

RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Graphic Abstract

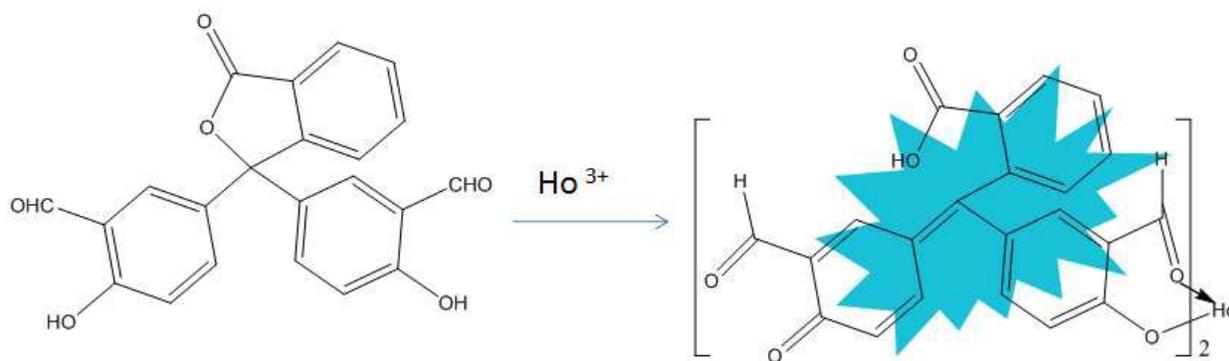
The Title:

A high selective and sensitive turn on fluorescent probe for detection of holmium ion and its bioimaging

List of Authors:

Yanqing Guo,^a Fangjun Huo,^b Caixia Yin^{*c}

The statement:



The phenolphthalein aldehyde was synthesized and used to an turn-on fluorescent probe for the detection of holmium ion (Ho^{3+}), which is one of lanthanide ions in HEPES:DMF = 1:1 (V/V pH=7.4) solution with an excellent selectivity and sensitivity for Ho^{3+} over other metal ions.

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

A high selective and sensitive turn on fluorescent probe for detection of holmium ion and its bioimaging

Yanqing Guo,^a Fangjun Huo,^b Caixia Yin,^{*c} Jin Kang,^c JianFang Li^b

Received (in XXX, XXX) Xth XXXXXXXXX 200X, Accepted Xth XXXXXXXXX 200X

DOI: 10.1039/b000000x

The phenolphthalein aldehyde was synthesized and used to a turn on fluorescent probe for the detection of holmium ion (Ho³⁺), which is one of lanthanide ions in HEPES:DMF = 1:1 (V/V pH=7.4) solution with an excellent selectivity and sensitivity for Ho³⁺ over other metal ions.

Rare earth (RE) is an important non-renewable resources. With the development of science, it was made various demands for materials. Application of rare earth elements, from their use of homogeneous property, extends to the use of single rare earth speciality, and has gone deep into all areas of modern technology. Because rare earth element (REE) has the special properties to promote crop production and improve crop quality, the area of application of compound fertilizer of rare earth element is more and more widely. Generally, low concentrations of REEs is present in soil, plant, water, and atmosphere, but REEs can accumulate in such environments following anthropogenic inputs because of the low mobility of these elements¹⁻³. As the products containing rare earths continuously into the environment, long-term risks to plants, animals, the environment and human health has attracted widespread attention at home and abroad scientific community.

Holmium is one of lanthanide ions, however, it is 20 times more abundant than silver⁴. There is a growing trend in the uses of holmium element, due to the fact that it is studied to produce catalysts, polish glass or use for biological applications⁵⁻⁷. Lanthanides do not partake in any metabolic process. However, due to the similarity of lanthanides ionic radii to calcium ion, possessing a higher charge, they exhibit a high affinity for the Ca²⁺ sites on biological molecules and a stronger binding to water molecules⁷⁻¹⁰. The bioaccumulation of the holmium ions in the body can be a threat to the liver. Nowadays, it is known that holmium causes damages to the cell membranes of the water animals with negative effects on the reproduction and the nervous systems^{7,8,10-12}. This makes the detections of Ho greatly important. Conventional methods of Ho ion analysis have included neutron activation analysis, ICP-MS, ICP-AES and isotope dilution mass spectrometry¹³⁻¹⁸. In recent years, it was reported a number of opcode and membrane sensor for selective determination of Ho³⁺^{4,10,18-25}. However, the fluorescent probe to detect Ho³⁺ has not been reported.

Herein, we report a novel fluorescent probe based on phenolphthalein aldehyde to selective detect Ho³⁺ in medium of near neutral pH value. And the recognition ability of probe for Ho³⁺ was studied. The result showed that this probe works well at physiological pH and has a high selectivity and sensitivity for Ho³⁺ over other metal ions. These desirable attributes render the sensor suitable for detection of Ho³⁺. Furthermore, the system is used to bioimaging.

The synthesis of probe is summarized in Scheme 1. To a stirred TFA (10 mL) on ice was added phenolphthalein (636.64 mg, 2 mmol) and hexamethylenetetraamine (403.7 mg, 2.88 mmol). The solution was allowed to warm to room temperature, and then it was refluxed for 8h. The excess TFA was removed under vacuum. 30 mL water was then added to the remaining solution, and the mixture was warmed at 60 °C whilst stirring for another 30 min. Upon cooling on ice, a pale-yellow solid were precipitated out from the solution which were collected by filtration. Then the crude product was purified by chromatography on a silica gel column, and give the desired product as a white solid (57 % yield). ¹H NMR (300 MHz, 25 °C, DMSO-*d*₆): δ 11.04 (bs, 2H), 10.22 (bs, 2H), 7.94 (d, 1H, J = 7.2 Hz), 7.84 (m, 2H, J = 20.7 Hz), 7.68 (q, 1H, J = 14.4Hz), 7.51 (s, 2H), 7.43 (d, 2H, J = 8.7Hz), 7.02 (d, 2H, J = 8.7Hz); ¹³C NMR (75 MHz, DMSO-*d*₆): δ 189.9, 168.1, 160.3, 150.5, 134.6, 134.2, 130.7, 129.4, 125.7, 125.1, 123.6, 121.1, 117.5, 89.3; ESI-MS [M-H]⁻ 373.25; Elemental analysis (calcd %) for C₂₂H₁₄O₆: C, 70.59; H, 3.77; Found: C, 70.38; H, 3.89. (Fig. S1, ESI[†])

The recognition ability of the probe was investigated by the fluorescence spectra. The selectivity towards the metal ions is one of the most important criteria for probe design. The effect of a wide range of environmentally and physiologically active metal ions was investigated for probe using the fluorescence spectra of solutions containing probe and the metal ions (100 equiv.) in the HEPES:DMF = 1:1 (V/V pH=7.4) solution. As shown in Fig. 1, whereas metal ions such as Cu²⁺, Cr²⁺, Gd³⁺, Ni²⁺, Co²⁺, VO²⁺, La³⁺, Pb²⁺, Zr⁴⁺, Mn²⁺, Zn²⁺, Ru³⁺, Yb³⁺, Cd²⁺, Sm³⁺, Bi³⁺, Ce⁴⁺, Er³⁺, Ca²⁺, Mg²⁺, Sn²⁺, Fe³⁺, Dy³⁺, Pr³⁺, Tb³⁺, Pm³⁺, Lu³⁺, Tm³⁺, Nd³⁺ and Eu³⁺ do not result in any apparent changes in fluorescence intensity, there is notable change when Ho³⁺ is involved (λ_{ex} = 365 nm) (Fig. S2, ESI[†]). Fig. 1 shows that fluorescence optical density at 480 nm when various metal ions are added and fluorescence color change.

The probe could specifically recognize Ho³⁺ was explained as following: peripheral electron shell configuration of Ho³⁺ is 4f¹⁰, which results in the abundant level structures, as well as some high metastable state. The nice isolation between energy levels prompts Ho³⁺ having abundant electron transition²⁶. Its peculiar energy level structure inducing the bigger gap between 3k8, 5s2 and adjacent low level makes non-radiative relaxation be not easy to happen. What's more, absorption peak of Ho³⁺ is very rich, which resulted from two way: one is low energy photon is easy to excite Ho³⁺ ions, another is other rare earth with sensitized effect realizes inversion of high level particle number to obtain the strong blue-green glow^{27, 28}. Green glow is corresponding to the energy level transition of ⁵S₂ (⁵F₄)—⁵I₈ of Ho³⁺ spectral term. The fluorescence of Dy³⁺ is easy to be quenched by high-energy

vibration of O–H bond in water molecules²⁹.

A titration experiment of the probe for Ho³⁺ was also carried out on fluorescence spectrophotometer. The probe displayed no emission at 480 nm. With addition and gradual increasing amount of Ho³⁺ to the HEPES/DMF = 1:1 (V/V, pH=7.4) solution containing probe, a significant increase in fluorescence intensity at 480 nm was observed with an excitation wavelength 365 nm. When 260 μM Ho³⁺ was added to the solution of the probe (20 μM), a more than 25-fold increase in fluorescence intensity at 526 nm was observed (Fig. 2) with a change of fluorescence quantum yield from $\phi_{\text{probe}} = 0.001$ to $\phi_{\text{probe-Ho}^{3+}} = 0.256$. In addition, with increasing Cu²⁺ concentration, the absorption peaks at 330 gradually decreased and 380 nm increased. One isosbestic points was noted at 348 nm indicating the formation of a new species (Fig. S3, ESI†). According to linear Benesie Hildebrand expression, the measured intensity $[1/(A - A_0)]$ at 380 nm varied as a function of $1/[\text{Ho}^{3+}]$ in a linear relationship ($R = 0.99797$) (Fig. S4, ESI†), indicates formation of 1:1 stoichiometry between Ho³⁺ and probe. The association constant of probe with Ho³⁺ was calculated to be $1.08 \times 10^7 \text{ M}^{-1}$.

The effect of pH on the fluorescence properties of the system was investigated for pH values of 2.0–13.0. The results are shown in Fig. 3. Over the pH 2.0–8.0, the free probe exhibits no fluorescence in the HEPES/DMF = 1:1 (V/V, pH=7.4) solution. It is obvious that the fluorescent signal of the probe increases significantly with increasing pH from 8.0 to 12.0. However, the intensity of the probe exhibits a non-ignorable decrease at pH 13.0. Meanwhile, the fluorescence signal for the probe solution containing Ho³⁺ increased sharply from pH 4.0 to 8.0, and then decreased from pH 8.0 to 11.0. The fluorescence intensity of the probe solution containing Ho³⁺ has a little increases at pH 12.0 compared with at pH 11.0. Holmium exists in the form of Ho(OH)₃ precipitate due to the strong basicity of pH 13.0 which result in the fluorescence decrease of probe with addition of Ho³⁺. Therefore, considering pH and fluorescence intensity, a pH of 7.4 was selected for the analytical system.

The effect of reaction time on the fluorescence emission of the system was also studied in the presence of 10 equiv. of Ho³⁺ and the results are shown in Fig. 4. The fluorescence signal for the system increases gradually with increasing reaction time, and the addition of Ho³⁺, under the selected reaction conditions, was rapid. Therefore, a 4 min reaction time was selected for the following experiments.

To investigate the detection limit of probe for Ho³⁺, probe (20 μM) was treated with various concentrations of Ho³⁺ (0–260 μM) and the relative emission intensities at 480 nm were plotted as a function of the Ho³⁺ concentration (Fig. S5, ESI†). The fluorescence intensity of probe is linearly proportional to the Ho³⁺ concentrations, and the detection limit, based on the definition by IUPAC ($C_{\text{DL}} = 3 \text{ Sb/m}$)³⁰, was found to be 0.055 μM indicating this probe is sensitive enough to monitor Ho³⁺ compared with other method (Table 1)^{18,19,23,25}.

The reaction mechanism of the present system was also studied. We presumed that the fluorescence increase could be attributed to ring-open after the coordination of probe with Ho³⁺. Namely, because of the probe with hydroxyl and aldehyde groups (their O atoms as hard bases), easily coordinating with Ln ions as hard acids and resulting in the ring open, increased the conjugate structure of intramolecular, which lead to the system became strong fluorescence. The identification of coordination product in the ESI-MS analysis made it possible to propose the signaling mechanism: a peak at $m/z = 984.00$, corresponding to $[\text{probe-Ho} + 4\text{H}_2\text{O} + \text{H}]^+$, is clearly observed (Fig. S6, ESI†). The proposed

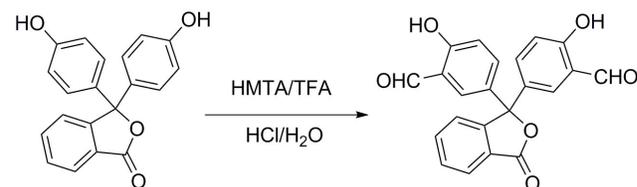
detection mechanism and the structures of the probes, both with and without the addition of Ho³⁺, are shown in Scheme 2.

The ability of probe reacting with Ho³⁺ within living cells was also evaluated by laser confocal fluorescence imaging using a Leica TCS SP5 laser scanning microscope. The optical window at the yellow channel (450–550 nm) was chosen as a signal output. As shown in Fig. 5a, HepG2 cells incubated with 20 μM probe for 30 min at 37 °C and washed 3 times with PBS showed no fluorescence. In a further experiment it was found that HepG2 cells displayed green fluorescence when the cells were first incubated with 20 μM of probe for 30 min at 37 °C and washed 3 times with PBS, then incubated with 100 μM HoCl₃ and washed 3 times with PBS (Fig. 5b). These cell experiments show the good cell-membrane permeability of probe, and it can thus be used to mark Ho³⁺ within living cells.

In summary, we have developed a fluorescent probe for the detection of Ho³⁺ by turn-on fluorescence over other analytes in aqueous solution. This probe is based on a phenolphthalein derivative which has a high selectivity and sensitivity for Ho³⁺ over other analytes in HEPES: DMF = 1:1 (V/V pH=7.4). Furthermore, the system is used to bioimaging.

The work was supported by the National Natural Science Foundation of China (No. 21102086, 21472118), the Shanxi Province Science Foundation for Youths (Nos. 2012021009-4 and 2013011011-1), the Shanxi Province Foundation for Returnee (No. 2012-007), the Taiyuan Technology Star Special (No. 12024703), the Program for the Top Young and Middle-aged Innovative Talents of Higher Learning Institutions of Shanxi (TYMIT, No. 2013802), Talents Support Program of Shanxi Province (No. 2014401) and CAS Key Laboratory of Analytical Chemistry for Living Biosystems Open Foundation (No. ACL201304).

Inserting Graphics



Scheme 1 Synthesis of probe..

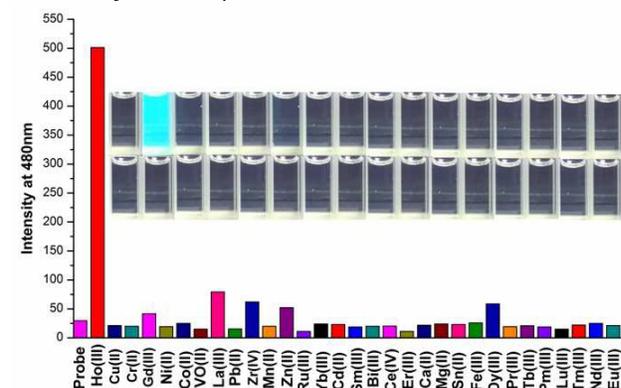


Fig. 1. Optical density two-dimensional graph of the probe at 480 nm, respectively upon the addition of several metal ions (top: from left to right, probe, Ho³⁺, Cu²⁺, Cr²⁺, Gd³⁺, Ni²⁺, Co²⁺, VO²⁺, La³⁺, Pb²⁺, Zr⁴⁺, Mn²⁺, Zn²⁺, Ru³⁺, Yb³⁺, Cd²⁺; bottom: from right to left, Sm³⁺, Bi³⁺, Ce⁴⁺, Er³⁺,

Ca²⁺, Mg²⁺, Sn²⁺, Fe³⁺, Dy³⁺, Pr³⁺, Tb³⁺, Pm³⁺, Lu³⁺, Tm³⁺, Nd³⁺ and Eu³⁺); Inset: a color change photograph for Ho³⁺ and the other metal ions under illumination with a 365 nm UV lamp.

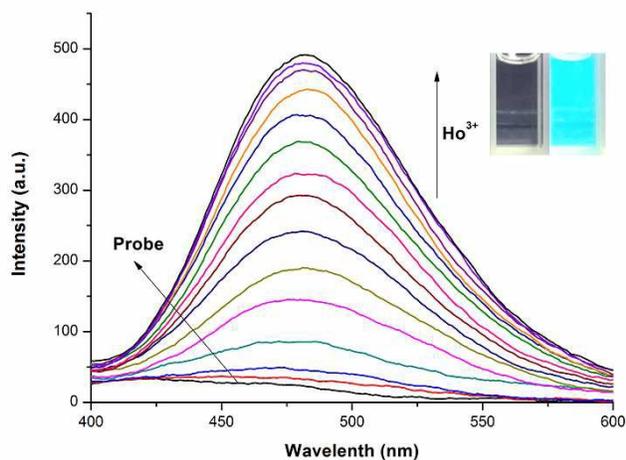


Fig. 2. Fluorescence spectral changes of probe (20 μM) in 10 mmol/L HEPES buffer /DMF (v/v=1:1, pH 7.4) (λ_{ex} = 365 nm, slit: 5 nm/5 nm) upon addition of Ho³⁺; Ho³⁺ was added gradually with [Ho³⁺] = 0–260 μM; Inset: a color change photograph for Ho³⁺ under illumination with a 365 nm UV lamp.

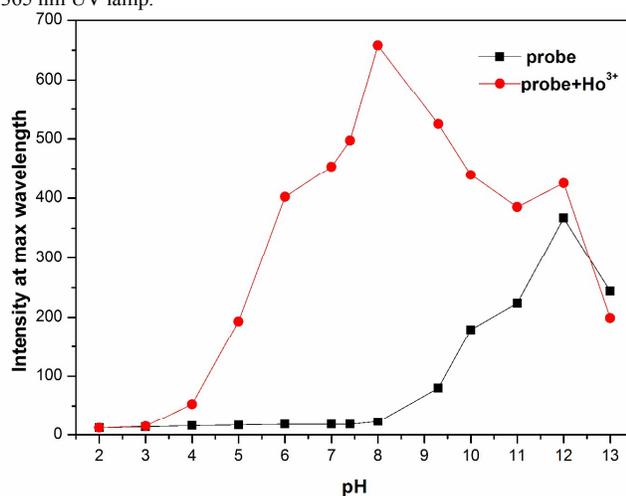


Fig. 3. Effect of pH on the fluorescence intensity of the probe and probe+Ho³⁺ (260 μM).

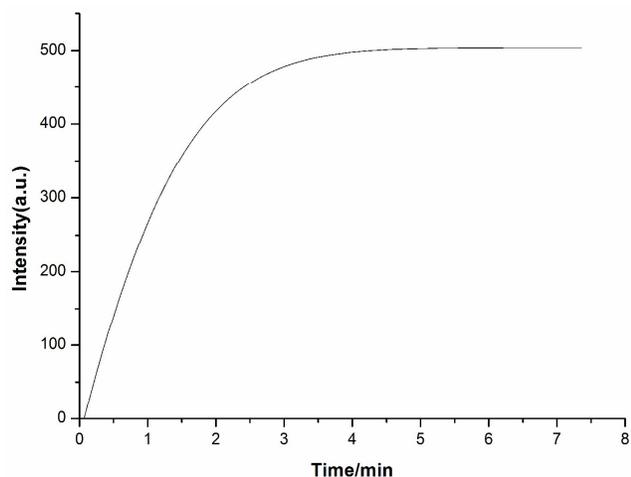
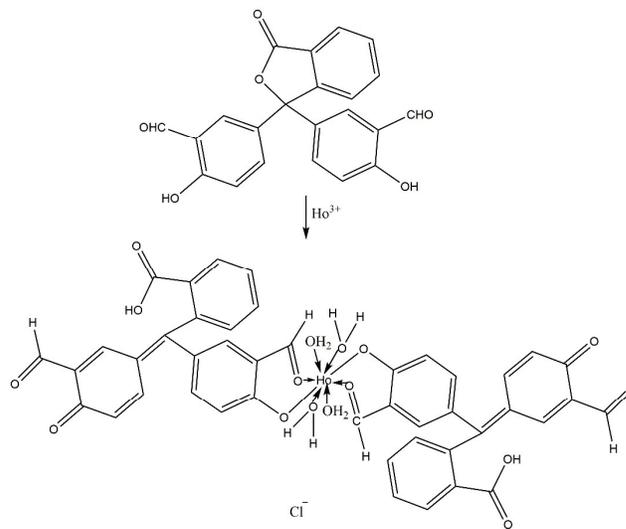


Fig. 4. Reaction time profiles of probe (20 μM) and Ho³⁺ (260 μM) Table 1. A comparison table about the detection methods for Ho³⁺.

	method	analyte	detection limit
ref (18)	membrane sensor	Ho ³⁺	0.75 μM
ref (19)	Potentiometric Membrane Sensor	Ho ³⁺	0.63 μM
ref (23)	polymeric membrane sensor	Ho ³⁺	5 μM
Ref (25)	membrane sensor	Ho ³⁺	0.62 μM
this work	Fluorescent probe	Ho ³⁺	0.055 μM



Scheme 2. The mechanism of chemosensor.

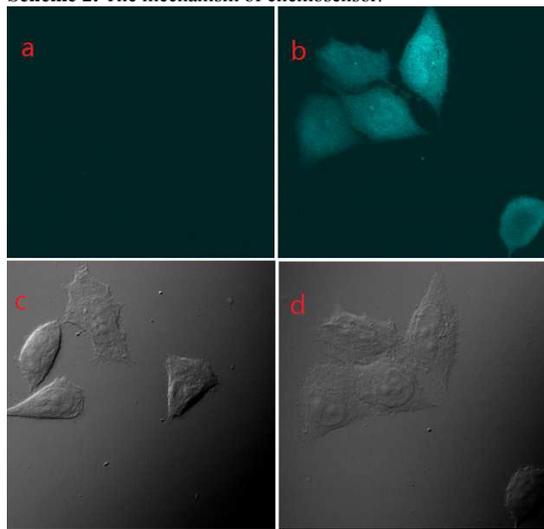


Fig. 5. Confocal fluorescence images in HepG2 cells. (a) Fluorescence image of HepG2 cells with adding probe (20 μM) and its bright field image (c); (b) Fluorescence image of HepG2 cells incubated with 20 μM probe for 30 min at 37 °C, then incubated with 100 μM HoCl₃ for 30 min at 37 °C and its bright field image (d)

Notes and references

- ³⁰ ^a College of Chemistry and Chemical Engineering, Jinzhong University, Yuci 030619, China.
^b Research Institute of Applied Chemistry, Shanxi University, Taiyuan, 030006, China.
^c Institute of Molecular Science, Shanxi University, Taiyuan 030006, China. Fax: +86 351 7011022; Tel: +86 351 7011022.
³⁵ E-mail: yincx@sxu.edu.cn

Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/b000000x/

- 5 1. X. D. Cao, X. R. Wang and G. W. Zhao, *Chemosphere*, 2000, **40**, 23-28.
2. L. D. Aquino, M. Morgana, M. A. Carboni, M. Staiano, M. V. Antisari, M. Re, M. Lorito, F. Vinale, K. M. Abadi and S. L. Woo, *Soil Biol. Biochem.*, 2009, **41**, 2406-2413.
3. S. Zhang, X. Shan, *Environ. Pollut.*, 2001, **112**, 395-405.
- 10 4. H. A. Zamani, A. Zanganeh-Asadabadi, M. Rohani, M. S. Zabihi, J. Fadaee, M. R. Ganjalic, F. Faridbodc and S. Meghdadi, *Mater. Sci. Eng. C*, 2013, **33**, 984-988.
5. J. Kabalin, P. Gilling and M. Fraundorfer, *J. Clin. Laser Med. Surg.*, 1998, **16**, 21-27.
- 15 6. O. R. Kirk and F. D. Othmer, 'Encyclopedia of Chemical Technology', Wiley, New York, 1982, **19**, 851.
7. V. S. Sastri, J. C. G. Bunzli, V. R. Rao, G. V. S. Rayudu and J. R. Perumareddi, Modern Aspects of Rare Earths and Their Complexes, Elsevier publication, Elsevier, Amsterdam, 2003.
- 20 8. S. P. Fricker, *Chem. Soc. Rev.*, 2006, **35**, 524-533.
9. K. Wang, Y. Cheng, X. Yang and R. Li, *Met. Ions Biol. Syst.*, 2003, **40**, 707-751.
10. F. Faridbod, M. R. Ganjali, B. Larijani, M. Hosseini and P. Norouzi, *Mater. Sci. Eng. C*, 2010, **30**, 555-560.
- 25 11. M. R. Ganjalialia, M. Hosseini, A. Karimi, H. Haji-Hashemi, M. Salavati-Niasarid and P. Norouzi, *Spectrochim. Acta Part A*, 2014, **121**, 224-229.
12. O. Vicente, A. Padro', L. Martinez, R. Olsina and E. Marchevsky, *Spectrochim. Acta Part B*, 1998, **53**, 1281-1287.
- 30 13. R. S. Houk, V. A. Fassel, G. D. Reach and H. J. Svec, *Anal. Chem.*, 1980, **52**, 2283-2289.
14. R. Al-Merey and H. J. M. Bowen, *J. Radioanal. Nucl. Chem.*, 1991, **153**, 221-234.
15. M. Anbu, T. P. Rao, C. S. P. Iyer, and A. D. Damodaran, *Chem. Anal.*, 1996, **41**, 781-785.
- 35 16. N. X. Wang, W. Jiang, Z. K. Si and Z. Qi, *Mikrochim. Acta*, 1997, **126**, 251-255.
17. J. Li, S. Liu, X. Mao, P. Gao and Z. Yan, *J. Electroanal. Chem.*, 2004, **561**, 137-142.
- 40 18. M. R. Ganjali, H. Shams, F. Faridbod, L. Hajiaghababaei and P. Norouzi, *Mater. Sci. Eng. C*, 2009, **29**, 1380-1386.
19. M. R. Ganjali, R. Nematii, F. Faridbod, P. Norouzi and F. Darviche, *Int. J. Electrochem. Sci.*, 2008, **3**, 1288-1298.
20. M. R. Ganjali, P. Norouzi, M. Adib and A. Ahmadalinezhad, *Anal. Lett.*, 2006, **39**, 1075-1086.
- 45 21. M. R. Ganjali, S. Rasoolipour, M. Rezapour, P. Norouzi, M. Amirnasrb and S. Meghdadi, *J. Braz. Chem. Soc.*, 2006, **17**, 1211-1216.
22. H. A. Zamani, M. R. Ganjali, P. Norouzi and S. Meghdadi, *J. Appl. Electrochem.*, 2007, **37**, 853-859.
- 50 23. M. R. Ganjali, S. Rasoolipoura, M. Rezapoura, P. Norouzi, M. Amirnasr and S. Meghdadi, *Sens. Actuators B*, 2006, **119**, 89-93.
24. H. A. Zamani, R. Fatemeh, N. R. Fatemeh, A. Ali, I. Alihossien, R. G. Mohammad, F. Farnoush and S. N. Masoud, *Chin. J. Chem.*, 2011, **29**, 1523-1528.
- 55 25. H. Zamani, *Chinese Chem. Lett.*, 2011, **22**, 201-204.
26. Z. M. Wang, D. R. Yuan, Y. S. Yin, G. Su, *Opt. Mater.*, 2007, **29**, 663-666.
27. W. Li, Z. W. Liu, S. G. Xiao, *Nat. Sci. J. Xiangtan Univ.*, 2002, **24**, 29-32.
- 60 28. S. H. Sai, C. M. Nie, X. M. He, *J. Univ. South China (Sci. & Tech.)*, 2007, **21**, 42-45.
29. N. Zhang, w. Z. Shi, S. H. Tang, *Henan Sci.*, 2003, **21**, 408-410.
30. Y. B. Ding, X. Li, T. Li, W. H. Zhu and Y. S. Xie, *J. Org. Chem.*, 2013, 65 **78**, 5328-5338.