# Chemical Science

## Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemicalscience

# **Chemical Science**

### ARTICLE

Cite this: DOI: 10.1039/x0xx00000x Received 00th ZZZ 2014, Accepted 00th ZZZ 2014 DOI: 10.1039/x0xx00000x www.rsc.org/

## Well Plate Circular Dichroism Reader for the Rapid Determination of Enantiomeric Excess

P. Metola,<sup>*a*</sup> S.M. Nichols,<sup>*b*</sup> B. Kahr,<sup>*b*\*</sup> and E.V. Anslyn<sup>*a*\*</sup>

Circular dichroism (*CD*) spectropolarimeters typically employ one photoelastic modulator. However, spectropolarimeters employing two or even four modulators are more versatile and can be used to subvert common measurement errors arising from imperfectly isotropic samples or sample holders. Small linear anisotropies that can cause large errors in *CD* measurement can be associated with multi-well sample holders. Thus, high-throughput *CD* analyses in multiwell plates have not yet been demonstrated. One such application is the determination of enantiomeric excess of a library of reaction products. Herein, a spectropolarimeter employing four photoelastic modulators and a translation stage was used to determine the enantiomeric excess of a family of chiral amine complexes much more rapidly than could be achieved with a robotic fluid injection system. These experiments are proof of concept for high-throughput *CD* analysis. In practice, commercially available glass bottomed well plates are sufficiently strain free such that a simple instrument with just one photoelastic modulator and a vertical optical train should be able to deliver the *CD* without special considerations given herein. On the other hand, polystyrene well plates cannot be used in this way.

#### Introduction

Chiral HPLC and GC are the most common methods for determining enantiomeric excess (ee), but they are serial and slow.<sup>1-5</sup> Demand for simple, fast assays for the determination of ee has grown over the last several years driven by the development of high-throughput screening (HTS) protocols for reaction development using parallel synthesis<sup>6-10</sup> in which reagent or catalyst ligands and/or reaction conditions are varied systematically.<sup>11-21</sup> Parallel synthesis has become popular with the advent of micro-well plates, micro-reactors, mini-block reactors, and automated liquid handlers.<sup>22,23</sup> Given the synthetic efficiency of parallel syntheses, a limiting step in asymmetric catalyst discovery is the determination of ee. Unfortunately, a fast CD plate reader that can deliver ee in a high-throughput platform has not been demonstrated. By contrast, Leung and Anslyn utilized colorimetric indicator displacement assays<sup>28</sup> that require only 1 to 10 minutes to read a 96-well plate with a simple UV-vis spectrophotometer, bona fide high-throughput screens that can analyze thousands of samples per day.<sup>24,2</sup>

The design and construction of an apparatus able to perform the sampling operations and read a 96-well plate may bring *CD* into the realm of HTS, a boon to the synthesis community because a growing number of *ee* assays involve chiroptical measurement.<sup>29</sup> For example, the groups of Anslyn and Wolf have developed and tested *CD* methods for the analysis of chiral analytes including amino acids<sup>30</sup>, alcohols,<sup>31</sup> amino

alcohols<sup>32,33</sup> carboxylates<sup>34</sup> and amines.<sup>35-38</sup> Biological/biochemical sciences may benefit likewise. *CD* spectroscopy is sensitive to the conformational analysis of peptides, proteins, and nucleic acids.<sup>39,40</sup> *CD* can also be used to study dynamic processes such as binding interactions between a macromolecule and small molecule drugs, dyes, guests, and ligands.<sup>39-41</sup>

Manufacturers of *CD* spectropolarimeters have attempted to streamline the acquisition of data by designing auto-samplers or robotic injectors,<sup>42</sup> but they only marginally speed measurement; *CD* "robots" are not true plate readers but automated flow injectors, drawing each sample and injecting it sequentially. Moreover, they can be sensitive to non-aqueous media, malfunctioning when handling common organic solvents.<sup>43</sup>

Why is a HTS-CD spectrometer not currently available? Presumably, this is because multi-well plates are not necessarily optically isotropic. They can be burdened with strain linear birefringence (*LB*), i.e. refractive index variance. Parasitic *LB* has plagued measurements of the *CD* of anisotropic media for decades.<sup>44-50</sup> Moreover, menisci -- non-normal interfaces -- in well plates can introduce Fresnel linear dichroism (*LD*) into the optical system. *LB* and *LD* can be much larger than circular birefringence (*CB* or optical rotation) and *CD*. The minor chiroptical perturbation to the polarization state of light in anisotropic media can be of the same order of

magnitude as parasitic ellipticities from imperfect samples, containers, and optical components.

#### **Experimental Strategy**

#### Instrument Design

*CD* spectropolarimeters typically modulate between left and right circular polarization states by using a photoelastic modulator (PEM) placed before the sample. The differential transmission is recorded. But the measured *CD* can be grossly affected by the admixture of linear polarization in the input light and its coupling to linear anisotropies of the medium (in solid samples) or the container. Multiple photoelastic modulators (PEMs) provide a strategy for separating and eliminating contributions to the change in the polarization of light due to parasitic linear anisotropies that have traditionally given rise to artifactual chiral signatures.<sup>58,59</sup>

Most polarization techniques rely on measuring particular elements of the sample Mueller matrix, the linear operator that contains all of the information about the polarization transforming properties of an analyte.<sup>60</sup> Increasingly complex samples spread the desired optical effects over all 16 elements of the Mueller matrix.<sup>45</sup> In 2011, we introduced a polarimeter using four PEMs (4-PEM spectropolarimeter or simply "4-PEM" in brief) that measures simultaneously all Mueller matrix elements without reconfiguration, thus avoiding errors associated with moving optical components.<sup>58,59</sup> A schematic is shown in Figure 1. This instrument is fast, sensitive, and accurate. It will be used here as a high-speed automated plate reader.



Figure 1. Four photoelastic modulator (4-PEM) spectropolarimeter. A: light source, B: monochromators, C: mirrors, D: linear polarizers, E, F: photoelastic modulators set at  $0^{\circ}$  (E) and  $45^{\circ}$  (F). G: Sample holder, H: photomultiplier tube.

In our instrument, light from a Xe arc lamp coupled to a monochromator (290 nm to 850 nm) was normally incident on the sample. The polarization state generator and the polarizer state analyzer are each composed of a linear polarizer and two PEMs (Hinds Instruments) operating at a different frequencies. The PEMs were set at relative orientations of  $\pm 45^{\circ}$ . The advantage of this setup over other Mueller matrix polarimeters is that measurements are obtained without any moving parts. The transmitted light was detected with a photomultiplier. The 4-PEM measures the 16 parameters of a normalized Mueller matrix at each wavelength.

A 96-well glass bottom plate (In Vitro Scientific, P96-1.5H-N, with a MatTek Corporation top cover glass, PCS10872-1.0) was mounted on two linear actuators (Newport) with a range of only three wells in the vertical and horizontal directions, thus only a  $3\times3$  block of wells could be measured without repositioning the plate. The cover glass was secured with elastic bands because our polarimeter has a horizontal light path. There

Page 2 of 5

#### Chemical System

With the goal of testing the performance of a multiple PEM polarimeter in HTS measurements, we chose a validated *CD* assay for *ee* of  $\alpha$ -chiral amines.<sup>35</sup> Aldehyde **1** reacts with the a chiral amine (4-7) to give a bidentate imine **2**. Fe(II), in turn, binds three equivalents of that imine to give several helical octahedral complexes represented as **3**, whose handedness is



Scheme 1. Two-step assembly that leads to the CD-active  $Fe^{(II)}$  complex used to report the *ee* of chiral amines **4-7**.

dictated by the chirality of the amine (Scheme 1). We determined the enantioenrichment of amine 4-7 by measuring *CD* in the strong metal-to-ligand charge transfer bands that arise upon formation of complex  $3^{.35}$ . This reaction is fast, simple, relies on commercially available reagents and can be analyzed *in situ*.

#### **Results and Discussion**

#### **Mueller Matrix Polarimetry**

A Mueller matrix is a real valued  $4 \times 4$  matrix that represents an optical element or sample. Stokes-Mueller calculus tracks the transformation of the state of polarization of light passing through the 4-PEM and the sample.

The Mueller matrix of an optically active solution,  $(M_{sol})$  in (1), has only two parameters where  $\alpha = CD$ ,  $\beta = CB$ , and  $\alpha^2 + \beta^2 + \gamma^2 = 1$ . It has been shown previously that a Mueller matrix which introduces small, parasitic linear anisotropies (*LB* and *LD* less than ~0.5 rad) and negligible depolarization can be expressed as the explicit matrix ( $M_{anis}$ ) in (1).<sup>61</sup> Here, *LB'* and *LD'* refer to a set of orthogonal axes at ±45° with respect to the instrument coordinate system.

In the absence of linear anisotropies, CD can be taken directly from either the  $M_{03}$  or  $M_{30}$  elements. Commercial CD

**Chemical Science** 

spectrometers typically measure  $M_{03}$  using a single PEM positioned before the sample. However, even small ellipticities introduced by the sample holder will perturb the measured Mueller matrix. To evaluate the Mueller matrix with anisotropy introduced into sample holder before or after the sample, i.e. closer to or further from the light source, we analyzed the

adapted to simple UV-vis HTS. However, there is no direct way to verify the *CD* measurements with a single PEM, while for two (or in this case four) PEMs,  $M_{03}$ ,  $M_{30}$ , and  $M_{33}$  can be monitored simultaneously to increase data reliability. Additionally, two PEMs deliver other elements of the Mueller matrix that can be used to estimate the magnitude of the linear

$$M_{sol} = \begin{bmatrix} 1 & 0 & 0 & \alpha \\ 0 & \gamma & \beta & 0 \\ 0 & -\beta & \gamma & 0 \\ \alpha & 0 & 0 & 1 \end{bmatrix}, \ M_{anis} = \begin{pmatrix} 1 + \frac{1}{2}(LD^{2} + LD^{-2}) & -LD & -LD' & \frac{1}{2}(LBLD' + LB'LD) \\ -LD & 1 + \frac{1}{2}(LD^{2} + LB'^{2}) & \frac{1}{2}(LBLB' + LDLD') & LB' \\ -LD' & \frac{1}{2}(LBLB' + LDLD') & 1 + \frac{1}{2}(LD'^{2} + LB^{2}) & -LB \\ -\frac{1}{2}(LBLD' + LB'LD) & -LB' & LB & 1 - \frac{1}{2}(LB^{2} + LB'^{2}) \end{pmatrix}$$

#### respective matrix products $M_{sol}M_{anis}$ or $M_{anis}M_{sol}$ .

As explained in the supporting information, regardless of the sample position, analysis of the matrix products shows that *CD* can be approximated from the elements of measured Mueller matrix as  $CD = (M_{03}+M_{30})/(1+M_{33})$ . For very small linear anisotropies,  $M_{33}$  approaches 1, and the correction for *CD* approaches the simple average of  $M_{03}$  and  $M_{30}$ . For large linear anisotropies,  $M_{03}$  and  $M_{30}$  will be very different from one another, and the correction fails. Figure 2 illustrates the deviations from the artifact-free *CD* that result from measurement of only  $M_{03}$  or  $M_{30}$  in the presence of modest anisotropy.



Figure 2. CD errors when using only one PEM. A 0.02 M solution of Cu(II) tartrate measured without (artifact-free CD) and with (all other spectra) a birefringent piece of adhesive tape placed after the solution cuvette using the 4-PEM.

We emulated a well-plate measurement of a 0.02 M Cu(II)-*D*-tartrate solution in a quartz cuvette placed just before a transparent birefringent material (a piece of adhesive tape). The complete *M* was measured with the 4-PEM.  $M_{03}$  and  $M_{30}$  by themselves did not match the *CD* measured directly in the glass cuvette without the interfering linearly birefringent tape. But,  $(M_{03}+M_{30})/(1+M_{33})$  matched quite well.

Using the In Vitro Scientific glass bottomed well plates, in practice  $M_{03}$  and  $M_{30}$  were equal and either could be taken as the *CD*. This implies that glass bottomed well plates, at least from this manufacturer, can be used along with a single photoelastic modulator. That is a HTS *CD* instrument could well be fashioned from a commercial device with a vertical optical train and a high speed scanning stage of the type already

anisotropies.<sup>62,63</sup> We caution that polystyrene well plates are unsuitable for the measurement of CD as the perturbing LB is much too large. Such a plate was analyzed with the 4-PEM and the heterogeneities in the optical properties were captured with an imaging Mueller matrix microscope described previously.<sup>64</sup>

#### **Enantiomeric Excess Determination in Well Plates**

Amines 4-7 in Fe complexes in Scheme 1 were analyzed by the 4-PEM. CD spectra were measured (Figure 3) for solutions spanning the enantiomeric composition between 100% *R* configuration and 100% *S* configuration. A calibration curve was built from  $M_{03}$  values (Figure 4) and fit to a line in the case of 4. The  $R^2$  value was 0.998.

While the calibration curve in Figure 4 is a straight line for 4, they are not generally linear because the *CD* represents mixtures of stereoisomers at intermediate concentrations. The curves in the supporting material are generally sigmoidal and fit to  $3^{rd}$  order polynomials. See supporting information.



Figure 3. CD spectra for the assembly with 4.



Figure 4. Calibration curve for the assembly with 4 at 540 nm.

Once the calibration curves that relate *CD* signal intensity to *ee* for each amine were obtained, a number of "unknown" samples were analyzed from new stock solutions of products. The *CD* values were converted to values of *ee* using the calibration curves previously established. The calculated values of *ee* were compared to the actual values of the solutions measured. The *ee* errors ranged between |5-9|%. These are errors in chemistry (weighing reagents and volumes) and are consistent with those found when measuring such samples individually and fall within the range of accuracy that is expected for a HTS method.<sup>28</sup>

#### Conclusions

With a spectropolarimeter carrying more than one photoelastic modulator, CD was measured rapidly in a multi-well plate format. The technology has been tested with an existing sensing ensemble and the results are consistent with previous data. There should be few obstacles to HTS-CD in high quality well plates with glass interfaces.

#### Acknowledgements

AE gratefully acknowledges the financial support from the National Institutes of Health (GM077437) and the Welch Foundation (F-1151). BK thanks the National Science Foundation (DMR-1105000) and SMN is grateful for a National Science Foundation Predoctoral Fellowship (DGE-12342536).

#### Notes and references

<sup>a</sup> Department of Chemistry, The University of Texas at Austin, Austin, Texas, 78712, USA. Email: anslyn@austin.utexas.edu

<sup>b</sup> Department of Chemistry and Molecular Design Institute, New York University, New York, New York 10003, USA. Email: bart.kahr@nyu.edu

Electronic Supplementary Information (ESI) available: details of experimental conditions, CD and *ee* calculations, Mueller matrix data. See DOI: 10.1039/b000000x/

- 1 G. Alves, A. Fortuna, A. Falcao, *Trends Chromatogr.* 2008, 4, 1-10.
- 2 J. P. Stambuli, J. F. Hartwig, Curr. Opin. Chem. Biol. 2003, 7, 420.
- 3 K. D. Shimizu, M. L. Snapper, A. H. Hoveyda, *Chem-Eur J.* 1998, 4, 1885.
- 4 S. H. Shabbir, C. J. Regan, E. V. Anslyn, Natl. Acad. Sci. USA 2009, 106, 10487.
- 5 M. T. Reetz, Angew. Chem. Int. Edit. 2002, 41, 1335.
- 6 J. F. Traverse, M. L. Snapper, Drug Discov. Today. 2002, 7(19), 1002-12.
- 7 C. Jaekel, R. Paciello, Chem. Rev. 2006, 106(7), 2912-42.
- 8 C. Gennari, U. Piarulli, Chem. Rev. 2003, 103(8), 3071-100.
- 9 D. Wahler, J.-L. Reymond, Curr. Opin. Biotechnol. 2001, 12(6), 535-44.
- 10 J. P. Stambuli, J. F. Hartwig, Curr. Opin. Chem. Biol. 2003, 7(3), 420-6.
- 11 J. G. de Vries, L. Lefort, Chem. Eur. J. 2006, 12(18), 4722-34.
- 12 K. Ding, Chem. Commun. 2008, 8, 909-21.
- 13 L. Lefort, J. A. F. Boogers, A. H. M. de Vries, J. G. de Vries, *Top. Catal.* 2006, 40(1-4), 185-91.
- 14 A. J. Minnaard, B. L. Feringa, L. Lefort, J. G. de Vries, Acc. Chem. Res. 2007, 40(12), 1267-77.
- 15 W. Seiche, B. Breit, Catalysts with Chiral Self-assembling Ligands in Phosphorus Ligands in Asymmetric Catalysis, Ed. A. Börner, Wiley-VCH, Weinheim 2008.

- 16 K. Sakthivel, W. Notz, T. Bui, C. F. Barbas, J. Am. Chem. Soc. 2001, 123(22), 5260-7.
- 17 L. Lefort, J. A. F. Boogers, A. H. M. de Vries, J. G. de Vries, Org. Lett. 2004, 6(11), 1733-5.
- 18 B. L. Feringa, Acc. Chem. Res. 2000, 33(6), 346-53.
- 19 O. Pamies, M. Dieguez, Chem. Eur. J. 2008, 14(3), 944-60.
- 20 P. P. Pescarmona, J. C. Van der Waal, I. E. Maxwell, T. Maschmeyer, *Catal. Lett.* 1999, 63(1,2), 1-11.
- 21 R. H. Crabtree, Chem. Commun. 1999, 17, 1611-6.
- 22 D. G. Powers, D. L. Coffen, Drug Discov. Today 1999, 4(8), 377-83.
- 23 R. B. C. Jagt, P. Y. Toullec, E. P. Schudde, J. G. De Vries, B. L. Feringa, A. J. Minnaard, J. Comb. Chem. 2007, 9(3), 407-14.
- 24 T. M. Tarasow, S. L. Tarasow, B. E. Eaton, *Nature* 1997, **389**(6646), 54-7
- 25 M. T. Reetz, K. M. Kuhling, S. Wilensek, H. Husmann, U. W. Hausig, M. Hermes, *Catal. Today* 2001, **67**(4), 389-96.
- 26 J. M. Takacs, J. Han, Org. Lett. 2004, 6(18), 3099-102.
- 27 S. Bromidge, P. C. Wilson, A. Whiting, *Tetrahedron Lett.* 1998, 39(48), 8905-8.
- 28 D. Leung, E. V. Anslyn, J. Am. Chem. Soc. 2008, 130(37), 12328-12333.
- 29 C. Wolf, K. W. Bentley, Chem. Soc. Rev. 2013.
- 30 M. W. Ghosn, C. Wolf, Tetrahedron, 2010, 66(23), 3989-3994.
- 31 L. You, J. S. Berman, E. V. Anslyn, Nat. Chem. 2011, 3(12), 943-948.
- 32 M. W. Ghosn, C. Wolf, J. Am. Chem. Soc. 2009, 131(45), 16360-16361.
- 33 D. P. Iwaniuk, K. W. Bentley, C. Wolf, *Chirality*, 2012, 24(7), 584.
- 34 L. A. Joyce, M. S. Maynor, J. M. Dragna, G. M. da Cruz, V. M. Lynch, J. W. Canary, E. V. Anslyn, J. Am. Chem. Soc. 2011, 133(34), 13746-13752.
- 35 J. M. Dragna, G. Pescitelli, L. Tran, V. M. Lynch, E. V. Anslyn, L. Di Bari, J. Am. Chem. Soc. 2012, 134(9), 4398-4407.
- 36 P. Metola, E. V. Anslyn, T. D. James, S. D. Bull, Chem. Sci. 2011, 3(1), 156-161.
- 37 M. W. Ghosn, C. Wolf, *Tetrahedron*, 2011, **67**(36), 6799-6803.
- 38 D. P. Iwaniuk, C. Wolf, Org. Lett. 2011, 13(10), 2602-2605.
- 39 N. Berova, K. Nakanishi, R. W. Woody, *Circular dichroism:* principles and applications. New York: Wiley-VCH, 2000.
- 40 N. Purdie, H. G. Brittain, Analytical applications of circular dichroism. Elsevier, 1993.
- 41 M. Eriksson, B. Nordén, Meth. Enzymol. 2001, 340, 68-98.
- 42 Jasco ASU-800 Autosampler
- 43 In our experience, the tubing used is not compatible with acetonitrile or halogenated solvents, and thus the robotic handler rapidly plugs with the use of these solvents. Additionally, the threshold for concentration tolerated by the plastic tubes before clogging occurs is low enough that it is another cause of concern.
- 44 K. W. Hipps, G. A. Crosby, J. Phys. Chem. 1979, 83, 555-562.
- 45 J. Schellman, H. P. Jensen, Chem. Rev. 1987, 87, 1359-1399.
- 46 J. Michl, E. W. Thulstrup, Spectroscopy with Polarized Light, VCH: New York, 1986.
- 47 R. Kuroda, T. Harada, Y. Shindo, *Rev. Sci. Instrum.* 2001, 72, 3802-3810.
- 48 M. F. Maestre, G. C. Salzman, R. A. Tobey, C. Bustamante, *Biochemistry*, 1985, 24, 5152-5157.
- 49 J. R. L. Moxon, A. R. Renshaw, J. Phys.: Condens. Matter, 1990, 2, 6807-6836.
- 50 M. Kremers, H. Meekes, J. Phys. D: Appl. Phys. 1995, 28, 1212-1224.
- 51 F. Arago, Mem. Inst. Fr. 1811, 1, 93-134.
- 52 J. K. O'Loane, Chem. Rev. 1980, 80, 41-61.
- 53 J. Applequist, Am. Sci. 1987, 75, 59-68.
- 54 W. Kaminsky, Rep. Prog. Phys. 2000, 63, 1575-1640.

**Chemical Science** 

- 55 B. Kahr, O. Arteaga, Chemphyschem, 2012, 16, 79-88.
- 56 J. Kobayashi, T. Asahi, S. Takahashi, A. M. Glazer, J. Appl. Crystallogr. 1988, 21, 479-484.
- 57 O. Arteaga, A. Canillas, G. E. Jellison, Jr. Appl. Opt. 2009, 48, 5307-5317.
- 58 O. Arteaga, J. Freudenghtal, B. Wang, S. Nichols, B. Kahr, *Chim. Oggi.* 2012, **30**, 31-36.
- 59 O. Arteaga, J. Freudenthal, B. Wang, B. Kahr, *Appl. Opt.* 2012, **51**, 6805-6817.
- 60 D. Goldstein, D. H. Goldstein. *Polarized Light, revised and expanded.* Vol. 83. CRC Press, 2003.
- 61 O. Arteaga, Ph.D. Thesis, University of Barcelona, 2011.
- 62 G. E. Jellison, Jr.; F. A. Modine, Appl. Opt. 1997, 36, 8184-8189.
- 63 G. E. Jellison, Jr.; F. A. Modine, Appl. Opt. 1997, 36, 8190-8198.
- 64 A. G. Shtukenberg, J. Freudenthal, B. Kahr, J. Am. Chem. Soc. 2010, 132, 9341-9349.