46 128 **2.4 Sample preparation**

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48 129 Urine, bean sprout and soil were collected for analysis. Urine sample was taken from a
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50 130 healthy male person. In order to remove the reducing substances such as ascorbic acid and
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52 131 glutathione, the 50 mL urine sample (contain 0.1 g Na₂CO₃) was heated at 60°C for 10 min.
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55 132 An appropriate amount of standard IAA solution was added into the 10 mL treated urine
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4 133 sample. The spiked samples were extracted with CH_2Cl_2 for three times after stayed for 30
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6 134 min. Then the distilled water was added for analysis after the organic phases were collected
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9 135 and drying. The spiked urine sample with concentration of 10.0, 20.0, 40.0 $\text{ng}\cdot\text{mL}^{-1}$ were
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11 136 directly analyzed. The experimental were approved by the Ethics Committee of Sun Yat-sen University,
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14 137 and all the procedures were performed in accordance with the Regulations for the Administration of
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16 138 Affairs Concerning Experimental. Informed consent was obtained for the experimentation with human
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19 139 urine.

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21 140 The bean sprout was purchased from the market. 500 g bean sprout were grinded and
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24 141 centrifuged. The obtained liquid was filtered by using filter paper and stored in refrigerator
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26 142 overnight. A suitable volume filtrate was set into a flask and mixed with IAA standard
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29 143 solutions. It was mixed thoroughly and kept for 0.5 h. The extraction was performed in 25 mL
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31 144 CH_2Cl_2 for three times after stayed for 0.5 h. The distilled water was added for analysis after
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34 145 the organic phases were collected and evaporate to dryness. The beans sprout without IAA
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36 146 standard solutions was regarded as blank sample. Three different concentration levels of IAA
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39 147 were spiked into the samples with the concentration of 0.050, 0.100 and 0.200 $\text{mg}\cdot\text{Kg}^{-1}$.

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41 148 The soil sample was collected from the lawn in the campus. And allowed it to air dry.
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44 149 The dry soil was ground into powder and sieving. An appropriate amount of standard IAA
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46 150 solution was added into the 2.5 g soil sample. The spiked samples were oscillation extracted
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49 151 with CH_2Cl_2 for three times after stayed for 30 min. The extract liquor endured vacuum
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51 152 distillation and ultrasonic pretreatment. Then the distilled water was added for analysis after
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54 153 the organic phases were removed. The spiked urine sample with concentration of 0.50, 1.25,
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56 154 2.50 $\text{mg}\cdot\text{Kg}^{-1}$ were directly analyzed.
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155 3. Results and Discussion

156 3.1 Kinetics curve of the CL reaction of DPA-H₂SO₄-IAA

157 In the primary experiment, the emitted CL was based on the oxidation reaction of IAA by
158 DPA in the acid medium. In the batch mode, a typical intensity versus times response
159 curvewas used to describe the CL reaction. The response curve depended on the experimental
160 factors such as reagent concentrations. So the experimental parameters were kept constant, the
161 intensity time curve of IAA-H₂SO₄-DPA was recorded to study the kinetic characteristic of
162 the CL reaction in **Fig.2**. The CL intensity peak appeared within 0.8 s since the DPA was
163 injected into the mixture solution of H₂SO₄ and IAA (or blank). The CL signals would
164 decrease to baseline within 32 s. As seen from the **Fig. 2**, the CL reaction of IAA-DPA-H₂SO₄
165 was a quick reaction obviously. The kinetic curve indicated the CL system was rapid and
166 sensitive enough and suitable for the analysis of IAA.

168 3.2 Optimization of the experiment procedure

169 3.2.1 The effect of DPA solution

170 The CL was emitted from the oxidation reaction of IAA by DPA in acid medium. As the
171 only oxidant, the concentration of DPA was important. To test the effect of DPA solution, a
172 range of concentrations of DPA (5×10^{-5} mol·L⁻¹ to 6×10^{-4} mol·L⁻¹) was investigated. As
173 shown in **Fig.3**, the CL intensity was increased along with the increase of the DPA
174 concentration in a low-concentration range, and reached the maximum at 4.5×10^{-4} mol·L⁻¹.
175 When the concentration was over 4.5×10^{-4} mol·L⁻¹, the CL intensity decreased probably
176 because higher concentration of DPA caused self-absorption. Hence 4.5×10^{-4} mol·L⁻¹ was
177 optimal concentration and was selected in the subsequent work.

178 3.2.2 The effect of acid medium

179 In the prime experiment, the CL intensity involved DPA in acid medium could be
180 enhanced in presence of IAA. The influence of the common acid such nitric acid,
181 hydrochloric acid, poly phosphoric acid, sulfuric acid and phosphoric acid were investigated.
182 The results show that the CL intensity of DPA-IAA in the sulfuric acid was maximum and
183 stable. When the other conditions were fixed, the effect of H₂SO₄ concentration was test over
184 the range 0.1 mol·L⁻¹ to 4.0 mol·L⁻¹. As the result in **Fig. 4** demonstrated, the relative CL
185 intensity was increased alone with an increase of the H₂SO₄ concentration in a concentration
186 range (<1.0 mol·L⁻¹), and reached a maximum; when the concentration was over 1.0 mol·L⁻¹,
187 the CL intensity decreased obviously with an increase of the H₂SO₄ concentration. One
188 possible explanation for our observation may be that enhanced baseline noise resulted from
189 released heat when the H₂SO₄ was at higher concentrations. Consequently, the optimal H₂SO₄
190 concentration was 1.0 mol·L⁻¹.

191 3.2.3 The effect of flow rate

192 Within a given variable range of 2.0-6.0 mL·min⁻¹, the CL system was investigated. The
193 result showed that the relative CL intensity increased with the increasing of flow rate in the
194 range of 2.0-4.0 mL·min⁻¹, no obvious increasing of the relative CL intensity was observed
195 when further increase flow rate to 6.0 mL·min⁻¹. And high flow rate may lead to high
196 background, decrease of signal-to-noise ratio and poor reproducibility. Based on the points
197 above, we used 4.0 mL·min⁻¹ in the whole experiment.

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199 3.3 Analytical characteristic of the FI-CL method

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4 200 Under the optimum experimental conditions, the calibration graph of change of CL
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6 201 intensity, ΔI , against IAA concentration was linear in the range of 1.0×10^{-10} g·mL⁻¹ to 1.0×10^{-8}
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8 202 g·mL⁻¹ with a detection limit of 7.7×10^{-11} g·mL⁻¹ (3σ). The regression equation was ΔI
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11 203 $=66.69+411.15c$ (c being the IAA concentration (ng·mL⁻¹)). The relative standard deviation
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13 204 was 0.5% for 1.0×10^{-9} g·mL⁻¹ IAA ($n=11$). The result was comparable and even better than
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16 205 most of those obtained from the other methods reported in literatures, as shown in **Table 1**.
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18 206 The proposed method displayed higher sensitivity and a wider linear range.

21 207 **3.4 Influence of coexisting foreign species**

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25 208 Under the optimum experimental conditions, the influence on CL intensity of some
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27 209 foreign coexisting component in human urine, bean sprouts and soil samples was examined.
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30 210 The experiments were carried out by comparing with the intensities obtained with and without
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32 211 the potentially interfering substances added (**Table 2**). The results indicated that under the
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34 212 experimental conditions of 1 ng·mL⁻¹ IAA and a given relative error (less than 5%), the
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36 213 interference from common component and ions can not be ignored, especially uric acid and Cl⁻,
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38 214 that is probably because they affected the formation of luminescence intermediates. So it
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41 215 was necessary to do pretreatment work before the detection of urine sample.

42 216 **3.5 Analytical applications in the determination of real sample**

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50 217 According to the procedure detailed in the experimental section, the proposed method was
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52 218 applied to the determination of IAA in human urine, bean sprouts and soil. And recovery
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54 219 studies were carried out on real samples for the evaluation of the validity of the proposed
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57 220 method. The No.1 results of each matrix in **Table 3** were further compared with that obtained
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4 221 by HPLC method. As shown in the table **Table 3**, the recovery values for the three samples
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6 222 were in the range of 87.0-118.1% with the RSDs of 0.6 - 2.7%. The HPLC value for the three
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9 223 matrix were 42.8 ± 0.3 , 0.102 ± 0.006 and N.D. respectively, which agreed well with the results
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11 224 obtained by the proposed method.

12 13 225 **3.6 Mechanism studies of the CL reaction**

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17 226 To explore the mechanism of the CL reaction, the ultraviolet visible absorption spectra
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19 227 and fluorescence spectra of the proposed system were recorded.

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22 228 The ultraviolet visible absorption spectra of $\text{IAA}-[\text{Ag}(\text{HIO}_6)_2]^{5-}-\text{H}_2\text{SO}_4$ is shown in **Fig.**
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24 229 **5**. DPA had a characteristic absorption peak at 357 nm. After adding sulfuric acid to DPA
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26 230 solution, the color of DPA gradually faded away, and intensity of the characteristic absorption
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28 231 at 357 nm decreased, which demonstrated reactions took place between sulfuric acid and DPA.
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31 232 When IAA mixed with DPA or DPA-sulfuric acid, the spectra of IAA is obviously different
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33 233 from that obtained from simple superpose the spectra of IAA and DPA or DPA-sulfuric acid.
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36 234 It was obvious that reactions have taken place between IAA and DPA.

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41 235 The fluorescence spectra of $\text{IAA}-[\text{Ag}(\text{HIO}_6)_2]^{5-}-\text{H}_2\text{SO}_4$ is shown in **Fig. 6**. Both sulfuric
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43 236 acid and DPA were non-fluorescent in the wavelength range of 300-450 nm, but the
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45 237 matrix spectral peak of DPA existed. However, after the two compounds were mixed, the
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47 238 matrix spectral peak of DPA was greatly changed. Based on these spectra, we could conclude
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49 239 that DPA had reacted with sulfuric acid. IAA could generate characteristic fluorescence
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51 240 emission under ultraviolet excitation and the maximum excitation and emission
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53 241 wavelength were observed at 289 nm and 361 nm respectively. The fluorescence spectra of
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4 242 the mixture containing IAA and sulfuric acid showed a decline in fluorescence emission
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6 243 intensity, but no significant shift was observed. It suggested that there was no chemical
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9 244 reaction took place between sulfuric acid and IAA. When IAA mixed with DPA, the
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11 245 fluorescence emission intensity of IAA decreased and significant shifts were observed, it can
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13 246 be inferred that a chemical reaction occurred between sulfuric acid and IAA. When sulfuric
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16 247 acid, IAA and DPA were mixed together, the emission peak at 361 nm disappeared and a new
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18 248 fluorescence emission with much lower intensity arised at wavelength of 385 nm, which
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21 249 proved that intensive reaction had take place among the three compounds. Based on the
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23 250 discussion above, the possible chemiluminescence mechanism of IAA-[Ag(HIO₆)₂]⁵⁻-H₂SO₄
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26 251 system could be described as follows(**Fig. 7**).

252 **4. Conclusion**

253 In this study, a novel FI-CL method for the determination of IAA was reported based on
254 its luminescence enhancement of diperiodatoargentate (III) in sulfuric acid. The enhanced CL
255 systems proposed has been applied for the determination of IAA at the 10 ng·mL⁻¹ level in
256 human urine, mung bean sprouts and soil samples with satisfactory results. With rapidity, low
257 detection limit, high sensitivity, good linearity, this approach showed great potential for the
258 trace determination of IAA in biological samples in the routine use. The CL reaction
259 mechanism was discussed briefly.

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4 307 **Figure Captions**

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7 308 **Fig. 1 Schematic diagram of flow injection CL analysis system**

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9 309 P, peristaltic pump; V, injection valve; M, mixtured position; F, spiral glass flow cell; PMT,
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11 310 photomultiplier tube; W, waste; PC, personal computer.

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14 311 a, IAA solution or blank solution; b, H₂SO₄ solution; c, DPA solution.

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18 312 **Fig. 2 Kinetic curves of IAA-[Ag(HIO₆)₂]⁵⁻-H₂SO₄**

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20 313 [Ag(HIO₆)₂]⁵⁻: 4.5×10⁻⁴ mol·L⁻¹; IAA: 1.0×10⁻⁸ g·mL⁻¹; H₂SO₄: 1.0 mol·L⁻¹.

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24 314 **Fig. 3 Effect of DPA concentration on chemiluminescence intensity**

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26 315 H₂SO₄: 0.01mol·L⁻¹; IAA: 5.0×10⁻⁸ g·mL⁻¹.

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30 316 **Fig. 4 Effect of H₂SO₄ concentration on chemiluminescence intensity**

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32 317 DPA: 4.5×10⁻⁴ mol·L⁻¹, IAA: 5.0×10⁻⁸ g·mL⁻¹.

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36 318 **Fig. 5 UV-vis spectra of different systems**

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38 319 (1)H₂SO₄; (2) [Ag(HIO₆)₂]⁵⁻; (3)H₂SO₄+ [Ag(HIO₆)₂]⁵⁻; (4)[Ag(HIO₆)₂]⁵⁻+ IAA;

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40 320 (5)H₂SO₄+ IAA; (6)IAA; (7)H₂SO₄+ [Ag(HIO₆)₂]⁵⁻+ IAA.

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43 321 H₂SO₄: 0.05 mol·L⁻¹; IAA: 1×10⁻⁵ g·mL⁻¹; [Ag(HIO₆)₂]⁵⁻: 5.0×10⁻⁵ mol·L⁻¹.

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47 322 **Fig. 6 Fluorescence emission spectra of IAA-[Ag(HIO₆)₂]⁵⁻-H₂SO₄ system**

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49 323 (1)H₂SO₄; (2)IAA; (3)[Ag(HIO₆)₂]⁵⁻; (4)H₂SO₄+IAA; (5)[Ag(HIO₆)₂]⁵⁻+IAA;

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51 324 (6)H₂SO₄+ [Ag(HIO₆)₂]⁵⁻; (7)H₂SO₄+ [Ag(HIO₆)₂]⁵⁻+ IAA.

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55 325 **Fig. 7 Possible mechanism of IAA-[Ag(HIO₆)₂]⁵⁻-H₂SO₄ system**

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327 **Tables**

328 Table 1 Comparison of analytical method reported in literatures

Method	Matrix	Linear range (ng·mL ⁻¹)	Detection Limit (ng·mL ⁻¹)	Reference
ED	cinnamomum camphora, prunus yedoensis Mats, firmiana simplex.	17.52-1226.4	8.75	1
CE-UV	banana, cabbage, cucumber.	2-500	0.67	5
FL	fruit juice.	45-640	12	10
HPLC-UV	coconut juice.	10-2000	1.3	15
HPLC-FL	cucumber, tomato.	0.18-35.08	0.063	11
LC-MS/MS	pyropia haitanensis, laminaria japonica.	5-500	0.105	18
MIM-SPR	peach, rosa, crape myrtle.	(1.75-87.60)×10 ⁻⁴	0.35×10 ⁻⁴ (Peach)	26
FI-CL	human urine, mung bean sprouts, soil.	0.10-10	0.077	This work

329 ED: electrochemical detection

330 FL: fluorescence

331 CL: chemiluminescence

332 MIM-SPR: Molecular Imprinting Monolayer Techniques on a Surface Plasmon Resonance Sensor.

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334 Table 2 The interference of some common foreign species on the determination of
 335 $1.0 \times 10^{-9} \text{ g} \cdot \text{mL}^{-1}$ IAA

Foreign species	Tolerance limits	Foreign species	Tolerance limits
Cl ⁻	0.1	uric acid	0.1
CO ₃	100	glucose	1
NO ₃	1	starch	10
Ac ⁻	1000	urea	100
EDTA	1	vitamin C	0.05
Co ²⁺	0.1	Ni ²⁺	100
Na ⁺	200	Zn ²⁺	100
NH ₄ ⁺	1	Cu ²⁺	10000
Fe ³⁺	0.5	Fe ²⁺	0.1
Mg ²⁺	1	Ni ²⁺	1

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341 Table 3 The results of IAA in human urine, mung bean sprouts and soil samples(n = 3)

Samples	No.	Detected	Added	Found	Recovery (%)	RSD (%)
human urine (ng·mL ⁻¹)	1		10.0	55.8	117.1	1.5
		44.1 ± 0.9	20.0	65.6	107.8	2.5
			40.0	85.9	104.5	1.5
	2		10.0	102.6	103.5	1.3
		92.2±1.1	20.0	115.5	116.4	2.6
			40.0	134.1	104.7	2.0
Mung bean sprouts (mg·Kg ⁻¹)	1		0.050	0.148	94.6	2.4
		0.101± 0.003	0.100	0.199	97.6	2.2
			0.200	0.298	98.4	1.2
	2		0.050	0.160	92.4	2.7
		0.114±0.004	0.100	0.210	95.7	1.3
			0.200	0.288	87.0	2.1
soil (mg·Kg ⁻¹)	1		0.50	0.59	118.8	1.1
		N.D.	1.25	1.45	115.9	1.0
			2.50	2.54	101.8	0.7
	2		0.50	0.57	113.1	1.4
		N.D.	1.25	1.27	101.3	0.6
			2.50	2.35	94.1	1.2

342 N.D. not detected

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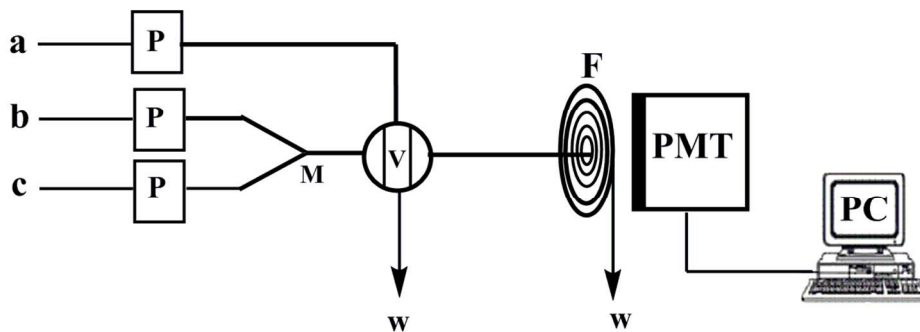


Fig.1
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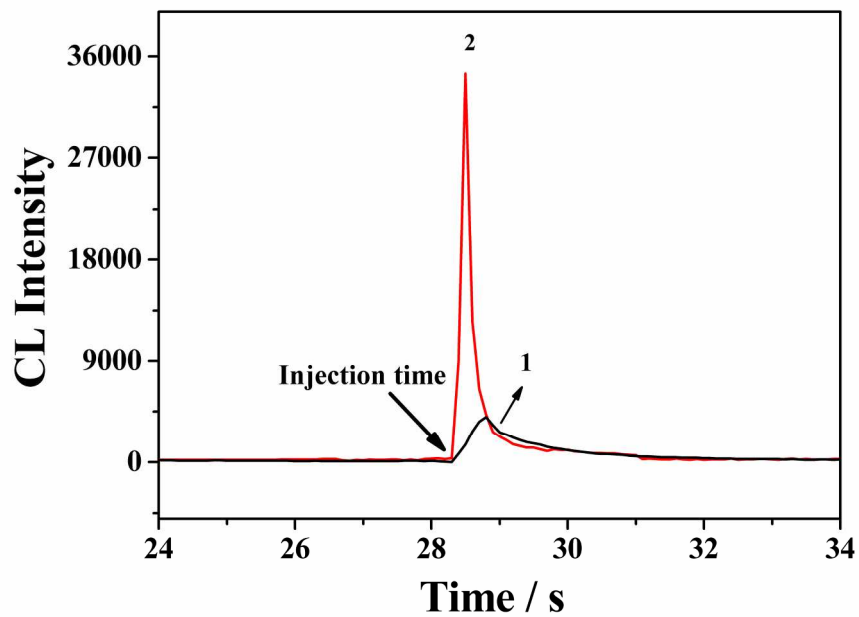


Fig.2
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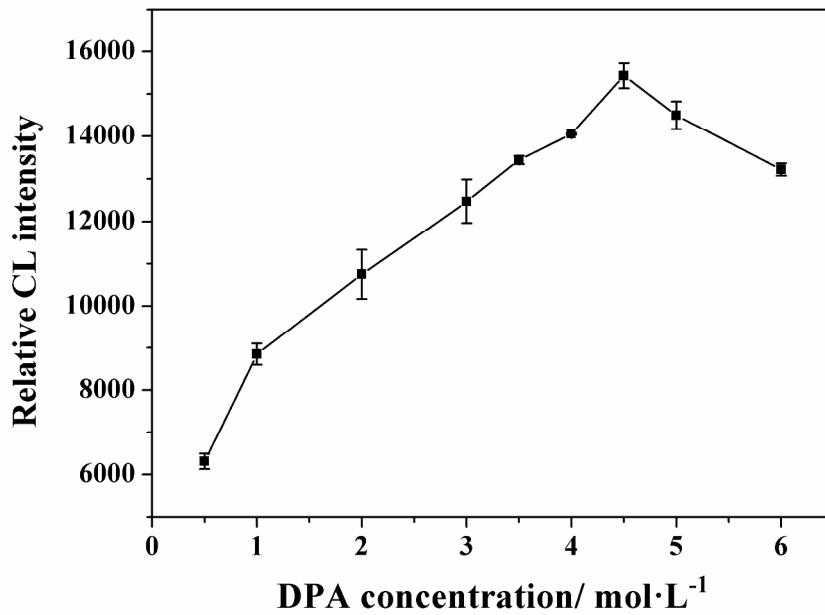


Fig.3
508x359mm (150 x 150 DPI)

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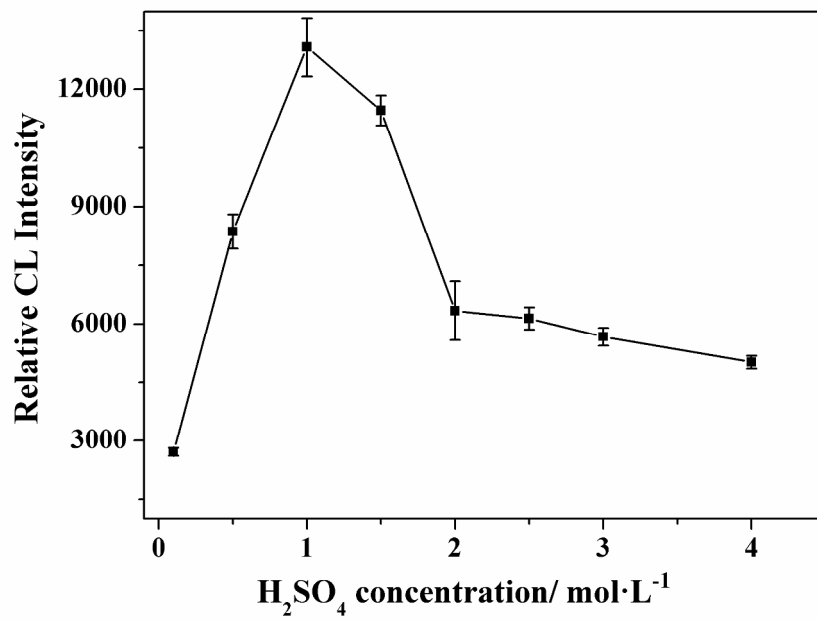


Fig.4
508x359mm (150 x 150 DPI)

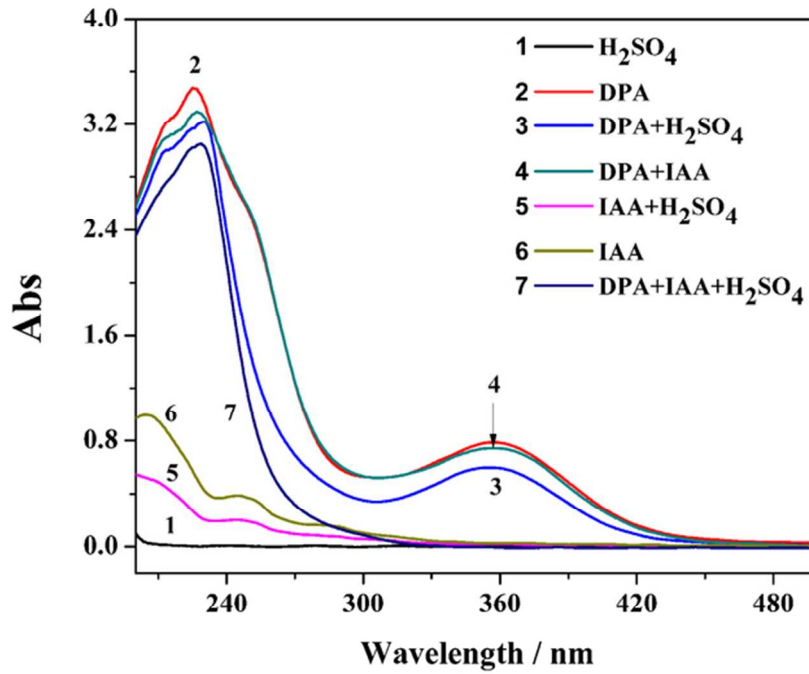


Fig.5
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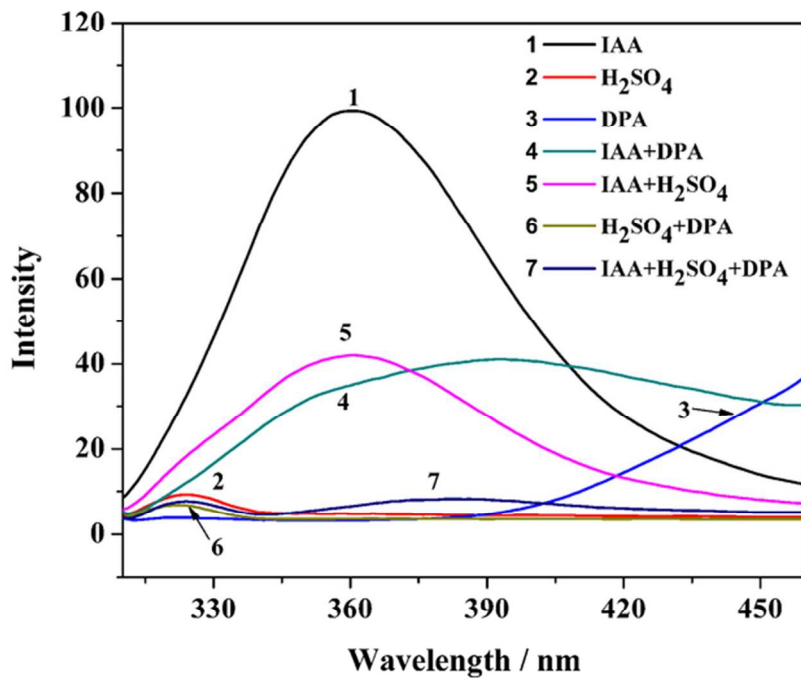


Fig.6
65x50mm (300 x 300 DPI)

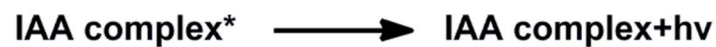
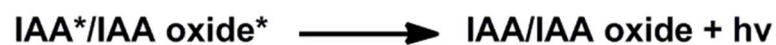
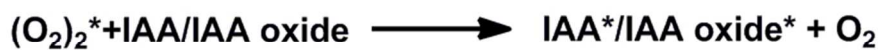
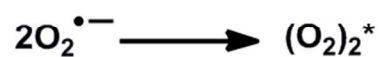
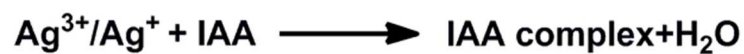
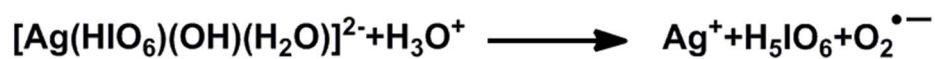
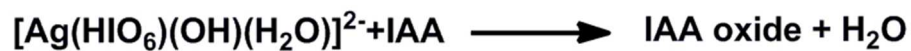
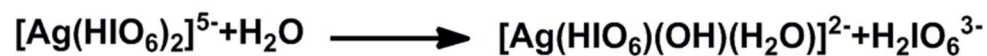


Fig.7
71x62mm (300 x 300 DPI)