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1           **Supramolecular solvent microextraction of gold prior to its**  
2           **determination by microsample injection system coupled with**  
3           **flame atomic absorption spectrometry \***

4  
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7  
8           **Abstract**

9           A supramolecular solvent based liquid-liquid microextraction (SsLLME) system for  
10          gold was developed prior to its microsample injection system coupled with flame atomic  
11          absorption spectrometric (MS-FAAS) determination. 1,3,4-thiadiazole-2,5-dithiol was used as  
12          complexing agent to obtain hydrophobic complex. The analytical factors affecting the  
13          microextraction efficiency, such as pH, type and volume of supramolecular solvent, amount  
14          of complexing agent, ultrasonication and centrifuge time and sample volume were  
15          investigated. The limit of detection (LOD), enhancement factor (EF) and relative standard  
16          deviation (RSD) of the method was  $1.5 \mu\text{g L}^{-1}$ , 51 and 4.2 %, respectively. The accuracy of  
17          method was checked by analysis of CDN-GS-3D Gold Ore certified reference material and  
18          addition-recovery tests. The method was successfully applied for the determination of gold in  
19          environmental samples.

20          **Keywords:** Supramolecular solvent, Microextraction, Gold, Flame atomic absorption  
21                           spectrometry.

22          \*This study is a part of PhD thesis of Erkan Yilmaz

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24

## 25 1. Introduction

26 Gold is the most important noble metal, due to its usage in jewelry, pharmaceutical  
27 application and industry, etc.<sup>1-4</sup> Concentrations of gold in real samples like environmental and  
28 pharmaceutical materials are usually below the detection limit of the flame atomic absorption  
29 spectrometer to be determined directly, due to insufficient sensitivity and effects of the matrix  
30 interference.<sup>1, 5, 6</sup> Hence, under these circumstances, a simple, sensitive and selective  
31 separation and preconcentration procedure method for determination of trace gold in real  
32 samples were required prior to flame atomic absorption spectrometric determination.<sup>1, 2, 5-7</sup>  
33 Various separation and enrichment methods such as including solid phase extraction (SPE),<sup>1, 8</sup>  
34 cloud point extraction,<sup>9, 10</sup> liquid-liquid extraction,<sup>11</sup> coprecipitation, etc.<sup>12</sup> have been  
35 developed by the scientists. However these methods have some disadvantages like high  
36 consumption of toxic organic solvents and generate secondary toxic waste, difficult and  
37 tedious operation and time consuming.<sup>13-17</sup> In order to eliminate these disadvantages, small-  
38 scale preconcentration-separation methods called microextraction including dispersive liquid-  
39 liquid microextraction,<sup>13, 14</sup> hollow fiber liquid phase microextraction,<sup>18</sup> solid phase  
40 microextraction<sup>17</sup> and ultrasound-assisted emulsification of solidified floating organic drop  
41 microextraction<sup>19</sup> have been developed for preconcentration and separation of gold from  
42 various media.

43 A new mode of microextraction technique named supramolecular solvent based liquid-  
44 liquid microextraction (SsLLME) has been developed for preconcentration and separation of  
45 organic and inorganic species.<sup>20-24</sup> Supramolecular solvents (Ss) are obtained from amphiphile  
46 solutions by two well-defined self-assembly global processes. The processes occur on two  
47 scales, nano and molecular by the external effects like pH, temperature and electrolyte  
48 concentration of sample, type and volume of solvent. First, amphiphiles aggregate to obtain  
49 supramolecular assemblies, like reverse micelles or vesicles or aqueous, in a homogenous

50 solution and then, coacervate in a second step and give water-immiscible phase that separate  
51 from the bulk solution.<sup>20-24</sup> In microextraction studies, the interactions between analytes in  
52 water phase and extraction phase are important to increase extraction efficiency.<sup>21, 25</sup> Hence,  
53 the selection of suitable extraction medium is important step. The supramolecular solvents  
54 have a lot of interaction area such as hydrophobic, dispersion forces and hydrogen bonding  
55 for organic analytes and metal-ligand complex.<sup>20-24</sup> The interaction areas in supramolecular  
56 assemblies provide high extraction capability and short extraction time. The preparation of  
57 supramolecular solvents at room temperature using conventional, cheap and harmless  
58 chemicals is the most important advantage.<sup>20-24</sup> The supramolecular solvent used was formed  
59 reverse micelles of 1-Decanol in the nano- and microscale regimes dispersed in a continuous  
60 phase of tetrahydrofuran (THF): water. The basis of supramolecular solvent based  
61 microextraction method is hydrophobic and  $\pi$ -cation interactions and the formation of  
62 hydrogen bonds between analyte and supramolecular solvent phase. The hydrophobic  
63 character and water immiscibility of certain supramolecular solvents allow their use in solvent  
64 extraction of hydrophobic compounds.

65 In the present study, the supramolecular solvent based liquid-liquid microextraction  
66 method (SsLLME) for the preconcentration and separation of gold in environmental samples  
67 was developed prior to its microsample injection-flame atomic absorption spectrometric  
68 determination.

69

70

## 71 **2. Experimental**

### 72 **2.1. Chemicals and solutions**

73 All solutions were prepared with reverse osmosis purified water (18.2 M $\Omega$  cm,  
74 Millipore). Except if otherwise stated all chemicals used were analytical reagent grade and

75 were used without further purification. Extraction solvents (1-Decanol, Undecanol) and THF  
76 were purchased from Merck (Darmstadt, Germany). Decanoic acid was purchased from  
77 Sigma-Aldrich (St. Louis, MO, USA). Concentrated (36 % (v/v)) HCl (Merck, Darmstadt,  
78 Germany) and 65% HNO<sub>3</sub> (E. Merck, Darmstadt, Germany) were used.

79 The model solutions of gold(III) was established using standard solutions prepared by  
80 dilution from 1000 mg L<sup>-1</sup> gold(III) stock solution (E. Merck, Darmstadt, Germany). Gold  
81 Ore CDN-GS-3D certified reference material (CDN Resource Laboratories Ltd. Canada) was  
82 also used. A solution of 0.1% (m/v) of 1,3,4-thiadiazole-2,5-dithiol (Aldrich, USA) was daily  
83 prepared in ethanol. In order to adjust of pH of sample solutions, Buffer solutions were  
84 prepared by using combination of salts and solutions as follows: phosphate buffer solution  
85 (pH 2.0–4.0, sodium dihydrogen phosphate/phosphoric acid), acetate buffer solution (pH 5.0-  
86 6.0 ammonium acetate/acetic acid), phosphate buffer solution (pH 7.0-7.5 sodium dihydrogen  
87 phosphate/disodium hydrogen phosphate).

88

## 89 **2.2. Instrumental**

90 A Perkin-Elmer Model 3110 flame atomic absorption spectrometer (Norwalk, CT,  
91 USA) including a gold hollow cathode lamp (operated conditions as follows: wavelength  
92 242.8 nm, spectral band width: 0.7 nm and lamp current: 15.0 mA). Air–acetylene flame was  
93 used for absorbance measurements. The continuous aspiration mode was used for  
94 determination of extracted gold concentration in last volume. The samples were introduced  
95 into the nebulizer of the FAAS by using a home made micro-injection unit including Teflon  
96 funnel which connected to the nebulizer with capillary tubing.<sup>25, 26</sup> 100 µl of the samples was  
97 injected to the micro-injection unit by using Eppendorf pipette and peak heights were  
98 measured as signals. In order to produce supramolecular solvent, an ultrasonic water bath  
99 (Norwalk, CT, USA) was used. A Sartorius PT-10 model pH meter with glass-electrode was

100 used for pH adjustments of sample solutions (Sartorius Co., Goettingen, Germany). The  
101 centrifugation was performed using an ALC PK 120 model centrifuge (Buckinghamshire,  
102 England).

103

### 104 **2.3. Supramolecular solvent based liquid–liquid microextraction procedure**

105 A graphical diagram of the SLSDE-ILDLLME protocol was shown in Fig. 1. A 10  
106 mL of sample solution containing 0.5  $\mu\text{g}$  Au(III), 0.05 mg 1,3,4-thiadiazole-2,5-dithiol and 2  
107 mL of pH 6 buffer solution were placed into a 50-mL polypropylene centrifuge tube. Then,  
108 the extraction solution, which prepared by mixing 50  $\mu\text{L}$  1-decanol and 200  $\mu\text{L}$  THF, was  
109 injected in to the sample solution and the tube was capped. The mixture was kept in an  
110 ultrasonic bath for 3 min and the supramolecular solvent, made up 1-Decanol dispersed in  
111 THF: water spontaneously formed. The obtained mixture was centrifuged at 4000 rpm for 3  
112 min to accelerate the complete separation of the water and extraction phase. The  
113 supramolecular solvent phase situated at the top of the water phase because of its lower  
114 density than water, was taken by using micropipette and its volume completed to 250  $\mu\text{l}$  with  
115 ethanol.

116

117 **(Fig 1. here)**

118

### 119 **2.4. Analysis of real samples**

120 The supramolecular solvent based liquid–liquid microextraction procedure was  
121 applied to gold ore samples and rock samples obtained from different cities of Turkey (Ordu,  
122 Samsun, Nigde and Erzincan) Turkey. The ore and rock samples were homogenized with an  
123 agate homogenizer and dried at 80  $^{\circ}\text{C}$  for 24 hour.

124 Ore (0.1 g) and rock samples and 0.15 g of certified reference material CDN-GS-3D  
125 Gold Ore was weighed, placed into a 100-mL beaker, and digested with 30 mL of aqua regia  
126 at 95 °C until semidried masses were obtained. The resulting mixture again was mixed with  
127 30 mL of aqua regia and evaporated almost to dryness. 5 mL of purified water was added to  
128 the residue. The suspension was filtered through a blue band filter paper (Macherey-Nagel,  
129 Düren, Germany), and the insoluble part was washed with purified water. Then, the SsLLME  
130 procedure given in section 2.3 was applied to the samples.

131 The suggested method was applied to tap from Canakkale and well water from Sivas,  
132 Turkey. The water samples were filtered through a Millipore® cellulose membrane filter  
133 (0.45 µm pore size) and the suggested SsLLME procedure (Section 2.3) was applied to the 7.0  
134 mL of water samples.

135

136

### 137 **3. Results and discussion**

138 In order to obtain quantitative recoveries of gold(III), the microextraction procedure  
139 was optimized for various analytical parameters. The conditions for preconcentration of gold  
140 ions were selected using model solutions.

141

#### 142 **3.1. Influence of pH**

143 pH of the working media is most critical parameter on the supramolecular solvent  
144 based liquid phase microextraction studies to obtain the supramolecular solvent and  
145 subsequent extraction of metal–ligand complex.<sup>27</sup> The effect of pH of the sample solution on  
146 the extraction efficiency of gold(III) were investigated in the pH range 2.0–7.5. The results  
147 given in Fig. 2 reveal that the best recovery can be achieved when the pH of sample solution  
148 was adjusted at 6.0. It is because of the neutral character of Au(III)-1,3,4-thiadiazole-2,5-

149 dithiol complex in this interval and the type of interactions governing its solubilisation in the  
150 supramolecular solvent phase. Therefore, pH 6.0 as working pH was chosen for the  
151 subsequent experiments.

152

153 **(Fig 2. here)**

154

### 155 **3.2. Type of supramolecular solvent**

156 The high extraction capability of the supramolecular solvent was consequence of both  
157 the power of analyte–extractant interactions and the special structure of the aggregates  
158 making it up.<sup>21, 28</sup> To obtain the best supramolecular solvent phase, three supramolecular  
159 solvent including decanoic acid-THF, 1-decanol–THF and undecanol- THF were used in this  
160 study. The recovery % values of gold (III) by using 1-decanol-THF, undecanol-THF and  
161 decanoic acid-THF were 100±2, 53±0 and 63±6, respectively. 1-decanol-THF supramolecular  
162 solvent was used as extraction phase for the further study.

163

### 164 **3.3. Influence of the volume ratio of 1-decanol and THF**

165 The effect of the 1-decanol/THF volume ratio on the recovery of gold was also  
166 studied. To study the effect of varying the volume of supramolecular solvent on the efficiency  
167 of the extraction of gold from sample solution, the volume of 1-decanol was changed between  
168 50 and 300  $\mu\text{L}$  and the THF was kept constant at 600  $\mu\text{L}$ . Fifty  $\mu\text{L}$  volume of 1–decanol was  
169 adequate for quantitative recovery of gold. The effect of the volume of THF on the extraction  
170 recovery was also studied. Different volumes of THF in the range of 100–600  $\mu\text{L}$  with  
171 addition of 50  $\mu\text{L}$  of 1-Decanol were examined. The results illustrated in Fig. 3 reveal that  
172 after 200  $\mu\text{L}$ , the recoveries stayed quantitative and consequently, 200  $\mu\text{L}$  of THF was  
173 selected as the optimum volume.

174

175 **(Fig 3. here)**

176

177 **3.4. Effect of complexing agent amount**

178 The extraction efficiency depends on the hydrophobicity of the complexing agent  
179 which provides solubility of metal complex in supramolecular solvent phase.<sup>21</sup> For this  
180 purpose, 1,3,4-thiadiazole-2,5-dithiol was used as complexing agent because of the highly  
181 hydrophobic nature of its gold complex. Amount of complexing agent has a critical effect on  
182 the quantitative recoveries of the analytes in liquid phase microextraction.<sup>29</sup> Thus, it is highly  
183 important to establish the minimal complexing agent amount that leads to total complex  
184 formation while getting the highest recovery. The effect of amount of 3,4-thiadiazole-2,5-  
185 dithiol was also investigated in the range of 0.0-0.5 mg. The results are depicted in Fig. 4. A  
186 amount of 0.05 mg of 3,4-thiadiazole-2,5-dithiol was selected for the further experiments.

187

188 **(Fig 4. here)**

189

190 **3.5. Effect of ultrasonication and centrifuge time**

191 It is known that ultrasonic radiation is an important way to increase the kinetics and  
192 extraction efficiency. The ultrasonic radiation increases the interactions between analyte in  
193 sample solution and extraction phase for mass transfer.<sup>30, 31</sup> In this study, an ultrasonic water  
194 bath was used for formation of supramolecular solvent made up of reverse micelles of 1-  
195 decanol dispersed in THF: water and extraction of Au(III)-1,3,4-thiadiazole-2,5-dithiol  
196 complex. The effect of ultrasonication time on extraction efficiency was examined in the  
197 range of 2–4 minute. Three minute of ultrasonication time was enough to obtain quantitative

198 recovery. To spend minimum time, 3 minute of ultrasonication time was used for further  
199 work.

200 The influence of centrifugation time on the extraction efficiency of Au(III) in the  
201 developed procedure were examined at 4000 rpm between 1 and 9 min. The quantitative  
202 recovery was obtained with 3 minute of centrifuge time.

203

### 204 **3.6. Study of interferences**

205 The effects of matrix ions on extraction efficiency and on the selectivity of the  
206 developed method were also investigated by using the optimum conditions. A 10 mL model  
207 solution, which including 0.5  $\mu\text{g}$  of Au(III) and different amounts of other ions, was prepared  
208 and the suggested extraction method was applied to the solution. The obtained results are  
209 shown in Table 1. The tolerance limits of matrix ions were taken as that value which caused a  
210 deviation of more than  $\pm 5\%$  in the recovery. Under the optimum conditions, no interference  
211 was observed from most of the ions tested except for  $\text{Fe}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Ni}^{2+}$  and  $\text{Mn}^{2+}$  ions. In order  
212 to eliminate the interference effects of the ions, EDTA,  $\text{NH}_4\text{F}$ , KSCN, ascorbic acid and citric  
213 acid was checked. In the presence of KSCN, ascorbic acid and citric acid, the extraction of  
214 gold(III) was not possible. But, the interference effect of the ions can be eliminated by using  
215 25 mg of EDTA and 75 mg of  $\text{NH}_4\text{F}$  as a masking agent.<sup>3</sup>

216

217 **(Table 1. here)**

218

### 219 **3.7. Effect of the sample volume**

220 The effect of sample volume on the extraction of Au(III) was investigated by using  
221 10–40 mL of model solution keeping other conditions constant. It was found that the recovery

222 of gold up to 15 mL of the sample solution were quantitative. Hence, a preconcentration  
223 factor of 60 was obtained using a final volume of 0.25 mL.

224

### 225 **3.8. Analytical figures of merit**

226 The analytical features of the developed method were evaluated under the optimum  
227 conditions. The whole preconcentration procedure could take about 10 min. It provides to  
228 simultaneously treat as many samples as can be placed in the centrifuge. Calibration curve  
229 constructed was  $A = 4 \times 10^{-3} + 0.834C$  with a correlation coefficient ( $R^2 = 0.996$ ). Where A is  
230 the absorbance and C is the gold concentration in  $\mu\text{g L}^{-1}$ . The limits of detection (LOD), which  
231 is defined as  $C_{\text{LOD}} = 3S_d/m$ , (where  $C_{\text{LOD}}$ ,  $S_d$  and  $m$  are the limit of detection, standard  
232 deviation of the eleven blank, and slope of a calibration graph, respectively) was  $1.5 \mu\text{g L}^{-1}$ .  
233 The limit of quantification (LOQ) was found as  $4.95 \mu\text{g L}^{-1}$  which calculated from the ratio of  
234 ten times the standard deviation of the eleven blank solutions to the slope of the calibration  
235 curve.

236 The preconcentration factor (PF), defined as the ratio of model solution to last volume  
237 was 60. The enhancement factor (EF), defined the slope ratio of calibration curve after and  
238 before preconcentration, was 51. The relative standard deviation (RSD) obtained from the  
239 microextraction procedure of 10 replicates of 10 mL solution containing  $50 \mu\text{g L}^{-1}$  Au(III)  
240 was 4.2 %.

241 The consumptive index is the volume of samples necessary to attain a unit of  
242 preconcentration factor. The consumptive index (CI) can be found for practical purposes as:  
243  $CI = V_s / EF$  where  $V_s$  is the volume of sample solution (mL) consumed to achieve the EF  
244 value.<sup>32</sup> The CI was found 0.29.

245

246

### 247 **3.9. Applications of the method**

248 To prove the accuracy and applicable of the method, the developed microextraction  
249 method was applied to the determination of gold in CDN-GS-3D Gold Ore certified reference  
250 material. The obtained results of reference material in Table 2 agreed with the certified  
251 values.

252

253 **(Table 2. here)**

254

255 The accuracy of the suggested method was also verified by spiking various amounts of  
256 gold(III) in water and ore samples (Table 3). A good agreement was obtained between the  
257 added and recovered gold amounts. The all results obtained show that the suggested  
258 microextraction method was confidential and matrix independence for determination of gold  
259 in wide range of samples. The present SsLLME procedure was applied to the determination of  
260 gold in ore and rock samples obtained from Nigde and Erzincan (Table 4).

261

262 **(Table 3. here)**

263 **(Table 4. here)**

264

### 265 **3.10. Comparison with other preconcentration techniques**

266 The SsLLME- MS-FAAS method was compared with the other preconcentration  
267 methods used for the determination of gold in real samples in Table 5. The developed  
268 SsLLME- MS-FAAS method is environmental friendly with low usage of toxic organic  
269 solvents. The comparative data of different analytical characteristics showed that the obtained  
270 low detection limit and high enhancement factor of the proposed microextraction

271 methodology are much better than other reported preconcentration methods for gold  
272 determinations in real samples.

273

274 **(Table 5. here)**

275

276

#### 277 **4. Conclusion**

278 This study describes a novel and simple supramolecular solvent based liquid-liquid  
279 microextraction for preconcentration and separation of trace amounts of gold in ore samples  
280 prior to its determination by microsample introduction-flame atomic absorption spectrometry  
281 (FAAS). The important properties of the developed method: (I) it offers a simple and rapid  
282 alternative to conventional sample preconcentration methods, (II) decrease the consumption  
283 of toxic organic solvents and amount of secondary toxic waste, (III) It is free from matrix  
284 interference which are associated with gold in its natural occurrence. The suggested method  
285 was successfully applied for determination of low concentrations of gold in water, rock and  
286 ore samples with good accuracy and precision.

287

288

#### 289 **Acknowledgements**

290 The authors are grateful for the financial support of the Unit of the Scientific Research  
291 Projects of Erciyes University (FBD-2014-4991) (Kayseri, Turkey).

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296 **References**

- 297 1. M. Tuzen, K. O. Saygi and M. Soylak, *J. Hazard. Mater.*, 2008, 156, 591–595.
- 298 2. H. B. Senturk, A. Gundogdu, V. N. Bulut, C. Duran, M. Soylak, L. Elci and M. Tufekci, *J.*  
299 *Hazard. Mater.*, 2007, 149, 317–323.
- 300 3. G. S. Kamble, S. S. Kolekar, S. H. Han and M. A. Anuse, *Talanta*, 2010, 81, 1088-1095.
- 301 4. M. H. Sorouraddin, M. Saadati and A. Aghaei, *Monatsh. Chem.*, 2011, 142, 439–445.
- 302 5. D. Afzali, A. Mostafavi and M. Mirzaei, *J. Hazard. Mater.*, 2010, 181, 957–961.
- 303 6. M. Karimi, V. Amani, F. Aboufazeli, H. R. L. Z. Zhad, O. Sadeghi and E. Najafi, *Journal*  
304 *of Chemistry*, Article ID 142845 (2013) <http://dx.doi.org/10.1155/2013/142845>
- 305 7. M. Soylak and E. Yilmaz, *Atom. Spectrosc.*, 2013, 34, 15-19.
- 306 8. E. Mladenova, I. Karadjova and D. L. Tsalev, *J. Sep. Sci.*, 2012, 35, 1249-1265.
- 307 9. S. Tong, Q. Jia, N. Song, W. Zhou, T. Duan and C. Bao, *Microchim. Acta*, 2011, 172, 95-  
308 102.
- 309 10. W.S. El-Naggar, T.A. Lasheen, E.S.A. Nouh and A.K. Ghonaim, *Cent. Eur. J. Chem.*,  
310 2010, 8, 34-40.
- 311 11. J. Wu, H. Fu, B. J. Li and X.S. Zhu, *Spectrosc. Spect. Anal.*, 2011, 31, 260-262.
- 312 12. M. Soylak and M. Tuzen, *J. Hazard. Mater.*, 2008, 152, 656-661.
- 313 13. A. Iraj, D. Afzali and A. Mostafavi, *Int. J. Environ. An. Ch.*, 2013, 93, 315-324.
- 314 14. M. Shamsipura and M. Ramezani, *Talanta*, 2008, 75, 294–300.
- 315 15. H. Ashkenani and M. A. Taher, *Microchem. J.* 2012, 103, 185–190.
- 316 16. I. D. L. Calle, F. P. Pereira, N. Cabaleiro, I. Lavilla and C. Bendicho, *Talanta*, 2011, 84,  
317 109–115.
- 318 17. J. Hassan, M. Shamsipur and M. H. Karbasi, *Microchem. J.*, 2011, 99, 93–96.
- 319 18. C. J. Zeng, L. F. Tang, *Anal. Lett.*, 2013, 46, 1442-1453.
- 320 19. S. Tajik and M. A. Taher, *Microchim. Acta*, 2011, 173, 249-257.

- 321 20. S. G. Fonseca, A. B. Gómez, S. Rubio and D. P. Bendito, *J. Chromatogr.*, 2010, 1217A,  
322 2376-2382.
- 323 21. E. Yilmaz and M. Soylak, *Talanta*, 2014, 126, 191-195.
- 324 22. Q. Yang, W. Su, X. Y. Jiang and X. Q. Chen, *Int. J. Environ. An. Ch.*, 2014, 94, 812-821.
- 325 23. F. Rezaei, Y. Yamini, M. Moradi and B. Daraei, *Anal. Chim. Acta*, 2013, 804, 135-142.
- 326 24. A. Moral, M. D. Sicilia and S. Rubio, *Anal. Chim. Acta*, 2009, 650, 207–213.
- 327 25. E. Yilmaz and M. Soylak, *Talanta*, 2013, 116, 882–886.
- 328 26. M. Soylak and E. Yilmaz, *Desalination*, 2011, 275, 297–301.
- 329 27. Z. H. Li, J. X. Chen, M. S. Liu and Y. L. Yang, *Anal. Methods*, 2014, 6, 2294-2298.
- 330 28. M. Moradi, Y. Yamini, M. Tayyebi and H. Asiabi, *Anal. Bioanal. Chem.*, 2013, 405,  
331 4235-4243.
- 332 29. F. Shah, E. Yilmaz, T. G. Kazi, H. I. Afridi and M. Soylak, *Anal. Methods*, 2012, 4 4091-  
333 4095.
- 334 30. A. Bordagaray, R. G. Arrona and E. Millan, *Food Anal. Method.*, 2014, 7, 1195-1203.
- 335 31. P. Reboredo-Rodriguez, L. Rey-Salgueiro, J. Regueiro, C. Gonzalez-Barreiro and B.  
336 Cancho-Grande, *Food Chem.*, 2014, 150, 128-136.
- 337 32. P. X. Baliza, L. S. G. Teixeira and V. A. Lemos, *Microchem. J.*, 2009, 93, 220–224.
- 338 33. S. Tong, Q. Jia, N. Song, W. Zhou, T. Duan and C. Bao, *Microchim. Acta*, 2010, 172, 95-  
339 102.
- 340 34. S. Tajik and M. A. Taher, *Microchim. Acta*, 2011, 173, 249–257.
- 341 35. V. N. Bulut, C. Duran, Z. Biyiklioglu, M. Tufekci and M. Soylak, *Geostandard. Geoanal.*  
342 *Res.*, 2011, 35, 471-483.

343

344

345

346

347 **Figure captions**

348 Figure 1. Graphical representation of supramolecular solvent based liquid-liquid  
349 microextraction method (SsLLME)

350 Fig. 2. Effect of the pH on the extraction efficiency of Au(III) (N=3, 1-Decanol volume= 100  
351  $\mu\text{L}$ , THF volume=600  $\mu\text{L}$ , Amount of complexing agent= 0.1 mg).

352 Fig. 3. Effect of the volume of THF on the extraction efficiency of Au(III) (N=3, pH= 6.0, 1-  
353 Decanol volume=50  $\mu\text{L}$ , Amount of complexing agent= 0.1 mg).

354 Fig. 4. Effect of the amount of 1,3,4-thiadiazole-2,5-dithiol on the extraction efficiency of  
355 Au(III) (N=3, pH= 6.0, 1-Decanol volume= 50  $\mu\text{L}$ , THF volume=200  $\mu\text{L}$ ).

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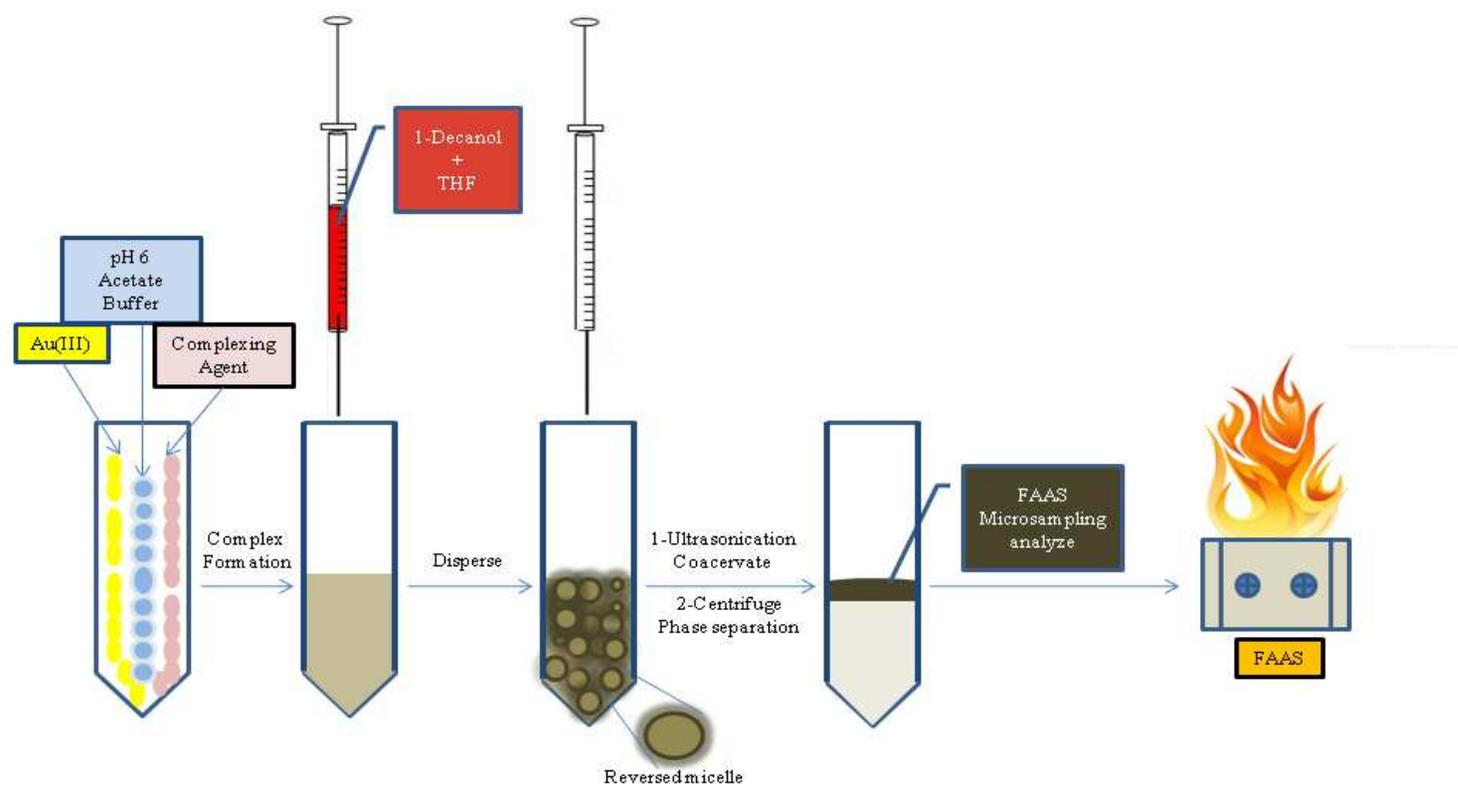


Fig. 1.

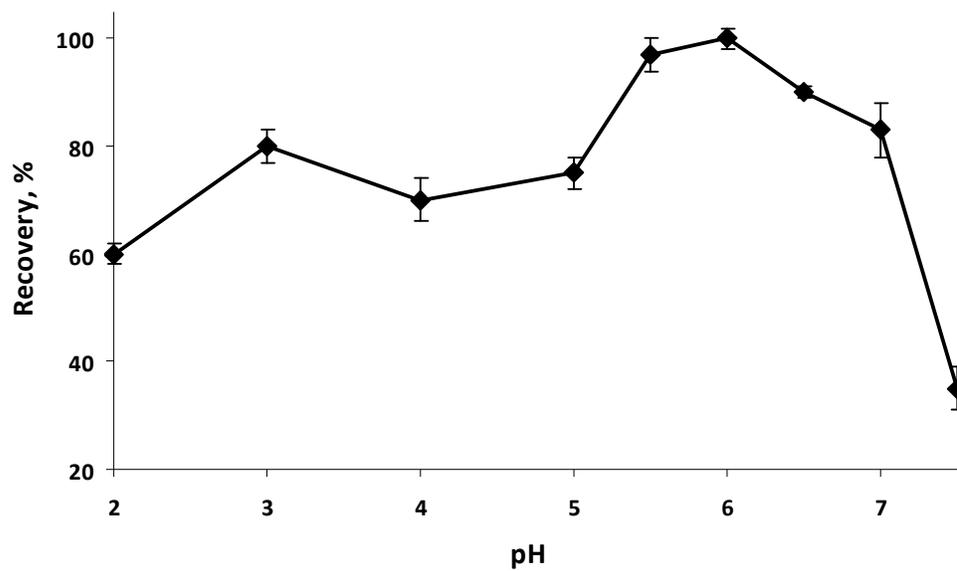


Fig. 2.

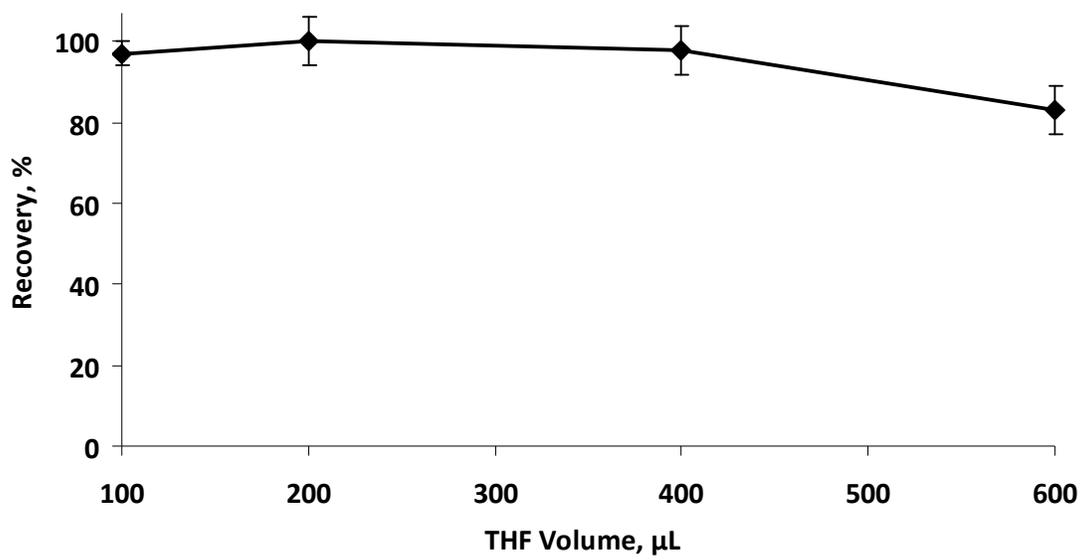


Fig. 3.

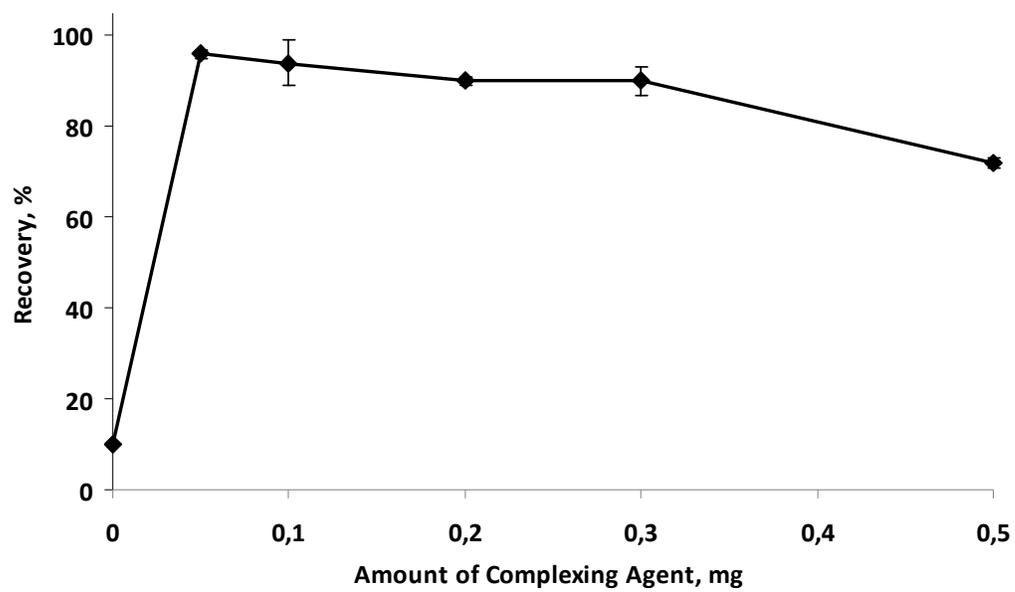


Fig. 4.

**Table 1.** Effect of some matrix ions on the extraction efficiency of Au(III) (N=3).

Ion	Added as	Concentration, mg L <sup>-1</sup>	Recovery, %
Na <sup>+</sup>	NaNO <sub>3</sub>	50000	97±5
K <sup>+</sup>	KCl	50000	99±5
Mg <sup>2+</sup>	Mg(NO <sub>3</sub> ) <sub>2</sub> .6H <sub>2</sub> O	500	103±4
Ca <sup>2+</sup>	Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O	500	95±5
Zn <sup>2+</sup>	Zn(NO <sub>3</sub> ) <sub>2</sub> .6H <sub>2</sub> O	5	100±3
Cr <sup>3+</sup>	Cr(NO <sub>3</sub> ) <sub>3</sub> .9H <sub>2</sub> O	10	96±3
Pd <sup>2+</sup>	Pd(NO <sub>3</sub> ) <sub>2</sub>	2.5	97±1
Co <sup>2+</sup>	Co(NO <sub>3</sub> ) <sub>2</sub> .6H <sub>2</sub> O	2.5	99±4
Cl <sup>-</sup>	NaCl	77000	97±5
Fe <sup>3+</sup>	Fe(NO <sub>3</sub> ) <sub>3</sub> .9H <sub>2</sub> O	100 <sup>a</sup>	95±0
Cu <sup>2+</sup>	Cu(NO <sub>3</sub> ) <sub>2</sub> .3H <sub>2</sub> O	50 <sup>b</sup>	99±2
Mn <sup>2+</sup>	Mn(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O	50 <sup>b</sup>	98±0
Ni <sup>2+</sup>	Ni(NO <sub>3</sub> ) <sub>2</sub> .6H <sub>2</sub> O	10 <sup>b</sup>	100±1

<sup>a</sup> Masked with 75 mg NH<sub>4</sub>F.

<sup>b</sup> Masked with 25 mg EDTA.

**Table 2.** The analysis results of CDN-GS-3D Gold ore certified reference material (N: 5).

Certified Value ( $\mu\text{g g}^{-1}$ )	Found ( $\mu\text{g g}^{-1}$ )	Recovery, %
$3.41 \pm 0.25$	$3.38 \pm 0.2$	99

<sup>a</sup> Mean  $\pm$  standard deviation.

**Table 3.** Addition and recovery test for microextraction of Gold(III) in water and ore samples (n=5).

	Added, $\mu\text{g}$	Found, $\mu\text{g}$	Recovery, %
Tap water from Canakkale	0.0	<sup>a</sup> <LOQ	-
	2.0	2.08 $\pm$ 0.00 <sup>b</sup>	104
	4.0	4.04 $\pm$ 0.18	100
Well water from Sivas	0.0	<LOQ	-
	0.75	0.77 $\pm$ 0.05	102
	1.5	1.47 $\pm$ 0.08	98
Ore sample from Ordu	0.0	<LOQ	-
	0.75	0.78 $\pm$ 0.07	103
	1.5	1.48 $\pm$ 0.07	99
Ore sample from Samsun	0.0	<LOQ	-
	1.0	1.04 $\pm$ 0.03	104
	2.0	1.90 $\pm$ 0.03	95
Ore sample from Ordu-2	0.0	<LOQ	-
	0.75	0.72 $\pm$ 0.02	96
	1.5	1.51 $\pm$ 0.10	100

<sup>a</sup>LOQ: Limit of quantification.

<sup>b</sup>mean  $\pm$  standard deviation.

**Table 4.** The application of presented method for gold level of ore and rock samples (N= 5).

Sample	Concentration, $\mu\text{g g}^{-1}$
Ore Sample from Nigde-1	$1.73 \pm 0.16$
Ore Sample from Nigde-2	$3.35 \pm 0.25$
Rock Sample from Erzincan-1	$0.35 \pm 0.06$
Rock Sample from Erzincan-2	<sup>a</sup> <LOQ
Rock Sample from Erzincan-3	<LOQ

<sup>a</sup> LOQ: limit of quantification.

<sup>b</sup> Mean  $\pm$  standard deviation.

**Table 5.** Comparison of the IL-DMME with other methods for the determination of gold

Method	LOD ( $\mu\text{g L}^{-1}$ )	PF	Samples	Ref.
SPE-FAAS <sup>a</sup>	1.61	31	Water, soil, sediment	[1]
SPE-FAAS <sup>a</sup>	16.6	200	Water, soil, ore	[2]
CPE-FAAS <sup>b</sup>	3.8	16	ore	[33]
USAE-SFODME-FAAS <sup>c</sup>	0.45	34.8	Pharmaceutical, water	[34]
LLE- spectrophotometric <sup>d</sup>	0.5	200	Water, Ore	[35]
ILME-FAAS <sup>e</sup>	3.4	40	Water, soil	[7]
SsLLME-FAAS <sup>f</sup>	1.5	60	Water, Ore	This work

<sup>a</sup>Solid phase extraction-Flame atomic absorption spectrometry

<sup>b</sup>Cloud point extraction- Flame atomic absorption spectrometry

<sup>c</sup>Ultrasound-assisted emulsification of solidified floating organic drop microextraction- Flame atomic absorption spectrometry

<sup>d</sup>Liquid liquid extraction- UV-VIS spectrometry

<sup>e</sup>Ionic liquid microextraction-Flame atomic absorption spectrometry

<sup>f</sup>Supramolecular solvent based liquid-liquid microextraction-Flame atomic absorption spectrometry