

RSC Advances



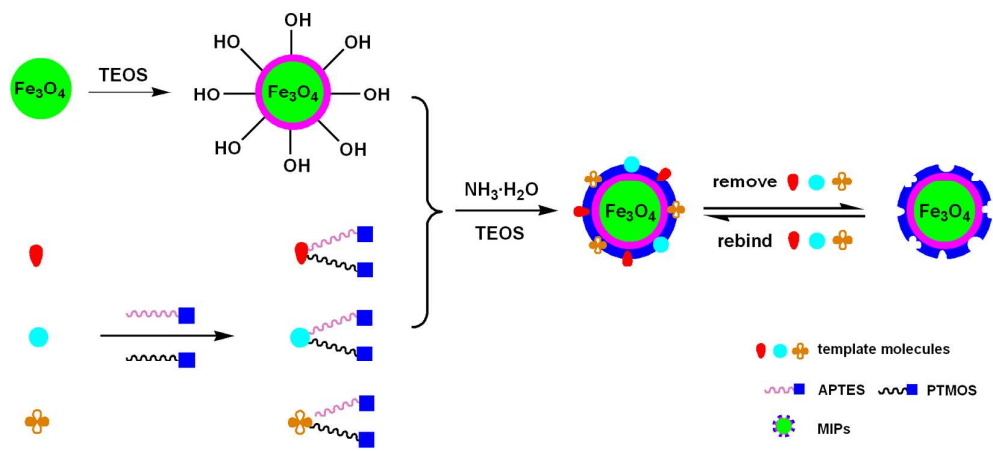
This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

A novel type of superparamagnetic molecularly imprinted polymers introducing unique concept of multi-template for specific separation and determination of three endocrine disrupting compounds (EDCs) simultaneously.



Novel magnetic multi-template molecularly imprinted polymers for specific separation and determination of three endocrine disrupting compounds simultaneously in environmental water samples

Ruixia Gao,*^a Yi Hao,^{a,b} Siqi Zhao,^a Lili Zhang,^{a,b} Xihui Cui,^{a,b} Dechun Liu,^c Yuhai Tang*^{a,b} and Yuansuo Zheng^a

^a *Institute of Analytical Science, School of Science, Xi'an Jiaotong University, Xi'an 710049, China.*

^b *College of Pharmacy, Xi'an Jiaotong University, Xi'an 710061, China.*

^c *Department of Hepatobiliary Surgery, First Hospital of Xi'an Jiaotong University Xi'an 710061, China*

* Corresponding authors: Tel.: +86 29 82655399; fax: +86 29 82655399.

E-mail: ruixiagao@mail.xjtu.edu.cn (R. Gao); tyh57@mail.xjtu.edu.cn (Y. Tang).

Abstract

In order to improve the practical applied value of molecularly imprinted polymers, a novel concept of multiple templates was introduced to prepare an original type of magnetic molecularly imprinted polymers. The magnetic multi-template molecularly imprinted polymers were obtained by selecting silica-coated magnetic nanoparticles as supporters, three endocrine disrupting compounds (17 β -estradiol (E2), estriol (E3), and diethylstilbestrol (DES)) as the multi-template, and two kinds of silane coupling agents (3-aminopropyltriethoxysilane (APTES) and phenyltrimethoxysilane (PTMOS)) as bifunctional monomers for simultaneously specific recognition of E2, E3, and DES. The as-synthesized polymers possessed homogeneous imprinting shells, stable crystalline phase, and super-paramagnetic property. Meanwhile, the imprinted nanomaterials displayed not only extraordinarily fast kinetics, but also satisfactory adsorption capacity, as well as favorable selectivity. More importantly, the prepared polymers exhibited similar recognition performance with that of physical mixture of three single-template polymers, but the synthetic procedure of the former was simplified with significant reduction in both preparation time and solvent consumption. In addition, the imprinted nanoparticles were applied as a specific adsorbent coupled with HPLC-UV for rapid isolation and determination of E2, E3, and DES simultaneously. The limits of detection (LODs) of proposed method for three target estrogens of E2, E3, and DES were 0.27, 0.19, and 0.08 ng mL⁻¹, respectively, which were lower than that obtained by some other sample pretreatment methods followed by HPLC-UV analysis. Furthermore, the developed method was

successfully applied for detection of multiple aimed estrogens in environmental water
24 samples with satisfactory recoveries in the range of 92.3-98.6%.

Introduction

Endocrine disrupting compounds (EDCs) are recognized as a diverse group of
27 substances that interfere with central regulatory functions in human and wildlife by
antagonizing or mimicking the effects of endogenous hormones.¹⁻³ EDCs can
bio-accumulate in the food chain and disrupt the synthesis, secretion, transport, and
30 elimination of natural hormones in the body of humans and animals.^{4,5} With the rapid
development of human life, the cosmetics, contraceptives, and hormone replacement
therapy drugs use increase dramatically, which together with their metabolites and
33 transformation products enter the human living environment, particularly the aquatic
environment.⁶⁻⁸ There has been much evidence manifesting that the accumulation of
the natural or synthetic EDCs may cause feminization of fish and raise many diseases
36 such as breast and prostate cancers of humans.^{9,10} What's more, some works reported
that EDCs could lead to slow development in infants and inferior quantity and quality
of human sperm.^{11,12} Therefore, accurate and reliable analysis of EDCs in complex
39 environmental matrices at low concentration levels is of importance for human health
and reproduction. The most commonly used methods for the determination of EDCs
are immunological methods^{13,14} and chromatography hyphenated technology
42 including GC-MS^{15,16} and LC-MS.^{17,18} Immunological methods are usually highly
selective for aimed compound but the instability and complicated production of
natural antibodies limit their applications to some extent. Chromatography

45 hyphenated methods can effectively detect estrogens, however, the complex
pretreatment process is required and the routine sample preparation technique
generally lacks selectivity.¹⁹ To solve these problems, several novel pretreatment
48 approaches using molecularly imprinted polymers (MIPs) as an efficient alternative
solid phase extractant for separation of EDCs have been established.^{20, 21}

MIPs are artificial polymers that possess specific recognition sites complementary
51 to the shape, size, and functional groups of predetermined analyte molecule. The
advantages of MIPs over natural receptors include ease of preparation, low cost, and
high mechanical stability, which have led them to be applied in a variety of fields,
54 such as chromatography,²² chemical sensors,²³ catalysis,²⁴ and solid-phase
extraction.²⁵ Among these applications, MIPs used as sorbents for the solid phase
extraction procedure are of desirableness for the selective removal or determination of
57 pollutants from matrix samples. When Fe₃O₄ magnetic nanoparticles act as nano-sized
supporters, the resultant MIPs not only have strong special recognition and adsorption
ability, but also can be easily removed from the adsorbed solution in the presence of
60 an external magnetic field.²⁶ Therefore, Fe₃O₄ magnetic nanoparticles have been
introduced to combine with the surface imprinting technique.

Currently, most of the imprinted polymers were prepared involving single template,
63 whose recognition sites only for one template molecule, and could not exhibit high
affinity and selectivity for a family of analytes. For another, the detection of only one
analyte in complex real samples is of little practical value. To address these problems
66 and expand the applicability of MIPs, some MIPs using two or multiple templates

were prepared to shorten analytical time and economize the cost.^{27,28} Although this purpose can also be achieved by physically mixing the individually imprinted polymer particles, this method requires for synthesizing several polymers and demands on much time. Though there were many publications reporting on the separation/enrichment of EDCs based on MIPs by using a single estrogen as template,^{29,30} to the best of our knowledge, magnetic multi-template molecularly imprinted polymers for simultaneous determination of three EDCs has not been reported.

In this work, we prepared magnetic multi-template molecularly imprinted polymers combining the merits of surface imprinting technique, sol-gel approach, and magnetic separation for simultaneously selective recognition E2, E3, and DES. The morphology and composition of the resulting products were characterized by TEM, FT-IR, VSM, and XRD. The obtained imprinted polymers exhibited fast kinetics, excellent specific recognition, and favorable selective affinity towards three templates, and also had satisfactory regeneration potential. The adsorption capabilities of the resulting polymers were also compared with the mixture of three imprinted polymers using E2, E3, and DES as single template respectively. The resultant imprinted nanomaterials were coupled with HPLC-UV for the determination of three EDCs using spiked environmental water samples at trace concentration levels.

Experimental

87 Chemicals and reagents

Tetraethoxysilane (TEOS), APTES, and PTMOS were purchased from Alfa Aesar Chemical Company. Ferric chloride hexahydrate ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$), anhydrous sodium acetate (NaOAc), ethylene glycol, ethanol, methanol, polyvinyl pyrrolidone (PVP), ammonium hydroxide (25%), and acetic acid were from Xi'an Chemicals Ltd. Estrone (E1), E2, E3, DES, bisphenol A (BPA), bisphenol F (BPF), and 17α -ethynylestradiol (EE2) (**Fig.1**) were obtained from Sigma. The highly purified water ($18.0 \text{ M}\Omega \text{ cm}^{-1}$) was obtained from a WaterPro water system (Axlwater Corporation, TY10AXLC1805-2, China) and used throughout the experiments. Environmental water samples were collected from local lake and river and filtered through $0.22 \mu\text{m}$ filters, stored in precleaned glass bottles. All reagents used were of at least analytical grade.

99 **Instruments and HPLC analysis**

A Tecnai G2 T2 S-TWIN transmission electron microscope and an SS-550 scanning electron microscopy (SEM) were used to examine TEM and SEM images of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$. The infrared spectra were recorded with Nicolet AVATAR-330 Fourier transform infrared (FT-IR) spectrometer, the magnetic properties were measured with a vibrating sample magnetometer (VSM) (LDJ 9600-1, USA), and the crystalline phases were characterized by a Rigaku D/max/2500v/pc (Japan) X-ray diffractometer with a $\text{Cu K}\alpha$ radiation.

The HPLC analyses were performed on a Hitachi L-2130 HPLC system including a binary pump and a variable wavelength UV detector. A Shimadzu VP-ODS C18 ($5 \mu\text{m}$ particle size, $150 \text{ mm} \times 4.6 \text{ mm}$) analytical column was used for analyte separation.

The mobile phase was methanol-H₂O (60/40, v/v) delivered at a flow rate of 1.0 mL
111 min⁻¹. The injection volume was 20 μL, and the column effluent was monitored at 280
nm.

Synthesis of Fe₃O₄@Multi-MIPs and reference polymers

114 The monodispersed Fe₃O₄ and the silica-modified Fe₃O₄ (denoted as Fe₃O₄@SiO₂)
were prepared as our previous work.²⁶ The multi-template imprinted polymers
(denoted as Fe₃O₄@Multi-MIPs) were prepared *via* a sol-gel process. 30 mg of each
117 E2, E3, and DES were dissolved in 20 mL of ethanol, and mixed with 100 μL of
APTES and 100 μL of PTMOS. The mixture was stirred for 2 h to form
template-monomer complex. Then, 0.2 g of Fe₃O₄@SiO₂, 400 μL of TEOS, and 600
120 μL of ammonium hydroxide (25%) were added to the reaction mixture, which was
allowed to proceed for 6 h. The obtained products were rinsed with highly purified
water until the supernatant was neutral, and the mixture of methanol/HAc (95:5, v/v)
123 was added as an eluent to remove the templates at room temperature. The resulting
imprinted polymers were collected by an external magnetic field and repeatedly
washed with highly purified water, and then dried under vacuum. In parallel,
126 Fe₃O₄@E2-MIPs, Fe₃O₄@E3-MIPs, Fe₃O₄@DES-MIPs, and Fe₃O₄@NIPs were
prepared by the same procedure with adding the template E2, E3, DES, and in the
absence of the template EDCs, respectively.

129 Binding experiments of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs

To investigate the adsorption kinetics of the obtained imprinted polymers, 20 mg of
Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs were mixed with 10 mL of ethanol solution of

132 three EDCs at a concentration of $30 \mu\text{g mL}^{-1}$, and the mixture was incubated at regular
time intervals from 1 min to 6 min at room temperature. Then, the polymers were
separated by a magnet and the concentrations of three EDCs in the supernatant were
135 determined by HPLC-UV. The adsorption capacity (Q , mg g^{-1}) for three EDCs bound
to the imprinted polymers was calculated as following equation:

$$Q = \frac{(C_0 - C_1)V}{W} \quad \mathbf{1}$$

138 Where C_0 and C_1 ($\mu\text{g mL}^{-1}$) represent the initial and the free solution concentration
of EDCs, respectively. V (mL) represents the volume of the solution and W (mg)
represents the weight of the polymer.

141 To determine the isothermal adsorption capacity of the adsorbents, 20 mg of
 $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ were added to 10 mL of ethanol solution at
varied initial concentration ($0.50\text{-}50 \mu\text{g mL}^{-1}$) of three EDCs. Then, the
144 $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ were isolated by a magnet and three EDCs residues in the
supernatant were determined by HPLC-UV.

To evaluate the specific recognition of the imprinted nanomaterials, 10 mL of
147 mixed standard solution of E2, E3, DES, E1, EE2, BPA, and BPF at initial
concentration of $30 \mu\text{g mL}^{-1}$ was incubated with 20 mg $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ or
 $\text{Fe}_3\text{O}_4@\text{NIPs}$ for 4 min, then the separation and detection procedures were conducted
150 as described earlier in the isothermal adsorption experiment.

Recognition performance of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and reference polymers

To compare the templates recognition of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ with reference polymers
153 ($\text{Fe}_3\text{O}_4@\text{NIPs}$, $\text{Fe}_3\text{O}_4@\text{E2-MIPs}$, $\text{Fe}_3\text{O}_4@\text{E3-MIPs}$, $\text{Fe}_3\text{O}_4@\text{DES-MIPs}$, and the

156 $\text{Fe}_3\text{O}_4@\text{E2-MIPs}/\text{Fe}_3\text{O}_4@\text{E3-MIPs}/\text{Fe}_3\text{O}_4@\text{DES-MIPs}$ (1:1:1, w/w/w) mixture
(denoted as M-MIPs), 20 mg of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and reference polymers were
incubated with the mixture of E2, E3, and DES respectively at a concentration of 30
 $\mu\text{g mL}^{-1}$ in 10 mL of ethanol at room temperature for 4 min. Then the extraction and
detection procedures were conducted as described earlier adsorption experiment.

159 **Reproducibility and reusability of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$**

20 mg of six batches of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ prepared on different
days were added to the solution of three EDCs in 10 mL of ethanol at a concentration
162 of 30 $\mu\text{g mL}^{-1}$ to investigate their reproducibility. After incubation for 4 min at room
temperature, the supernatants and polymers were separated by a magnetic field and
the bound amounts of three EDCs were measured by HPLC-UV.

165 To estimate the reusability of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$, 20 mg of
polymers were added to the solution of three EDCs in 10 mL of ethanol at a
concentration of 30 $\mu\text{g mL}^{-1}$ and incubated at room temperature for 4 min. Then,
168 $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ were removed by a magnet and the bound
amounts of three EDCs were quantified by HPLC. The recovered $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$
and $\text{Fe}_3\text{O}_4@\text{NIPs}$ were washed with a mixture of methanol/HAc (95:5, v/v) till we
171 ensured complete removal of residual three EDCs in the polymers and washed with
highly purified water for several times, then dried under vacuum at 40 °C, and then
reused in succeeding adsorption-desorption cycles.

174 **Analysis of three EDCs in environmental water samples**

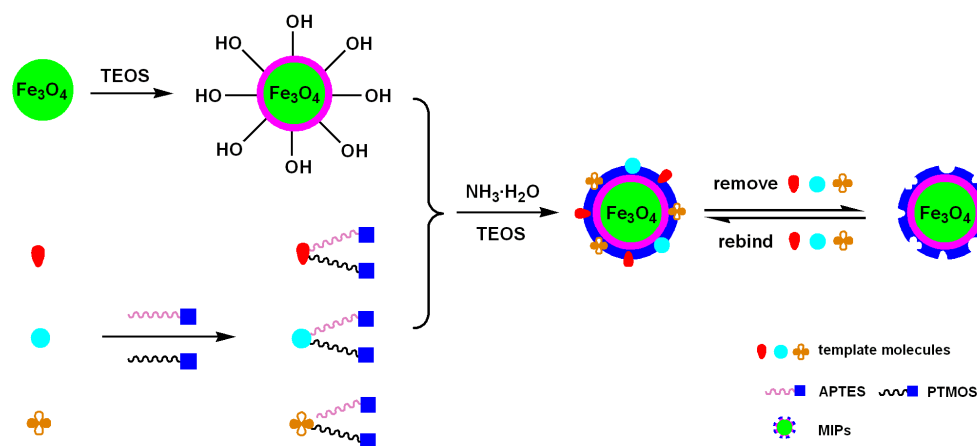
The lake and river water samples collected from the Xing Qing Park and Weihe River (Xi'an, China) were spiked with E2, E3, and DES at three levels (0.5, 1.0, and 3.0 ng mL⁻¹). Then, the samples were filtered by 0.22 μm membrane filters without any other pretreatment and stored in the precleaned glass bottles at room temperature until they were analyzed. 100 mg of Fe₃O₄@Multi-MIPs or Fe₃O₄@NIPs were added to 100 mL of the lake and river water samples containing the three EDCs, respectively. After 4 minutes of incubation on an oscillator, the Fe₃O₄@Multi-MIPs or Fe₃O₄@NIPs were isolated by a magnet and the supernatant solution was discarded. The Fe₃O₄@Multi-MIPs or Fe₃O₄@NIPs which absorbed target molecules were eluted with a mixture of methanol/HAc (95:5, v/v) solution, and then the elution was collected and evaporated to dry under a stream of nitrogen. Finally, the residue of the elution was dissolved in 0.5 mL of methanol and measured by HPLC-UV.

Results and discussion

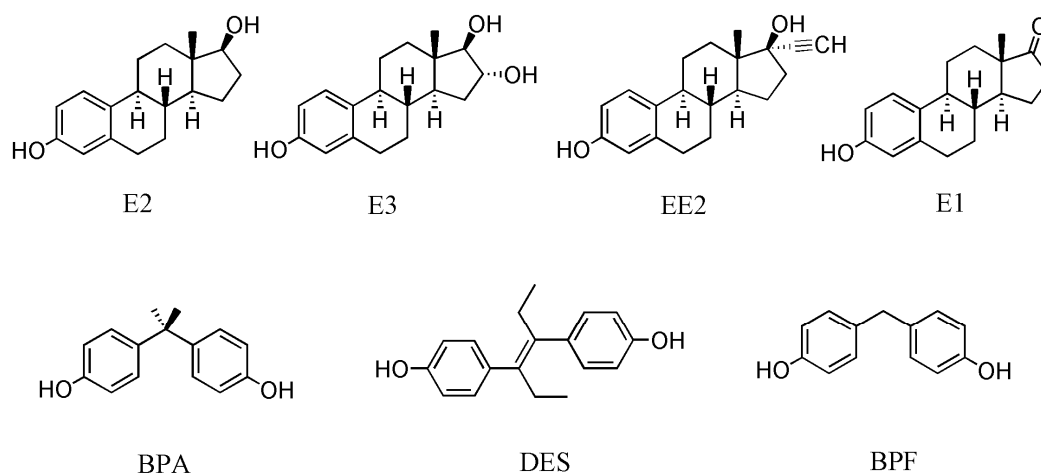
Synthesis of Fe₃O₄@Multi-MIPs

The general scheme for the synthesis of multi-template imprinted magnetic nanomaterials based on surface imprinting technique, sol-gel process, and magnetic supporter was illustrated in **Fig. 1**. The formation of the Fe₃O₄@Multi-MIPs included four major procedures: (1) Fe₃O₄ NPs were synthesized by a modified solvothermal method, and whose surfaces were transformed to silica shells using TEOS through a sol-gel process. (2) The template-monomer complex was obtained with E2, E3, and DES as multi-templates, APTES and PTMOS as bifunctional monomers. (3) The templates-monomers complex was anchored on the surface of Fe₃O₄@SiO₂ using

ammonium hydroxide as the catalyst through another sol-gel process. (4) A thin MIP
 198 layer which had specific recognition cavities for E2, E3, and DES was formed after the
 removal of the multiple templates.



201 Fig. 1. Scheme of the synthetic route for Fe_3O_4 @Multi-MIPs.



204 Fig. 2. Molecular structures of EDCs used in this study.

The molecular recognition capability of Fe_3O_4 @Multi-MIPs is greatly affected by
 the functional monomers which can create the molecular recognition sites through
 207 interacting with the template molecules and maintain their shape, size, and location of
 functionalities in the polymer network after the templates are removed. In this study,

two kinds of silane coupling agents as co-monomers were selected to produce the
210 imprinted binding sites according to the structures and features of E2, E3, and DES
(**Fig.2**). One is APTES which can form hydrogen bonds between its amino groups and
hydroxyl groups of the templates. The other is PTMOS which can establish π - π
213 interactions with the templates *via* the phenyl groups of the two.

The effect of the volume ratios of APTES and PTMOS on the imprinting
performance of Fe_3O_4 @Multi-MIPs and Fe_3O_4 @NIPs was investigated. As shown in
216 **Table 1**, we found that the recognition ability of Fe_3O_4 @Multi-MIPs towards E3/DES
was more susceptible to the volume ratio of APTES/PTMOS. When inspecting the
structures of E3 and DES, E3 possesses three -OH, which could have better ability
219 than other two templates to form multiple hydrogen bonds with the -NH₂ of APTES,
while DES holds two phenyl groups and may have stronger capacity to establish π - π
interactions with PTMOS. We found that the volume ratio of APTES and PTMOS was
222 1:1 which had the best adsorption ability to all three templates, not only in adsorption
capacity (Q), but also in imprinting factor (IF). This might result from the coordinate
effect of hydrogen bonds and π - π interactions which might achieve to the best
225 condition between functional monomers and the template molecules when APTES
and PTMOS were in the same volume. Therefore, the optimized volume ratio of
APTES to PTMOS was confirmed as 1:1.

228 To achieve a satisfactory recognition quality, the binding performance of
 Fe_3O_4 @Multi-MIPs and Fe_3O_4 @NIPs with different amounts of functional monomers
ranging from 50 μL to 150 μL was evaluated and listed in **Table 1**. It was observed

231 obviously that the Q and IF increased as the amounts of functional monomers
increased from 50 μL to 100 μL , due to the augment of the amounts of the recognition
cavities on the thin MIP layers. However, a decrease of Q and IF was appeared with
234 the increasing of the amounts of functional monomers from 100 μL to 150 μL ,
because agglomeration of the functional monomers would happen if whose amount
was excess, and led to a reduction in the number of recognition cavities. The
237 experimental results revealed that 100 μL of APTES and 100 μL of PTMOS were
appropriate in this work.

Characterization of prepared nanomaterials

240 The morphology of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ was characterized
by SEM (**Fig. S1**) and TEM. As shown in **Fig. 3A**, the uncoated Fe_3O_4 nanoparticles
were relatively homogeneous and well-dispersed with a mean diameter of about 200
243 nm. After coating with a thin SiO_2 shell, the diameter of $\text{Fe}_3\text{O}_4@\text{SiO}_2$ increased to
about 240 nm (**Fig. 3B**), corresponding to a 10 nm thick SiO_2 layer covered on Fe_3O_4 .
The SiO_2 shell was uniformly coated on Fe_3O_4 magnetic core for all the samples,
246 which could prevent oxidation of magnetic core and provide a biocompatible and
hydrophilic surface. In addition, some interesting functionalities could be simply
linked to the surface of $\text{Fe}_3\text{O}_4@\text{SiO}_2$ through sol-gel reactions. The images in **Fig. 3C**
249 displayed the formation of core-shell structured $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ after the
template-monomer complex was anchored on the surface of $\text{Fe}_3\text{O}_4@\text{SiO}_2$ *via* another
sol-gel process under the optimized synthetic conditions. The SEM image (**Fig. S1C**)
252 revealed a relatively uniform size distribution and the shell of as-prepared product

was thickened to 15 nm appeared in TEM image (**Fig. 3C**), indicating that a MIP layer with thickness of about 5 nm was attached on the $\text{Fe}_3\text{O}_4@\text{SiO}_2$ successfully. The thin MIP layers would be effective to the mass transport between solution and the surface of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$.

The EDS analysis was also conducted to confirm the existence of imprinted polymer. For the Fe_3O_4 , only Fe and O signals appeared, in accordance with its elemental composition (Fig. S2A). The signal of Si emerged and percentage of each element changed greatly after coating with SiO_2 (Fig. S2B). The emergence of the signal of N in Fig. S2C provided additional evidence for the formation of imprinted shell on the surface of $\text{Fe}_3\text{O}_4@\text{SiO}_2$.

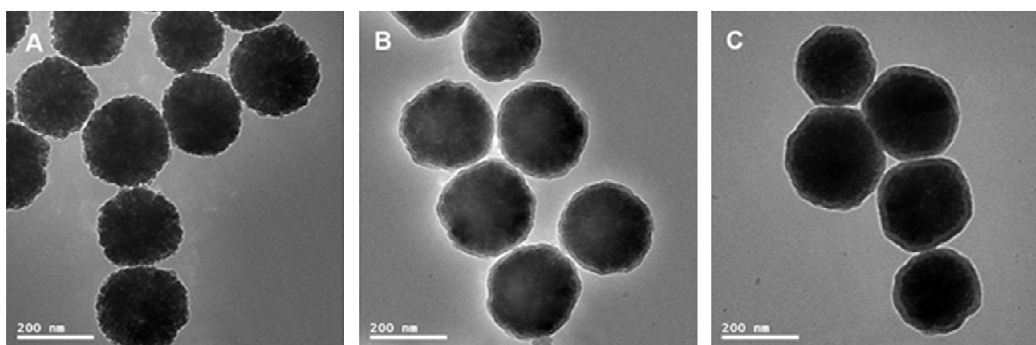


Fig. 3. TEM images of Fe_3O_4 (A), $\text{Fe}_3\text{O}_4@\text{SiO}_2$ (B), and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ (C).

The FT-IR spectra provided a direct proof for synthetic process of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$. The characteristic peaks of the stretch of Fe-O group for Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ were all observed around 580 cm^{-1} (**Fig. 4**). The absorption peaks at 3442 cm^{-1} and 1640 cm^{-1} were assigned to stretching vibration and bending vibration of O-H (**Fig. 4A**), suggesting hydroxyl groups on the surface of Fe_3O_4 .³¹ In comparison with the curve of pure Fe_3O_4 , a strong peak at approximately 1100 cm^{-1} attributed to the stretching vibration of Si-O-Si could be

observed in the curve of $\text{Fe}_3\text{O}_4@\text{SiO}_2$ (**Fig. 4B**), which indicated the formation of
273 silica film on the surface of Fe_3O_4 . The typical peak at 3442 cm^{-1} arising from N-H
stretching vibration might overlap with that of O-H stretching vibration,³² but the
relatively high intensity of the peak and bending vibration of N-H at 1562 cm^{-1} still
276 revealed the contribution of $-\text{NH}_2$ groups. The absorbance at 1430 cm^{-1} was ascribed
to the stretch of $\text{Si}-\text{C}_6\text{H}_5$ (**Fig. 4C**). The existence of these characteristic peaks derived
from APTES and PTMOS proved that the MIP layer was successfully coated on
279 $\text{Fe}_3\text{O}_4@\text{SiO}_2$.

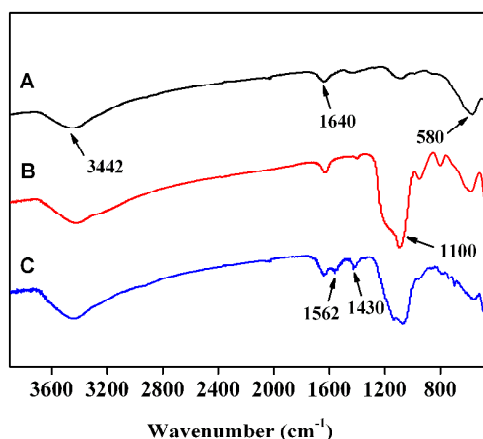


Fig. 4. FT-IR spectra of Fe_3O_4 (A), $\text{Fe}_3\text{O}_4@\text{SiO}_2$ (B), and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ (C).

282 The X-ray diffraction (XRD) patterns of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and
 $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ were displayed in **Fig. 5**. In the 2θ range of 10° - 80° , six
representative peaks of Fe_3O_4 ($2\theta = 30.38^\circ$, 35.58° , 43.14° , 53.48° , 57.08° , and 62.66°)
285 were detected for the three samples. The peak positions at the corresponding 2θ values
were indexed as (220), (311), (400), (422), (511), and (440), respectively, which
matched well with the database of magnetite in the JCPDS-International Center for
288 Diffraction Data (JCPDS Card: 19-629) file. The XRD patterns showed that the

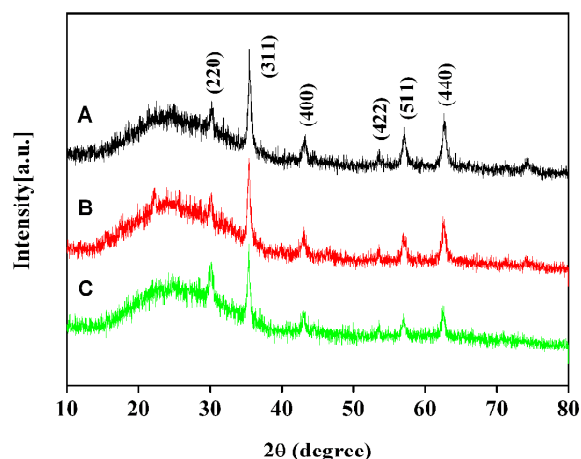
synthetic nanoparticles were highly crystalline materials and the crystalline of magnetite remained unchanged during the preparation of Fe₃O₄@Multi-MIPs.

291 **Table 1**

Effect of volume ratios and amounts of functional monomers on the imprinting performance of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs.^a

	Analytes	Q_{MIP} (mg g ⁻¹)	Q_{NIP} (mg g ⁻¹)	IF
$V_{\text{APTES}}/V_{\text{PTMOS}}$				
1:0	E2	2.16	0.99	2.18
	E3	4.29	1.03	4.16
	DES	2.63	0.87	3.02
0:1	E2	1.76	0.73	2.41
	E3	3.89	1.35	2.88
	DES	4.96	2.87	1.72
1:2	E2	1.91	0.89	2.15
	E3	3.64	1.05	3.47
	DES	5.42	2.16	2.51
1:1	E2	3.45	0.91	3.79
	E3	5.76	1.12	5.14
	DES	6.44	2.21	2.91
2:1	E2	2.54	0.87	2.92
	E3	4.58	1.08	4.24
	DES	4.67	2.19	2.13
$V_{\text{APTES}}/PTMOS$ (μL)				
50	E2	2.08	0.83	2.51
	E3	3.57	1.01	3.53
	DES	5.08	1.89	2.69
75	E2	2.76	0.87	3.17
	E3	4.89	1.08	4.53
	DES	5.72	1.97	2.90
100	E2	3.45	0.91	3.79
	E3	5.76	1.12	5.14
	DES	6.44	2.21	2.91
125	E2	2.83	0.89	3.18
	E3	4.65	1.06	4.39
	DES	5.91	2.15	2.75
150	E2	2.41	0.92	2.62
	E3	4.22	1.09	3.87
	DES	5.24	2.17	2.41

294 ^a In this experiment, 20 mg of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs were incubated with the mixture of E2, E3, and DES at a concentration of 30 μg mL⁻¹ in 10 mL of ethanol for 4 min at room temperature.

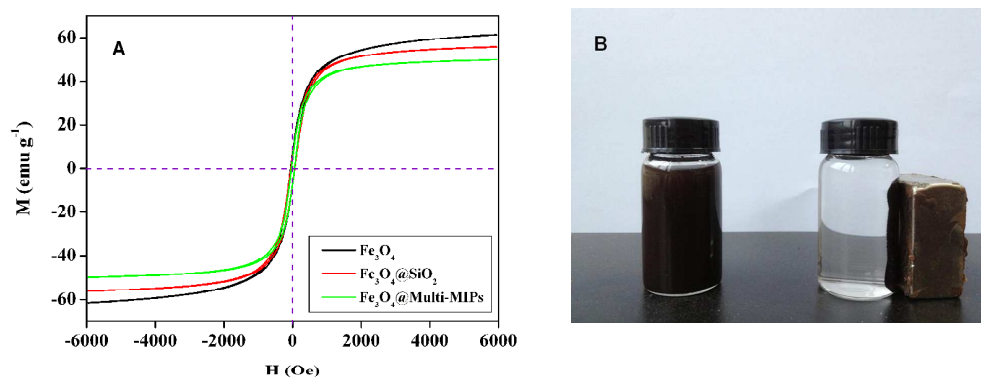


297

Fig. 5. XRD patterns of Fe_3O_4 (A), $\text{Fe}_3\text{O}_4@\text{SiO}_2$ (B), and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ (C).

The magnetic properties of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ were studied using a vibrating sample magnetometer at room temperature, and the plots of magnetization *versus* magnetic field (M-H loop) were illustrated in **Fig. 6A**. The remanent magnetizations of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ were close to zero, suggesting that the three samples were all superparamagnetic. The saturation magnetization of $\text{Fe}_3\text{O}_4@\text{SiO}_2/ \text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ was reduced to 55.99/50.15 emu g^{-1} in comparison with 61.51 emu g^{-1} of Fe_3O_4 . The decrease in magnetic saturation of $\text{Fe}_3\text{O}_4@\text{SiO}_2/\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ might be attributed to the increased mass of modified silane/MIP shells on the surface of $\text{Fe}_3\text{O}_4/\text{Fe}_3\text{O}_4@\text{SiO}_2$. The two reduced saturation magnetization values were all about 5 emu g^{-1} , which was rather small, demonstrating that the modified silane/MIP layers on the surface of $\text{Fe}_3\text{O}_4/\text{Fe}_3\text{O}_4@\text{SiO}_2$ were quite thin. The thin MIP shells would be effective to adsorb and desorb template molecules between solution and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$. **Fig. 6B** displayed the separation and redispersion process of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$. The $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ can be attracted to the wall of the vial rapidly, and the dispersion

became clear and transparent within several seconds in the presence of an external
 315 magnetic field. When the external magnetic field was absent, a dark homogeneous
 dispersion could be observed. The superparamagnetism of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$
 prevented the polymers from aggregating and enabled rapid redispersion after the
 318 magnetic field was taken away, which was very useful for their application.



324 Fig. 6. Magnetization curves of Fe_3O_4 , $\text{Fe}_3\text{O}_4@\text{SiO}_2$, and $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ (A); The separation
 and redispersion of a solution of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ in the absence (left) and presence (right) of
 an external magnetic field (B).

327 Adsorption kinetics of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$

The adsorption kinetics of E2, E3, and DES onto $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and
 $\text{Fe}_3\text{O}_4@\text{NIPs}$ were investigated. As presented in **Fig. 7A**, the imprinted nanomaterials
 330 displayed a fast adsorption rate. The adsorption capacity increased rapidly in the first
 3 min and almost reached equilibrium after 4 min for all of three different templates.
 It took shorter time to reach adsorption saturation than that of some other surface
 333 imprinting technologies for EDCs,^{33,34} which demonstrated that three EDCs
 approached the surface imprinting cavities of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ easily. The thin and
 uniform MIP shells of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ provided efficient mass transport, thus

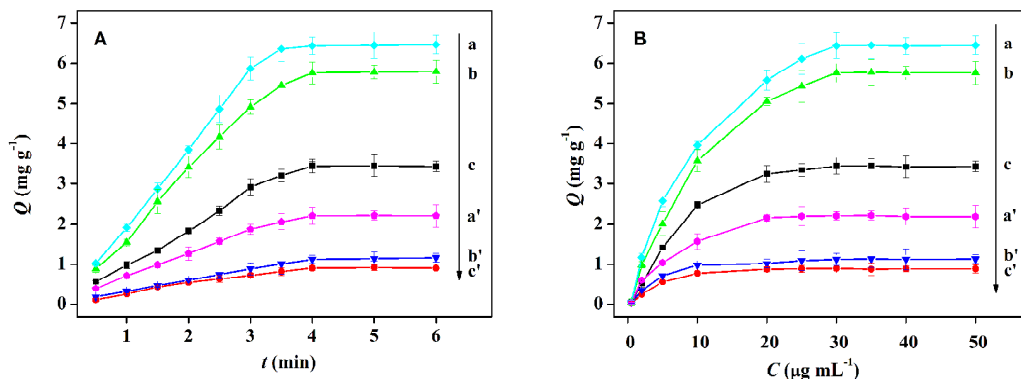
336 overcoming some drawbacks of traditionally packed imprinted materials³⁵ and
non-thin imprinted nanomaterials.³⁶

Adsorption isotherms of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs

339 The isothermal adsorption of E2, E3, and DES onto Fe₃O₄@Multi-MIPs and
Fe₃O₄@NIPs were evaluated in the initial concentration range of 0.50-50 µg mL⁻¹,
and the corresponding results were shown in **Fig. 7B**. It could be seen that the
342 adsorption capacity of Fe₃O₄@Multi-MIPs increased rapidly along with the increase
of initial concentration and reached adsorption saturation when the initial
concentration was above 30 µg mL⁻¹. The amounts of the three EDCs bound to
345 Fe₃O₄@Multi-MIPs were higher than that of Fe₃O₄@NIPs at the same initial
concentration, which suggested that the imprinted recognition cavities played
important roles in the process of the three EDCs binding to Fe₃O₄@Multi-MIPs. To
348 estimate the binding properties of the imprinted nanomaterials, the saturation binding
data were further processed by the Langmuir equation, which was expressed as
following equation:

$$351 \quad \frac{C_e}{Q_e} = \frac{C_e}{Q_{\max}} + \frac{1}{Q_{\max} K_L} \quad 2$$

Where Q_e (mg g⁻¹) is the amount of template molecules bound to
Fe₃O₄@Multi-MIPs at equilibrium, Q_{\max} (mg g⁻¹) is the apparent maximum
354 adsorption capacity, C_e (µg mL⁻¹) is the free analytical concentration at equilibrium,
and K_L (L mg⁻¹) is the Langmuir constant. The values of K_L and Q_{\max} can be
calculated from the slope and intercept of the linear plotted in C_e/Q_e versus C_e .



357 Fig. 7. Adsorption kinetics (A) and isotherms (B) of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ to DES (a, a'), E3 (b, b'), and E2 (c, c'), respectively.

Langmuir model fitted the equilibrium data rather well from the deduction of linear correlation which were greater than 0.99 (**Table S1**). From the slopes and intercepts of the straight lines obtained, the values of apparent maximum adsorption capacities of E2, E3, and DES were 3.74 mg g^{-1} , 6.02 mg g^{-1} , and 6.89 mg g^{-1} for $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$, and 0.99 mg g^{-1} , 1.22 mg g^{-1} , and 2.40 mg g^{-1} for $\text{Fe}_3\text{O}_4@\text{NIPs}$, which were close to the maximum amounts of adsorption obtained from experimental results (**Fig. 7B**). It can be concluded that the adsorption of E2, E3, and DES onto $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ might be all monolayer adsorption.³⁷ Furthermore, the adsorption was occurred uniformly on the active binding sites of the surface of the resultant imprinted nanomaterials and no further adsorption could take place at the site once a template molecule occupied this site.³⁶

Specific recognition of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$

In order to measure the specificity of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ to E2, E3, and DES, four other EDCs (E1, EE2, BPA, and BPF) were selected as analogues. The adsorption performance of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$ to the mixture of E2,

E3, DES, E1, EE2, BPF, and BPA at a concentration of 30 $\mu\text{g mL}^{-1}$ in 10 mL of
375 ethanol was investigated.

The imprinting factor (IF) and selectivity coefficient (SC) were usually used to
evaluate the specific recognition property of the imprinted materials towards template
378 molecules and structurally related compounds. The IF and SC were calculated by the
following formulas:

$$IF = \frac{Q_{\text{MIP}}}{Q_{\text{NIP}}} \quad 3$$

$$381 \quad SC = \frac{IF_t}{IF_a} \quad 4$$

Where Q_{MIP} and Q_{NIP} (mg g^{-1}) represent the adsorption capacity of EDCs for
 $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{NIPs}$, IF_t and IF_a are the imprinting factors for
384 templates and analogues.

As presented in **Table 2**, the Q and IF of E2, E3, and DES for $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$
were higher than those of four other EDCs, indicating that $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ had a
387 relatively higher affinity for the template molecules than their analogues. Although
the structures of E1, EE2, BPA, and BPF all contain hydroxyl and phenyl functional
groups (**Fig. 2**) which are capable of forming hydrogen bonding and π - π interaction
390 with $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$, the imprinted nanomaterials still exhibited high recognition
for the template molecules. This could be explained as that the template molecules
were imprinted on the MIP layers during the preparation of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$, after
393 removal of the template molecules, the complementarity of cavities in MIP layers to
the templates both in size and shape were established. Furthermore, we surprisingly

found that the *IF* of E2, E3, and DES in the mixture of seven EDCs were higher than
396 those in the mixture solution of three EDCs. This phenomenon was probably ascribed
to the fact that EDCs competed with each other much more drastically in seven EDCs
than in three EDCs mixture, which led to the decrease of adsorption of three EDCs in
399 both Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs. However, since the specific recognition
sites existed on the MIP layers were complementary in shape, size, and spatial
arrangement to template molecules, the template molecules had advantage in
402 occupying the binding sites over three other EDCs. The adsorption of seven EDCs on
Fe₃O₄@NIPs was non-specific, so every EDCs had the equal opportunity to be
adsorbed. This resulted in that the adsorption capacities of template molecules on
405 Fe₃O₄@NIPs in the seven EDCs mixed solution were smaller than those of the
Fe₃O₄@NIPs in the three EDCs mixed solution. Besides, the Fe₃O₄@Multi-MIPs
exhibited higher specificity for templates when compared with duo-molecularly
408 imprinted polymer prepared by Xia et al.²⁷ These results demonstrated satisfactory
imprinting efficiency of the present method.

Reproducibility and reusability of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs

411 The reproducibility of the obtained imprinted magnetic nanomaterials was
investigated by using six batches of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs prepared
on different days. Five independent replicates were measured for each batch. The
414 average adsorption capacity of the six batches of Fe₃O₄@Multi-MIPs and
Fe₃O₄@NIPs for E2, E3, and DES were 3.45, 5.76, and 6.44 mg g⁻¹, and 0.91, 1.12,
and 2.21 mg g⁻¹, respectively. The relative standard deviation (RSD) of every batch

417 was listed in **Table S2**. The results exhibited that the reproducibility of six batches of
Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs were all satisfactory with a RSD less than
8.6%.

420 The reusability of polymers was considered to have a great cost benefit on
extending their utilization. Based on this, the adsorption-desorption cycle was
repeated six times by using the same Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs to
423 evaluate their stability. The change of the amounts of adsorbed three template EDCs
after six regeneration cycles, as shown in **Fig. 8**, indicated that the adsorption
capacities of Fe₃O₄@Multi-MIPs still maintained at almost steady values of 93.2%,
426 94.1%, and 92.6% for E2, E3, and DES, whereas the binding amounts of
Fe₃O₄@NIPs remained almost unchanged. The decreased affinity adsorption of the
Fe₃O₄@Multi-MIPs might be ascribed to the fact that some recognition sites in the
429 network of imprinted polymers could be jammed after regeneration or destructed after
rewashing. On the other hand, the adsorption affinity of Fe₃O₄@NIPs was nonspecific
and the effect of washing steps could be negligible. The regeneration experiments of
432 Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs verified the favorable stability of the
nanomaterials prepared in this work.

Template recognition of Fe₃O₄@Multi-MIPs and reference polymers

435 We evaluated template recognition of Fe₃O₄@Multi-MIPs in parallel experiments
with the reference polymers for E2, E3, and DES in a mixture solution. The *Q* and *IF*
of the EDCs absorbed by the polymers were summarized in **Table 3**. It can be seen
438 that Fe₃O₄@E2-MIPs, Fe₃O₄@E3-MIPs, and Fe₃O₄@DES-MIPs all had high

adsorption to their corresponding template. In addition, compared with $\text{Fe}_3\text{O}_4@\text{NIPs}$, $\text{Fe}_3\text{O}_4@\text{E2-MIPs}$ exhibited higher adsorption capacity to E3 and $\text{Fe}_3\text{O}_4@\text{E3-MIPs}$ also absorbed higher amount to E2, which may be attributed to the resemblance of E2 and E3, particularly in their functional groups and structures. $\text{Fe}_3\text{O}_4@\text{E2-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{E3-MIPs}$ both absorbed less DES, and $\text{Fe}_3\text{O}_4@\text{DES-MIPs}$ absorbed less E2 and E3, which may be because of definite structural differences between DES and E2/E3, resulting in worse matching of $\text{Fe}_3\text{O}_4@\text{E2-MIPs}$ and $\text{Fe}_3\text{O}_4@\text{E3-MIPs}$ to DES and $\text{Fe}_3\text{O}_4@\text{DES-MIPs}$ to E2/E3. These results demonstrated that the imprinted recognition cavities played important roles in the process of EDCs adsorption. The recognition capacity of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and M-MIPs for three EDCs was similar with the order of $\text{DES} > \text{E3} > \text{E2}$. This may be due to the three EDCs containing different functional groups, DES has two phenyl groups and two hydroxyl groups, E3 contains one phenyl group and three hydroxyl groups, and E2 holds one phenyl group and two hydroxyl groups. After the comprehensive comparison of the experimental results, we can deduced that π - π interactions played a more important role in the recognition process compared with hydrogen bonding interactions. Although the adsorption effects of $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ and M-MIPs for three EDCs were similar, $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ prepared using three EDCs as the mixed-template molecules not only made the extraction procedure to be carried out in a single step for three EDCs, but also simplified the synthesis and processing procedures with distinct reduction in both preparation time and solvent consumption.

462 **Table 2**

The adsorption capacities, imprinting factors, and selectivity coefficients of templates and analogues for Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs.^a

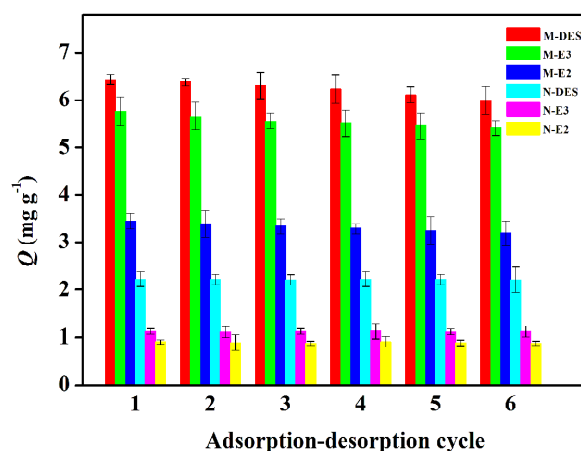
Analytes	Q_{MIP} (mg g ⁻¹)	Q_{NIP} (mg g ⁻¹)	IF	SC		
				SC_1^b	SC_2^c	SC_3^d
E2	2.98	0.93	3.20	—	—	—
E3	5.23	0.82	6.38	—	—	—
DES	5.97	1.05	5.69	—	—	—
E1	1.87	0.741	2.52	1.27	2.53	2.26
EE2	1.52	0.683	2.23	1.43	2.86	2.55
BPA	0.945	0.512	1.85	1.73	3.45	3.08
BPF	1.29	0.718	1.80	1.78	3.54	3.16

465 ^a In this experiment, 20 mg of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs were incubated with the mixture of E2, E3, DES, E1, EE2, BPA, and BPF at a concentration of 30 μg mL⁻¹ in 10 mL of ethanol for 4 min at room temperature.

^b $SC_1 = IF_{\text{E2}} / IF_{\text{a}}$.

468 ^c $SC_2 = IF_{\text{E3}} / IF_{\text{a}}$.

^d $SC_3 = IF_{\text{DES}} / IF_{\text{a}}$.



471 Fig. 8. Reusability of Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs.

Method performance

Under the optimal conditions, the experimental parameters were investigated to
 474 validate the proposed method (**Table 4**). The relatively high correlation coefficients
 ($R^2 > 0.9995$) of the Fe₃O₄@Multi-MIPs solid-phase extraction coupled with HPLC
 method was achieved in the range of 0.3-100.0 ng mL⁻¹ for three steroids estrogens
 477 (E2, E3, and DES). The limits of detection (LODs) and the limits of quantitation

(LOQs) for three steroids estrogens, defined as signal-to-noise ratio of 3 and 10, were in the range of 0.08-0.27 and 0.29-0.94 ng mL⁻¹, proving that the proposed method was sensitive. To investigate the precision, relative standard deviations (RSDs) of intra- and inter-day tests were measured at three different concentrations of 1.0, 10.0, and 100.0 ng mL⁻¹. RSDs of intra- and inter-day precisions were obtained in the range of 2.3-4.5% and 3.2-5.9%, which demonstrated that the developed method had a satisfactory reproducibility.

Table 3

The adsorption capacities and imprinting factors of Fe₃O₄@Multi-MIPs and reference polymers.^a

Polymers	<i>Q</i> (mg g ⁻¹)			<i>IF</i>		
	E2	E3	DES	E2	E3	DES
Fe ₃ O ₄ @NIPs	0.912	1.12	2.21	—	—	—
Fe ₃ O ₄ @E2-MIPs	9.87	3.49	2.57	10.8	3.12	1.16
Fe ₃ O ₄ @E3-MIPs	2.84	11.5	2.63	3.11	10.3	1.19
Fe ₃ O ₄ @DES-MIPs	1.57	2.13	13.52	1.72	1.90	6.12
Fe ₃ O ₄ @Multi-MIPs	3.45	5.76	6.44	3.78	5.14	2.91
M-MIPs	3.82	5.38	6.57	4.19	4.80	2.97

^a In this experiment, 20 mg of Fe₃O₄@Multi-MIPs and reference polymers were incubated with the mixture of E2, E3, and DES at a concentration of 30 µg mL⁻¹ in 10 mL of ethanol for 4 min at room temperature.

Table 4

The performance parameters of the proposed method (*n* = 6).

Compounds	Linearity range (ng mL ⁻¹)	<i>R</i> ²	RSD (%)		LODs (ng mL ⁻¹)	LOQs (ng mL ⁻¹)
			Intra-day	Inter-day		
E2	1.0-100	0.9996	3.8-4.5	4.3-5.9	0.27	0.94
E3	0.8-100	0.9997	2.7-3.9	3.5-4.8	0.19	0.72
DES	0.3-100	0.9998	2.3-3.5	3.2-3.6	0.08	0.29

Different methods for determination of steroids estrogens were summarized briefly in **Table 5**. As can be seen, the present approach had lower LODs than those of other reported methods followed by HPLC-UV analysis,^{21,38} and possessed the merits of solvent-saving and easy separation of sorbent after extraction under an external

magnetic field compared with the liquid-liquid-liquid micro-extraction and non-magnetic molecularly imprinted solid-phase extraction, respectively. Meanwhile, the developed method based on using Fe_3O_4 @Multi-MIPs as solid extractant had comparable LODs in comparison with the method of UPLC-MS-MS,¹⁷ proving that the pretreatment in this work was much effective and selective. Furthermore, although the LODs obtained in this work was slightly higher than that of GC-MS analysis method,³⁹ the one more step of derivatization was needed before GC-MS analysis, which was complex and hard to control. Through the comparison, we could conclude that the method developed in this work was simple, time-saving, reliable, effective, and sensitive.

Table 5

Comparison of LODs with other published methods for the determination of estrogens.

Analytes	Extraction method	Analytical system	LODs (ng mL ⁻¹)	Reference
E2, E3, DES	MISPE ^a	HPLC-UV	1.00-8.00	[21]
E2, E3, DES	HF-LLLME ^b	HPLC-UV	0.11-0.66	[38]
E2, E3	ZIF-8-MSPE ^c	UPLC-MS-MS	0.05	[17]
E2, E3, DES	—	GC-MS	0.0034-0.0042	[39]
E2, E3, DES	Fe_3O_4 @Multi-MIPs-SPE	HPLC-UV	0.08-0.27	this work

^a MISPE: Molecularly imprinted solid-phase extraction.

^b HF-LLLME: Hollow fiber liquid-liquid-liquid microextraction.

^c ZIF-8-MSPE: Zeolitic imidazolate framework-8 micro-solid-phase extraction.

Analysis in environmental water samples

To evaluate the accuracy and potential application of the developed method that uses Fe_3O_4 @Multi-MIPs as the SPE adsorbents coupled with HPLC-UV for selective separation and detection of E2, E3, and DES in environmental water samples. The environmental water samples spiked with three levels of E2, E3, and DES (0.5, 1.0, and 3.0 ng mL⁻¹) were analyzed (**Table 6**). The recoveries of river water and lake

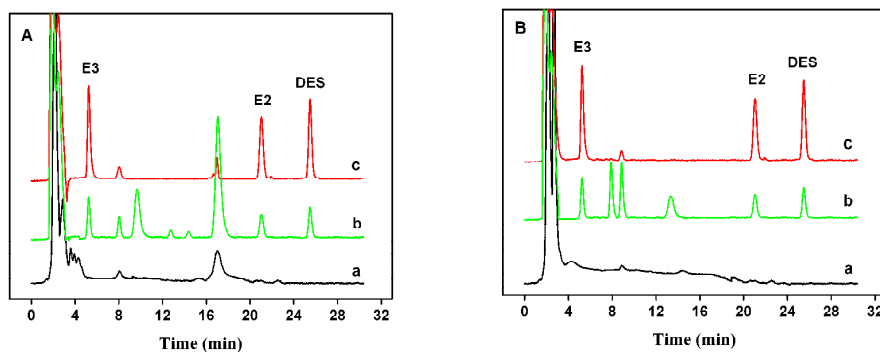
water samples were ranged from 93.4 to 98.6% and 92.3 to 98.2%, respectively. The RSD was less than 6.5%. The chromatograms of water samples spiked with E2, E3, and DES at the concentration of 3.0 ng mL⁻¹, and the elution of adsorbed Fe₃O₄@Multi-MIPs and Fe₃O₄@NIPs to E2, E3, and DES washed with a mixture of methanol/HAc (95:5, v/v) were exhibited in **Fig. 9**. The peaks of E2, E3, and DES could not be observed from chromatograms of the spiked river and lake water samples (**Fig. 9a**). Through the preconcentration of spiked river or lake water samples by Fe₃O₄@Multi-MIPs, and washing by methanol/HAc (95:5, v/v), the peaks of E2, E3, and DES appeared and other interference peaks were almost completely eliminated. (**Fig. 9c**). The results demonstrated that Fe₃O₄@Multi-MIPs offered a simple method for simultaneously selective enrichment and efficient separation of E2, E3, and DES in spiked environmental water samples, while presenting an extremely high anti-interference ability compared with Fe₃O₄@NIPs (**Fig. 9b**). These results revealed that Fe₃O₄@Multi-MIPs could be directly applied for simultaneously selective isolation and determination of E2, E3, and DES in environmental water samples.

Table 6

Recoveries of Fe₃O₄@Multi-MIPs binding E2, E3, and DES for spiked lake and river water samples (n=5).

Analytes	Lake Water						River Water					
	0.50 ng mL ⁻¹		1.0 ng mL ⁻¹		3.0 ng mL ⁻¹		0.50 ng mL ⁻¹		1.0 ng mL ⁻¹		3.0 ng mL ⁻¹	
	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)
E2	92.3	6.3	93.7	4.8	95.1	3.2	93.4	6.5	94.1	5.4	96.5	2.8
E3	93.2	5.1	94.9	4.2	97.4	2.4	93.8	5.3	95.6	3.2	98.1	2.1
DES	93.6	4.5	95.5	3.6	98.2	1.9	94.2	4.4	96.3	3.7	98.6	1.6

537



540 Fig.9. Chromatograms of lake (A) and river (B) water samples. (a) Samples spiked with templates
at the concentration of 3.0 ng mL^{-1} , elution of (b) $\text{Fe}_3\text{O}_4@\text{NIPs}$ and (c) $\text{Fe}_3\text{O}_4@\text{Multi-MIPs}$ after
the polymers adsorbing 100 mL of lake or river water spiked samples, respectively.

543 Conclusions

In this study, an innovative method was developed to prepare the multi-template
imprinted polymers by combining the advantages of surface imprinting technique,
546 sol-gel approach, and magnetic separation for the specific extraction of three EDCs
simultaneously. The prepared imprinted nanomaterials exhibited fast kinetics,
excellent recognition performance, and favorable selective affinity towards three
549 templates, and also had regeneration potential for reuse at least six times without
significant degradation in the binding property. Meanwhile, the resulting products
were used as absorbents coupled with HPLC-UV for specific isolation and detection
552 of three EDCs from environmental water samples concurrently. The proposed method
showed the features of simplicity, sensitivity and reliability. At the same time, good
recoveries and low LODs were obtained. Our research findings suggested that
555 multi-templates imprinted polymers can be triumphantly utilized for efficient and
selective removal of a class of pollutants from environmental matrices.

Acknowledgements

558 The authors are grateful for financial support from the National Natural Science
Foundation of China (No. 21305107), the Fundamental Research Funds for the
Central Universities (Nos. 08143081, 08142034), and China Postdoctoral Science
561 Foundation (No. 2014M562388).

References

1. D. W. Kolpin, E. T. Furlong, M. T. Meyer, E. M. Thurman, S. D. Zaugg, L. B. Barber and H.
564 T. Buxton, *Environ. Sci. Technol.*, 2012, **36**, 1202-1211.
2. I. Robinson, G. Junqua, R. V. Coillie and O. Thomas, *Anal. Bioanal. Chem.*, 2007, **387**,
1143-1151.
- 567 3. J. B. Gadd, L. A. Tremblay and G. L. Northcott, *Environ. Pollut.*, 2010, **158**, 730-736.
4. A. Vallejo, A. Usobiaga, M. Ortiz-Zarragoitia, M. P. Cajaraville, L. A. Fernández and O.
Zuloaga, *Anal. Bioanal. Chem.*, 2010, **398**, 2307-2314.
- 570 5. D. Fatta-Kassinos, I. K. Kalavrouziotis, P. H. Koukoulakis and M. I. Vasquez, *Sci. Total
Environ.*, 2011, **409**, 3555-3563.
6. M. S. Souza, P. Hallgren, E. Balseiro and L. A. Hansson, *Environ. Pollut.*, 2013, **178**,
573 237-243.
7. M. Kostich, R. Flick and J. Martinson, *Environ. Pollut.*, 2013, **178**, 271-277.
8. M. Grassi, L. Rizzo and A. Farina, *Environ. Sci. Pollut. Res.*, 2013, **20**, 3616-3628.
- 576 9. V. Okoh, A. Deoraj and D. Roy, *Biochim. Biophys. Acta*, 2011, **1815**, 115-133.
10. W. Y. Hu, G. B. Shi, D. P. Hu, J. L. Nelles and G. S. Prins, *Mol. Cell. Endocrinol.*, 2012, **354**,
63-73.

- 579 11. P. A. Fowler, M. Bellingham, K. D. Sinclair, N. P. Evans, P. Pocar, B. Fischer, K. Schaedlich,
J. S. Schmidt, M. R. Amezaga, S. Bhattacharya, S. M. Rhind and P. J. O'Shaughnessy, *Mol.*
Cell. Endocrinol., 2012, **355**, 231-239.
- 582 12. C. Stouder and A. Paoloni-Giacobino, *Reproduction*, 2011, **141**, 207-216.
13. C. P. Silva, D. L. D. Lima, R. J. Schneider, M. Otero and V. I. Esteves, *J. Environ. Manage.*,
2013, **124**, 121-127.
- 585 14. S. K. Garnayak, J. Mohanty, T. V. Rao, S. K. Sahoo and P. K. Sahoo, *Aquaculture*, 2013,
392-395, 148-155.
15. G. Saravanabhavan, R. Helleur and J. Hellou, *Chemosphere*, 2009, **76**, 1156-1162.
- 588 16. C. M. Lu, M. T. Wang, J. Mu, D. C. Han, Y. P. Bai and H. Q. Zhang, *Food Chem.*, 2013, **141**,
1796-1806.
17. Y. H. Wang, S. G. Jin, Q. Y. Wang, G. H. Lu, J. J. Jiang and D. R. Zhu, *J. Chromatogr. A*,
591 2013, **1291**, 27-32.
18. T. M. Penning, S. H. Lee, Y. Jin, A. Gutierrez and I. A. Blair, *J. Steroid Biochem. Mol. Biol.*,
2010, **121**, 546-555.
- 594 19. E. Tahmasebi, Y. Yamini, M. Moradi and A. Esrafil, *Anal. Chim. Acta*, 2013, **770**, 68-74.
20. Z. K. Lin, W. J. Cheng, Y. Y. Li, Z. R. Liu, X. P. Chen, C. J. Huang, *Anal. Chim. Acta*, 2012,
720, 71-76.
- 597 21. C. D. Zhao, X. M. Guan, X. Y. Liu and H. X. Zhang, *J. Chromatogr. A*, 2012, **1229**, 72-78.
22. Q. C. Zhang, X. H. Xiao, G. K. Li, *Talanta*, 2014, **123**, 63-70.
23. B. Wu, Z. H. Wang, Z. H. Xue, X. B. Zhou, J. Du, X. H. Liu and X. Q. Lu, *Analyst*, 2012,
600 **137**, 3644-3652.

24. Y. Guo and T. Y. Guo, *Chem. Commun.*, 2013, **49**, 1073-1075.
25. T. Madrakian, M. Ahmadi, A. Afkhami and M. Soleimani, *Analyst*, 2013, **138**, 4542-4549.
- 603 26. A. Zengin, E. Yildirim, U. Tamerb and T. Caykara, *Analyst*, 2013, **138**, 7238-7245
27. X. L. Xia, E. P. C. Lai and B. Örmeci, *Environ. Sci. Pollut. Res.*, 2013, **20**, 3331-3339.
28. Y. P. Duan, C. M. Dai, Y. L. Zhang and L. Chen, *Anal. Chim. Acta*, 2013, **758**, 93-100.
- 606 29. Y. M. Ren, J. Yang, W. Q. Ma, J. Ma, J. Feng and X. L. Liu, *Water Res.*, 2014, **50**, 90-100.
30. J. Zhang, L. L. Wang and Y. T. Han, *J. Sep. Sci.*, 2013, **36**, 3486-3492.
31. S. Mohapatra, N. Pramanik, S. Mukherjee, S. K. Ghosh and P. Pramanik, *J. Mater. Sci.*, 2007,
- 609 **42**, 7566-7574.
32. Q. W. Peng, J. Gan, S. F. Wang, L. B. Kong, G. R. Chen, Y. X. Yang and G. J. Huang, *Ind. Eng. Chem. Res.*, 2013, **52**, 7713-7717.
- 612 33. Z. K. Lin, Q. Y. He, L. T. Wang, X. D. Wang, Q. X. Dong and C. J. Huang, *J. Hazard. Mater.*, 2013, **252-253**, 57-63.
34. S. Wang, R. Y. Wang, X. L. Wu, Y. Wang, C. Xue, J. H. Wu, J. L. Hong, J. Liu and X. M. Zhou, *J. Chromatogr. B*, 2012, **905**, 105-112.
- 615 35. J. Gañán, A. Gallego-Picó, R. M. Garcinuño, P. Fernández-Hernando, S. Morante, I. Sierra and J. S. Durand, *Anal. Bioanal. Chem.*, 2012, **403**, 3025-3029.
- 618 36. X. Kong, R. X. Gao, X. W. He, L. X. Chen and Y. K. Zhang, *J. Chromatogr. A*, 2012, **1245**, 8-16.
37. F. F. Chen, X. Y. Xie and Y. P. Shi, *J. Chromatogr. A*, 2013, **1300**, 112-118.
- 621 38. B. B. Chen, Y. L. Huang, M. He and B. Hu, *J. Chromatogr. A*, 2013, **1305**, 17-26.

39. N. Migowska, M. Caban, P. Stepnowski and J. Kumirska, *Sci.Total Environ.*, 2012, **441**,

77-88.

624