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34 **Abbreviation**

35 **D1:** NaOH delignification

36 **D2:** NaClO₂ delignification

37 **O1:** TEMPO mediated oxidation with NaClO₂ at pH 7

38 **O2:** TEMPO mediated oxidation with NaClO at pH 10-11

39 **HPH:** Disintegration using a high pressure homogenizer (10 passes at 600 Bar)

40 **HSB:** Disintegration using a high speed blender for 20 min

41

42 **1. Introduction**

43 Nanofibrillated cellulose (NFC) refers to nanosized cellulose fibrils formed by long, flexible
44 and entangled cellulose nanofibrils and is composed of bundles of elementary fibrils (or
45 cellulose microfibrils) separated by less-ordered regions. Depending on the plant species and
46 the mode of preparation, the lateral dimensions of the nanofibrils are of the order of 10 to 100
47 nm with lengths in the micrometer scale. This class of natural nanofibrils constitutes a real
48 breakthrough in cellulose-based materials and has become a topic of great interest in the last
49 decade¹. Their nanoscale dimensions, biodegradable character, cost effectiveness, high aspect
50 ratio, light weight and sustainability constitute an impetus for this increasing interest. All of
51 these attributes make nanosized cellulose very attractive for a broad range of applications
52 within the field of innovative materials^{2,3}. Furthermore, according to the current knowledge,
53 nanocellulose is classified as a non-toxic material⁴, completely biodegradable and without
54 adverse effects on the health or the environment. These benefits facilitate the use of
55 nanocellulose and eliminate safety concerns, commonly encountered, for mineral and carbon
56 nanofillers.

57 Given its exceptional high stiffness and strength along with its high capacity to build up a
58 rigid entangled network, NFC has gained considerable attention as one of the most promising
59 reinforcements in the realm of sustainable nanofillers with a broad range of potential
60 applications, such as for creating low-weight polymer-based composites^{5,6,7} strength
61 additives for paper^{8,9}, barrier packagings¹⁰ and adsorbent products¹¹.

62 NFC is produced by delaminating cellulosic fibers under an intense mechanical shearing
63 action in order to break up the cell walls and release the nanosized fibrils. In addition to the
64 high pressure homogenization (HPH) and micro-fluidization, which are the main methods
65 currently used for the production of NFC, other means of generating microfibrils were

66 reported in the literature such as grinding and ultrasound-assisted fibrillation. However,
67 irrespective of the disintegration process adopted, the production of NFC faces two major
68 problems, namely (1) the clogging of the pulp, when the pulp is pumped through high
69 pressure fluidizers/homogenizers, and (2) the high energy consumption needed for the
70 efficient delamination of the cell wall via multiple passes through the homogenizer. This
71 high-energy input is necessary in order to overcome the strong hydrogen interactions among
72 neighbouring microfibrils. Values ranging from 20 to 50 KWh/Kg have been reported¹².
73 Therefore, one of the most important challenges associated with the production of NFCs on
74 an industrial scale is to decrease the energy demand and facilitate the overall process.
75 Pretreatment is sometimes used to address this problem. Enzymatic, mechanical or chemical
76 pretreatment has shown to heavily decrease the energy demand¹³. This latter approach turned
77 out to be one of the most efficient processes to facilitate the break up of the fibre network and
78 release the microfibrils through an electrostatic repulsion and osmotic effect¹⁴. In this context,
79 the TEMPO-mediated oxidation is the prevalent method to generate carboxylic groups in a
80 controlled way. An obvious correlation between the carboxyl content and the ease of
81 fibrillation was pointed out in several publications¹⁵. Furthermore, the chemical composition
82 of the cellulose source, namely in terms of the hemicellulose content, was found to play a key
83 role in the efficiency of the nanofibrillation process. The higher the hemicellulose content, the
84 easier is the nanofibrillation aptitude of the fibres¹⁶.

85 Although woody fibres remain the main source for the production of NFC, in practice, any
86 resource of cellulose fibres could also be used. Specifically, agricultural crops and by-
87 products, such as wheat straw, rice straw, rapeseed and corn stalks are the most abundant
88 resources on earth and are an underutilized source of cellulose that are generally allowed to
89 decompose in the fields or are burned for energy production. This class of biomass has
90 received increasing attention in recent years as an alternative resource for the extraction of
91 cellulose fibers. Several studies have been concerned with the extraction of NFC from
92 agricultural crops^{17,18,19} and all adopted a high consuming energy device, making NFC costly
93 to produce. In comparison with wood, agricultural crops have shorter growth cycles, do not
94 compete with the supply of wood and have a lower lignin content, making the delignification
95 process easier. Triticale (*Triticosecale* Wittmack) which is a hybrid crop developed by
96 crossing wheat (*triticum*) and rye (*secale*) is widely used as an industrial crop because of the
97 higher yield in grain and biomass (straw), compared with the other cereal crops. As such, the
98 utilization of triticale can increase the available biomass for industrial use without increasing
99 competition with food production for agricultural land.

100 In the present work triticale straws were used as a starting material to produce NFC, adopting
101 high-pressure homogenization and conventional high-speed blender for the disintegration
102 process. The main emphasis is to highlight how the delignification process and the fibre
103 activation can affect the ease of the nanofibrillation process. Particularly, it will be shown that
104 under specific conditions of the delignification process and fibres pre-treatment, the
105 nanofibrillation process can be implemented by simply using a conventional high-speed
106 blender.

107 **2. Experimental section**

108 *2.1 Samples*

109 Triticale straws were harvested at maturity by the end of June. After further drying, the straws
110 were ground to a coarse powder and crude fibres were Soxhlet extracted for 12 h , using first a
111 solvent mixture composed of toluene/ethanol (60/40 v/v) and then hot water (70°C) for 1 h to
112 remove pectin and sand.

113 The pulping procedure for the crude fibres was carried out as follows:

114 *2.2.1 Delignification processes*

115 *Soda pulping procedure (designated here as D₁)*

116 The extracted biomass was added to water (solid content 10 wt%) and then pulped with a 5
117 wt% NaOH solution for 2 h at 70-80 °C under mechanical stirring. This treatment was repeated
118 three times until the fibres were well individualized. The ensuing fibres were subsequently
119 filtered and rinsed with distilled water and twice bleached with NaClO₂ to remove the residual
120 lignin.

121 *NaClO₂/Acetic acid pulping procedure (designated here as D₂)*

122 The NaClO₂/AA pulping process was carried out as follows: Five grams of dry soxhlet
123 extracted biomass were added to water and mixed to form suspension at a solid content 10
124 wt%. Then, 0.5 g of sodium chlorite (NaClO₂) and 0.5 ml of acetic acid per gram of dry
125 biomass were added, and the suspension was kept under mechanical stirring at a temperature of
126 70 °C for 6 h without removal of any liquor. Fresh charges of sodium chlorite and acetic acid
127 were added to the reaction every 1.5 h for up to 6 h.

128 The pulp yield is calculated through equation 1:

$$129 \quad \text{Pulp Yield \%} = \frac{[m_w(1 - MC/100)]}{m_d} \times 100 \quad (1)$$

130 where, m_w , m_d and MC are the weight of the wet biomass recovered, the weight of the dry
131 sample used and MC the moisture content of the recovered solids, respectively.

132 2.2.2 Bleaching procedure

133 The bleaching treatment was carried out at 70 °C for 1 h at pH 4.8. The solution was composed
134 of equal parts of aqueous chlorite (1.7 wt% NaClO₂ in water) and an acetate buffer (27 g
135 NaOH and 75 mL glacial acetic acid diluted to 1 L of distilled water).

136 2.3 Chemical composition

137 The determination of the basic chemical composition was conducted following TAPPI standard
138 protocols. (TAPPI T 257 cm-02). Samples were first submitted to Soxhlet extraction with
139 ethanol/toluene and water. Then the chemical contents were determined using the following
140 methods, Ash (Tappi T 211 om-93) extractive (Tappi T264 om-07), Klason lignin (Tappi T222
141 om-88), and hemicelluloses (Tappi T249-cm-85).

142 2.4 Fibre length measurements

143 Fibre length, width and curl index of the pulps and the length and width of the particles were
144 measured by image analysis using a MorFi Lab equipment.

145 2.5 TEMPO-mediated oxidation

146 The TEMPO-mediated oxidation was carried out at pH 7 and 10, using NaClO₂ and NaClO as
147 oxidizing agents, respectively. These two methods denominated TEMPO-NaClO-NaClO₂ and
148 TEMPO-NaBr-NaClO were implemented, as described below:

149 *TEMPO-NaClO-NaClO₂ (O1): Cellulose fibres (2 g) were dispersed in a 0.05 M sodium
150 phosphate buffer (200 mL, pH 7) solution, containing TEMPO (40 mg). Sodium chlorite and
151 the 2 M (1.5 g) sodium hypochlorite solution 12° (4 mL) were added to the flask, which was
152 stoppered and stirred at 500 rpm and 60 °C for 12 h. The oxidation was stopped by adding 50
153 mL of ethanol, and the oxidized fibres were filtered and washed twice using deionised water.

154 *TEMPO-NaBr-NaClO (O2): Cellulose fibres (2 g) were suspended in 200 mL water.
155 TEMPO (30 mg) and NaBr (250 mg) were added to the suspension. Then 50 mL of a

156 commercial NaClO solution (12°) was added dropwise to the cellulose suspension at a
157 temperature around 5°C, kept constant throughout the oxidation reaction. The pH was
158 maintained around 10 by the continuous addition of a 0.1 M aqueous solution of NaOH. The
159 oxidation was stopped by adding ethanol (20 mL) and the pH was adjusted to 7 by adding 0.1
160 M HCl.

161 2.6 Carboxyl content

162 The carboxyl content of the oxidised cellulose was determined using conductimetric titration,
163 as described elsewhere¹⁴.

164 2.7 Fibrillation process

165 *High pressure homogenization (HPH)*: NFC was prepared from the delignified pulp by
166 pumping through a GEA Homogenizer processor (NS1001L PANDA 2 K-GEA, Italy). The
167 homogenization was conducted in two steps. Firstly, the fibre suspension at a concentration of
168 1.5 wt% was passed several times through thin slits at a pressure of 300 bar (4350 Psi) until the
169 suspension turned to a gel. Then, the fibrillation was pursued by further passes at a pressure of
170 600 bar (8700 Psi).

171 *Fibrillation using a conventional high speed blender (HSB)*: the fibres in a water suspension at
172 a concentration of 2 wt% were disintegrated during 15-20 min in a “one-step operation” using
173 a domestic high speed blender (MOULINEX 400 W) with a constant running speed of 11 000
174 rpm, in which the blades rotate in a recessed section at the bottom of a container of 1 L
175 capacity.

176 2.8 Yield in nanofibrillated cellulose

177 Centrifugation of a diluted NFC suspension was shown to be an efficient means to separate the
178 unfibrillated materials¹⁴ from those partially fibrillated. The protocol was carried out as
179 follows: a dilute suspension with about 0.1 wt% solid content (Sc) was centrifuged at 4500 rpm
180 for 20 min to separate the nanofibrillated material (in supernatant fraction) from the non-
181 fibrillated or partially fibrillated ones, which settle down. Then, the sediment fraction was
182 dried to a constant weight at 90 °C. The yield was calculated from Eq.2

$$183 \quad \text{Yield \%} = \left(1 - \frac{\text{weight of dried sediment}}{(\text{weight of diluted sample} \times \%Sc)} \right) \times 100 \quad (2)$$

184 The results represented the average values of three replications.

185 2.9 Field-emission scanning electron microscopy (FE-SEM)

186 A Weiss SEM was used to obtain images by capturing secondary electrons emitted from the
187 surface of a NFC sample, prepared from a drop of the NFC suspension (with a solid content
188 about 0.05 wt%) deposited on the surface of a silicon wafer and coated with a thin carbon
189 layer, applied by ion sputtering with a thickness limited to 2 to 3 nm. To ensure a good image
190 resolution without any damage to the samples during the analysis, the acceleration voltage was
191 maintained at a relatively low range (2-5 kV).

192 2.10 Determination of the crystalline index

193 The crystallinity was evaluated from an X-ray diffraction (XRD) pattern obtained using a
194 BRUKER AXS diffractometer (Madison, WI) with a Cu-K α radiation, generated at 30 kV and
195 an incident current of 100 mA. The (2 θ) angular region from 5° to 40° was scanned by steps of
196 0.05° using a step time of 10 s. The crystalline index (CrI) was calculated by equation 3 using
197 the diffraction intensities of the crystalline structure and that of the amorphous fraction,
198 according to the method of Segal et al²⁰:

$$199 \quad CrI\% = \left[\frac{I_{002} - I_{am}}{I_{002}} \right] \times 100 \quad (3)$$

200 where I_{002} is the maximum intensity of the (002) diffraction peak, taken at 2 θ between 22° and
201 23° for cellulose I, and I_{am} is the intensity of the amorphous diffraction peak taken at 2 θ
202 between 18° and 19° for cellulose I.

203 Scherrer's equation was used to calculate the crystallite size, T (nm), perpendicular to the
204 (200) plane for cellulose I crystals:

$$205 \quad T = \frac{K\lambda}{\beta \cdot \cos\theta} \quad (4)$$

206 Where K is a dimensionless shape factor and usually taken to be 0.9, λ (1.54 Å) is the X-ray
207 wavelength, β is the line broadening at half the maximum intensity (FWHM), in radians, and
208 θ is the diffraction angle

209 2.11 Energy consumption

210 The instrument used for the energy consumption measurement was a power analyser from
211 SCOMECA DIRIS A20 which is an integrating tool that measures precisely the total amount of

212 electrical energy, the voltage and the current intensity supplied to the electrical equipment in a
213 given period of time.

214 *2.12. Nanocomposites processing*

215 Commercial acrylic latex obtained by the copolymerization of styrene (S) and butyl
216 acrylate (BuA) was used as a matrix. The size of the polymer particles was around 150 nm
217 and the solid content 50 wt%. The glass–rubber transition temperature (T_g) of this poly(S-co-
218 BuA) copolymer was about -10°C .

219 The NFC gel was mixed with the latex in order to obtain nanocomposite films with
220 weight fraction of cellulose ranging from 0% to 15%. After stirring for 1h, the mixture was
221 cast in a Teflon mould and stored at 40°C until water evaporation was completed. A
222 transparent to translucent film, depending on the NFC content, was obtained with a thickness
223 in the range of 300 to 400 μm .

224 *2.13. Tensile tests*

225 The non-linear mechanical behaviour of the films was analyzed using an Instron testing
226 machine in tensile mode, with a load cell of 100 N working at a strain rate of 10 mm/min at
227 25°C . The specimens were obtained using a cutting device.

228 *2.14. Transparency measurement*

229 The transparency of neat acrylic film and nanocomposite films was measured at
230 wavelengths from 200 to 800 nm using a UV-visible spectrometer (Lambda 35, Perkin-
231 Elmer). The transmission spectra of the films were recorded using air as reference.

232 The following abbreviations were used to design the different means of treatment performed
233 during the production of NFC:

234

235 **3. Results and discussion**

236

237 **3.1 Pulp characterization**

238 The typical feature of the stem anatomy was analyzed by SEM observation of transverse
239 sections of the stem. As shown in Figure 1, several different cell elements, namely
240 parenchyma, vascular tissue, epidermis, and the fibre cells can be perceived. These micro-
241 structural aspects are related to different functions in the plant. The parenchyma cell acts as

242 carbohydrate storage, the vascular tissues provide long distance transport and the structural
243 support and elementary fibres inside the fibre bundles supply the mechanical support. Most of
244 the fibres displayed a large lumen width, in the range of 10 to 20 μm and the thickness of the
245 cell walls layed between 2 to 5 μm , depending on their position in the stalk, the outer cells
246 being thicker than the inner ones.

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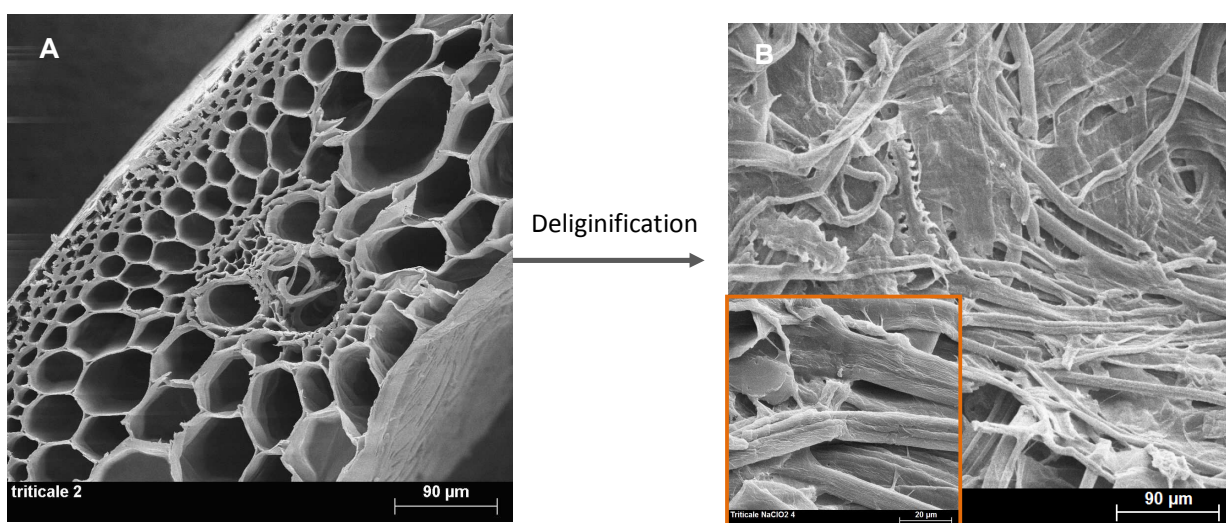
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256 Figure 1 : SEM observation of (A) cross-section of triticale straw, and (B) of the ensuing fibres
257 after the delignification process.

258

259 Given the lower lignin content in the crops residues, along with their more open structure
260 compared to those of hard- and softwoods, milder pulping conditions with no sulfur processes
261 can be applied for the lignin extraction.

262 The chemical composition of native triticale straw, NaOH and NaClO_2 extracted fibres are
263 given in Table 1. Native triticale was composed of 39, 31, 21 and 5 cellulose, hemicelluloses,
264 lignin and ash, respectively. This composition is typical of agricultural crops residue²¹. Ash
265 was mainly composed of silica, as confirmed by XRD (Figure 2). NaClO_2 delignification led
266 to the effective removal of lignin, with the highest yield around 65 %. The ensuing pulp was
267 white, without any necessity to implement a bleaching treatment. By means of the soda
268 pulping process, a brown pulp was obtained with a yield of around 45 %. The specificity of
269 each delignification method accounts for the pulp characteristics. Actually, contrary to the
270 soda pulping method known to remove lignin as well as a high fraction of hemicelluloses, the

271 NaClO₂ method is more selective for lignin²² and preserves most of the hemicelluloses in the
272 biomass²³.

273 **Table 1.** Chemical composition (wt %) and properties of the agricultural by-products used here.

Constituents	Native	NaOH	NaClO ₂
	triticale straw	extracted	extracted
Cellulose	39±1	84±2	71±2
Hemicellulose	31±1	16±1	29±1
Lignin	21±1	-	-
Ash	5±0.5	-	-
Hot water extractible	4		
Properties			
Pulp yield	-	45±3	65±3
Fibres length (µm)	-	840±50	700±50
Fibres width (µm)	-	22±4	21±4
CrI	48±2	60±2	74±2

274 * Solubility in hot water in %

275 ** as % of dry matter.

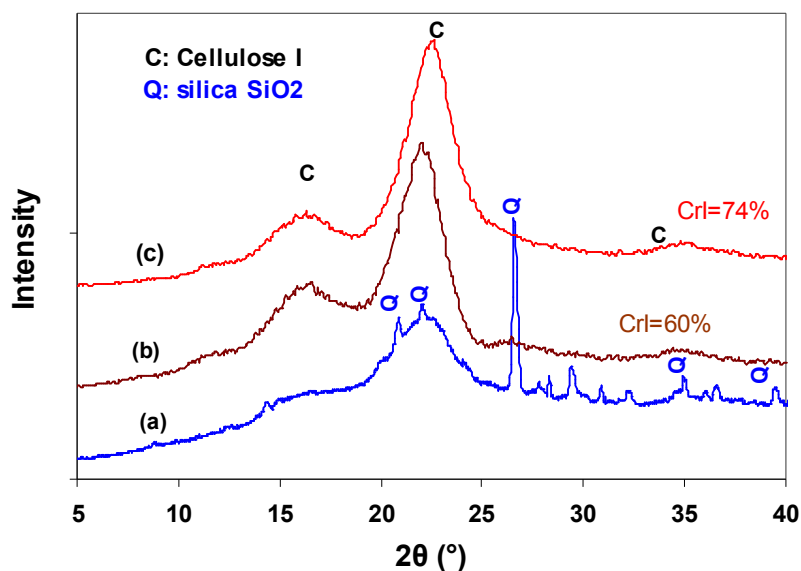
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277 The peaks at 2θ values of 15.2°, 16.7°, and 23.1° corresponding to (101), (101 (10 $\bar{1}$)), (002)
278 planes, respectively, are the diffraction peaks of cellulose I structure (JCPDS. No. 03-0226).

279 The XRD characterization revealed the presence of the main characteristic peaks of cellulose I
280 (JCPDS. No. 03-0226) at 2θ values of 15.2°, 16.7°, and 23.1° corresponding to (101), (10 $\bar{1}$),
281 (002) planes, respectively, for all studied samples, before and after the extraction procedure,
282 indicating the preservation of the native crystalline structure even in the presence of the NaOH
283 treatment. After the lignin extraction, the crystallinity index increased due to the partial
284 removal of the amorphous hemicelluloses and lignin. In addition, lower crystallinity indexes
285 were found when the lignin extraction was implemented with the NaClO₂ procedure, which is
286 in accordance with the highest content of hemicellulose in these samples. The XRD revealed
287 the presence of silica (SiO₂) (JCPDS. No. 33-116) on the triticale straw, which was completely

288 removed after the delignification treatment. This presence accounts for the relatively high ash
289 content of triticale compared to woody plants. The presence of silica is typical of agricultural
290 crops residue. It is accumulated in the form of SiO_2 by transport of water-soluble silicic acid
291 from the soil to the plant tissues through the roots. This element is of significance in the life of
292 plants and the performance of crops. During the sclerification of the cell wall, $\text{Si}(\text{OH})_4$
293 undergoes condensation to give the Si-O-Si oligomers that further grow to form SiO_2
294 nanoparticles. Silicic acid also acted as a cross-linking agent between the lignin and the
295 carbohydrate²⁴, via complexations with phenolic acids and the hydroxyl groups of
296 hemicelluloses and cellulose.

297



298

299 Figure 2 : XRD patterns for (a) neat triticale straw, (b) NaOH delignified, and (c) NaClO_2 delignified
300 pulps.

301

302 The morphology of the fibres extracted from the crop residues exhibited typical cells already
303 observed in the cross-section of the stems (Figure 1B). In addition to the fibres, other non-
304 fibrous cell elements, such as vessels and parenchyma cells, could be distinguished. The
305 fibrous cells were collapsed to approximately 10-30 μm in width and characterized by thin cell
306 walls about 2-5 μm thick. The vessels and collenchyma cells were larger than other cells, with
307 a rectangular shape approximately 100–300 μm long and 40–50 μm wide. The mean length

308 and width of the delignified fibres, determined from MORPHI analysis, were around 500-600
309 μm and 20 μm , respectively.

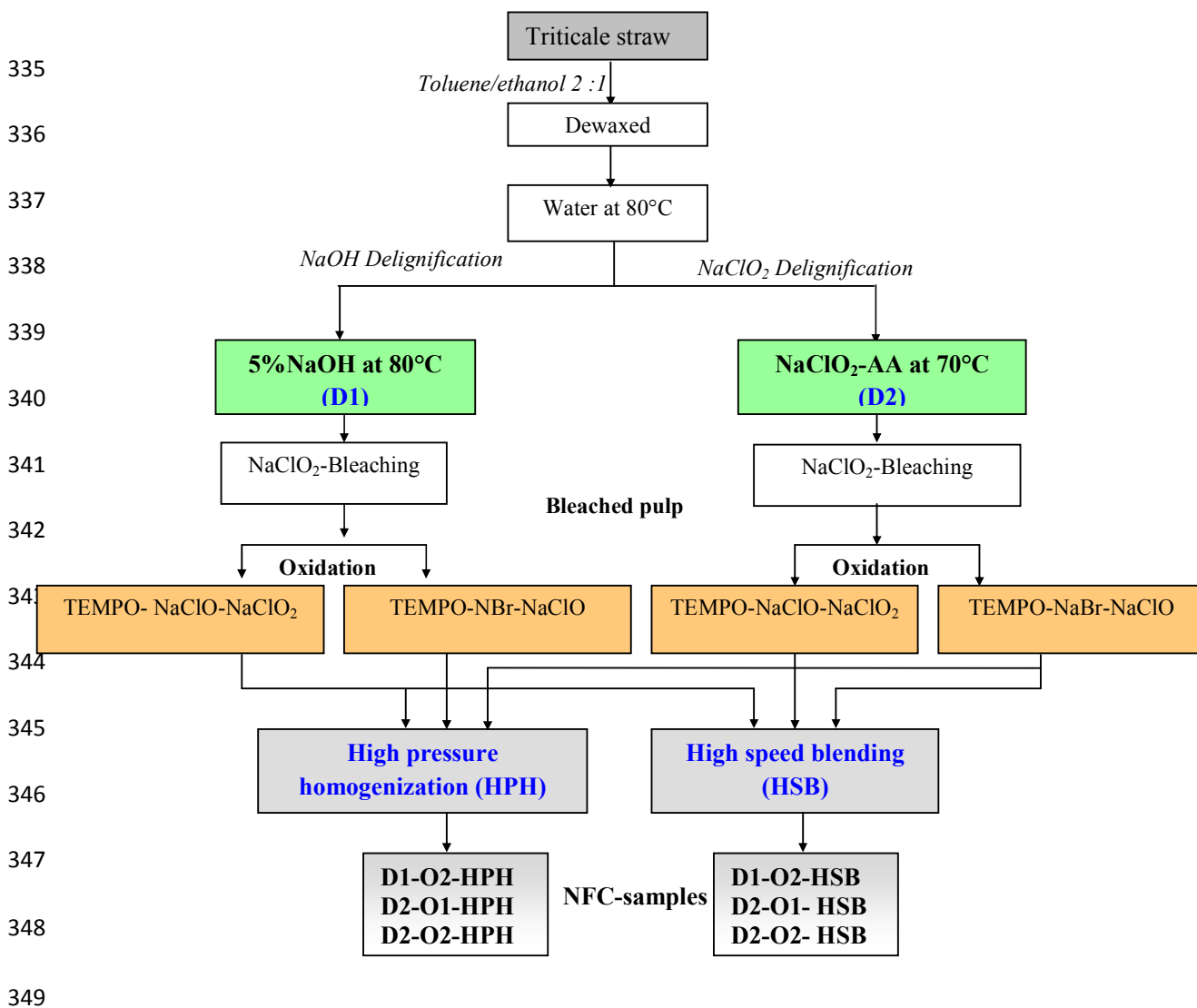
310 **3.2 Nanofibrillation behaviour of the fibres**

311 A high-pressure homogenizer (HPH) and a high-speed blender (HSB) were used as mechanical
312 devices to breakup the cell wall and release the cellulose microfibrils. The former process is
313 the conventional approach commonly adopted to produce NFC, but with a high-energy input.
314 The latter was employed here as a low energy demand method to produce NFC. For both
315 approaches, the fibres were first submitted to a TEMPO-mediated oxidation pre-treatment to
316 bring the carboxyl content up to 500 $\mu\text{mol/g}$ and to facilitate the defibrillation process. The
317 oxidation pretreatment aims at generating carboxylic groups, whose ionization facilitates the
318 fibrillation and the break-down of the cell wall of the fibres through different effects, namely:
319 (a) the oxidation generates negative charges that bring forth repulsive forces between
320 microfibrils within the cell wall; (b) the oxidation favours the hydration and swelling of the
321 fibres, making them more flexible; (c) the oxidation loosens the primary and S1 cell walls,
322 making the S2 layer more accessible and more prone to fibrillation during the homogenization
323 process; and finally (d), the oxidation results in chain scission in the amorphous zones, creating
324 defaults within the fibre cell wall, which facilitates the mechanical fibrillation.

325 The oxidation was carried out either at neutral or basic pH (viz. the O1 and O2 modes) using
326 NaClO_2 and NaClO as oxidizing agents, respectively. Although both methods are specific to
327 the selective oxidation of the C6 primary alcohol groups into aldehydes and/or carboxylic acid
328 groups, they affect differently the degree of polymerization²⁵ (DP) of the oxidized cellulose.
329 The former (O1) led to a significant decrease of more a factor of five, whereas the latter left
330 essentially unchanged.

331 To clearly distinct between the different NFC samples, a chart diagram showing all the steps
332 adopted to generate NFC from triticale straw, with their corresponding abbreviations is given
333 in scheme 1.

334

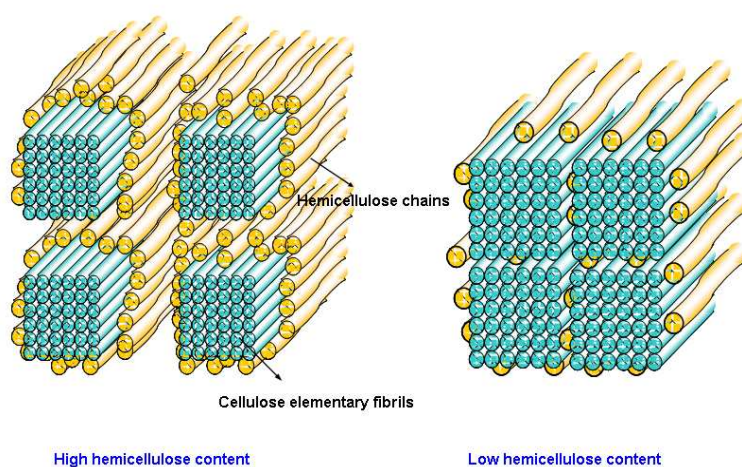


350 **Scheme 1:** Illustrative scheme of the various treatments performed on the triticale straw for
 351 the preparation of NFC samples by a high pressure homogenization (HPH) and a high speed
 352 blender (HSB).

353 The change in the fibrillation yield and in the transparency degree at 700 nm according to the
 354 pulping mode, the oxidation method and the disintegration mode are given in Figure 3. The
 355 highest fibrillation yield, exceeding 95%, was achieved using HPH starting from fibres
 356 delignified via NaClO_2 (D2-O2-HPH and D2-O1-HPH). This means that when a high content
 357 in hemicelluloses was left in the fibres after the delignification and bleaching treatments, the
 358 fibres were effectively fibrillated into NFC via HPH, irrespective of the oxidation mode.
 359 However, a slightly higher transparency degree of the NFC gel was observed when the
 360 O_2 oxidation route was adopted (Transparency close to 94% for the O_2 oxidation against 84%
 361 for O1). However, a decrease in the fibrillation yield as well as in the transparency were noted

362 when NaOH delignified mode was used, even when the HPH disintegration mode was
363 adopted. Interestingly, when lignin extraction was performed via NaClO_2 mode, it was possible
364 to disintegrate cellulose fibers into NFC using a conventional HSB. Both of the two oxidation
365 routes (O2 or O1) led to a high fibrillation yield (around 88%) and good transparency (samples
366 D2-O2-HSB; D2-O1-HSB). On the other hand, when the delignification was carried out with
367 NaOH, a translucent NFC gel was obtained with a fibrillation yield that do not exceed 50%
368 fibrillation yield, even after prolonged disintegration in the HSB for more than 30 min for
369 (sample, D1-O2-HSB). The fibrillation yield further decrease to about 20% when the NaOH
370 delignified fibers were oxidized via O1 route (D1-O1-HSB). This means that fibers became
371 hard to be converted into NFC via the HSB when the lignin extraction was performed with
372 NaOH, especially when the TEMPO-mediated oxidation was carried out at neutral pH (O1
373 route).

374 The easier fibrillation ability of NaClO_2 delignified fibres is explained by the higher residual
375 hemicelluloses content left in the fibres that contributed to reduce the interaction via hydrogen
376 bonding among cellulose microfibrils within the cell wall. The key role of hemicelluloses in
377 the fibrillation process has been highlighted in our previous work²⁶. Thanks to their amorphous
378 and highly hydrated character, the hemicelluloses surrounding the microfibrils behave as a
379 protective colloid preventing the microfibrils from coming close together to self-associate into
380 larger aggregates through hydrogen bonding. A schematic view of the assembly mode of
381 cellulose microfibrils according to the delignification route is given in Scheme2.

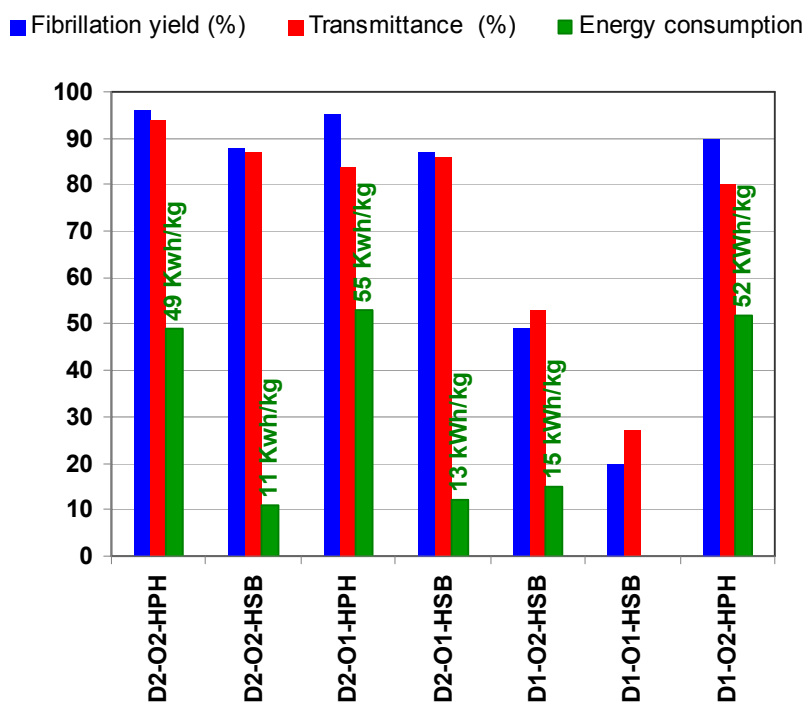


Scheme 2: Schematic illustration of the microfibrils assembly according to the content of hemicellulose within the fibers

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383

384 The low cell wall thickness of triticale fibres is another reason likely to facilitate the break-up
385 of the cell wall via HSB mechanical disintegration.



386

387 **Figure 3:** Evolution of the fibrillation yield, transmittance of the NFC gel and energy consumption (per
388 Kg of dry NFC) during the production of NFC following the different routes adopted in this study.

389

390 The difference in the optical aspect of the NFC gel was further highlighted from the digital
391 photos showing the aspect of the NFC gel produced from triticale via the methods discussed
392 above (Figure 4). The most transparent NFC suspensions were those where the delignification
393 was implemented using D2 (NaClO₂/acetic acid) process. Both of the HPH and the high speed
394 blender (HSB) modes of disintegration led to NFC suspensions with a higher transparency
395 level, which is indicative of highly nanofibrillated cellulose. In contrast, the NaOH
396 delignification (D1) mode led to an opaque NFC suspension, namely when HPH was used for
397 the disintegration (D1-O2-HSB). The difference in the optical transparency of the NFC gel is
398 essentially due to the difference in the nanofibrillation extent of the fibers. The presence of

399 partially fibrillated materials with a width larger than 100 nm led inevitably to a huge drop in
400 the transparency of the NFC gel because of light scattering at water-fibrils discontinuity.

401

402

403

404 **Figure 4:** visual aspect of NFC from triticale at a solid content of 1wt%, according to the
405 production mode.

406

407

408 Although the energetic input during the nanofibrillation process is a key issue in the
409 cost production of NFC, the quantification of energy input according to the fibers
410 pretreatment has been reported only in a few papers^{27,28}. In addition, it is important to keep
411 in mind that energy consumption should be defined with respect to a given degree of
412 fibrillation or a set value of gel-transparency. This is an important point, since the change
413 from a liquid to a gel-like form of the fibers suspension did not imply the conversion of the
414 whole cellulose fibers into nanofibrillated material. A nanofibrillation extent of over 90%
415 requires necessarily a high-energy input to completely breakup the cell wall and release the
416 cellulose microfibrils or bundles of cellulose microfibrils. The energy consumption for the
417 different approach adopted in the present work was also included in Figure 3. Starting from
418 NaClO₂/AA delignified fibres, the energy input necessary to attain a fibrillation yield of
419 around 90% was in the range of 40-45 kWh/kg of dry NFC with HPH and decreased to about
420 9-12 kWh/kg when HSB was used. This corresponds to a 80% reduction in energy input. The
421 need to raise the pressure up to 500-700 bars during the HPH processing is the origin of the
422 high energy consumption of this disintegration mode.

423

424 **3.3 Morphology of NFC**

425

426 The morphology of the different samples of NFC prepared via different approaches
427 was analysed using FE-SEM, performed on the supernatant fraction (Figure 5). NFC
428 obtained by the (D2-O1-HPH) and (D2-O1-HSB) routes were composed of individualized
429 fibrils with a width of 20-30 nm and a length exceeding 2 μm. If one takes into account the
430 coating metallisation layer necessary for FE-SEM observation to be within a 2-3 nm

431 thickness, then the ultimate width should be in the range of 15-20 nm and the aspect ratio
432 exceed 100. It turns out that the width of the NFC was much higher than the average thickness
433 of cellulose crystallites which is around 3.5 nm, as calculated according to the Scherrer
434 equation. This means that the ensuing nanofibrils were, in turn, formed by bundles of
435 elementary cellulose fibrils, regardless of whether HPH or HSB were used for the
436 disintegration process.

437 NFC produced via (D2-O2-HSB) or (D2-O2-HPH) exhibited quite the same width, but looked
438 shorter than those produced via (D2-O1-HSB) or (D2-O1-HPH). Their length was lower than
439 1 μm , which led to an aspect ratio of about 50. The difference in the NFC length according to
440 the oxidation mode (neutral vs basic pH) was explained by the higher extent of the degree of
441 polymerization reduction when the oxidation was carried out under basic conditions. Indeed,
442 it was reported that the TEMPO-NaBr-NaClO at pH 10 led to a remarkable cleavage of
443 polysaccharides as a result of the β -elimination promoted by the presence of C6-aldehydes
444 formed as an intermediate structure, and through a radical scission resulting from the
445 formation of hydroxyl radicals during the oxidation process²⁹. The decrease in the degree of
446 polymerization during the oxidation process led inevitably to a fall in the length of the
447 cellulose microfibrils. This correlation can be understood if one assumes that during the
448 oxidation process, the amorphous domains are more prone to oxidation than the crystallite
449 domains, due to their higher accessibility. Given the ultrastucture of the microfibrils being
450 composed of alternating crystallite and disordered amorphous domains regularly distributed
451 along the microfibrils axis, the resulting breakup in the chains within the amorphous domains
452 led inevitably to a shortening in the microfibrils length. Similar results were reported by
453 Benhamou et al³⁰ revealing a reduction in the fibrils length, as the carboxyl content increased
454 and the oxidation was carried out at basic pH.

455 NFC produced from NaOH-delignified pulps by HSB (D1-O2-HSB) showed in addition to
456 the nanosized thin fibrils with width lower than 20 nm, not fully disintegrated microfibrils
457 with a width in the range of 100-200 nm. Presumably, energy generated with the HSB process
458 was not sufficient to completely breakup the hydrogen-bonding network holding the cellulose
459 microfibrils. We infer that the removal of a high fraction of hemicelluloses during the pulping
460 process with NaOH, brought the microfibrils into closer contact, making intermolecular
461 hydrogen bonding more cohesive.

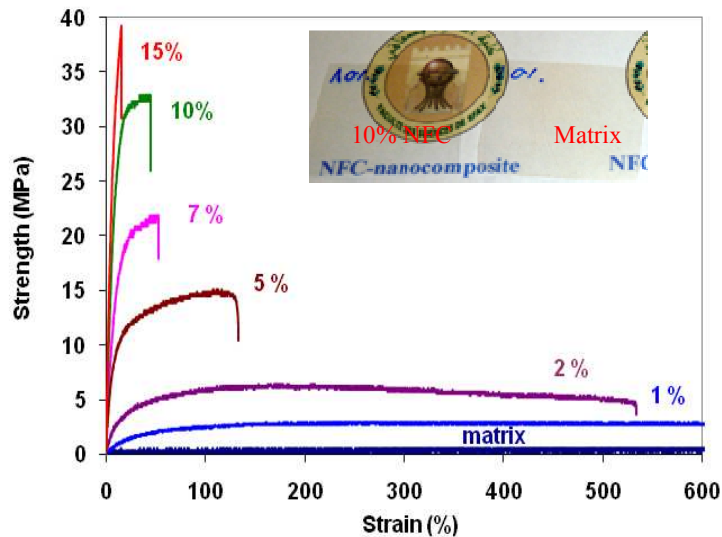
462 *Figure 5: FE-SEM images of NFC produced from tritical straw via high pressure*
463 *homogenization and high speed blender.*

464

465 3.4. Reinforcing potential of NFC

466 To investigate the reinforcing potential of NFC from triticale produced by the different
467 approaches, nanocomposite films were prepared via solvent-casting and tested using non-
468 linear tensile tests performed at room temperature. A ductile water-borne acrylic matrix with a
469 T_g around -10°C was chosen as a matrix in order to reach the limit strength without premature
470 breaking of the sample due to excessive rigidity. Typical stress-strain curves obtained from
471 tensile tests for NFC-based nanocomposite films are shown in Figure 6. The addition of NFC
472 led to the steady enhancement in the tensile modulus as well as in the tensile strength.
473 However, as shown in Figure 7, the increment was dependent on the preparation route of
474 NFC. The highest reinforcing potential was associated with D2-O1-HPH (NaClO₂
475 delignification, oxidation at pH 7 and disintegration via HPH), followed by D2-O2-HPH, D2-
476 O1-HSB and D2-O2-HSB. For instance, at 10 wt% NFC content, the respective increment in
477 the tensile modulus/tensile strength were 160/27, 79/11, 70/11, and 22 time/5 time that of the
478 neat matrix.

479



480

481 **Figure 6:** Typical stress–strain curves for nanocomposite films prepared from NFC
 482 produced via D2-O1-HPH

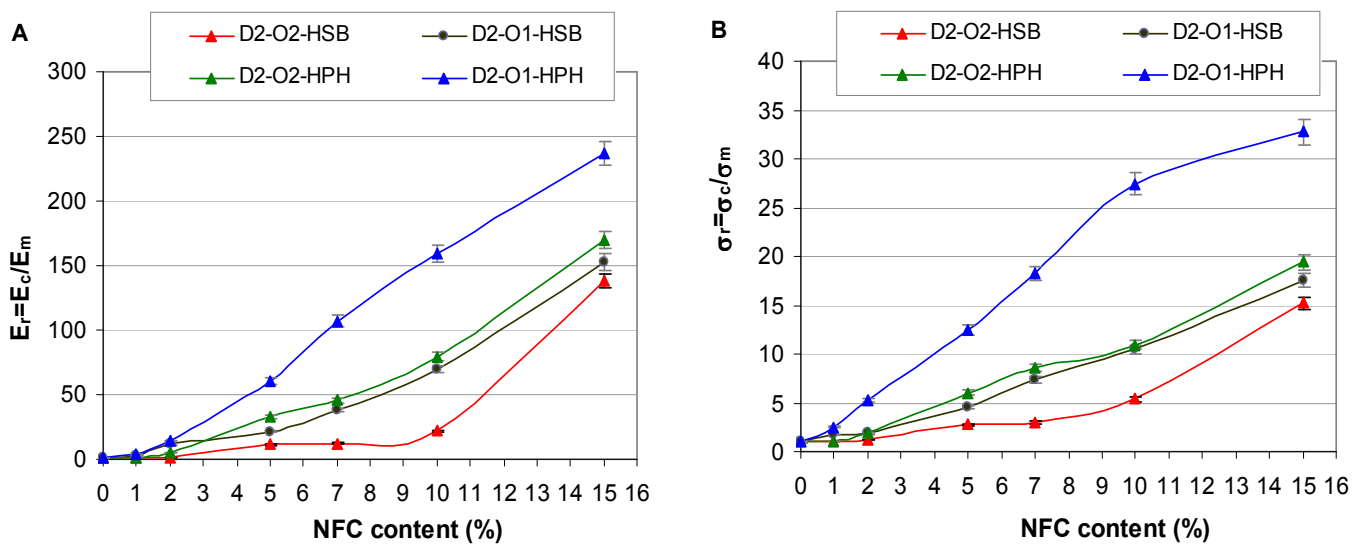
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495 **Figure 7:** increment in (A) the tensile modulus, and (B) the tensile strength of
 496 nanocomposite films according to the NFC content and the mode of their production.

497

498 The stronger reinforcing potential of NFC produced by the D2-O1-HPH approach was
 499 attributed to the consequence of the higher length of the cellulose nanofibrils, as confirmed by
 500 the FE-SEM observations. The higher length, along with the narrow thin width of the NFC
 501 (about 20 nm as shown in Fig 5) produced through this approach, led to a higher aspect ratio
 502 that is more favourable to generate an entangled network. The set-up of an interconnected
 503 network held-up by strong hydrogen bonding is known to be a prerequisite in order to take
 504 advantage of the unusual enhancement of the stiffness and strength in the nanocellulose
 505 reinforced nanocomposites³¹. The concept of percolation originally adopted for rod-like
 506 cellulose nanocrystals was shown to be valid for NFC⁵. Accordingly, the tensile modulus of
 507 the nanocomposite is simply the product of the modulus of percolating filler network and
 508 the volume fraction of the percolating filler phase (equation 5)

$$509 \quad E_c = \psi E_r \quad (5)$$

510 Where ψ can be written as:

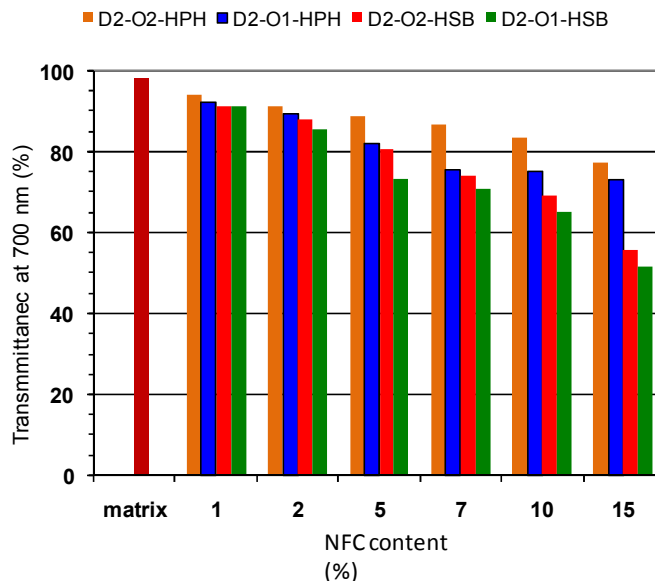
$$511 \quad \psi = 0 \quad \text{For } \phi < \phi_p \quad (6)$$

$$512 \quad \psi = \phi \cdot \left(\frac{\phi - \phi_p}{1 - \phi_p} \right)^b \quad \text{For } \phi \geq \phi_p \quad (7)$$

513 Where ψ , ϕ and b are the volume fraction of the percolating network, the total volume
 514 fraction of the nanofiller and the critical exponent, respectively. Since the percolation
 515 threshold is inversely proportional to the aspect ratio of the dispersed objects, the higher the
 516 aspect ratio, the stronger is the reinforcing potential of NFC .

517 Interestingly, it can be seen that NFC produced via HSB (D2-O1-HSB) exhibited nearly the
 518 same reinforcing potential than that prepared via HPH (D2-O2-H) over the whole range of
 519 NFC loading. Here again, the higher aspect ratio of NFC from the (D2-O1-HSB) process
 520 compared to that from the (D2-O2-HPH) counterpart, accounts for the upholding of the
 521 reinforcing efficiency of the NFC, even though a high-speed blender was used for the
 522 disintegration process. This result is important to highlight since the production of NFC using
 523 a conventional high speed blender instead of the high pressure homogenizer or microfluidizer
 524 usefully adopted, contributed to make easier the scale-up production of NFC not only in term
 525 of plant facility but also on the basis of the energy cost. The drop in the reinforcing potential
 526 for NFC produced through (D2-O1-HSB) was likely due to its lower aspect ratio and lower
 527 fibrillation yield.

528 3.5. Optical Properties of the Nanocomposite Films



529

530

Figure 7. (a) Transmittance at 700 nm of nanocomposite film vs NFC content and according to their production mode.

531

532

533 Generally, in addition to the mechanical reinforcing effect provided by the inclusion of
 534 cellulose-based nanofiller in a polymer matrix, the optical transparency of the polymeric
 535 matrix is another aspect which is often aimed to be preserved. In general, for nanocomposite
 536 materials, the reduction in the transparency is caused by light scattering against the randomly
 537 dispersed particles, brought about by the discontinuity between the refractive index of the
 538 matrix and that of the nanofiller. The critical factor controlling the transparency of
 539 nanocomposites is the width of the nanofiller or, more specifically, the effective scattering
 540 cross-sectional area and its dispersion level within the host polymer matrix. In order to
 541 evaluate the optical transparency of the nanocomposites for different nanoparticle contents,
 542 the transmittance at 700 nm was used, and the results are shown in Figure 7a. The film
 543 transmittance was normalized to a 200 μm -thickness using the Beer–Lambert law, in order to
 544 avoid the effect of a small fluctuation in the film thickness. The addition of NFC led to a
 545 steady drop in the transparency of the film, namely over 7% NFC loading, where a ~20%
 546 decrease in the transmittance was noted. However, the decrease in the transparency depended
 547 on the NFC origin and the following order was noted D2-O2-HPH>D2-O1-HPH>D2-O2-
 548 HSB>D2-O1-HSB. The highest transparency was observed for nanocomposites with NFC
 549 produced via HPH, which was probably the consequence of a more effective fibrillation

550 degree brought about by the HPH. In fact, since the scattering intensity is proportional to the
551 third power of particle size, the presence of partially fibrillated material with a size within the
552 micron scale, even in a low proportion, led inevitably to a fall in transparency. However, it is
553 interesting to note that over the whole range of NFC contents, a higher transmittance was
554 noted for NFC from D2-O2-H, compared to that from D2-O1-HPH. This means that a longer
555 NFC induced more scattering than a shorter one. The same trend was noted for NFC produced
556 via HSB (D2-O2-HSB>D2-O1-HSB). This behavior is attributed to the higher tendency of
557 longer cellulose nanofibrils to generate a bonded area through entanglement and cross-section
558 contact with a higher thickness, once the water was removed and a film formed. The resulting
559 bonded area brought therefore more scattering than individual nanofibrils.

560 **4. Conclusion**

561 Nanofibrillated cellulose from tritical straw was produced using both a high pressure
562 homogenization and a conventional high speed blender. Alkaline and NaClO₂/acetic acid
563 pulping process was adopted to remove lignin and a TEMPO-mediated oxidation at two pH
564 was carried out to bring the carboxyl content up to 500 μmol/g and facilitated the fibrillation
565 process.

566 The delignification mode and the oxidation route were shown to control the fibrillation extent
567 and the energy consumption during the disintegration process. The highest energy
568 consumption was noted when the alkaline pulping process was adopted. The content of the
569 residual hemicellulose left within the cellulose fibers after lignin removal was considered as
570 key parameters controlling the fibrillation process and the energy consumption. The
571 transparency degree of the NFC gel and the morphology of the cellulose nanofibrils were also
572 dependent of the delignification and the disintegration mode. When the NaClO₂/acetic acid
573 pulping process and TEMPO-oxidation at neutral pH were implemented, long individual
574 nanofibrils with width around 20 nm and length exceeding several μm were obtained via the
575 high pressure homogenization as well as the high speed blender. The oxidation at basic pH led
576 to a shorter nanofibrils resulting from the DP reduction during this chemical pretreatment.
577 The change in the nanofibrils morphology according to the delignification/disintegration
578 modes affected equally the reinforcing potential of the NFC. NFC produced from
579 NaClO₂/acetic acid delignified pulp using high pressure homogenization exhibited a
580 reinforcing potential about two folds higher than that produced from high speed blender.

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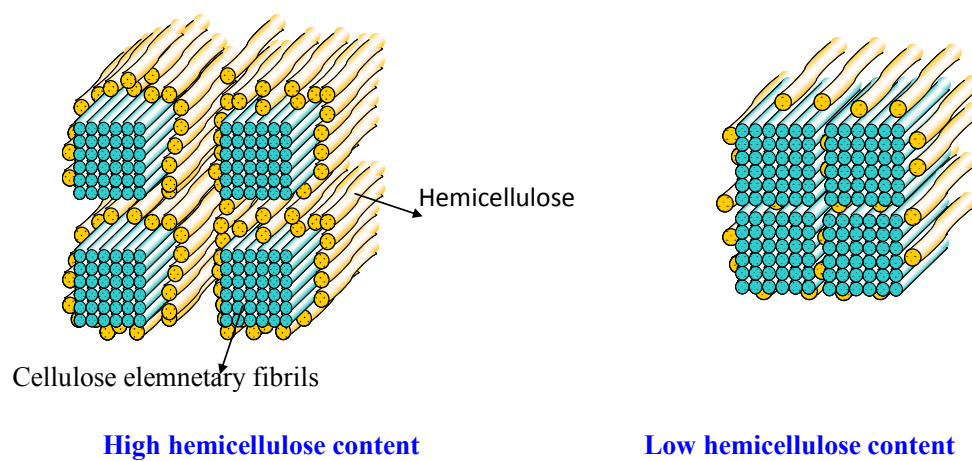
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Scheme 2: Schematic illustration of the microfibrils assembly according to the content of hemicellulose within the fibers

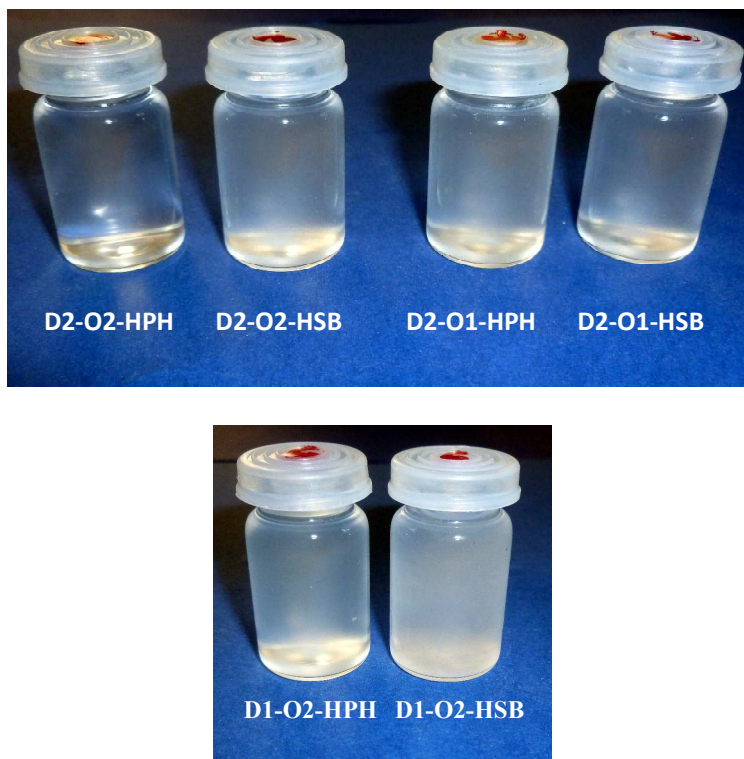


Figure 4: visual aspect of NFC from triticale at a solid content of 1wt%, according to the production mode.

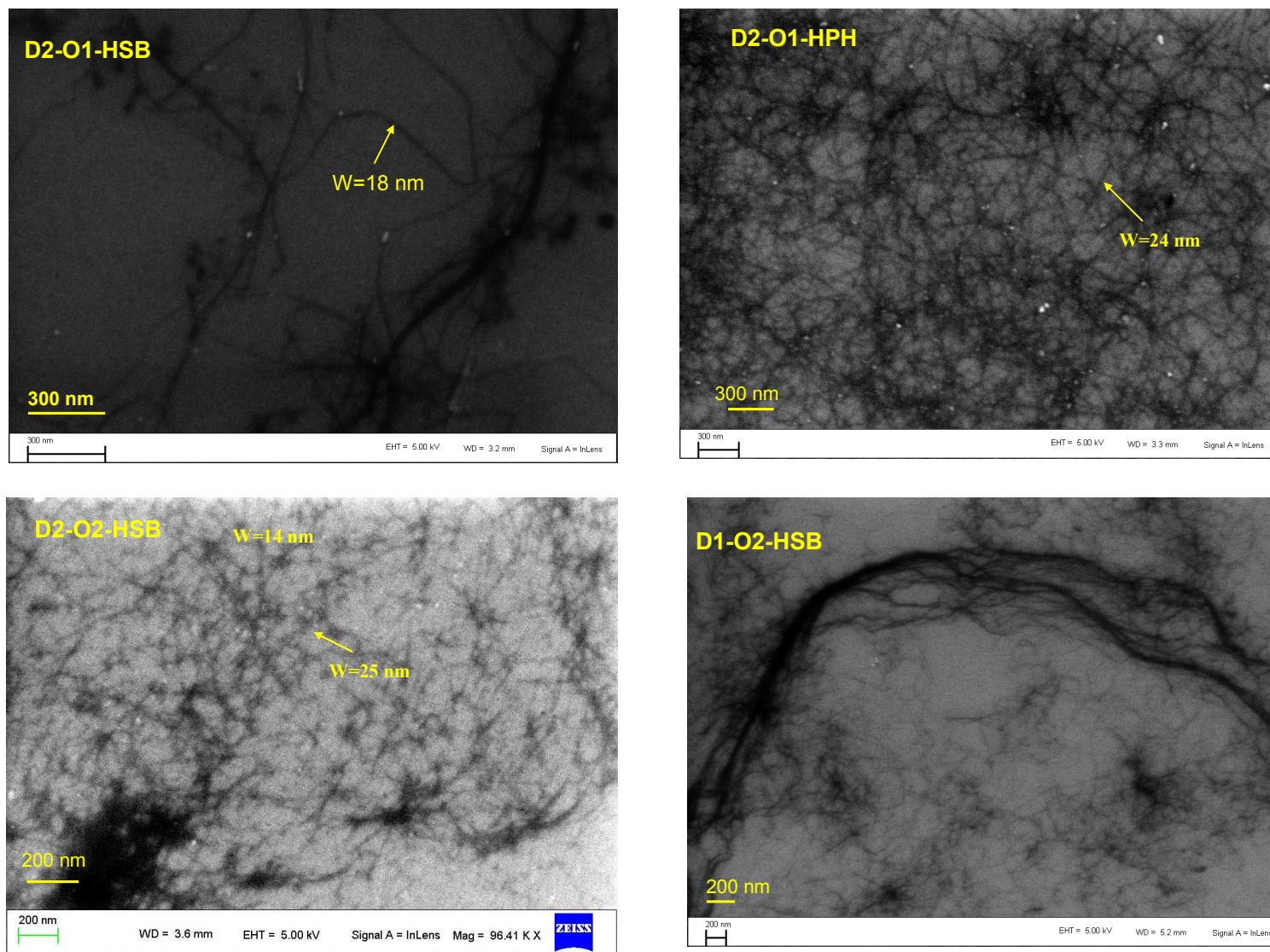


Figure 5: FE-SEM images of NFC produced from tritical straw via high pressure homogenization and high speed blender