

RSC Advances

Expedient synthesis of a pentasaccharide related to the Ospecific polysaccharide of Escherichia coli O117:K98:H4 strain

Journal:	RSC Advances
Manuscript ID:	RA-ART-09-2014-010538.R1
Article Type:	Paper
Date Submitted by the Author:	27-Oct-2014
Complete List of Authors:	Misra, Anup; Bose Institute, Molecular Medicine Division Bhaumik, Ishani; Bose Institute, Division of Molecular Medicine

SCHOLARONE[™] Manuscripts

Expedient synthesis of a pentasaccharide related to the *O*-specific polysaccharide of *Escherichia coli* O117:K98:H4 strain

Ishani Bhaumik,^a and Anup Kumar Misra^{*a}



A convenient synthetic strategy has been developed for the synthesis of a pentasaccharide related to the O-specific polysaccharide of *Escherichia coli* O117:K98:H4 strain using sequential glycosylations of functionalized monosaccharide moieties.

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxx

ARTICLE TYPE

Expedient synthesis of a pentasaccharide related to the *O*-specific polysaccharide of *Escherichia coli* O117:K98:H4 strain[†]

Ishani Bhaumik,^a and Anup Kumar Misra*^a

Received (in XXX, XXX) Xth XXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX 5 DOI: 10.1039/b000000x

A convenient synthetic strategy has been developed for the synthesis of a pentasaccharide related to the O-specific polysaccharide of *Escherichia coli* O117:K98:H4 strain using sequential glycosylations of functionalized monosaccharide moieties. Application of a one-pot reaction condition for two glycosylations and *in situ* PMB group removal reduced the number of reaction steps significantly. All

¹⁰ glycosylation reactions were stereoselective with satisfactory yield.

Introduction

Escherichia coli (*E. coli*) strains are found in the human gastrointestinal tract as commensal organisms. However, in many occasions they acquire virulence properties in a host with poor ¹⁵ immunity.¹ Several number of infections in human reported till

- date caused by pathogenic *E. coli*. *E. coli* infections are classified in three general clinical symptoms such as, urinary tract infections (UTI),² septicaemia³ or meningitis⁴ and diarrheal infections.⁵ Several strains of *E. coli* have been identified which
- ²⁰ are associated with each clinical symptoms. *E. coli* O117:K98:H4 strain has been found to cause septicaemia in children.⁶ It also produces verocytotoxin (VT) and causes diarrhoea particularly in travellers.⁷ This strain is also occasionally responsible for the acute urinary tract infections in woman.⁸ The structure of the *O*-
- ²⁵ specific polysaccharide present in the cell wall of *E. coli* O117:K98:H4 strain has been reported by Ruth Leslie *et al.*, which is composed of D-galactosamine, D-glucose, D-galactose and L-rhamnose.⁹ Emergence of multidrug resistant bacterial strains is serious concern for controlling bacterial infections using
- ³⁰ antibiotics. Since cell wall polysaccharides are involved in various stages of bacterial infections to the host, they have been used in the development of vaccines for the long term protections from the infectious diseases.¹⁰ Conventionally, glycoconjugate vaccines are prepared by isolating the polysaccharides from the
- ³⁵ bacterial cell wall and coupling with a career protein.^{10,11} However isolation of the oligosaccharides from the bacterial cells with adequate purity and structural integrity is quite tedious. On the contrary, chemical synthesis could provide oligosaccharide fragments with appropriate structure and purity. In the recent past
- ⁴⁰ several successful attempts were made in the development of vaccine candidates using synthetic oligosaccharide fragments for the preparation of glycoconjugate derivatives by conjugating with a carrier protein.¹²⁻¹⁴ In an ongoing program focusing the concise chemical synthesis oligosaccharides related to the bacterial cell
- ⁴⁵ wall,¹⁵ a linear synthetic strategy for the synthesis of a pentasaccharide related to the *O*-specific polysaccharide of *E. coli* O117:K98:H4 is presented herein.

→4)-β-D-GalpNAc-(1→3)-α-L-Rhap-(1→4)-α-D-Glcp-(1→4)-β-D-Galp-(1→3)-α-D-GalpNAc-(1→

50 Structure of the repeating unit of the O-specific polysaccharide of E. coli O117:K98:H4.

Results and discussion

The strategy for the synthesis of the pentasaccharide as *p*methoxyphenyl (PMP) glycoside (1) involves sequential ⁵⁵ glycosylations of suitably functionalized monosaccharide intermediates (Figure 1). The selection of the PMP group at the anomeric protection of the reducing end of the pentasaccharide could provide the option for the conjugation of the pentasaccharide with an appropriate protein or aglycon moiety ⁶⁰ after oxidative removal of the PMP group.¹⁶ As per the requirement of the synthetic strategy, monosaccharide intermediates **2**, **3**,¹⁷ **4**,¹⁸ **5**¹⁹ and **6**²⁰ were prepared in good yield from the commercially available reducing sugars using earlier reported reaction conditions. A number of recently developed ⁶⁵ reaction methodologies have been used in the synthesis of the target pentasaccharide.



Figure 1: Structure of the synthesized pentasaccharide with its synthetic intermediates.

- *p*-Methoxyphenyl 2-azido-4,6-*O*-benzylidene-2-deoxy- α -Ds galactopyranoside (7)²¹ (prepared from D-galactal) was treated with sodium cyanoborohydride²² in the presence of HCl in Et₂O to give 3,4-dihydroxy derivative which on treatment with triethyl orthoacetate in the presence of *p*-toluenesulfonic acid²³ followed by acid hydrolysis of the orthoester furnished *p*-methoxyphenyl
- ¹⁰ 4-*O*-acetyl-2-azido-6-*O*-benzyl-2-deoxy- α -D-galactopyranoside (2) in 72% over all yield. Nitrosyl tetrafluoroborate (NOBF₄)²⁴ mediated stereoselective glycosylation of compound 2 with trichloroacetimidate derivative 3 furnished disaccharide derivative 8 in 79% yield, which was characterized by NMR
- ¹⁵ spectroscopy [signals at δ 5.34 (d, J = 3.5 Hz, H-1_A), 4.67 (d, J = 8.0 Hz, H-1_B) in the ¹H NMR and at δ 101.2 (C-1_B), 98.7 (C-1_A) in the ¹³C NMR spectra]. Direct conversion of acetoxy groups in compound **8** into benzyloxy group using benzyl bromide and solid sodium hydroxide²⁵ afforded compound **9** in 90% yield.
- ²⁰ Regioselective ring opening²² of the benzylidene acetal in compound 9 using sodium cyanoborohydride in the presence of HCl in Et₂O gave 4-hydroxylated disaccharide acceptor 10 in 75% yield. In order to confirm the regioselective ring opening of the benzylidene acetal, compound 10 was conventionally
- $_{25}$ acetylated using acetic anhydride and pyridine and subjected to the NMR spectral analysis. Appearance of a broad singlet at δ 5.71 in the ¹H NMR spectrum of the acetylated compound confirmed the downfield shift of 4-hydroxy group of the Dgalactopyranosyl moiety after acetylation and hence formation of
- ³⁰ compound **10**. NOBF₄ mediated stereoselective 1,2-*cis* glycosylation²⁴ of compound **10** with trichloroacetimidate derivative **4** furnished trisaccharide derivative **11** in 75% yield together with the other isomeric product (~5%), which was separated by column chromatography. Formation of 1,2-*cis*
- ³⁵ linkage in compound **11** was confirmed by NMR spectroscopy (signals at δ 5.34 (d, J = 3.5 Hz, H-1_A), 4.90 (d, J = 3.0 Hz, H-1_C), 4.70 (d, J = 8.0 Hz, H-1_B) in ¹H NMR and at δ 105.5 (C-1_B), 100.6 (C-1_C), 99.3 (C-1_A) in ¹³C NMR spectra]. Treatment of compound **11** with perchloric acid on silica (HClO₄-SiO₂)^{26,27} in
- ⁴⁰ acetonitrile furnished a trisaccharide diol derivative, which was selectively benzoylated at the primary hydroxyl group using benzoyl cyanide²⁸ and pyridine to give trisaccharide acceptor **12** in 76% yield in two steps (Scheme 1).



Scheme 1: Reagents and conditions: (a) NaBH₃CN, HCI-Et₂O, THF, MS-3Å, 5 °C, 1 h; (b) CH₃C(OEt)₃, *p*-TsOH, DMF, room temperature, 2 h, then H₂O, room temperature, 30 min, 72%; (c) 3, NOBF₄, CH₂Cl₂, - 20 °C, 45 min, 79%; (d) benzyl bromide, NaOH, TBAB, THF, room
temperature, 5 h, 90%; (e) NaBH₃CN, HCI-Et₂O, THF, MS-3Å, 5 °C, 1 h, 75%; (f) 4, NOBF₄, CH₂Cl₂-Et₂O (1:4 v/v), - 10 °C, 30 min, 75%; (g) HCIO₄-SiO₂, CH₃CN, room temperature, 25 min; (h) benzoyl cyanide, pyridine, CH₂Cl₂, 0 °C, 4 h, over all 76%.

Compound 13 was synthesised in a 3-step-one-pot-sequence ss starting from trisaccharide acceptor 12. Thus, acceptor 12 and 3-PMB protected L-fucosyl donor 5 were reacted in the presence of NIS and HClO₄-SiO₂ at low temperature giving the expected tetrasaccharide intermediate. Slowly raising the reaction temperature in the reaction vessel initiated the hydrolysis 60 (HClO₄-SiO₂) of the PMB group and produced the desired tetrasaccharide acceptor. The reaction mixture was once again cooled and a mixture of galactosamine donor 6 and fresh NIS was

added to eventually furnish pentasaccharide 13 in 65 % overall vield. Formation of compound 13 was unambiguously confirmed by NMR spectroscopy [signals at δ 5.52 (d, J = 7.5 Hz, H-1_E), 5.44 (d, J = 3.5 Hz, H-1_A), 5.08 (br s, H-1_D), 5.00 (d, J = 3.0 Hz, $_{5}$ H-1_C), 4.77 (d, J = 8.0 Hz, H-1_B) in ¹H NMR and δ 105.8 (C-1_B), 99.3 (C-1_c), 99.1 (C-1_A), 98.1 (C-1_E), 97.2 (C-1_D) in ¹³C NMR spectra]. Carrying out three reactions in one pot set up significantly reduced the number of steps. The PMB ether acted as an in situ removable temporary protecting group for the ¹⁰ hydroxy functionality.³⁰ The pentasaccharide derivative **13** was subjected to a sequence of reactions consisting of (a) treatment with hydrazine hydrate³¹ followed by acetylation using acetic anhydride and pyridine for the conversion of phthalimido group into acetamido group; (b) treatment with thioacetic $acid^{32}$ to 15 convert azido group to acetamido group; (c) removal of benzyl ethers and benzylidene acetal under a catalytic transfer hydrogenation condition using triethylsilane and 20%Pd(OH)-C³³ and finally (d) saponification using sodium methoxide³⁴ to furnish desired pentasaccharide PMP glycoside 1 in 52% over all 20 yield. NMR spectrum of compound 1 unambiguously supported its structure [signals at δ 5.51 (d, J = 3.5 Hz, H-1_A), 4.94 (d, J = $3.5 \text{ Hz}, \text{H-1}_{\text{C}}$), $4.88 \text{ (br s, H-1}_{\text{D}}$), $4.64 \text{ (d, } J = 8.0 \text{ Hz}, \text{H-1}_{\text{B}}$), 4.62

5.5 Hz, H-1_C), 4.88 (or s, H-1_D), 4.04 (d, J = 8.0 Hz, H-1_B), 4.02 (d, J = 7.5 Hz, H-1_E) in ¹H NMR and at δ 104.9 (C-1_B), 103.3 (C-1_E), 100.4 (C-1_D), 100.0 (C-1_C), 97.2 (C-1_A) in ¹³C NMR spectra] ²⁵ (Scheme 2).



Scheme 2: Reagents and conditions: (a) 5, NIS, HCIO₄-SiO₂, CH₂Cl₂, MS-4Å, – 45 °C, 30 min; then 10 °C, 30 min; then 6, NIS, – 30 °C, 1 h, 65%; (b) NH₂NH₂·H₂O, EtOH, 80 °C, 8 h; (c) acetic anhydride, pyridine, 30 room temperature, 1 h; (d) CH₃COSH, pyridine, room temperature, 18 h; (e) Et₃SiH, 20% Pd(OH)₂-C, CH₃OH, room temperature, 24 h; (f) 0.1 M CH₃ONa, CH₃OH, room temperature, 4 h, over all 52%.

Conclusions

In conclusion, a straightforward linear synthesis of a ³⁵ pentasaccharide has been developed applying a one pot reaction condition for two stereoselective glycosylation reactions and removal of PMB group *in situ*. High stereoselective outcome was observed in most of the glycosylation reactions. Thioglycosides and glycosyl trichloroacetimidate derivatives have been used in

⁴⁰ the glycosylation reactions using recently developed reaction conditions.

Experimental

General methods

- All reactions were monitored by thin layer chromatography over ⁴⁵ silica gel coated TLC plates. The spots on TLC were visualized by warming ceric sulphate (2% Ce(SO₄)₂ in 2N H₂SO₄) sprayed plates in hot plate. Silica gel 230-400 mesh was used for column chromatography. NMR spectra were recorded on Brucker Avance 500 MHz using CDCl₃ as solvent and TMS as internal reference ⁵⁰ unless stated otherwise. Chemical shift value is expressed in δ ppm. The complete assignment of proton and carbon spectra was carried out by using a standard set of NMR experiments, e.g. ¹H NMR, ¹³C NMR, ¹³C DEPT 135, 2D COSY and 2D HSQC etc. MALDI-MS were recorded on a Bruker Daltonics mass ⁵⁵ spectrometer. Optical rotations were recorded in a Jasco P-2000 spectrometer. Commercially available grades of organic solvents of adequate purity are used in all reactions.
- *p*-METHOXYPHENYL 4-O-ACETYL-2-AZIDO-6-O-BENZYL-2-DEOXY-α-D-GALACTOPYRANOSIDE (2): To 60 a solution of compound 7 (2 g, 5.01 mmol) in dry THF (15 mL) were added MS-3Å (2 g) and NaBH₃CN (1.8 g, 28.64 mmol) and the reaction mixture was stirred at 0 °C for 20 min. To the cold reaction mixture was added drop wise HCl in Et₂O (~ 10 mL) till the solution became acidic (pH \sim 2) and the reaction mixture was 65 allowed to stir at 5 °C for 1 h. The reaction mixture was filtered through a Celite bed and the filtering bed was washed with CH₂Cl₂ (100 mL). The combined filtrate was successively washed with satd. NaHCO3 and water, dried (Na2SO4) and concentrated. The crude product was purified over SiO₂ using 70 hexane-EtOAc (3:1) as eluent to give 3,4-diol derivative. To a solution of the diol derivative in anhydrous DMF (10 mL) were added CH₃C(OEt)₃ (3 mL, 16.36 mmol) and p-TsOH (250 mg) and the reaction mixture was allowed to stir at room temperature for 2 h. After complete consumption of the starting material
- ⁷⁵ (TLC; hexane:EtOAc 3:1), H_2O (10 mL) was added to the reaction mixture and it was stirred at room temperature for 30 min. The solvents were removed under reduced pressure and the crude product was purified over SiO₂ using hexane-EtOAc (4:1) as eluent to give pure compound **2** (1.6 g, 72%). White solid;
- ⁸⁰ m.p. 64-65 °C [EtOH]; $[\alpha]_D^{25}$ -11.7 (*c* 1.0, CHCl₃); IR (KBr): 3027, 2363, 2110, 1713, 1589, 1218, 1042, 760 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.24-7.14 (m, 5 H, Ar-H), 6.96 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.70 (d, *J* = 9.0 Hz, 2 H, Ar-H), 5.37 (d, *J* = 2.5 Hz, 1 H, H-4), 5.34 (d, *J* = 3.0 Hz, 1 H, H-1), 4.42 (d, *J* = 11.5
- ⁸⁵ Hz, 1 H, PhCH₂), 4.35 (dd, J = 10.5, 3 Hz, 1 H, H-3), 4.32 (d, J = 11.5 Hz, 1 H, PhCH₂), 4.25-4.22 (m, 1 H, H-5), 3.67 (s, 3 H, OCH₃), 3.48 (dd, J = 10.5, 3.0 Hz, 1 H, H-2), 3.43-3.41 (m, 2 H, H-6_{ab}), 2.01 (s, 3 H, COCH₃); ¹³C NMR (125 MHz, CDCl₃): δ 171.2 (COCH₃), 156.6-114.6 (Ar-C), 98.3 (C-1), 73.4 (PhCH₂),
- $_{90}$ 70.5 (C-4), 68.5 (C-5), 68.0 (C-6), 67.4 (C-3), 60.1 (C-2), 55.5 (OCH₃), 20.7 (COCH₃); ESI-MS: 466.1 [M+Na]⁺; Anal. Calcd. for C₂₂H₂₅N₃O₇ (443.17): C, 59.59; H, 5.68; found: C, 59.46; H, 5.85.
- *p*-METHOXYPHENYL (2,3-DI-*O*-ACETYL-4,6-*O*-95 BENZYLIDENE-β-D-GALACTOPYRANOSYL)-(1→3)-4-*O*-ACETYL-2-AZIDO-6-*O*-BENZYL-2-DEOXY-α-D-

GALACTOPYRANOSIDE (8): A solution of compound **2** (1.3 g, 2.93 mmol) and compound **3** (1.6 g, 3.22 mmol) in anhydrous CH_2Cl_2 (20 mL) was cooled to - 20 °C under argon. To the

cooled reaction mixture was added NOBF₄ (0.4 g, 3.42 mmol) and the reaction mixture was allowed to stir at same temperature for 45 min. The reaction mixture was diluted with CH₂Cl₂ (100 mL) and the organic layer was successively washed with satd. ⁵ NaHCO₃ and water, dried (Na₂SO₄) and concentrated. The crude product was purified over SiO₂ using hexane-EtOAc (5:1) as eluent to give pure compound **8** (1.8 g, 79%). White solid; m.p. 164-165 °C [EtOH]; $[\alpha]_D^{25}$ + 147.6 (*c* 1.0, CHCl₃); IR (KBr): 3027, 2935, 2366, 2100, 1755, 1378, 1218, 1042, 761 cm⁻¹; ¹H

- ¹⁰ NMR (500 MHz, CDCl₃): δ 7.43-7.16 (m, 10 H, Ar-H), 6.98 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.66 (d, *J* = 9.0 Hz, 2 H, Ar-H), 5.46 (d, *J* = 3.5 Hz, 1 H, H-4_A), 5.36 (s, 1 H, PhC*H*), 5.34 (d, *J* = 3.5 Hz, 1 H, H-1_A), 5.30 (dd, *J* = 7.5 Hz each, 1 H, H-2_B), 4.84 (dd, *J* = 8.5, 3.5 Hz, 1 H, H-3_B), 4.67 (d, *J* = 8.0 Hz, 1 H, H-1_B), 4.39-4.36 (m, 15 2 H, PhC*H*₂), 4.28-4.20 (m, 4 H, H-3_A, H-4_B, H-5_A, H-6_{aB}), 3.88-
- 3.85 (m, 1 H, H-6_{bB}), 3.70 (dd, J = 10.5, 3.5 Hz, 1 H, H-2_A), 3.66 (s, 3 H, OCH₃), 3.54 (dd, J = 10.5, 4.0 Hz, 1 H, H-6_{aA}), 3.43-3.38 (m, 2 H, H-5_B, H-6_{bA}), 2.03, 2.02, 2.00 (3 s, 9 H, 3 COCH₃); ¹³C NMR (125 MHz, CDCl₃): δ 170.6, 170.4, 169.1 (COCH₃), 155.7-
- ²⁰ 114.6 (Ar-C), 101.4 (PhCH), 101.2 (C-1_B), 98.7 (C-1_A), 75.2 (C-4_B), 73.4 (PhCH₂), 73.2 (C-5_A), 71.9 (C-3_A), 69.7 (C-4_A), 69.5 (C-6_A), 69.4 (C-3_B), 68.6 (C-2_B), 68.4 (C-6_B), 66.5 (C-5_B), 59.2 (C-2_A), 55.5 (OCH₃), 20.8 (2 C, 2 COCH₃), 20.7 (COCH₃); MALDI-MS: 800.2 [M+Na]⁺; Anal. Calcd. for C₃₉H₄₃N₃O₁₄
 ²⁵ (777.27); C, 60.23; H, 5.57; found: C, 60.09; H, 5.76.
- *p*-METHOXYPHENYL (2,3-DI-*O*-BENZYL-4,6-*O*-BENZYLIDENE- β -D-GALACTOPYRANOSYL)-(1 \rightarrow 3)-2-AZIDO-4,6-DI-*O*-BENZYL-2-DEOXY- α -D-
- **GALACTOPYRANOSIDE (9):** To a solution of compound **8** 30 (1.6 g, 2.06 mmol) in THF (25 mL) were added benzyl bromide (2.5 ml, 21.02 mmol), powdered NaOH (2 g, 50 mmol) and TBAB (100 mg) and the reaction mixture was allowed to stir at room temperature for 5 h. The reaction mixture was diluted with CH₂Cl₂ (100 mL) and the organic layer was washed with water,
- ³⁵ dried (Na₂SO₄) and concentrated. The crude product was purified over SiO₂ using hexane-Et₂O (8:1) as eluent to give pure compound **9** (1.7 g, 90%). Colorless oil; $[\alpha]_D^{25} + 137$ (*c* 1.0, CHCl₃); IR (neat): 3432, 3030, 2929, 2100, 1640, 1500, 1457, 1360, 1218, 1099, 1056, 697 cm⁻¹; ¹H NMR (500 MHz, CDCl₃):
- ⁴⁰ δ 7.37-7.11 (m, 25 H, Ar-H), 7.00 (d, J = 9.0 Hz, 2 H, Ar-H), 6.69 (d, J = 9.0 Hz, 2 H, Ar-H), 5.41 (s, 1 H, PhCH), 5.38 (d, J = 3.5 Hz, 1 H, H-1_A), 5.12 (d, J = 11.0 Hz, 1 H, PhCH₂), 5.01 (d, J= 11.0 Hz, 1 H, PhCH₂), 4.76 (d, J = 7.5 Hz, 1 H, H-1_B), 4.73-4.61 (m, 4 H, 2 PhCH₂), 4.35-4.25 (m, 3 H, H-3_A, PhCH₂), 4.20
- ⁴⁵ (d, J = 12.5 Hz, 1 H, H-6_{aB}), 4.16-4.13 (m, 1 H, H-5_A), 4.03 (d, J = 3.5 Hz, 1 H, H-4_B), 3.95 (d, J = 12.5 Hz, 1 H, H-6_{bB}), 3.84 (t, J = 7.5 Hz each, 1 H, H-2_B), 3.79-3.77 (m, 1 H, H-2_A), 3.67 (s, 3 H, OCH₃), 3.55 (dd, J = 10.0, 3.0 Hz, 1 H, H-3_B), 3.50-3.45 (m, 2 H, H-6_{abA}), 3.35-3.34 (m, 1 H, H-5_B); ¹³C NMR (125 MHz, CDCl₃):
- ⁵⁰ δ 155.5-114.6 (Ar-C), 105.2 (C-1_B), 101.5 (PhCH), 99.3 (C-1_A), 79.0 (C-3_B), 78.6 (C-2_B), 77.1 (C-4_B), 76.8 (C-4_A), 75.4 (PhCH₂), 75.1 (PhCH₂), 73.9 (C-3_A), 73.2 (PhCH₂), 72.4 (PhCH₂), 70.3 (C-5_A), 69.2 (C-6_A), 69.0 (C-6_B), 66.3 (C-5_B), 59.4 (C-2_A), 55.5 (OCH₃); MALDI-MS: 944.3 [M+Na]⁺; Anal. Calcd. for ⁵⁵ C₅₄H₅₅N₃O₁₁ (921.38): C, 70.34; H, 6.01; found: C, 70.18; H, 6.20.

p-METHOXYPHENYL (2,3,6-TRI-*O*-BENZYL- β -D-GALACTOPYRANOSYL)-(1 \rightarrow 3)-2-AZIDO-4,6-DI-*O*-

BENZYL-2-DEOXY-α-D-GALACTOPYRANOSIDE (10): Το

⁶⁰ a solution of compound **9** (1.6 g, 1.73 mmol) in dry THF (20 mL) were added MS-3Å (3 g) and NaBH₃CN (0.8 g, 12.73 mmol) and the reaction mixture was stirred at 0 °C for 20 min. To the cold reaction mixture was added drop wise HCl in Et₂O (\sim 7 mL) till the solution became acidic (pH \sim 2) and the reaction mixture was

- ⁶⁵ allowed to stir at 5 °C for 1 h. The reaction mixture was filtered through a Celite bed and the filtering bed was washed with CH₂Cl₂ (100 mL). The combined filtrate was successively washed with satd. NaHCO₃ and water, dried (Na₂SO₄) and concentrated. The crude product was purified over SiO₂ using ⁷⁰ hexane-EtOAc (6:1) as eluent to give pure compound **10** (1.2 g, 75%). Colorless oil; $[\alpha]_D^{25} + 67$ (*c* 1.0, CHCl₃); IR (neat): 3526,
- 2927, 2100, 1500, 1374, 1217, 1022, 697 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.32-7.10 (m, 25 H, Ar-H), 6.98 (d, J = 9.0 Hz, 2 H, Ar-H), 6.68 (d, J = 9.0 Hz, 2 H, Ar-H), 5.7 (d, J = 3.0 Hz, 1
- ⁷⁵ H, H-1_A), 4.99 (d, J = 11.0 Hz, 1 H, PhC H_2), 4.89 (d, J = 11.0 Hz, 1 H, PhC H_2), 4.66 (d, J = 7.5 Hz, 1 H, H-1_B), 4.65-4.57 (m, 4 H, PhC H_2), 4.43-4.40 (m, 2 H, PhC H_2), 4.35-4.24 (m, 3 H, H-3_A, PhC H_2), 4.15 (br s, 1 H, H-4_A), 4.13-4.10 (m, 1 H, H-5_A), 3.93 (d, J = 2.5 Hz, 1 H, H-4_B), 3.78-3.75 (m, 1 H, H-2_A), 3.73-3.70 (m, 1
- ⁸⁰ H, H-6_aA), 3.68 (s, 3 H, OCH₃), 3.67-3.57(m, 3 H, H-2_B, H-3_B, H-6_bA), 3.49-3.45 (m, 2 H, H-5_B, H-6_aA), 3.42-3.38 (m, 1 H, H-6_bA); ¹³C NMR (125 MHz, CDCl₃): δ 155.5-114.6 (Ar-C), 105.2 (C-1_B), 99.2 (C-1_A), 80.5 (C-3_B), 78.9 (C-2_B), 76.9 (C-4_B), 76.5 (C-4_A), 75.2 (PhCH₂), 74.7 (PhCH₂), 73.7 (PhCH₂), 73.2 ⁸⁵ (PhCH₂), 73.1 (C-3_A), 72.8 (PhCH₂), 70.3 (C-5_A), 69.3 (C-6_A), 69.1 (C-6_B), 66.8 (C-5_B), 59.5 (C-2_A), 55.4 (OCH₃); MALDI-MS:

946.3 $[M+Na]^+$; Anal. Calcd. for C₅₄H₅₇N₃O₁₁ (923.40): C, 70.19; H, 6.22; found: C, 70.05; H, 6.38.

- p-METHOXYPHENYL(2,3-DI-O-BENZYL-4,6-O-90 BENZYLIDENE-α-D-GLUCOPYRANOSYL)-(1→4)-(2,3,6-TRI-O-BENZYL-β-D-GALACTOPYRANOSYL)-(1→3)-2-AZIDO-4,6-DI-O-BENZYL-2-DEOXY-α-D-
- **GALACTOPYRANOSIDE (11):** A solution of compound **10** (1 g, 1.08 mmol) and compound **4** (0.7 g, 1.18 mmol) in anhydrous ⁹⁵ CH₂Cl₂-Et₂O (10 mL; 1:4 v/v) was cooled to – 10 °C under argon. To the cooled reaction mixture was added NOBF₄ (150 mg, 1.28 mmol) and the reaction mixture was allowed to stir at same temperature for 30 min. The reaction mixture was diluted with CH₂Cl₂ (100 mL) and the organic layer was successively ¹⁰⁰ washed with satd. NaHCO₃ and water, dried (Na₂SO₄) and concentrated. The crude product was purified over SiO₂ using hexane-EtOAc (5:1) as eluent to give pure compound **11** (1.1 g, 75%). Colorless oil; $[\alpha]_D^{25} + 65$ (*c* 1.0, CHCl₃); IR (neat): 3408, 3037, 2935, 2117, 1597, 1503, 1459, 1392, 1346, 1245, 1217, 1177, 1085, 1044, 998, 918, 697 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.38-6.97 (m, 42 H, Ar-H), 6.68 (d, *J* = 9.0 Hz, 2 H,
- CDCl₃): δ 7.38-6.97 (m, 42 H, Ar-H), 6.68 (d, J = 9.0 Hz, 2 H, Ar-H), 5.38 (s, 1 H, PhC*H*), 5.34 (d, J = 3.5 Hz, 1 H, H-1_A), 4.98 (d, J = 11.5 Hz, 1 H, PhC*H*₂), 4.91 (d, J = 11.5 Hz, 1 H, PhC*H*₂), 4.90 (d, J = 3.0 Hz, 1 H, H-1_C), 4.80 (d, J = 11.5 Hz, 1 H, 110 PhC*H*₂), 4.72 (d, J = 11.5 Hz, 1 H, PhC*H*₂), 4.70 (d, J = 8.0 Hz, 1
- H, H-1_B), 4.68-4.45 (m, 6 H, PhC H_2), 4.35-4.19 (m, 5 H, H-3_A, PhC H_2), 4.17-4.06 (m, 4 H, H-4_A, H-5_A, H-6_{abC}), 4.03-3.96 (m, 2 H, H-4_B, H-5_C), 3.82 (t, J = 8.5 Hz each, 1 H, H-4_C), 3.78 (t, J = 8.5 Hz each, 1 H, H-2_A), 3.67 (s, 3 H, CH) = 2.55 Hz each, 1 H, H-2_A), 3.55 Hz each, 1 H, H-2_A), 3.55 Hz each, 1 H, H
- ¹¹⁵ OCH₃), 3.58-3.52 (m, 2 H, H-3_B, H-6_{aA}), 3.47-3.40 (m, 4 H, H-5_B, H-6_{bA}, H-6_{abB}), 3.38-3.30 (m, 2 H, H-2_C, H-3_C); ¹³C NMR

(125 MHz, CDCl₃): δ 155.4-114.6 (Ar-C), 105.5 (C-1_B), 101.0 (PhCH), 100.6 (C-1_C), 99.3 (C-1_A), 82.8 (C-5_C), 80.6 (C-2_B), 79.7 (C-3_B), 79.3 (C-3_C), 78.4 (C-4_C), 77.1 (C-4_A),76.9 (C-4_B), 76.7 (C-5_B), 76.6 (C-5_C), 75.8 (C-3_A), 75.0 (PhCH₂), 74.9 (PhCH₂), 5 74.8 (PhCH₂), 74.2 (PhCH₂), 73.2 (PhCH₂), 73.1 (PhCH₂), 73.0 (PhCH₂), 72.0 (PhCH₂), 72.0 (PhCH₂), 72.0 (PhCH₂), 73.0 (PhCH₂

- $\begin{array}{l} (PhCH_2), \ 70.3 \ (C-5_A), \ 69.8 \ (C-6_A), \ 69.4 \ (C-6_B), \ 68.0 \ (C-6_C), \ 63.0 \\ (C-2_C), \ 59.3 \ (C-2_A), \ 55.5 \ (OCH_3); \ MALDI-MS: \ 1376.5 \ [M+Na]^+; \\ Anal. \ Calcd. \ for \ C_{81}H_{83}N_3O_{16} \ (1353.58): \ C, \ 71.82; \ H, \ 6.18; \\ found: \ C, \ 71.70; \ H, \ 6.35. \end{array}$
- ¹⁰ *p*-METHOXYPHENYL (6-*O*-BENZOYL-2,3-DI-*O*-BENZYL-α-D-GLUCOPYRANOSYL)-(1→4)-(2,3,6-TRI-*O*-BENZYL-β-D-GALACTOPYRANOSYL)-(1→3)-2-AZIDO-4,6-DI-*O*-BENZYL-2-DEOXY-α-D-

GALACTOPYRANOSIDE (12): To a solution of compound **11** ¹⁵ (1 g, 0.74 mmol) in CH₃CN (20 mL) was added HClO₄-SiO₂ (0.3 g) and the reaction mixture was stirred at room temperature for 25 min. The reaction mixture was filtered and concentrated under reduced pressure to give the 4,6-diol derivative. A solution of the diol derivative in CH₂Cl₂ (10 mL) was cooled to 0 °C. To the

- ²⁰ cooled reaction mixture were added pyridine (1 mL) and benzoyl cyanide (150 mg, mmol) and the reaction mixture was allowed to stir for 4 h at same temperature. The reaction mixture was poured into water and extracted with CH₂Cl₂ (50 mL). The organic layer was washed with satd. NaHCO₃, dried (Na₂SO₄) and
- ²⁵ concentrated. The crude product was purified over SiO₂ using hexane-EtOAc (7:1) as eluent to give pure compound **12** (770 mg, 76%). Colorless oil; $[\alpha]_D^{25} + 62$ (*c* 1.0, CHCl₃); IR (neat): 3370, 3036, 2929, 2116, 1739, 1600, 1488, 1454, 1365, 1340, 1236, 1217, 1100, 1049, 999, 918, 666 cm⁻¹; ¹H NMR (500 MHz,
- ³⁰ CDCl₃): δ 8.01-6.68 (m, 44 H, Ar-H), 5.36 (d, J = 3.0 Hz, 1 H, H-1_A), 5.05-4.95 (m, 2 H, PhCH₂), 4.89 (br s, 1 H, H-1_C), 4.79-4.76 (m, 1 H, PhCH₂), 4.70-4.69 (m, 1 H, PhCH₂), 4.68 (d, J = 3.0 Hz, 1 H, H-1_B), 4.68-4.60 (m, 2 H, PhCH₂), 4.60-4.55 (m, 2 H, PhCH₂), 4.55-4.49 (m, 2 H, PhCH₂), 4.35-4.30 (m, 3 H, H-3_A,
- ³⁵ PhC H_2), 4.30-4.20 (m, 4 H, H-4_A, H-5_A, PhC H_2), 4.13-4.11 (m, 2 H, H-5_C, H-6_aC), 4.02-3.98 (m, 2 H, H-4_B, H-4_C), 3.77-3.75 (m, 3 H, H-2_A, H-2_B, H-6_bC), 3.67 (s, 3 H, OC H_3), 3.65-3.64 (m, 1 H, H-6_aB), 3.56 (m, 2 H, H-3_B, H-6_aA), 3.45-3.43 (m, 3 H, H-5_B, H-6_bA, H-6_bB), 3.35-3.33 (m, 2 H, H-2_C, H-3_C); ¹³C NMR (125
- ⁴⁰ MHz, CDCl₃): δ 167.0 (PhCO), 155.4-114.6 (Ar-C), 105.7 (C-1_B), 99.7 (C-1_C), 99.3 (C-1_A), 81.5 (C-2_B), 80.3 (C-2_C), 80.2 (C-3_C), 78.6 (C-4_C), 77.3 (C-4_A), 76.7 (C-4_B), 75.5 (C-3_A), 75.3 (PhCH₂), 74.9 (2 C, 2 PhCH₂), 73.8 (PhCH₂), 73.2 (2 C, C-5_A, PhCH₂), 73.1 (2 C, 2 PhCH₂), 70.6 (C-5_B), 70.3 (2 C, C-3_B, C-
- $_{45}$ 5_C), 69.3 (C-6_A), 67.9 (C-6_B), 63.0 (C-6_C), 59.3 (C-2_A), 55.5 (OCH₃); MALDI-MS: 1392.5 [M+Na]⁺; Anal. Calcd. for C₈₁H₈₃N₃O₁₇ (1369.57): C, 70.98; H, 6.10; found: C, 70.82; H, 6.28.

p-METHOXYPHENYL (3-O-ACETYL-4,6-O-50 BENZYLIDENE-2-DEOXY-2-PHTHALIMIDO-β-D-GALACTOPYRANOSYL)-(1→3)-(2-O-ACETYL-4-O-BENZYL-α-L-RHAMNOPYRANOSYL)-(1→4)-(6-O-BENZOYL-2,3-DI-O-BENZYL-α-D-GLUCOPYRANOSYL)-(1→4)-(2,3,6-TRI-O-BENZYL-β-D-

55 GALACTOPYRANOSYL)-(1→3)-2-AZIDO-4,6-DI-O-BENZYL-2-DEOXY-α-D-GALACTOPYRANOSIDE (13): A solution of compound 12 (700 mg, 0.51 mmol), compound 5 (250 mg, 0.54 mmol) and MS-4Å (2 g) in anhydrous CH₂Cl₂ (10 mL)

was cooled to -45 °C under argon. To the cooled reaction 60 mixture were added NIS (130 mg, 0.58 mmol) and HClO₄-SiO₂ (100 mg) and it was allowed to stir at same temperature for 30 min. After consumption of the starting materials (TLC; hexane-EtOAc, 2:1) the temperature of the reaction mixture was raised to 10 °C and stirred for 30 min. After formation of a new spot in 65 TLC (hexane-EtOAc, 2:1) again the reaction mixture was cooled to - 30 °C. To the cooled reaction mixture were added a solution of compound 6 (240 mg, 0.50 mmol) in CH₂Cl₂ (2 mL) and NIS (115 mg, 0.51 mmol) and the reaction mixture was allowed to stir at - 30 °C for another 1 h. The reaction mixture was filtered 70 through a Celite bed and the filtering bed was washed with CH₂Cl₂ (50 mL). The combined organic layer was successively washed with 5% Na2S2O3, satd. NaHCO3 and water, dried (Na₂SO₄) and concentrated. The crude product was purified over SiO₂ using hexane-EtOAc (5:1) as eluent to give pure compound 75 **13** (685 mg, 65%). Colorless oil; $[\alpha]_D^{25}$ + 14 (*c* 1.0, CHCl₃); IR (neat): 3089, 2866, 1722, 1625, 1524, 1377, 1242, 1176, 1097, 1076, 989, 698 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 8.10-6.76 (m, 58 H, Ar-H), 5.52 (d, J = 7.5 Hz, 1 H, H-1_E), 5.50 (br s, 1 H, $H-2_{D}$), 5.44 (d, J = 3.5 Hz, 1 H, $H-1_{A}$), 5.37 (s, 1 H, PhCH), 5.14 ⁸⁰ (t, J = 9.0 Hz each, 1 H, H-3_E), 5.12-5.10 (m, 1 H, PhCH₂), 5.08 (br s, 1 H, H-1_D), 5.00 (d, J = 3.0 Hz, 1 H, H-1_C), 4.92-4.79 (3 d, $J = 11.0 \text{ Hz each}, 3 \text{ H}, \text{PhC}H_2$, 4.77 (d, $J = 8.0 \text{ Hz}, 1 \text{ H}, \text{H-1}_B$), 4.75-4.72 (m, 2 H, H-2_E, PhCH₂), 4.66-4.50 (m, 4 H, 2 PhCH₂), 4.46-4.37 (m, 4 H, PhCH₂, H-6_{aE}), 4.34-4.28 (m, 5 H, H-3_A, H-85 6_{aC}, H-5_A, PhCH₂), 4.25-4.17 (m, 5 H, H-4_A, H-5_C, H-4_E, PhCH₂), 4.10-4.01 (m, 4 H, H-4_B, H-4_C, H-4_D, H-6_{bE}), 3.89-3.81 (m, 6 H, H-3_D, H-2_A, H-2_B, H-6_{bC}, H-5_D, H-5_E), 3.74 (s, 3 H, OCH₃), 3.64-3.58 (m, 2 H, H-3_B, H-6_{aA}), 3.55-3.52 (m, 3 H, H-6_{aB}, H-6_{bA}, H-5_B), 3.45-3.42 (m, 1 H, H-6_{bB}), 3.38-3.35 (m, 2 H, H-2_C, H-3_C), $_{90}$ 2.01, 1.86 (2 s, 6 H, 2 COCH₃), 0.89 (d, J = 4.0 Hz, 3 H, CCH₃);

⁹⁰ 2.01, 1.86 (2 s, 6 H, 2 COCH₃), 0.89 (d, J = 4.0 Hz, 3 H, CCH₃); ¹³C NMR (125 MHz, CDCl₃): δ 172.0, 171.0 (2 COCH₃), 168.0, 167.0 (PhthCO), 166.0 (PhCO), 155.5-114.6 (Ar-C), 105.8 (C-1_B), 100.8 (PhCH), 99.3 (C-1_C), 99.1 (C-1_A), 98.1 (C-1_E), 97.2 (C-1_D), 81.3 (C-2_B), 79.8 (C-2_C), 79.7 (C-3_C), 79.6 (C-4_C), 79.3 ⁹⁵ (C-3_E), 79.0 (C-5_D), 78.7 (C-3_D), 77.6 (C-4_A), 76.8 (C-4_B), 76.1 (C-4_E), 75.3 (C-5_A), 75.1 (C-3_A), 75.0 (PhCH₂), 74.7 (PhCH₂), 74.3 (2 C, 2 PhCH₂), 73.8 (PhCH₂), 73.2 (PhCH₂), 73.0 (PhCH₂), 72.6 (C-2_D), 71.2 (PhCH₂), 70.3 (C-5_B), 69.4 (C-6_E), 69.3 (C-6_A), 69.0 (C-3_B), 68.7 (C-5_C), 67.9 (C-6_B), 67.8 (C-4_D), 65.8 (C-5_E), 100 62.6 (C-6_C), 59.3 (C-2_A), 55.5 (OCH₃), 51.1 (C-2_E), 21.0, 20.7 (2 C, COCH₃), 17.6 (CCH₃); MALDI-MS: 2091.7 [M+Na]⁺; Anal. Calcd. for C₁₁₉H₁₂₀N₄O₂₉ (2068.80): C, 69.04; H, 5.84; found: C, 68.86; H, 6.00.

p-METHOXYPHENYL (2-ACETAMIDO-2-DEOXY- β -D-105 GALACTOPYRANOSYL)-(1 \rightarrow 3)-(α -L-

RHAMNOPYRANOSYL)-(1→4)-(α-D-

GLUCOPYRANOSYL)-(1→4)-(β-D-

GALACTOPYRANOSYL)-(1→3)-2-ACETAMIDO-2-

DEOXY-α-D-GALACTOPYRANOSIDE (1): A solution of ¹¹⁰ compound **13** (600 mg, 0.29 mmol) and NH₂NH₂·H₂O (0.2 mL) in EtOH (10 mL) was stirred at 80 °C for 8 h. The solvents were removed under reduced pressure and a solution of the crude product in acetic anhydride-pyridine (2 mL, 1:1 v/v) was kept at room temperature for 1 h. The solvents were removed under ¹¹⁵ reduced pressure and the crude product was dissolved in pyridine (1 mL). Thioacetic acid (0.2 mL) was added to the reaction mixture and it was allowed to stir at room temperature for 18 h. The solvents were removed under reduced pressure and the crude product was passed through a short pad of SiO₂. To a solution of the *N*-acetylated product in CH₃OH (5 mL) was added 20% 5 Pd(OH)₂-C (100 mg) and Et₃SiH (1 ml, 6.26 mmol) and it was

- stirred at room temperature for 24 h. The reaction mixture was filtered through a Celite bed and the filtering bed was washed with CH₃OH (50 mL). The combined filtrate was concentrated under reduced pressure. A solution of the hydrogenolyzed
- ¹⁰ product in CH₃ONa (5 mL; 0.1 M in CH₃OH) was stirred at room temperature for 4 h. The reaction mixture was neutralized with Dowex 50W X8 (H⁺) resin, filtered and concentrated. The deprotected product was passed through a Sephadex LH-20 column using CH₃OH-H₂O (3:1) as eluent to give pure compound
- ¹⁵ **1** (150 mg, over all 52%). White powder; $[\alpha]_D^{25} + 54$ (*c* 0.5, H₂O); IR (KBr): 3436, 2942, 1619, 1400, 1157, 1096, 669 cm⁻¹; ¹H NMR (500 MHz, D₂O): δ 7.13 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.98 (d, *J* = 9.0 Hz, 2 H, Ar-H), 5.51 (d, *J* = 3.5 Hz, 1 H, H-1_A), 4.94 (d, *J* = 3.5 Hz, 1 H, H-1_C), 4.88 (br s, 1 H, H-1_D), 4.64 (d, *J*
- ²⁵ H-5_E, H-6_{abA}, H-6_{bB}, H-6_{abC}, H-6_{abE}), 3.63-3.60 (m, 2 H, H-2_B, H-4_C), 3.59-3.50 (m, 3 H, H-2_C, H-3_C, H-4_D), 2.06, 2.05 (2 s, 6 H, 2 COCH₃), 1.26 (d, J = 6.0 Hz, 3 H, CCH₃); ¹³C NMR (125 MHz, D₂O): δ 175.1, 175.0 (2 COCH₃), 155.2-115.0 (Ar-C), 104.9 (C-1_B), 103.3 (C-1_E), 100.4 (C-1_D), 100.0 (C-1_C), 97.2 (C-1_A), 80.0
- ³⁰ (C-3_D), 77.4 (C-3_A), 76.9 (C-4_C), 76.8 (C-4_B), 75.1 (C-4_E), 74.8 (C-5_E), 72.0 (C-3_B), 71.9 (C-3_C), 71.4 (C-2_B), 71.3 (C-2_C), 70.9 (C-2_D), 70.8 (2 C, C-3_E, C-4_D), 70.4 (C-5_C), 70.3 (C-5_D), 69.3 (C-5_A), 68.8 (C-5_B), 67.7 (C-4_A), 60.9 (2 C, C-6_A, C-6_E), 59.9 (C-6_C), 59.6 (C-6_B), 55.8 (OCH₃), 52.5 (C-2_E), 48.8 (C-2_A), 22.2, 22.0 (2
- $_{35}$ COCH3), 16.5 (CCH3); MALDI-MS: 1023.3 [M+Na]^+; Anal. Calcd. for $C_{41}H_{64}N_2O_{26}$ (1000.37): C, 49.20; H, 6.44; found: C, 49.00; H, 6.60.

Acknowledgements

I. B. thanks Council of Scientific and Industrial Research (CSIR), ⁴⁰ India for providing Junior Research Fellowship. This work was supported by CSIR, India (AKM) [Project No. 02(0038)/11/EMR-II]. The authors thank D. Dhara and M. Jana for their technical assistance.

Notes and references

- ⁴⁵ ^a Bose Institute, Division of Molecular Medicine, P-1/12, C.I.T. Scheme VII-M, Kolkata-700054, India; Fax: 91-33-2355 3886; Tel: 91 33-2569 3240; E-mail: <u>akmisra69@gmail.com</u>
- † Electronic Supplementary Information (ESI) available: [Copies of 1D and 2D NMR spectra of compounds 2, 8-13, and 1]. See 50 DOI: 10.1039/b000000x/
- (a) I. Ørskov, F. Ørskov, B. Jann, K. Jann, *Bacteriol. Rev.* 1977, 41, 667; (b) J. R. Johnson, A. L. Stell, *J. Infect, Dis.* 2000, 181, 261; (c) P. B. Eckburg, E. M. Bik, C. N. Bernstein, E. Purdom, L. Dethlefsen, M. Sargent, S. R. Gill, K. E. Nelson, D. A. Relman, *Science* 2005, 308, 1635.
- 2. J. R. Johnson, Clin. Microbiol. Rev. 1991, 4, 80.
- R. K. Selander, T. K. Korhonen, V. Vaisanen-Rhen, P. H. Williams, P. E. Pattison, D. A. Caugant, *Infect. Immun.* 1986, 52, 213.

- 4. E. Bingen, B. Picard, N. Brahimi, S. Mathy, P. Desjardins, J. Elion,
- E. Denamur, J. Infect. Dis. 1998, 177, 642.
 M. M. Levine, J. Infect. Dis. 1987, 155, 377.
 - 5. M. M. Levine, J. Inject. Dis. 1987, 155, 577. 6. P. Stenutz, A. Weintraub, G. Widmalm, EEMS M.
 - R. Stenutz, A. Weintraub, G. Widmalm, *FEMS Microbial. Rev.* 2006, 30, 382.
- B. Olesen, C. Jensen, K. Olsen, V. Fussing, P. Gerner-Smidt, F. Scheutz, Scand. J. Infect. Dis. 2005, 37, 288.
- B. Foxman, S. D. Manning, P. Tallman, R. Bauer, L. Zhang, J. S. Koopman, B. Gillespie, J. D. Sobel, C. F. Marrs, *Am. J. Epidemiol.* 2002, **156**, 1133.
- 9. M. Ruth Leslie, H. Parolis, L. A. S. Parolis, *Carbohydr. Res.* 2000, **323**, 103.
- 10. C. M. Taylor, I. S. Roberts, Contrib. Microbiol. 2005, 12, 55.
- (a) M. S. Artenstein, R. Gold, J. G. Zimmerly, F. A. Wyle, H. Schneider, C. Harkins, N. Eng. J. Med. 1970, 282, 417; (b) W. Zou, H. J. Jennings, Preparation of Glycoconjugate Vaccines, in
- 75 Carbohydrate-Based Vaccines and Immunotherapies (Z. Guo, G.-J. Boons, Eds.), 2009, John Wiley & Sons, Inc., Hoboken, NJ, USA, DOI: 10.1002/9780470473283.ch2.
 - (a) B. Kuberan, R. J. Linhardt, Curr. Org. Chem. 2000, 4, 653; (b) R. Roy, Drug Discovery Today: Technol. 2004, 1, 327.
- 80 13. R. Dagan, J. Poolman, C. A. Siegrist, Vaccine 2010, 28, 5513.
 - 14. J. F. G. Vliegenthart, FEBS Lett. 2006, 580, 2945.
- (a) T. Ghosh, A. Santra, A. K. Misra, *RSC Adv.* 2014, **4**, 54; (b) D. Dhara, R. K. Kar, A. Bhunia, A. K. Misra, *Eur. J. Org. Chem.* 2014, 4577; (c) M. Jana, A. Sau, A. K. Misra, *Tetrahedron: Asymm.* 2014, 5
 25, 632; (d) M. Jana, R. K. Kar, A. Bhunia, A. K. Misra, *RSC Adv.*
- 2014, 4, 37079.
 16. D. B. Werz, P. H. Seeberger, Angew. Chem. Int. Ed. Engl. 2005, 44, 6315.
- 17. S. Figueroa-Pérez, V. Vérez-Bencomo, *Carbohydr. Res.* 1999, **317**, 29.
- L. J. Liotta, R. D. Capotosto, R. A. Garbitt, B. M. Horan, P. J. Kelly, A. P. Koleros, L. M. Brouillette, A. M. Kuhn, S. Targontsidis, *Carbohydr. Res.* 2001, **331**, 247.
- 19. C. Mukherjee, A. K. Misra, Glycoconjugate. J. 2008, 25, 611.
- 95 20. R. Kumar, P. R. Maulik, A. K. Misra, *Glycoconjugate J.* 2008, 25, 511.
- 21. C. Mukherjee, A. K. Misra, Tetrahedron: Asymm. 2008, 19, 2746.
- 22. P. J. Garegg, H. Hultberg, Carbohydr. Res. 1981, 93, C10.
- 23. H. Paulsen, V. Rutz, I. Bockhansen, Liebigs Ann. Chem. 1992, 747.
- 100 24. A. Sau, A. Santra, A. K. Misra, Synlett 2012, 2341.
- 25. S. K. Madhusudan, G. Agnihotri, D. S. Negi, A. K. Misra, *Carbohydr. Res.* 2005, **340**, 1373.
- 26. A. K. Chakraborti, R. Gulhane, Chem. Commun. 2003, 1896.
- 27. G. Agnihotri, A. K. Misra, Tetrahedron Lett. 2006, 47, 3653.
- 105 28. S. A. Abbas, A. H. Haines, Carbohydr. Res. 1975, 39, 358.
- B. Mukhopadhyaya, B. Collet, R. A. Field, *Tetrahedron Lett.* 2005, 46, 5923.
- S. Bhattacharyya, B. G. Magnusson, U. Wellmar, U. J. Nilsson, J. Chem. Soc., Perkin Trans 1, 2001, 886.
- 110 31. H.-H. Lee, D. A. Schwartz, J. F. Harris, J. P. Carver, J. J. Krepinsky, *Can. J. Chem.* 1986, 64, 1912.
 - 32. Y. Nakahara, T. Ogawa, Carbohydr. Res. 1996, 292, 71.
 - 33. A. Santra, T. Ghosh, A. K. Misra, *Beilstein J. Org, Chem.* 2013, 9, 74.
- 115 34. G. Zemplén, Ber. Dtsch. Chem. Ges. 1926, 59, 1254.