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Design and synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones with C-3’ side chains as potent Met kinase inhibitors†

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Abstract

Pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold 4 with various C-3’ side chains were designed as potent Met kinase inhibitors. Structural optimization led to compounds 10, 20, and 22–24 that demonstrated subnanomolar IC₅₀ values in the biochemical assay. The potent compound 20 inhibited Met with IC₅₀ value of 0.37 nM and the proliferation of MKN45 cells with IC₅₀ of 0.22 µM. It suppressed Met autophosphorylation with the downstream signaling through Gab-1, PLC-γ, FAK, Akt, STAT3, and ERK in cell. Complete inhibition of STAT3 and ERK phosphorylation was observed in MKN45 cells treated with 20 at the concentration of 100 nM. A computation simulation study was performed to reveal the interaction of 20 with Met.
Introduction

Met is a transmembrane receptor tyrosine kinase expressed in normal and malignant cells. Upon binding to its natural ligand hepatocyte growth factor/scatter factor (HGF/SF), Met induces several signaling pathways responsible for cell proliferation, motility, migration, and survival.\textsuperscript{1-3} The cellular events play an important role in normal embryonic development and wound healing\textsuperscript{4} nevertheless overexpression of HGF and/or Met or activating mutation of Met can lead to cancer.\textsuperscript{5} As a result, Met is considered as a potential target for cancer therapy.\textsuperscript{6-8}

C-5 substituted pyrrole–indolin-2-ones SU11274 (1)\textsuperscript{9} and PHA665752 (2)\textsuperscript{10} are the prototype Met kinase inhibitors from in silico design before the three dimensional structure of Met was characterized (Figure 1). They are obtained using the homology model of Met built from the closely related FGFR1 kinase.\textsuperscript{11} The C-4’ amino–amido side chains of 1 and 2 were designed to modulate the pharmacokinetic properties\textsuperscript{12} and would provide additional interactions with the enzyme to enhance the activities. Though 1 and 2 are only used as the biology tool, subsequent scaffold hopping based on the structure of 2 led to PF-2341066 (3),\textsuperscript{13} the most advanced Met and ALK kinase inhibitor recently approved by FDA for the treatment of NSCLC (see Figure 1).

Figure 1

Based on the literatures reporting the discovery of pyrrole–indolin-2-ones as multikinase inhibitors for VEGFR-2, FGFR1, and PDGFR-β kinases, different inhibition profiles are observed with different positions of the side chains.\textsuperscript{14,15} For FGFR1 inhibition, moving the C-4’ side chain in a pyrrole–indolin-2-one to the C-3’ position provided compounds with 5- to 10-fold increased potency (see the structures of 1 and 2 in Figure 1). As compound 2 was designed from the Met homology model of FGFR1, we envisaged that pyrrole–indolin-2-ones with C-3’ side chains would also be potent Met inhibitors. Currently, the role of C-3’ side chain for Met inhibition has not been studied in detail.

In this study, we used pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-one of scaffold 4 as the template to study the role of C-3’ substituents for the inhibition of Met kinase. Except for the C-3’ substituents, scaffold 4 possesses a pyrrole-fused cyclohexanone moiety for patent purposes.\textsuperscript{16}
Subsequent optimization led to compounds 10, 20, and 22–24 that demonstrated subnanomolar IC\textsubscript{50} values for Met inhibition in the biochemical assay. A computation modeling was used to reveal the different interactions of the C-3’ side chain of 4 with Met in comparison with the C-4’ side chain of 2. We expected the structural information would contribute to the further design of new scaffolds against Met, as in the case of designing 3 from 1 and 2.

Results and discussion

Chemistry

Scheme 1 presents the synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7–12 of scaffold 4 bearing C-3’ carboxyl-, ethoxycarbonyl-, and amido-ethyl side chains (X = CO\textsubscript{2}H, CO\textsubscript{2}Et, and CONR\textsubscript{2}R\textsubscript{3} in 4, Figure 1). 2-Formylpyrrole 5, prepared according to our published procedures,\textsuperscript{17} was condensed with 5-[(2,6-dichlorobenzyl)sulfonyl]indolin-2-one (6) to provide compound 7 bearing a C-3’ carboxylethyl group in 89% yield. Esterification of 7 in EtOH in the presence of SOCl\textsubscript{2} provided 8 that possessed a C-3’ ethoxycarbylethyl group in 92% yield. Amidation of the carboxyl group in 7 by various amines using CDI as the coupling agent provided pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 9–12 bearing a C-3’ amido-ethyl group in 70–85% yields.

Scheme 1

For the synthesis of pyrrole–indolin-2-ones 20–24 bearing C-3’ aminopropyl side chains (X = CH\textsubscript{2}NR\textsubscript{2}R\textsubscript{3} in 4, Figure 1), pyroles 13a–e served as the starting material to react with phenylsulfonyl chloride (PhSO\textsubscript{2}Cl) in the presence of NaOH to provide the N\textsuperscript{1}-protected pyrrole 14a–e in 86–94% yields (Scheme 2). The carbonyl groups in 14a–e could be reacted with ethane-1,2-dithiol to give the dithiane 15a–e in 87–99% yields. Removal of phenylsulfonyl group in 15a–e under basic condition provided 16a–e in 96% to >99% yields, which was treated with LiAlH\textsubscript{4} in THF at room temperature to afford amine 17a–e in 60–96% yields. Introduction of a formyl group to the C-2 position of 17a–e was accomplished by use of Vilsmeier-Haack reaction to generate 18a–e in 71–97% yields. Deprotection of the dithianyl group using HgCl\textsubscript{2} afforded 19a–e in 66–93% yields, which was eventually condensed with 5-(2,6-dichlorobenzyl)sulfonyl]indolin-2-one (6) to
provide the corresponding 20–24 with aminopropyl groups at their C-3’ positions in 60–78% yields.

**Scheme 2**

*In vitro potency*

The IC$_{50}$ values of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7–12 and 20–24 for the inhibition of Met kinase and the proliferation of MKN45 cells (gastric carcinoma) that carry an amplified Met proto-oncogene$^{18}$ are presented in Table 1. We also tested the potencies of 7–12 and 20–24 for the inhibition of Aurora kinases for comparison due to pyrrole–indolin-2-ones possessing a C-5 bulky group and a C-3 carboxylethyl side chain were found to show Aurora kinase inhibitory activities in our previous study.$^{17}$ As shown in Table 1, the prototype pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-one 7 bearing a C-3’ carboxyethyl group displayed an IC$_{50}$ value of 30.40 nM for Met inhibition. However, it did not show significant antiproliferative activity for MKN45 cells (IC$_{50}$ > 10 µM). Compound 8, the ethyl ester analog to 7, showed a slightly improved potency (IC$_{50}$: 20.20 nM) as well as enhanced antiproliferative activity for MKN45 (IC$_{50}$: 3.25 µM). These two compounds did not significantly inhibit Aurora kinases except 7 inhibited Aurora A with an IC$_{50}$ value of 855 nM.

**Table 1**

For further improving the Met and MKN45 inhibitory activities, we evaluated compounds 9–12 with the C-3’ amidoethyl substituents. These compounds showed remarkable improvements for Met inhibition with IC$_{50}$ values less than 10 nM (IC$_{50}$ for 2: 9 nM).$^{19}$ The most potent amido compound 10 with a $\text{N,N-diethylaminoethylamino (Et}_2\text{NCH}_2\text{CH}_2\text{NH–)}$ substituent inhibited Met at subnanomolar concentration (IC$_{50}$: 0.15 nM) and showed antiproliferative activity for MKN45 cells at low micromolar concentration (IC$_{50}$: 2.34 µM). The N-methylpiperazinyl analog 12 displayed the most potent antiproliferative activity (MKN45 IC$_{50}$: 0.99 µM) among the amido compounds 9–12. Respective comparison of kinase and cell activities of 9 with 11 and 10 with 12 suggested that the NH group in the amido moiety would profit the enzymatic but not the cell activity. The difference in the cell activities of these compounds might come from their different cell permeability or inhibition profiles against other kinases, e.g., VEGFR-2, PDGFR, and FGFR1.$^{15}$
Pyrrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 20–24 bound with C-3’ aminopropyl groups showed enhanced activities both for Met kinase and MKN45 cells (Table 1). Except for 21 with a diethylamino group, these compounds inhibited Met at subnanomolar concentrations (IC<sub>50</sub>: 0.26–0.43 nM) and demonstrated better antiproliferative activities than their carboxyl, ester, and amido analogs (i.e. 7–12) on MKN45 cells with IC<sub>50</sub> values less than 1.0 µM. The promising results from the biochemical assay shown in Table 1 indicated that pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold 4 with C-3’ side chains also showed promising Met kinase inhibitory activity as their C-4’ analog 2. Furthermore, the use of the C-3’ ethyl ester, amido, and amino side chains also diminished the activity of the corresponding compounds against Aurora A and Aurora B kinases (IC<sub>50</sub> > 1.0 µM, Table 1) in comparison with the use of a C-3’ carboxyl side chain (compound 7). A great selectivity (~100- to 1000-fold) was thus achieved for Met over Aurora kinases for the compounds in Table 1.

Western blotting

As shown in Table 1, the great enhancement for the Met inhibition by compounds 20 and 22–24 did not reflect on their antiproliferative activities (MKN45 IC<sub>50</sub>: 0.22–0.82 µM). We then characterized the effect of potent compound 20 (MKN45 IC<sub>50</sub>: 0.22 µM) on Met signal transduction in MKN45 cells by Western blot, and the results are presented in Figure 2. Strong inhibition of tyrosine phosphorylation in the Met kinase domain (pY1234/1235) was observed with 20 at a concentration of 1.1 µM and the reduced level of phosphorylated Met (IC<sub>50</sub> ~ 0.1–1.1 µM) correlated to the antiproliferative IC<sub>50</sub> value of 20 (0.22 µM). The phosphorylation of Met docking protein Gab-1 was completely inhibited at the concentration of 0.4 µM. Compound 20 also potently inhibited the constitutive downstream signaling through PLC-γ, FAK, Akt, STAT-3, and Erk, which provided the evidence of Met inhibition by 20 in cell. Regarded as the downstream cellular effectors of Met inhibition, the phosphorylation of STAT3 and ERK in MKN45 cells was completely abolished by 20 at the concentration below 100 nM, showing better correlation to the results from biochemical assay than the antiproliferative activity.

**Figure 2**

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**Kinase profiling**

Since compound 20 shared a similar scaffold with the multikinase inhibitor sunitinib, we explored its inhibition profile against a panel of 451 kinases at the concentration of 1.0 µM on DiscoveRx KinomeScan scanMax platform. The results revealed that CAMKK1, CDKL2, CSF1R, DCAMKL3, FLT3 (D835Y), FLT3 (ITD), GRK1, GRK7, IRAK4, JAK2 (JH1 domain-catalytic), MAP4K2, MAP4K3, MAP4K4, MAP4K5, MET, MET(M1250T), MET(Y1235D), MST2, PDGFRB, RSK3 (Kin.Dom.1-N-terminal), TAOK1, TAOK2, TAOK3, TNK, TRKB, and YSK4 kinases were inhibited significantly by 20 with a percentage of control of < 10% (see the table in the Supporting Information).

**Molecular modeling**

To explore the interaction of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold 4 with Met, we modeled the binding mode of the potent compound 20 as well as reference compound 2 in Met (PDB code 2RFS) using docking simulations. After energy minimization, the binding poses of 20 and 2 were overlapped as illustrated in Figure 3. Obviously, 20 shared similar binding pose to 2 in Met. Both compounds formed two H-bonds from N^1–H and N^1'-H with Pro1158 and Met1160, respectively, and an H-bond from C=O with Met1160 in the hinge region. The oxygen atom of the C-5 sulfonyl group formed an H-bond with Asp1222, and the 2,6-dichlorophenyl fitted the hydrophobic pocket formed by Val1092, Tyr1230, and Gly1085 from the analysis of Ligplot. However, the side chain at the C-3’ position of 20 headed to the different region from the C-4’ amino–amido group in 2, and thus interacted with different residues in Met other than 2: 20 interacted with Ile1084 and 2 interacted with Tyr1159 and His1094 (Figure 3). The hydrophilic interaction of the C-3’ dimethylamino group might contribute the promising activity of 20.

**Figure 3**

**Conclusions**

Pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold 4 with a C-3’ side chain was designed as potent Met kinase inhibitors. Structural optimization led to compounds 10, 20, and 22–24 that showed subnanomolar IC<sub>50</sub> values in the biochemical assay. Computation modeling
revealed that the C-3’ side chain of 4 interacted with Ile1084 and would contribute to its promising activity. The potent compound 20 possessed an IC₅₀ value of 0.37 nM also inhibited the proliferation of MKN45 cells (IC₅₀: 0.22 µM). Cellular events including the inhibition of Met autophosphorylation and the downstream signaling through Gab-1, PLC-γ, FAK, Akt, STAT-3, and ERK were found on MKN45 cells treated with 20. Among the proteins, STAT3 and ERK were completely inhibited by 20 at the concentration of 100 nM.

**Experimental**

**Chemistry**

Reagents were used as purchased without further purification. 5-(2,6-Dichlorobenzyl)sulfonylindolin-2-one (6) was prepared according to literature procedures.²²,²³ Proton NMR spectra were recorded on a Bruker (500 MHz) spectrometer with CDCl₃ or DMSO-d₆ as solvent. Multiplicities are abbreviated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; J, coupling constant (hertz). Carbon-13 NMR spectra were obtained from a Bruker spectrometer (125 MHz) by use of CDCl₃ or DMSO-d₆ as solvent. Carbon-13 chemical shifts are referenced to the center peak of CDCl₃ (δ 77.0 ppm) or DMSO-d₆ (δ 39.5 ppm). ESI-MS spectra were recorded with an Applied Biosystems API 300 mass spectrometer. The purities of the compounds were greater than 95% as determined from reversed-phase HPLC. The synthesis of compounds 5, 13a–e, 17a–e, 18a–e, and 19a–e was followed our previously reported procedures.¹⁷

**General procedure for the synthesis of N-phenylsulfonylpyrroles 14a–e.** A CH₂Cl₂ solution (250 mL) containing NaOH suspension (0.10 mol, 5.0 equiv) was added with compound 13a–e (~0.020 mol, 1.0 equiv). The mixture was stirred at 0 °C for 30 min. A solution of phenylsulfonyl chloride (0.030 mol, 1.5 equiv) in CH₂Cl₂ (50 mL) was slowly added to the reaction mixture in a period of 30 min. After the addition was completed, the reaction mixture was stirred for additional 30 min at 0 °C and for 12 h at room temperature. The solution was added with H₂O (100 mL) and the organic layer was separated. The aqueous layer was re-extracted with CH₂Cl₂ (3 × 50 mL), and the combined organic layer was dried over MgSO₄(s) and concentrated under reduced pressure to give the desired 14a–e in 86–94% yields.
\(N,N\text{-Dimethyl-3-[4-oxo-1-(phenylsulfonyl)-4,5,6,7-tetrahydro-}1H\text{-indol-3-yl]propanamide (14a).}

Yield: 94%; \(^1\text{H} \text{NMR (CDCl}_3\text{) } \delta \text{ 7.85 (d, } J = 7.6 \text{ Hz, 2 H), 7.66 (t, } J = 7.5 \text{ Hz, 1 H), 7.55–7.58 (m, 2 H), 7.06 (s, 1 H), 3.02 (s, 3 H), 2.89–2.98 (m, 7 H), 2.56–2.61 (m, 2 H), 2.36–2.41 (m, 2 H), 2.02–2.09 (m, 2 H); ESI-MS } m/z \text{ 374.7 (M + H)}.\)

\(N,N\text{-Diethyl-3-[4-oxo-1-(phenylsulfonyl)-4,5,6,7-tetrahydro-}1H\text{-indol-3-yl]propanamide (14b).}

Yield: 93%; \(^1\text{H} \text{NMR (CDCl}_3\text{) } \delta \text{ 7.85 (d, } J = 7.6 \text{ Hz, 2 H), 7.66 (t, } J = 7.5 \text{ Hz, 1 H), 7.54–7.57 (m, 2 H), 7.06 (s, 1 H), 3.29–3.38 (m, 4 H), 2.96 (t, } J = 6.8 \text{ Hz, 4 H), 2.54–2.59 (m, 2 H), 2.37–2.43 (m, 2 H), 2.03–2.10 (m, 2 H), 1.07–1.13 (m, 6 H); ESI-MS } m/z \text{ 402.9 (M + H)}.\)

\(3\text{-[3-Oxo-3-(pyrrolidin-1-yl)propyl]-1-(phenylsulfonyl)-6,7-dihydro-}1H\text{-indol-4(5H)-one (14c).}

Yield: 89%; \(^1\text{H} \text{NMR (CDCl}_3\text{) } \delta \text{ 7.85 (d, } J = 7.5 \text{ Hz, 2 H), 7.65 (t, } J = 7.5 \text{ Hz, 1 H), 7.53–7.56 (m, 2 H), 7.06 (s, 1 H), 3.38–3.45 (m, 4 H), 2.92–2.97 (m, 4 H), 2.50–2.55 (m, 2 H), 2.34–2.41 (m, 2 H), 2.02–2.09 (m, 2 H), 1.85–1.92 (m, 2 H), 1.77–1.84 (m, 2 H); ESI-MS } m/z \text{ 400.8 (M + H)}.\)

\(3\text{-[3-Morpholino-3-oxopropyl]-1-(phenylsulfonyl)-6,7-dihydro-}1H\text{-indol-4(5H)-one (14d).}

Yield: 89%; \(^1\text{H} \text{NMR (CDCl}_3\text{) } \delta \text{ (d, } J = 7.6 \text{ Hz, 2 H), 7.67 (t, } J = 7.5 \text{ Hz, 1 H), 7.56–7.59 (m, 2 H), 7.07 (s, 1 H), 3.55–3.65 (m, 8 H), 2.90–2.99 (m, 4 H), 2.57–2.62 (m, 2 H), 2.37–2.42 (m, 2 H), 2.04–2.11 (m, 2 H); ESI-MS } m/z \text{ 417.0 (M + H)}.\)

\(3\text{-[3-(4-Methylpiperazin-1-yl)-3-oxopropyl]-1-(phenylsulfonyl)-6,7-dihydro-}1H\text{-indol-4(5H)-one (14e).}

Yield: 86%; \(^1\text{H} \text{NMR (CDCl}_3\text{) } \delta \text{ 7.86 (d, } J = 7.5 \text{ Hz, 2 H), 7.67 (t, } J = 7.5 \text{ Hz, 1 H), 7.56–7.59 (m, 2 H), 7.06 (s, 1 H), 3.50–3.64 (m, 4 H), 2.90–2.98 (m, 4 H), 2.57–2.62 (m, 2 H), 2.33–2.41 (m, 6 H), 2.28 (s, 3 H), 2.04–2.10 (m, 2 H) ESI-MS: } m/z \text{ 430.0 (M + H)}.\)

**General procedure for the synthesis of dithianes 15a–e.** To a methanol solution containing compound 14a–e (~0.20 mol, 1.0 equiv) and ethane-1,2-dithiol (1.1 equiv) was added BF\(_3\)-OEt\(_2\) (0.25 equiv). The reaction mixture was stirred at room temperature for 45 min. The solution was mixed with 10% aqueous NaOH (50 mL) and concentrated under reduced pressure to remove the methanol. The solution was extracted with CH\(_2\)Cl\(_2\) (3 × 200 mL), and the organic layer was washed with H\(_2\)O, dried over MgSO\(_4\)(s), and concentrated under reduced pressure to give the desired 15a–e in 87–99% yields.
N,N-Dimethyl-3-[1’-(phenylsulfonyl)-1’,5’,6’,7’-tetrahydrospiro[[1,3]dithiolane-2,4’-indole]-3’-yl]propanamide (15a). Yield: 87%; \(^1^H\) NMR (CDCl\(_3\)) \(\delta 7.77\) (d, \(J = 7.5\) Hz, 2 H), 7.62 (t, \(J = 7.5\) Hz, 1 H), 7.51–7.54 (m, 2 H), 6.99 (s, 1 H), 3.42–3.50 (m, 2 H), 3.30–3.36 (m, 2 H), 3.06–3.11 (m, 2 H), 3.02 (s, 3 H), 2.97 (s, 3 H), 2.61–2.70 (m, 4 H), 2.15–2.19 (m, 2 H), 1.80–1.89 (m, 2 H); ESI-MS \(m/z\) 450.8 (M + H).^

N,N-Diethyl-3-[1’-(phenylsulfonyl)-1’,5’,6’,7’-tetrahydrospiro[[1,3]dithiolane-2,4’-indole]-3’-yl]propanamide (15b). Yield: 92%; \(^1^H\) NMR (CDCl\(_3\)) \(\delta 7.77\) (d, \(J = 7.6\) Hz, 2 H), 7.61 (t, \(J = 7.4\) Hz, 1 H), 7.50–7.54 (m, 2 H), 6.99 (s, 1 H), 3.44–3.51 (m, 2 H), 3.37–3.43 (m, 2 H), 3.30–3.36 (m, 4 H), 3.07–3.13 (m, 2 H), 2.61–2.69 (m, 4 H), 2.15–2.19 (m, 2 H), 1.82–1.88 (m, 2 H), 1.10–1.20 (m, 6 H); ESI-MS \(m/z\) 478.8 (M + H).^

3-[1’-(Phenylsulfonyl)-1’,5’,6’,7’-tetrahydrospiro[[1,3]dithiolane-2,4’-indole]-3’-yl]-1-(pyrrolidin-1-yl)propan-1-one (15c). Yield: 90%; \(^1^H\) NMR (CDCl\(_3\)) \(\delta 7.74–7.80\) (m, 2 H), 7.62 (t, \(J = 7.5\) Hz, 1 H), 7.51–7.54 (m, 2 H), 6.99 (s, 1 H), 3.39–3.53 (m, 6 H), 3.28–3.38 (m, 2 H), 3.05–3.15 (m, 2 H), 2.57–2.68 (m, 4 H), 2.12–2.22 (m, 2 H), 1.92–2.00 (m, 2 H), 1.79–1.90 (m, 4 H); ESI-MS \(m/z\) 477.0 (M + H).^

1-Morpholino-3-[1’-(phenylsulfonyl)-1’,5’,6’,7’-tetrahydrospiro[[1,3]dithiolane-2,4’-indole]-3’-yl]propan-1-one (15d). Yield: 99%; \(^1^H\) NMR (CDCl\(_3\)) \(\delta 7.77\) (d, \(J = 7.4\) Hz, 2 H), 7.62 (t, \(J = 7.4\) Hz, 1 H), 7.51–7.54 (m, 2 H), 6.99 (s, 1 H), 3.61–3.69 (m, 2 H), 3.50–3.54 (m, 2 H), 3.42–3.49 (m, 2 H), 3.30–3.37 (m, 2 H), 3.07–3.12 (m, 2 H), 2.61–2.71 (m, 4 H), 2.15–2.19 (m, 2 H), 1.83–1.87 (m, 2 H); ESI-MS \(m/z\) 492.9 (M + H).^

1-(4-Methylpiperazin-1-yl)-3-[1’-(phenylsulfonyl)-1’,5’,6’,7’-tetrahydrospiro[[1,3]dithiolane-2,4’-indole]-3’-yl]propan-1-one (15e). Yield: 96%; \(^1^H\) NMR (CDCl\(_3\)) \(\delta 7.77\) (d, \(J = 7.5\) Hz, 2 H), 7.62 (t, \(J = 7.5\) Hz, 1 H), 7.50–7.53 (m, 2 H), 6.99 (s, 1 H), 3.64–3.68 (m, 2 H), 3.50–3.54 (m, 2 H), 3.42–3.49 (m, 2 H), 3.30–3.37 (m, 2 H), 3.07–3.12 (m, 2 H), 2.61–2.72 (m, 4 H), 2.36–2.40 (m, 4 H), 2.31 (s, 3 H), 2.16–2.20 (m, 2 H), 1.82–1.88 (m, 2 H); ESI-MS \(m/z\) 506.0 (M + H).^

**General procedure for the synthesis of pyrroles 16a–e.** Compounds 15a–e (1.0 equiv) in a solution of 30% MeOH and 70% H\(_2\)O were added with NaOH (5.0 equiv). The reaction mixture was
heated at reflux for 3.0 h. The solution was neutralized with 3.0 N HCl and the resulting precipitate was collected to provide the desired 16a–e in 96% to >99% yields. The spectroscopic data of were consistence with our previous report.17

General procedure for the synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7 and 20–24. 2-Formylpyrroles 5 or 19a–e (~0.50 mmol, 1.0 equiv) was mixed with 5-(2,6-dichlorobenzyl)sulfonylindolin-2-one (6, 1.0 equiv) and piperidine (0.10 mL) in EtOH (5.0 mL). The reaction mixture was heated at reflux for 12 h. The resulting precipitate was filtered and washed with EtOH to provide the corresponding pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7 and 20–24 in 60–89% yields.

(Z)-3-2-[[5-(2,6-Dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene]methyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-3-yl)propanoic Acid (7). Yield: 89%; 1H NMR (DMSO-d6) δ 13.65 (s, 1 H), 12.11 (brs, 1 H), 11.52 (s, 1 H), 8.21 (d, J = 1.6 Hz, 1 H), 7.94 (s, 1 H), 7.46–7.52 (m, 3 H), 7.39 (dd, J = 8.7, 7.4 Hz, 1 H), 7.06 (d, J = 8.2 Hz, 1 H), 4.88 (s, 2 H), 3.20 (t, J = 7.6 Hz, 2 H), 2.93 (t, J = 6.1 Hz, 2 H), 2.52 (t, J = 7.6 Hz, 2 H), 2.40–2.44 (m, 2 H), 2.02–2.10 (m, 2 H); 13C NMR (DMSO-d6) δ 19.28, 21.83, 21.91, 33.87, 37.50, 56.89, 108.78, 114.70, 118.27, 118.77, 124.91, 124.99, 125.43, 126.16, 126.53, 127.81, 130.16, 131.02, 133.03, 135.51, 142.09, 147.59, 168.70, 172.97, 192.83; ESI-MS m/z 573.0 (M + H)+.

(Z)-5-(2,6-Dichlorobenzylsulfonyl)-3-([3-(dimethylamino)propyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-2-yl)methylene)indolin-2-one (20). Yield: 70%; 1H NMR (DMSO-d6) δ 13.51 (s, 1 H), 11.46 (brs, 1 H), 7.97 (s, 1 H), 7.42–7.57 (m, 3 H), 7.34–7.42 (m, 2 H), 7.07 (d, J = 6.9 Hz, 1 H), 4.87 (s, 2 H), 2.92–3.05 (m, 4 H), 2.35–2.45 (m, 4 H), 1.97–2.19 (m, 8 H), 1.57–1.72 (m, 2 H); 13C NMR (DMSO-d6) δ 19.73, 21.84, 21.96, 26.58, 37.56, 43.98, 56.37, 56.86, 108.88, 114.08, 117.69, 118.80, 124.90, 124.95, 125.04, 125.77, 126.62, 127.83, 130.17, 130.82, 134.51, 135.52, 142.03, 147.73, 168.67, 192.72; ESI-MS m/z 586.0 (M + H)+.

(Z)-5-(2,6-Dichlorobenzylsulfonyl)-3-([3-(diethylamino)propyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-2-yl)methylene)indolin-2-one (21). Yield: 78%; 1H NMR (DMSO-d6) δ 13.62 (s, 1 H), 8.11 (s, 1 H), 7.89 (s, 1 H), 7.43–7.56 (m, 3 H), 7.37–7.42 (m, 1 H), 7.07 (d, J = 8.10 Hz, 1 H), 8.47 (s, 2
\( \text{H}, 2.90–3.02 \text{ (m, 4 H)}, 2.32–2.47 \text{ (m, 8 H)}, 2.01–2.12 \text{ (m, 2 H)}, 1.65–1.71 \text{ (m, 2 H)}, 0.91 \text{ (t, } J = 7.1 \text{ Hz, 6 H}); \) \(^{13}\text{C NMR (DMSO-}{d_6}\) \( \delta 10.36, 21.02, 21.85, 21.98, 26.59, 37.59, 44.85, 50.24, 56.88, 108.86, 114.10, 117.77, 118.83, 124.97, 125.12, 125.48, 126.33, 126.51, 127.81, 130.15, 130.95, 134.42, 135.50, 142.04, 147.72, 168.70, 192.67; \) ESI-MS \( m/z \) 614.0 (M + H\(^+\)).

\((Z)-5-(2,6-\text{Dichlorobenzylsulfonyl})-3-\{4-\text{oxo}-3-[3-(\text{pyrrolidin-1-yl})\text{propyl}]\}-4,5,6,7-\text{tetrahydro-1H-indol-2-yl}\text{methylene}\text{indolin-2-one (22). Yield: 66%; } ^1\text{H NMR (DMSO-}{d_6}\) \( \delta 13.62 \text{ (s, 1 H)}, 11.52 \text{ (s, 1 H)}, 8.05 \text{ (s, 1 H)}, 7.86 \text{ (s, 1 H)}, 7.45–7.50 \text{ (m, 3 H)}, 7.34–7.41 \text{ (m, 1 H)}, 7.05 \text{ (d, } J = 8.2 \text{ Hz, 1 H)}, 4.85 \text{ (s, 2 H)}, 3.03 \text{ (t, } J = 6.8 \text{ Hz, 2 H)}, 2.93 \text{ (t, } J = 5.9 \text{ Hz, 2 H)}, 2.27–2.42 \text{ (m, 8 H)}, 2.01–2.09 \text{ (m, 2 H)}, 1.63–1.73 \text{ (m, 6 H}); \) \(^{13}\text{C NMR (DMSO-}{d_6}\) \( \delta 20.92, 21.84, 21.97, 22.17, 28.05, 37.58, 52.45, 53.26, 56.87, 108.78, 114.18, 117.66, 118.78, 124.93, 124.99, 125.04, 126.40, 126.51, 127.81, 130.17, 130.93, 134.71, 135.52, 142.03, 147.70, 168.70, 192.70; \) ESI-MS \( m/z \) 612.0 (M + H\(^+\)).

\((Z)-5-(2,6-\text{Dichlorobenzylsulfonyl})-3-\{3-(3-\text{morpholinopropyl})-4-\text{oxo}-4,5,6,7-\text{tetrahydro-1H-indol-2-yl}\text{methylene}\text{indolin-2-one (23). Yield: 72%; } ^1\text{H NMR (DMSO-}{d_6}\) \( \delta 13.64 \text{ (s, 1 H)}, 11.48 \text{ (brs, 1 H)}, 8.21 \text{ (s, 1 H)}, 7.88 \text{ (s, 1 H)}, 7.46–7.50 \text{ (m, 3 H)}, 7.33–7.41 \text{ (m, 1 H)}, 7.05 \text{ (d, } J = 8.2 \text{ Hz, 1 H)}, 4.86 \text{ (s, 2 H)}, 3.51–3.57 \text{ (m, 4 H)}, 3.02 \text{ (t, } J = 7.1 \text{ Hz, 2 H)}, 2.93 \text{ (t, } J = 6.0 \text{ Hz, 2 H)}, 2.21–2.42 \text{ (m, 8 H)}, 1.99–2.09 \text{ (m, 2 H)}, 1.67–1.74 \text{ (m, 2 H}); \) \(^{13}\text{C NMR (DMSO-}{d_6}\) \( \delta 21.12, 21.84, 21.97, 22.17, 28.05, 37.58, 52.45, 53.26, 56.87, 108.78, 114.18, 117.66, 118.78, 124.94, 124.99, 125.04, 126.40, 126.51, 127.81, 130.17, 130.93, 134.71, 135.52, 142.03, 147.70, 168.70, 192.70; \) ESI-MS \( m/z \) 628.0 (M + H\(^+\)).

\((Z)-5-(2,6-\text{Dichlorobenzylsulfonyl})-3-\{3-(4-\text{methylpiperazin-1-yl})\text{propyl})-4-\text{oxo}-4,5,6,7-\text{tetrahydro-1H-indol-2-yl}\text{methylene}\text{indolin-2-one (24). Yield: 60%; } ^1\text{H NMR (DMSO-}{d_6}\) \( \delta 13.67 \text{ (s, 1 H)}, 11.52 \text{ (brs, 1 H)}, 8.20 \text{ (s, 1 H)}, 7.85 \text{ (s, 1 H)}, 7.46–7.51 \text{ (m, 3 H)}, 7.37–7.42 \text{ (m, 1 H)}, 7.06 \text{ (d, } J = 8.2 \text{ Hz, 1 H)}, 4.88 \text{ (s, 2 H)}, 2.91–3.03 \text{ (m, 4 H)}, 2.22–2.44 \text{ (m, 10 H)}, 2.01–2.14 \text{ (m, 7 H)}, 1.70 \text{ (d, } J = 6.8 \text{ Hz, 2 H}); \) \(^{13}\text{C NMR (DMSO-}{d_6}\) \( \delta 21.22, 21.84, 21.97, 22.17, 26.07, 37.58, 52.34, 56.35, 56.90, 65.22, 108.81, 114.20, 118.05, 118.84, 124.94, 125.02, 125.10, 126.34, 126.46, 127.81, 130.16, 131.03, 134.94, 135.52, 142.02, 147.70, 168.72, 192.70; \) ESI-MS \( m/z \) 641.0 (M + H\(^+\)).
(Z)-Ethyl 3-(2-[[5-(2,6-dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene][methyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-3-yl]propanoate (8). To a solution of 7 (105.5 mg, 0.1840 mmole) in EtOH (2.0 mL) was added with SOCl₂ (0.10 mL). The reaction mixture was heated at reflux for 12 h. The solution was concentrated under reduced pressure. The residue was added with H₂O (5.0 mL) and heated. The solution was cooled to room temperature and the resultant solids were collected to give the desired 8 (101.8 mg, 0.1692 mmol) in 92% yield: ¹H NMR (DMSO-d₆) δ 13.68 (s, 1 H), 11.53 (brs, 1 H), 8.27 (dd, J = 1.1 Hz, 1 H), 7.95 (s, 1 H), 7.46–7.53 (m, 3 H), 7.34–7.42 (m, 1 H), 7.06 (dd, J = 8.2 Hz, 1 H), 4.88 (s, 2 H), 4.02 (q, J = 7.1 Hz, 2 H), 3.25 (t, J = 7.4 Hz, 2 H), 2.94 (t, J = 6.0 Hz, 2 H), 2.61 (t, J = 7.4 Hz, 2 H), 2.38–2.45 (m, 2 H), 2.01–2.11 (m, 2 H), 1.13 (t, J = 7.1 Hz, 3 H); ¹³C NMR (DMSO-d₆) δ 13.13, 19.12, 21.84, 21.88, 33.49, 37.47, 56.90, 58.83, 108.81, 114.87, 118.32, 118.77, 124.93, 124.99, 125.34, 126.23, 126.63, 127.81, 130.14, 131.08, 132.42, 135.50, 142.11, 147.61, 168.68, 171.27, 192.87; ESI-MS m/z 601.0 (M + H)⁺.

General procedure for the synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 9–12. Compound 7 (~1.0 g, 1.0 equiv) was suspended in CH₂Cl₂ (20 mL) and mixed with CDI (1.2 equiv). The reaction mixture was stirred at room temperature for 3.0 h. The solution was added with methylamine, N,N-diethylethane-1,2-diamine, pyrrolidine, or 1-methylpiperazine (2.0 equiv), and the reaction mixture was stirred at room temperature for 12 h. The solution was diluted with CH₂Cl₂ (200 mL), and washed with 0.1 N HCl (50 mL), saturated Na₂CO₃ (50 mL), and brine (50 mL). The organic layer was dried over anhydrous MgSO₄(s) and concentrated under reduced pressure to provide the desired 9–12 as yellow solids in 70–85% yields.

(Z)-3-(2-[[5-(2,6-Dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene][methyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-3-yl]-N-methylpropanamide (9). Yield: 85%; ¹H NMR (DMSO-d₆) δ 13.61 (s, 1 H), 11.49 (brs, 1 H), 8.22 (dd, J = 1.6 Hz, 1 H), 7.90 (s, 1 H), 7.72 (dd, J = 4.2, 4.2 Hz, 1 H), 7.46–7.53 (m, 3 H), 7.39 (dd, J = 8.7, 7.4 Hz, 1 H), 7.06 (d, J = 8.2 Hz, 1 H), 4.88 (s, 2 H), 3.22 (t, J = 7.3 Hz, 2 H), 2.94 (t, J = 6.1 Hz, 2 H), 2.48 (s, 3 H), 2.45–2.40 (m, 2 H), 2.37 (t, J = 7.3 Hz, 2 H), 2.03–2.10 (m, 2 H); ¹³C NMR (DMSO-d₆) δ 19.67, 21.85, 21.93, 24.44, 35.34, 37.54, 56.90, 108.80, 114.46, 118.06, 118.74, 125.00, 125.37, 126.23, 126.55, 127.81, 130.18, 131.10, 133.66, 135.51, 142.06,
(Z)-3-(2-[[5-(2,6-Dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene]methyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-3-yl)-N-[2-(diethylamino)ethyl]propanamide (10). Yield: 72%; $^1$H NMR (DMSO-$d_6$) δ 13.60 (s, 1 H), 11.49 (brs, 1 H), 8.19 (s, 1 H), 7.89 (s, 1 H), 7.64 (d, $J = 5.5$ Hz, 1 H), 7.48–7.54 (m, 3 H), 7.35–7.39 (m, 1 H), 7.06 (d, $J = 8.2$ Hz, 1 H), 4.87 (s, 2 H), 3.18–3.27 (m, 2 H), 2.89–3.03 (m, 4 H), 2.40–2.45 (m, 2 H), 2.37 (t, $J = 7.2$ Hz, 2 H), 2.28 (q, $J = 7.0$ Hz, 4 H), 2.22 (t, $J = 7.2$ Hz, 2 H), 2.01–2.12 (m, 2 H), 0.77 (t, $J = 7.0$ Hz, 6 H); $^{13}$C NMR (DMSO-$d_6$) δ 10.56, 19.74, 21.85, 21.93, 35.47, 35.93, 37.55, 45.51, 50.64, 56.89, 108.76, 114.41, 117.94, 118.73, 124.93, 125.05, 125.49, 126.34, 126.47, 127.80, 130.17, 131.13, 133.50, 135.52, 142.07, 147.50, 168.69, 170.21, 192.79; ESI-MS m/z 671.0 (M + H)$^+$. 

(Z)-5-(2,6-Dichlorobenzylsulfonyl)-3-((4-oxo-3-[3-oxo-3-(pyrrolidin-1-yl)propyl]-4,5,6,7-tetrahydro-1H-indol-2-yl)methylene)indolin-2-one (11). Yield: 82%; $^1$H NMR (DMSO-$d_6$) δ 13.64 (s, 1 H), 11.44 (s, 1 H), 8.22 (d, $J = 1.3$ Hz, 1 H), 7.94 (s, 1 H), 7.48–7.51 (m, 3 H), 7.35–7.42 (m, 1 H), 7.05 (d, $J = 8.2$ Hz, 1 H), 4.88 (s, 2 H), 3.19–3.39 (m, 6 H), 2.94 (t, $J = 5.9$ Hz, 2 H), 2.52–2.57 (m, 2 H), 2.40–2.45 (m, 2 H), 2.03–2.09 (m, 2 H), 1.79–1.86 (m, 2 H), 1.68–1.76 (m, 2 H); $^{13}$C NMR (DMSO-$d_6$) δ 19.35, 21.85, 21.93, 22.97, 24.65, 34.28, 37.54, 44.32, 44.83, 56.86, 108.75, 114.49, 118.19, 118.85, 124.99, 125.06, 125.41, 126.19, 126.48, 127.82, 130.15, 131.04, 133.81, 135.51, 142.04, 147.58, 168.70, 168.75, 192.86; ESI-MS m/z 626.0 (M + H)$^+$. 

(Z)-5-(2,6-Dichlorobenzylsulfonyl)-3-([3-[3-(4-methylpiperazin-1-yl)-3-oxopropyl]-4-oxo-4,5,6,7-tetrahydro-1H-indol-2-yl)methylene]indolin-2-one (12). Yield: 70%; $^1$H NMR (DMSO-$d_6$) δ 13.66 (s, 1 H), 11.51 (brs, 1 H), 8.21 (d, $J = 0.93$ Hz, 1 H), 7.91 (s, 1 H), 7.47–7.50 (m, 3 H), 7.35–7.42 (m, 1 H), 7.05 (d, $J = 8.2$ Hz, 1 H), 4.88 (s, 2 H), 3.37–3.45 (m, 4 H), 3.20 (t, $J = 7.6$ Hz, 2 H), 2.94 (t, $J = 6.0$ Hz, 2 H), 2.57 (t, $J = 7.6$ Hz, 2 H), 2.43 (t, $J = 6.0$ Hz, 2 H), 2.14–2.17 (m, 4 H), 2.12 (s, 3 H), 2.04–2.09 (m, 2 H); $^{13}$C NMR (DMSO-$d_6$) δ 19.85, 21.86, 21.93, 32.63, 37.53, 39.90, 43.75, 44.65, 53.31, 53.76, 56.89, 108.76, 114.63, 118.27, 118.89, 124.99, 125.39, 126.23, 126.54, 127.83, 130.17, 131.05, 133.42, 135.51, 142.06, 147.62, 168.70, 169.02, 192.98; ESI-MS m/z 655.0 (M + H)$^+$. 

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Biological and computational methods

Met and Aurora kinase assays

Inhibition of kinase activity by the test compound was measured by quantifying the amount of
$^{33}$P incorporation into the substrate in the presence of a test compound. Standard assay conditions
utilized 5 ng of recombinant Met kinase (Upstate, Cat. No. 14-526), 1 µg Poly Glu-Tyr (Sigma,
Product Number P0275), 100 µM ATP, 0.2 µCi $[^{33}$P]ATP (specific activity 3000 Ci/mmol,
PerkinElmer), 8 mM MOPS-NaOH (pH 7.0), and 1 mM EDTA in a total volume of 25 µL. Reaction
mixtures were incubated at 30 °C for 30 min and reactions stopped by addition of 3% phosphoric
acid; mixtures were harvested onto a 96-well GF/B UniFilter (PerkinElmer) using a unifilter
harvester (PerkinElmer), and counted with a TopCount microplate scintillation counter
(PerkinElmer). The IC$_{50}$ values of inhibitors were determined after assays were conducted at 3-fold
serially diluted concentrations of each compound in duplicate. The results were analyzed using linear
regression software (GraphPad Prism version 4; GraphPad Software Inc., San Diego, CA). For
Aurora assay, 1 ng of recombinant Aurora A or Aurora B kinase (Upstate Biotechnology), 2 µg
peptide substrate (tetra-LRRRLSLG, synthesized by Genesis Biotech Inc.), 10 µM ATP (for Aurora A)
or 40 µM ATP (for Aurora B), 0.2 µCi $[^{33}$P]ATP (specific activity 3000 Ci/mmol, PerkinElmer), 8
mM MOPS-NaOH (pH 7.0), and 1 mM EDTA in a total volume of 25 µL were utilized.

Cell proliferation assay

The antiproliferative activity of the compounds with respect to human gastric carcinoma cell,
MKN45 (Japanese Collection of Research Bioresources), was measured using the CellTiter96 assay
kit (Promega) following the manufacturer’s instruction. In brief, the cells were maintained in DMEM
containing 0.2% FBS and incubated at 37 °C in 5% CO$_2$ atmosphere. Cells were plated at a density
of 2500 cells/well of a 96-well plate for 24 h, then treated with different concentrations of the
compounds, and incubated for another 72 hours. At the end of the incubation, CellTiter 96 Aqueous
One Solution Reagent (Promega) was added and incubated for another 3.0 hours. Cell viability was
determined by measuring absorbance at 490 nm using EMax® microplate reader (Molecular
Devices). Data were processed and analyzed using GraphPad Prism version 4.
Western blot analysis

The effect of Met inhibitors on cellular protein phosphorylation was evaluated by Western blot analysis on MKN45 cells. Cells grown to subconfluency were grown overnight in 0.5% (FBS) and DMEM. Cells in low serum were treated with compound 20 in DMSO (20 mM stock) for 30 minutes. Drug-treated cells were then harvested and washed twice with PBS, and the cell pellets were boiled in 2× SDS sample buffer and subjected to SDS–PAGE and immunoblotting analysis. Western blots were done using standard procedures and probed with the following antibodies: anti-Met (Cell Signaling Technology), anti-phospho-Met (Tyr1234/1235), anti-phospho-Gab-1 (Tyr627, Thermo Scientific), anti-phospho-FAK (Tyr861, Thermo Scientific), anti-phospho-PLC-γ (Tyr783), anti-phospho-Akt (Ser473), anti-phospho-STAT3 (Tyr705), anti-phospho-p44/42 ERK mitogen-activated protein kinase (Thr202/Tyr204), and horseradish peroxidase-conjugated secondary antibodies (Cell Signaling).

Computational method

The three dimensional structure of Met complexed with 1 was obtained from the Protein Data Bank (entry: 2RFS). Hydrogen atoms were added, and water molecules co-crystallized with the protein were removed from the original structure using Accelrys Discovery Studio 2.5 (Accelrys, Inc.). The modified crystal structure of Met was docked by a genetic algorithm based program (GOLD) using the Chem scoring (CCDC Software Limited, Cambridge, U.K.). The genetic algorithm was executed at the default settings, and the active site radius is 10 Å from inhibitor complex for the docking study. Two hundred genetic algorithm (GA) runs were performed for docking compounds 2 and 20. GOLD was run to save up to best docking solutions for each ligands. The results were visually analyzed using PyMOL program (http://www.pymol.org/) and Ligplot.21

Acknowledgements

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Note and references

†Electronic Supplementary Information (ESI) available: Ligplot diagrams of the ATP binding site of
Met complexed with compounds 2 and 20, and the kinase profiling data of 20. See DOI: xxx/xxx


Table 1  The enzymatic and cellular activities of 5-(2,6-dichlorobenzyl)sulfonylpyrrole–indolin-2-ones 7–12 and 20–24 with different C-3’ side chains as Met kinase inhibitors

<table>
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<tr>
<th>compd</th>
<th>X</th>
<th>Met IC(_{50}) (nM)(^a)</th>
<th>MKN45 IC(_{50}) (µM)(^{a,b})</th>
<th>%inhibition at 1.0 µM</th>
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<tr>
<td>7</td>
<td>HO–C(\stackrel{3}{-})O</td>
<td>30.40</td>
<td>&gt;10</td>
<td>65.5%(^c) 9.4%</td>
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<td>8</td>
<td>EtO–C(\stackrel{1}{-})O</td>
<td>20.20</td>
<td>3.25 ± 0.41</td>
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<td>9</td>
<td>MeHN–C(\stackrel{1}{-})O</td>
<td>4.22</td>
<td>&gt;10</td>
<td>13.0% 6.9%</td>
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<tr>
<td>10</td>
<td>Et(_2)N–HN–C(\stackrel{2}{-})O</td>
<td>0.15</td>
<td>2.34 ± 0.13</td>
<td>10.8% 9.2%</td>
</tr>
<tr>
<td>11</td>
<td>N–C(\stackrel{1}{-})O</td>
<td>7.17</td>
<td>9.25 ± 0.32</td>
<td>2.2% 5.5%</td>
</tr>
<tr>
<td>12</td>
<td>N–N–C(\stackrel{1}{-})O</td>
<td>2.94</td>
<td>0.99 ± 0.10</td>
<td>8.7% 1.3%</td>
</tr>
<tr>
<td>20</td>
<td>Me(_2)N–C</td>
<td>0.37</td>
<td>0.22 ± 0.03</td>
<td>2.7% 34.3%</td>
</tr>
<tr>
<td>21</td>
<td>Et(_2)N–H(_2)–C</td>
<td>7.70</td>
<td>0.40 ± 0.05</td>
<td>1.9% 6.2%</td>
</tr>
<tr>
<td>22</td>
<td>N–C(\stackrel{1}{-})O</td>
<td>0.31</td>
<td>0.82 ± 0.08</td>
<td>4.2% 22.3%</td>
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<tr>
<td>23</td>
<td>N–H(_2)–N–C</td>
<td>0.43</td>
<td>0.47 ± 0.05</td>
<td>23.7% 35.4%</td>
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<tr>
<td>24</td>
<td>N–N–H(_2)–N–C</td>
<td>0.26</td>
<td>0.24 ± 0.02</td>
<td>6.0% 15.3%</td>
</tr>
<tr>
<td>2</td>
<td>–</td>
<td>≥7</td>
<td>–</td>
<td>–  –</td>
</tr>
</tbody>
</table>

\(^a\) The IC\(_{50}\) values were averaged from two independent dose–response curves. \(^b\) A 72-h cell proliferation assay. \(^c\) IC\(_{50}\): 855 nM. \(^d\) Obtained from literature\(^{19}\).
Figure 1 Structures of SU11274 (1), PHA665752 (2), PF-2341066 (3), and 5-(2,6-dichlorobenzyl)sulfonylpyrrole–indolin-2-one of scaffold 4.

Figure 2 Effect of 20 on Met phosphorylation and signal transduction in MKN45 cells.
Figure 3 Overlapping of the docked binding modes of compounds 2 and 20 in the ATP binding site of Met kinase. Compound 2 is shown as light blue sticks, 20 is shown as yellow sticks, hydrogen bonds are shown as dashed gray line, and the oxygen, nitrogen, and sulfur atoms are colored in red, blue, and orange, respectively. The C-3’ side chain of 20 faced the region around Ile1084 and the C-4’ side chain of 2 faced the region around Tyr1159 and His1094.

Scheme 1 Synthesis of 5-(2,6-dichlorobenzyl)sulfonylpyrrole–indolin-2-ones 7–12 bearing C-3’ carboxyl-, ethoxycarbonyl-, or amido-ethyl side chains as Met kinase inhibitors. Reagents and conditions: (a) piperidine, EtOH, reflux, 12 h, 89%; (b) SOCl₂, EtOH, 92%; (c) 7, amine, CDI, CH₂Cl₂, 70–85%.
Scheme 2 Synthesis of 5-(2,6-dichlorobenzyl)sulfonylpyrrole-indolin-2-ones 20–24 equipped with C-3’ aminopropyl side chain. Reagents and conditions: (a) PhSO₂Cl, NaOH, CH₂Cl₂, 86–94%; (b) ethane-1,2-dithiol, BF₃·OEt₂, MeOH, 87–99%; (c) NaOH, H₂O, MeOH, reflux, 96% to >99%; (d) LiAlH₄, THF, r.t., 60–96%; (e) POCl₃, DMF, 71–97%; (f) HgCl₂, CaCO₃, CH₃CN, H₂O, 66–93%; (g) 5-(2,6-dichlorobenzylsulfonyl)indolin-2-one (6), piperidine, EtOH, reflux, 60–78%.

20. NR₂ = NMe₂ (70%)
21. NR₂ = NEt₂ (78%)
22. NR₂ = 1-pyrrolidinyl (66%)
23. NR₂ = 4-morpholinyl (72%)
24. NR₂ = 4-methyl-1-piperazinyl (60%)
Graphical abstract

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