RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This Accepted Manuscript will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

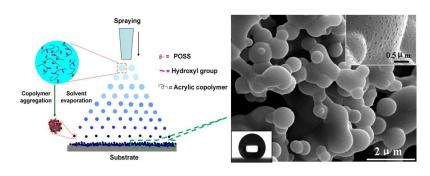
Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/advances

RSC Advances

A simple one step procedure to fabricate superhydrophobic polymer surface was successfully developed by spraying POSS-acrylic copolymer solution. The morphology of the superhydrophobic surface was made up of micron- and nano-scale spherical protrusions or particles. The hierarchical structure of the superhydrophobic surface was constructed by taking advantage of self-assembly of the POSS-acrylic copolymer in the spraying process.



Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxx

COMMUNICATIONS

One-step fabrication of superhydrophobic polymer surface from an acrylic copolymer containing POSS by spraying[†]

Hui Li,*^a Xiaoyun Zhao,^a Guohong Chu,^a Shuxiang Zhang,*^a and Xiaoyan Yuan^b

Received (in XXX, XXX) Xth XXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX 5 DOI: 10.1039/b000000x

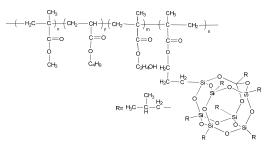
A superhydrophobic polymer surface with hierarchical structures is prepared by one-step spraying process with a POSS-acrylic random copolymer solution. The morphology of the superhydrophobic surface consists of micron/nano-scale spherical protrusions. The possible formation mechanism of the superhydrophobic surface is proposed. The as-prepared surface is of good superhydrophobicity and blood compatibility.

In the past decade, superhydrophobic surfaces have received ¹⁵ considerable attention in the field of materials and surface sciences, caused by their outstanding performance, such as in oilwater separation, and anti-icing applications, as well as in blood compatibility.¹⁻⁴ Serving as the best engineer and designer, nature forms unartificial superhydrophobic surfaces in many animals ²⁰ and plants such as water strider's legs and lotus leaves.⁵⁻⁶ Inspired by nature, a myriad of approaches, such as template synthesis, electrospinning, sol-gel method, casting and plasma etching, have been developed to fabricate superhydrophobic surfaces with water contact angle higher than 150° and low contact angle

- ²⁵ hysteresis.⁷⁻⁹ In general, to achieve superhydrophobic surfaces, a combination of low surface free energy materials and surface roughness is required. Thus far, a wide variety of materials including inorganic, organic and composite materials have been utilised to prepare superhydrophobic surfaces. Among them,
- ³⁰ polymeric materials have been considered to be appropriate substrates because of their unique and versatile properties.¹⁰ For example, Yuan et al. prepared superhydrophobic poly(vinyl chloride) (PVC) surfaces by the template and casting method,⁶ and Levkin et al. obtained a superhydrophobic porous poly(octyl
- 35 cyanoacrylate) coating by immersing a substrate coated with
- ^a School of Chemistry and Chemical Engineering, and Shandong Key Laboratory of Fluorine Chemistry and Chemical Engineering Materials, University of Jinan, Jinan 250022, China. Fax & Tel: 86 531 82767096;
- ⁴⁰ E-mail:chm_lih1@ujn.edu.cn, fhx_zhangsx@163.com. ^b School of Materials Science and Engineering, and Tianjin Key Laboratory of Composite and Functional Materials, Tianjin University, Tianjin 300072, China.
- [†]Electronic Supplementary Information (ESI) available: experimental ⁴⁵ section, characterization methods, FT-IR and ¹HNMR spectra and GPC curve of PAC, SEM image and the water contact angle of the surface of HPAC, Photographs of water droplets on the spray-coated different substrates, SEM image of the platelets adhesion on PVC surface. Number of the platelets adhesion on the different surfaces, See DOI:
- 50 10.1039/b00000x/

cyanoacrylate in an aqueous solution.¹¹ However, most of the techniques for the fabrication of superhydrophobic surfaces either involve multi-step processes and sometimes harsh conditions or are unsuitable for large-sized flat surfaces, which limit their 55 practical applications. А larger-scale fabrication of using superhydrophobic surfaces on substrates by polydivinylbenzene chalk was reported, but the surfaces of these substrates required a certain microstructure with rough mophology.12

Spray coating is a simple and well established technique in the modern surface-coating industry, that generates superhydrophobic coatings by spraying a low concentration suspension on flat substrates.¹³⁻¹⁶ Ogihara et al. reported, for example, a superhydrophobic coating by spraying an ethanol suspension of 65 SiO₂ nanoparticles on a paper substrate.¹⁷ However, based on this method, polymeric materials have rarely been used in the preparation of superhydrophobic coating by themselves. In a previous report, our group successfully fabricated superhydrophobic surfaces through spraying a micelle solution of ⁷⁰ a fluorinated acrylic copolymer.⁴ In this communication, we present a one-step spraying-coating process for the fabrication of a superhydrophobic surface with a Polyhedral oligomeric silsesquioxane(POSS) -acrylic random copolymer solution. Compared with the above-mentioned approaches, the copolymer 75 solution used herein is relatively low-priced, can be easily prepared, is translucent, and is applicable to multiple substrates. Of note, the superhydrophobic surface exhibits regular morphology and consists of micron- and nano-scale spherical protrusions or particles, leading to the formation of hierarchical 80 micro-nano structures. The as-prepared surface shows excellent superhydrophobicity and good blood compatibility. The possible formation mechanism of the superhydrophobic polymer surface is also proposed.



Scheme 1 The structure of the POSS-acrylic copolymer (PAC)

85

At first, a POSS-acrylic copolymer (PAC) was synthesized by free radical solution polymerization. The structure of PAC is illustrated in Scheme 1. Experiments are described in detail in the ESI[†] and the fourier transform infrared (FT-IR) and ¹H nuclear ⁵ magnetic resonance (NMR) spectra of PAC are shown in Fig.

sinagicite resonance (rivirk) spectra of FAC are shown in Fig. S1⁺ and Fig. S2⁺, respectively. As confirmed by gel permeation chromatography (GPC), the copolymer had a number-average molecular weight of 35,000 with a polydispersity index of 1.91. Moreover, only one peak appeared at the elution volume for GPC
¹⁰ (Fig. S3⁺), indicating copolymer formation.

Then the resultant PAC was dissolved in ethyl acetate to form a translucent solution containing 10 wt% of the copolymer. The diluted copolymer solution was deposited onto a copper net and dried at room temperature. The morphology of PAC self-15 assembly aggregates was characterised by transmission electron

- microscope (TEM) (Fig. 1). the copolymer was prone to aggregation, forming many nano-scale spherical particles with diameters of 20~40 nm after solvent evaporation. The self-assembly of the copolymer and surface mobility of the POSS ²⁰ moieties in the copolymer possibly contributed to sphere
- ²⁰ moleties in the copolymer possibly contributed to spher formation.¹⁸⁻²⁰

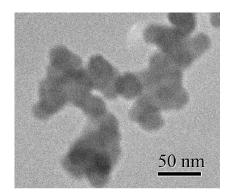


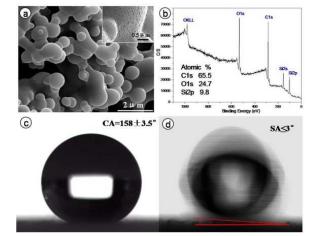
Fig. 1 TEM micrograph of the PAC self-assembly aggregates

After spraying the above-mentioned solution on a glass ²⁵ substrate and subsequent drying process, the copolymer formed a superhydrophobic POSS-acrylic copolymer (SPAC) surface. Meanwhile, the hydrophobic POSS-acrylic copolymer (HPAC) surface was also fabricated by spin coating for comparison (the preparation method of the HPAC surface is seen in the ESI†). ³⁰ The morphologies of the HPAC and SPAC surfaces were

- examined by scanning electron microscopy (SEM). The HPAC surface was smooth (Fig. S4†), and the static water contact angle (CA) on the HPAC surface was 106±2.3° (Fig. S4†, inset). However, the SPAC surface consisted of many connected miner and static antenna similar to the maximum scale miner and static stati
- ³⁵ micron-scale spherical protrusions, similar to the previous reported morphology,²¹ forming a rough structure (Fig. 2a). The high-magnification SEM image (Fig. 2a, inset) clearly showed numerous nano-scale outshoots on the surface of each micronscale spherical protrusion. The XPS spectrum of SPAC is shown
- ⁴⁰ in Fig. 2b, and the carbon, oxygen and silicone atomic percentages were estimated to be approximately 65.5%, 24.7% and 9.8%, respectively. There is a higher ratio of silicone to carbon in the XPS data (9.8%:65.5% \approx 0.149:1) than the theoretical value of the PAC molecule (\approx 0.036:1), suggesting ⁴⁵ that the migration of the POSS groups containing silicone to the
- coating surface, which is essential to obtain superhydrophobic

surfaces with low surface energy. As shown in the insets of Figs. 2c and 2d, the water CA on the SPAC surface could reach 158 ± 3.5° and the sliding angle (SA) was less than 3°, exhibiting ⁵⁰ outstanding superhydro- phobicity. Moreover, these large-scale superhydrophobic surfaces were easily formed on other substrates including paper, PVC film, steel mesh and aluminum plate (Fig. S5a-d†). Although the coated film exhibited a decrease in transmittance compared with the PVC film (Fig. S5e†), caused by ⁵⁵ the light scattering and diffuse reflection of the surface roughness,¹² the samples were visible after spray-coating the PAC solution (Fig. S5a and S5b†).

The possible formation mechanism for the superhydrophobic polymer surface prepared by spraying can be described as follows ⁶⁰ (Fig.3a). During spray coating, the polymer solution is atomised to large numbers of tiny liquid droplets with different particle sizes by the pressure at the spray gun nozzle. Fig. 3b shows the cumulative particle size distribution curve of liquid droplets (the



65 Fig. 2 SEM image (a) and XPS survey spectrum (b) and optical images of the water CA (c) and SA (d) of SPAC surface

test method is seen in the ESI⁺). The results indicate that approximately 50% of liquid droplets have sizes at or below 19.78µm, including approximately 10% of liquid droplets less 70 than 9.38µm, and 40% of those have sizes between 19.78 and 34.26µm. Before these tiny liquid droplets impact the substrate, the droplets gradually become smaller owing to the rapid evaporation of the solvent in air. Simultaneously, this process is accompanied by the self-assembly aggregation of the POSS-75 acrylic copolymer. The POSS functional groups can easily transfer to the air-copolymer solution interface (outside of the droplets) due to their self-aggregation behaviour and surface mobility,^{19,20} while the hydrophilic hydroxyl groups attached to the copolymers can migrate to the inside of droplets. This leads to ⁸⁰ the formation of submicron-sized polymeric particles. As a result, these particles stack and adhere together irregularly on the substrate when these particles hit the substrate surface, finally bringing about the formation of different-sized spherical protrusions of 500nm to 3µm (Fig. 2a). The surfaces of these 85 micron-scale spherical protrusions appear a significant amount of nano-scale outshoots because of the self-aggregates behaviour of POSS functional groups (Fig. 2a, inset), ultimately leading to the formation of hierarchical micro-nano structures. As indicated above, the possible morphologies of the relatively rough surface may be affected by a number of factors, such as the particle size distribution of liquid droplets, concentration of the polymer solution, evaporation rate of solvent, and self-assembly aggregates behaviour.

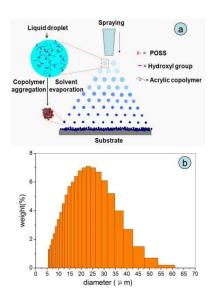


Fig. 3 Schematic representation of forming SPAC surface(a) and the cumulative particle size distribution curve of "liquid droplets" prepared by spraying(b), D_{10} =9.38µm, D_{50} =19.78µm, D_{90} =34.26µm.

In theory, superhydrophobic surfaces have been speculated to ¹⁰ possess good hemocompatibility.²² A widely accepted postulate in biomaterials science is that protein adsorption (especially, fibrinogen) is the first step in the blood-material interaction, followed by platelet adhesion and activation, leading to thrombus formation. ²³ Fig. 4 shows the amounts of bovine serum albumin

- ¹⁵ (BSA) and bovine fibrinogen (BFG) adsorbed on the HPAC and SPAC surfaces from their respective PBS solutions (1mg/ml). The SPAC surface had lower amounts of adsorbed BSA and BFG as compared with the HPAC surface. This result indicates that the SPAC surface could resist protein adsorption to some extent in ²⁰ accordance with the superhydrophobic surfaces reported by
- Shirtcliffe and Advincula.^{24, 25} Furthermore, the amount of BSA adsorbed on the SPAC surface decreased by approximately 27% as compared with that adsorbed on the HPAC surface; however, more importantly, the amount of adsorbed BFG decreased by 25 approximately 64%, which is deemed to be one of the major
- factors affecting platelet adhesion.^{26, 27}

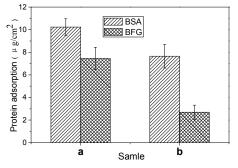


Fig. 4 BSA and BFG adsorbed amounts on the surfaces of HPAC (a) and SPAC (b)

It is well known that platelet adhesion and activation are considered as key events in blood coagulation.²⁷ In order to further evaluate the blood compatibility of the superhydrophobic surface, the platelet adhesion experiments on the HPAC and SPAC surfaces were also carried out. For comparison, similar 35 experiments were conducted on PVC surfaces. As shown in Fig. S6[†], numerous adhered platelets were spread out on the PVC surface, and pseudopodia were clearly observed, implying that they have been extremely activated. However, for HPAC surface, the number of adhered platelets dramatically decreased (~79%) as 40 compared with that on the PVC surface (Fig. S7⁺), and their morphology was maintained with almost no pseudopodia (Fig. 5a). More exhilaratingly, the adhering platelets nearly disappeared (Fig. S7[†]), and few pseudopodia as well as deformation were observed on the SPAC surface (Fig. 5b). This 45 should be attributed to the superhydrophobicity and the relative

low protein adsorption on the SPAC surface.^{22,24,28}

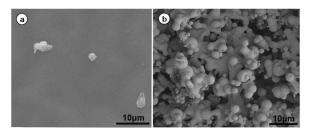


Fig. 5 SEM images of the platelets adhesion on the surfaces of HPAC (a) and SPAC (b)

In conclusion, we successfully developed a simple one-step procedure to fabricate superhydrophobic polymer surfaces through spraying a POSS-acrylic copolymer solution. The hierarchical structure of the superhydrophobic surface was constructed by taking advantage of the self-assembly of the 55 POSS-acrylic copolymer in the spraying process. The as-prepared surface by the spraying method presents excellent superhydrophobicity, outstanding anti-adhesion to platelet and protein adsorption resistance. Although the formed superhydrophobic polymer surface is not completely transparent, 60 this study describes a facile method for fabricating large-scale superhydrophobic surfaces with potential applications in blood compatibility, anti-icing, antifouling, and water repellency.

This work is financially supported by the National Natural ⁶⁵ Science Foundation of China (51103061).

Notes and references

- 1 C. R. Crick, J. A. Gibbins and I. P. Parki, *J. Mater. Chem. A*, 2013, 1, 5943-5948.
- 2 Z. Shi, I. Wyman, G. Liu, H. Hu, H. Zou and J. Hu, *Polymer*, 2013, 54, 6406-6414.
- 3 B. J. Privett, J. Youn, S. A. Hong, J. Lee, J. Han, J. H. Shin and M. H. Schoenfisch, *Langmuir*, 2011, 27 (15), 9597-9601.
- 4 H. Li, Y. Zhao and X. Yuan, Soft Matter, 2013, 9, 1005-1009.
- 5 X. Gao and L. Jiang, *Nature*, 2004, 432, 36
- 75 6 Z. Yuan, H. Chen and J. Zhang, Appl. Surf. Sci., 2008, 254, 1593-1598.
 - 7 K. Acatay, E. Simsek, C. O. Yang and Y. Z. Menceloglu, Angew. Chem. Int. Ed., 2004, 43, 5210-5213.
- 8 M. Hikita, K. Tanaka, T. Nakamura, T. Kajiyama and A. Takahara, 80 *Langmuir*, 2005, 21, 7299-7301.

- 9 X. Du and J. H. He, ACS Appl. Mater. Interfaces, 2011, 3, 1269-1277.
- 10 N. J. Shirtcliffe, G. McHale and M. I. Newton, *Polym. Sci. Part B: Polym. Phys.*, 2011, 49, 1203-1217.
- 11 X. Du, J. S. Li, L. X. Li and P. A. Levkin, J. Mater. Chem. A, 2013, 1, 1026-1029.
- 12 Y. L. Zhang, J. N. Wang, Y. He, Y. He, B. B. Xu, S. Wei and F. S. Xiao, *Langmuir*, 2011, 27, 12585-12590.
- 13 P. A. Levkin, F. Svec and J. M. J. Fréchet, Adv. Funct. Mater., 2009, 19, 1993-1998.
- 10 14 W. Wu, X. Wang, X. Liu and F. Zhou. ACS Appl. Mater. Interfaces, 2009, 1, 1656-1661.
 - 15 M. Stepien, J. J. Saarinen, H. Teisala, M. Tuominen, M. Aromaa, J. Kuusipalo, J. M. Mäkelä and M. Toivakka, *Appl. Surf. Sci.* 2011, 257, 1911-1917.
- 15 16 H. S. Hwang, N. H. Kim, S. G. Lee, D. Y. Lee, K. Cho and I. Park, ACS Appl. Mater. Interfaces, 2011, 3, 2179-2183.
 - 17 H. Ogihara, J. Xie, J. Okagaki and T. Saji, *Langmuir*, 2012, 28, 4605-4608
- 18 N. C. Escudé, Eugene and Y. X. Chen, *Chem. Mater*, 2009, 21, 5743-5753.
- 19 R. Y. Kannan, H. J. Salacinski, P. E. Butler and A. M. Seifalian, Accounts of Chemical Research, 2005, 38: 879-884.
- 20 S. T. Iacono, S. M. Budy, J. M. Mabry and D. W. Smith, *Macromolecules*, 2007, 40, 9517-9522.
- 25 21 X. Chen, P. Fei, K. A. Cavicchi, W. Yang and N. Ayres, *Colloid. Polym. Sci.*, 2014, 292, 477-484.
 - 22 T. L. Sun, H. Tan, D. Han, Q. Fu and L. Jiang, *Small*, 2005, 1, 959-963.
- 23 Z. Zhang, M. Zhang, S. Chen, T. A. Horbett, B. D. Ratner and S. Jiang, *Biomaterials*, 2008, 29, 4285-4291
- 24 R. B. Pernites, C. M. Santos, M. Maldonado, R. R. Ponnapati, D. F. Rodrigues and R. C. Advincula, *Chem. Mater.*, 2012, 24, 870-880.
- 25 Y. Koc, A. J. Mello, G. McHale, M. I. Newton, P. Roacha and N. J. Shirtcliffe, *Lab Chip*, 2008, 8, 582–586.
- 35 26 A. Wörz, B. Berchtold, K. Moosmann, O. Prucker and J. Rühe, J. Mater. Chem., 2012, 22, 19547-19561.
 - 27 L.R. Wang, H. Qin, S. Q. Nie, S. D. Sun, F. Ran and C. S. Zhao, Acta Biomater, 2013, 9, 8851-8863.
- 28 C. Mao, C. Liang, W. Luo, J. Bao, J. Shen, X. Hou and W. Zhao, J.
 40 Mater. Chem., 2009, 19, 9025-9029.