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### Graphical Abstract



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# ARTICLE TYPE

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# Fluorescent and antioxidant studies of effectively synthesized isochromenopyrrolone analogues

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An efficient strategy for the synthesis of 3-acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1H)ones, **4a-f** have been developed using Montmorillonite K10 as catalyst. The reaction proceeded from
acetyl acetone, substituted primary amines and ninhydrin *via* acidic media under solvent free condition.

<sup>10</sup> Compounds, **4a-c** exhibits green fluorescence under UV light and its fluorescent property in liquid state were investigated. All the synthesized compounds were subjected for the evaluation of free radical scavenging activity. Among all the tested compounds, **4a & 4c** exhibited good percentage inhibition in comparison with the standard ascorbic acid.

### 15 1 Introduction

Aromatic heterocyclic compounds are known to show a wide range of ecotoxic effects, e.g. acute toxicity, cytotoxicity,<sup>1</sup> photoinduced toxicity,<sup>2</sup> mutagenicity<sup>3</sup> and carcinogenicity.<sup>4</sup> New efficient methods for the synthesis of heterocycles are a major <sup>20</sup> interest of modern synthetic organic chemistry.<sup>5</sup> Among the

- <sup>20</sup> Interest of modern synthetic organic chemistry. Anong the heterocycles isocoumarin represents a foremost class of naturally occurring lactones that display a wide range of biological activities such as protease inhibitor properties, antifungal, cytotoxic, anti-allergic, antimicrobial and antimalarial effects.<sup>6-10</sup>
- <sup>25</sup> Isocoumarins are class of natural products with a broad spectrum of biological activity<sup>11, 12</sup> including antioxidative,<sup>13</sup> antiangiogenic,<sup>14</sup> antifungal,<sup>15, 16</sup> antiallergic and antimicrobial<sup>8-10</sup> activities. They are also important in medicinal chemistry as building blocks for the bioactive compounds.<sup>17, 18</sup> In addition to
- <sup>30</sup> its natural occurrence,<sup>19, 20</sup> various synthetic methodologies for construction of isocoumarins were disclosed.<sup>11</sup> Isocoumarins are useful synthetic precursors to other heterocyclic and carbocyclic compounds.<sup>21, 22</sup>

Free radicals oxidatively damage lipids, proteins and genomic <sup>35</sup> DNA integrity. They are widely recognized as the root cause of numerous degenerative diseases including cancer.<sup>23</sup> Antioxidants are potent scavengers of free radicals and serve as inhibitors of neoplastic processes.<sup>24</sup> Vegetables, fruits and their seeds are rich sources of vitamins C, E,  $\beta$ -carotene and also compounds that

- <sup>40</sup> might protect the organism against free radical induced injury and diseases.<sup>25, 26</sup> There has been an increasing interest in the use of natural antioxidants, such as tocopherols, flavonoids for the preservation of food materials.<sup>27</sup> These natural antioxidants avoid the toxicity problems which may arise from the use of
- <sup>45</sup> synthetic antioxidants, such as butylated hydroxy anisole (BHA), butylated hydroxy toluene (BHT) and propyl gallate (PG). Due to the complexity of antioxidant activity in foods, the evaluation of synthetic heterocyclic compounds recently widely focused.<sup>28</sup>

In this paper synthesis of isochromenopyrrolone analogues <sup>50</sup> using MK-10 under solvent free approach for organic synthesis is described which involves microwave exposure of neat reactants. The synthesis of 3-acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-ones, **4a-f** were carried out using MK-10 as catalyst was found to be more efficient with high yield compared to the <sup>55</sup> conventional synthesis method<sup>29</sup>.

### 2 Results and discussion

### 2.1 Chemistry

In this work, we have reported the synthesis of novel series of 3acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-<sup>60</sup> ones, **4a-f** using MK-10 as catalyst. The antioxidant activity and fluorescence properties of these 3-acetyl-2-methyl-1phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-ones, **4a-f** have also been reported.

The formation of **4a-f** was confirmed by FTIR, <sup>1</sup>H NMR, <sup>13</sup>C NMR and mass spectral analysis. FT-IR data of compound, **4a** shows absorption peak at 1462 cm<sup>-1</sup> indicates benzene C=Cstretching, peak at 1668 cm<sup>-1</sup> indicates alkene C=C stretching, peak at 1732 cm<sup>-1</sup> indicates C=O stretching, peaks at 3199-3057 cm<sup>-1</sup> indicates sp<sup>3</sup> C-H stretching. <sup>1</sup>H-NMR data of compound, 70 **4a** confirms the existence of one methyl of pyrrole ring, one methyl of acetyl group and two methyl of substituted amine groups at  $\delta$  1.04, 1.04, 2.32, 2.79 ppm respectively, two methylene protons of the ethyl groups at  $\delta$  2.26 and the seven aromatic protons exists between  $\delta$  6.2 to 8.36 ppm. <sup>13</sup>C-NMR 75 data of compound, **4a** confirms the existence of six aliphatic carbons at  $\delta$  12.26-31.30 ppm, sixteen aromatic carbons between  $\delta$  110.4-142.1 ppm and two carbonyl groups at  $\delta$  161.99 (-C=O of Isocoumarin ring) and 193.66 (-C=O of acetyl group).

### **80 2.2 Fluorescence studies**

For the fluorescence studies, we analyzed that the compound, **4c** exhibits a strong emission at 464 nm, the compound, **4b** shows emission at 462 nm which is on par to that of **4c** and the compound, **4a** shows weak emission at 456 nm.

### 2.3 Hydrogen peroxide scavenging assay

In the present study, Hydrogen peroxide scavenging assay was performed for isochromeno pyrrolones, **4a-f**. The absorbance of the standard ascorbic acid and the compounds, 4a-f was measured

- <sup>10</sup> at 230 nm and the results were tabulated in **Table-1**. The percentage inhibition of the target molecules, **4a-f** have been plotted against the various concentrations of the standard ascorbic acid and the test solutions (**Fig. 1**). All readings were triplicate to check the accuracy.
- 15 2.4 DPPH assay

In the present study, DPPH assay was performed for the Isochromeno pyrrolones, **4a-f**. The absorbance of the standard ascorbic acid, compounds, **4a-f** were measured at 520 nm and the results were tabulated in **Table-2**. The percentage inhibition of

20 the target molecules, have been plotted against the various concentrations of the standard ascorbic acid and the test solutions **4a-f (Fig. 2)**. All readings were triplicate to check the accuracy.

### **3** Experimental

45

- <sup>25</sup> Solvents and reagents were commercially sourced and used without further purification. Thin layered chromatography (TLC) was performed on preparative plates of silica gel. Visualization was made with UV chamber. Column chromatography was performed by using silica gel (60-120 mesh). Melting points
- <sup>30</sup> were measured on Elchem Microprocessor based DT apparatus using an open capillary tube and are corrected with standard benzoic acid. IR spectrum was obtained on a Nucon infrared spectrophotometer (KBR disc). UV-vis spectrum was obtained on UV-2550, Shimadzu Corporation, Kyoto, Japan. The NMR
- spectrum was recorded on a Bruker Avance III- 400 & 500 MHz spectrometer with Tetramethylsilane (TMS) as internal standard. Chemical shifts were reported in  $\delta$  units (ppm). GC-MS spectrum was recorded on a Perkin Elmer spectrophotometer, GC model: clarus 680, Mass Spectrometer model: clarus 600 (EI).
- <sup>40</sup> The fluorescence spectra were obtained on Hitachi F-7000 FL Spectrophotometer.

# 3.1 *General Procedure for the Synthesis of* 3-acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-one, *4f*.

### Method A: (E)-2-hydroxy-2-(2-oxo-4-(phenylimino)pentan-3yl)-1H-indene-1,3(2H)-dione, 3a

A mixture of acetyl acetone, **1** (1.1 mmol) and aniline, **2f** (1.1 mmol) were heated at 85 °C for 15-30 min. Then the reaction <sup>50</sup> mixture was poured into a solution of ninhydrin (1.1 mmol) in acetic acid (10 mL), stirred at room temperature until the disappearance of the reactants. The completion of the reaction was monitored by TLC. After the completion, the reaction mixture was cooled to room temperature and then quenched this

<sup>55</sup> mixture into crushed ice. The solid formed was allowed to stand for 30 min and filtered off to dryness. Compound **3f** was recrystallized by mixture of acetone and hexane (2:8), gave crystals of intermediate compound, **3**.

### 60 3-acetyl-2-methyl-1-phenylisochromeno[4,3-b]pyrrol-5(1H)one, 4f.

About 0.5 mL of Conc.H<sub>2</sub>SO<sub>4</sub> was added to a solution of 200 mg of compound, **3** in 10 mL of acetic acid. The reaction mixture was heated on water bath at 85 °C for about 30-45 min. The <sup>65</sup> initial pale green color of the solution was intensified to brown which indicates the conversion of product, **4a**. The completion of the reaction was monitored by TLC. Then the reaction mixture was cooled to room temperature and poured into crush ice with continuous stirring. Precipitated solid was allowed to stand for 30 min and filtered off to dryness. Then the solid obtained was purified by column chromatography using petroleum ether: ethyl acetate (8:2) as eluent to give the reddish brown crystals of isochromeno pyrrolones, **4a**.

### 75 Method-B: 3-acetyl-2-methyl-1-phenylisochromeno[4,3b]pyrrol-5(1H)-one, 4f.

Appropriated acetyl acetone, 1 (1.1 mmol), aniline, 2f (1.1 mmol), ninhydrin (1.1 mmol) followed by adding of MK-10 (10 mg) were introduced in an Erlen-meyer flask. This was subjected to microwave irradiation for sufficient intervals of time using resting intervalsof 1 min after every 30 s of irradiation. The reaction was monitored by TLC. After cooling, the reaction mixture became sticky and the sticky solid was then triturated to afford the product. The product was recrystallized by mixture of sacetone and hexane (2:8), gave crystals of compound, 4f. Optimization of 3-acetyl-2-methyl-1-phenylisochromeno[4,3-b]pyrrol-5(1*H*)-one, 4a-f has been done by comparing the yield of conventional and microwave irradiation methods. The results of the comparison were tabulated in Table – 3.

### 3-acetyl-1-(2,6-diethylphenyl)-2-methylisochromeno[4,3-b]

pyrrol-5(1*H*)-one, (4a). Pale brown solid, Mol. formula: C<sub>24</sub>H<sub>23</sub>NO<sub>3</sub>, Exact Mass: 373.1678, Mol.Wt.: 373.4443, yield 82 %, M.pt. 209-211 °C. FT-IR (KBr pellets, cm-1): 3388.93, 3199.91, 3057.17, 2966.52, 1732.08, 1668.43, 1462.04. 1H NMR (500MHz, CDCl3): δ= 8.35-8.36 (1H, dd, J<sub>1</sub> = 1.5 Hz J<sub>2</sub> = 1 Hz, -CH), 7.55-7.57 (1H, t, -CH), 7.34 (2H, m, -CH), 7.30 (1H, d, -CH), 7.28 (1H, d, -CH), 6.27-6.28 (1H, d, J = 5 Hz -CH), 2.79 (3H, s, -CH3), 2.32 (3H, s, -CH3), 2.23 (2×2H, m, 2-CH2) of Et, 100 1.01-1.05 (2×3H, t, 2-CH3) of Et. 13C NMR (125MHz, CDCl3): δ= 193.6, 161.9, 142.1, 141.0, 138.8, 134.9, 133.8, 131.7, 130.6, 130.5, 2×127.1, 2×125.6, 117.8, 117.7, 111.5, 110.4, 31.3, 2×23.7, 2×13.7, 12.2. Mass: m/z 373.21.

### 105 3-acetyl-1-(2,3-dimethylphenyl)-2-methylisochromeno[4,3-b]

**pyrrol-5(1***H***)-one, (4b).** Pale brown solid, Mol. formula:  $C_{22}H_{19}NO_3$ , Exact Mass: 345.1365, Mol.Wt.: 345.3912, yield 84%, M.pt. 202-204 °C. FT-IR (KBr pellets, cm<sup>-1</sup>) : 3201.83, 1761.65, 1664.57. <sup>1</sup>H NMR (500MHz, CDCl<sub>3</sub>):  $\delta$ = 1.85 (3H, s, -110 CH<sub>3</sub>), 2.36 (3H, s, -CH<sub>3</sub>), 2.43 (3H, s, -CH<sub>3</sub>), 2.77 (3H, s, -CH<sub>3</sub>), 6.28-6.30 (1H, d, *J* = 10 Hz, -CH), 7.14-7.16 (1H, d, *J* = 10 Hz, -CH), 7.33 (3H, m, -CH), 7.44-7.46 (1H, d, *J* = 10 Hz, -CH), 8.36-8.35 (1H, d, *J* = 5 Hz -CH). <sup>13</sup>C NMR (100MHz, CDCl<sub>3</sub>):  $\delta$ = 12.3, 13.8, 20.5, 31.4, 110.25, 117.8, 118.0, 125.7,126.1, 115 2×127.2, 130.8, 131.8, 131.9, 2×135.0, 135.5, 136.0, 139.2,

115 2×127.2, 130.8, 131.8, 131.9, 2×135.0, 135.5, 136.0, 139.2, 139.5, 162.1,193.9. GC-MS (*EI*): 345.26.

3-acetyl-1-benzyl-2-methylisochromeno[4,3-b]pyrrol-5(1H)-

- one (4c). Pale brown solid, Mol. formula:  $C_{21}H_{17}NO_3$ . Exact Mass: 331.1208, Mol.Wt.: 331.3646, yield 95%, M.pt.182 °C<sup>29</sup> (Lit. 182). FT-IR (KBr pellets, cm<sup>-1</sup>): 3194.12, 3126.61, 2968.45, 2877.79, 1722.43, 1666.50, 1612.49. <sup>1</sup>H NMR (500MHz, CDCl<sub>3</sub>):  $\delta$ = 2.67 (3H, s, -CH<sub>3</sub>), 2.74 (3H, s, -CH<sub>3</sub>), 5.57 (2H, s, -CH<sub>2</sub>), 7.07 (2H, d, *J* = 10 Hz, -CH), 7.34 (4H, m, -CH), 7.44-7.46 (1H, d, *J*=10 Hz, -CH), 7.54 (1H, m, -CH), 8.39 (1H, m, -
- <sup>10</sup> CH). <sup>13</sup>C NMR (125MHz, CDCl<sub>3</sub>):  $\delta$ = 11.4, 31.6, 48.6, 110.4, 111.6, 117.9, 118.8, 125.5, 126.7, 2×128.2, 129.3, 129.5, 130.4, 132.3, 135.1, 135.2, 139.1, 141.1, 161.9, 193.8. Mass: m/z 331.17.
- <sup>15</sup> 3-acetyl-2-methyl-1-m-tolylisochromeno[4,3-b]pyrrol-5(1*H*)-one (4d). Pale brown solid, Mol. formula: C<sub>21</sub>H<sub>17</sub>NO<sub>3</sub>, Exact mass: 331.1208, Mol.Wt.: 331.3646, yield 88 %, M.pt. 185-187
   <sup>o</sup>C. FT-IR (KBr pellets, cm<sup>-1</sup>): 2918.30, 2848.86, 1772.58, 1699.29, 1633.71. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>): δ= 2.48 (3H, s, -
- <sup>20</sup> CH<sub>3),</sub> 2.56 (3H, s, -CH<sub>3</sub>), 2.82 (3H, s, -CH<sub>3</sub>), 6.50-6.48 (1H, d, J = 8 Hz, -CH), 7.25 (1H, s, -CH), 7.35 (1H, s, -CH), 7.43-7.41 (1H, d, J = 8 Hz, -CH), 7.54 (1H, s, -CH), 7.59-7.58 (2H, d, J = 4Hz, -CH), 8.43-8.41 (1H, d, J = 8 Hz, 1H). <sup>13</sup>C NMR (100MHz, CDCl<sub>3</sub>):  $\delta = 12.6$ , 13.7, 20.5, 96.11, 112.5, 117.4, 117.5, 124.5,
- <sup>25</sup> 126.5, 126.8, 131.2, 131.6, 131.8, 133.3, 134.5, 136.0, 137.4, 139.0, 142.5, 163.2, 190.4. Mass: m/z 331.18.

### 3-acetyl-2-methyl-1-p-tolylisochromeno[4,3-b]pyrrol-5(1H)-

- one (4e). Pale brown solid, Mol. formula:  $C_{21}H_{17}NO_3$ , Exact <sup>30</sup> Mass: 331.1208, Mol.Wt.: 331.3646, yield 89 %, M.pt. 192-194 °C. FT-IR (KBr pellets, cm<sup>-1</sup>): 3199.91, 2966.52, 1786.08, 1670.35, 1606.70. <sup>1</sup>H NMR (500MHz, CDCl<sub>3</sub>):  $\delta$ = 2.40 (3H, s, CH<sub>3</sub>), 2.55 (3H, s, -CH<sub>3</sub>), 2.74 (3H, s, -CH<sub>3</sub>), 6.47-6.45 (1H, dd,  $J_I$  = 4.4 Hz,  $J_2$  = 1.4 Hz,-CH), 7.29 (3H, m, -CH), 7.38 (1H, m, -
- <sup>35</sup> CH), 7.44-7.42 (2H, d, J = 8 Hz, -CH), 8.35-8.33 (1H, d, J = 8 Hz, -CH). <sup>13</sup>C NMR (100MHz, CDCl<sub>3</sub>):  $\delta = 12.6$ , 13.7, 20.5, 96.11, 112.5, 117.4, 117.5, 124.5, 126.5, 126.8, 131.2, 131.6, 131.8, 133.3, 134.5, 136.0, 137.4, 139.0, 142.5, 163.2, 190.4. Mass: m/z 331.18.
- 40

### 3-acetyl-2-methyl-1-phenylisochromeno[4,3-b]pyrrol-5(1H)-

- one (4f). Pale brown solid, Mol. formula:  $C_{20}H_{15}NO_3$ , Exact Mass: 317.1052, Mol.Wt.: 317.3380, yield 85 %, M.pt. 245-247  $^{\circ}C^{29}$  (Lit. 248). FT-IR (KBr pellets, cm<sup>-1</sup>): 3057.17, 2918.30,
- <sup>45</sup> 1712.79, 1612.49, 1494.33. <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>): δ= 2.38 (3H, s, -CH<sub>3)</sub>, 2.73 (3H, s, -CH<sub>3</sub>), 6.36 (1H, s, -CH), 7.26 (1H, s, -CH), 7.31-7.29 (2H, t, d, J= 8 Hz, -CH), 7.44-7.42 (3H, d, J = 8 Hz, -CH), 7.64 (1H, s, -CH), 8.35-8.33 (1H, d, J = 8 Hz, -CH). <sup>13</sup>C NMR (100MHz, CDCl<sub>3</sub>): δ= 11.4, 31.6, 118.2, 124.0, 124.4,
- <sup>50</sup> 125.0, 125.9, 126.9, 127.3, 130.4, 130.7, 131.9, 132.0, 134.9, 135.7, 136.8, 139.0, 139.3, 190.4, 201.3. Mass: m/z 317.26.

### 3.2 Antioxidant assay

### 55 Preparation of stock solution

A solution of hydrogen peroxide (40 mM) was prepared in phosphate buffer saline (PBS) at pH 7.4. A stock solution (1 mg/ml) was prepared for the synthesized compounds, 4a-f in

methanol and subsequently diluted as 10, 20, 30, 50 & 100  $\mu L.$ 

### Hydrogen peroxide scavenging anti-oxidant activity

Antioxidant activity was measured by reported methodology.<sup>30</sup> For the present study the samples were prepared by taking 1 mg of the compounds, 4a-f dissolved in 1 mL of AR grade methanol. <sup>65</sup> The samples 10, 20, 30, 50 & 100 μL were taken in different test tubes to which 20 μL of hydrogen peroxide was added and made upto 2 mL using phosphate buffer and incubated at room temperature for 10 min. The absorbance of the incubated solutions and the blank solution (phosphate buffer without <sup>70</sup> hydrogen peroxide) were recorded against ascorbic acid. The absorbance was measured at 230 nm using a UV-vis Spectrophotometer. Radical Scavenging capacity (RSC) in percent was calculated by the following equation:

% of RSC (%) =  $[(A_b-A_s)/A_b] \times 100$ 

75 Where,

RSC = Radical Scavenging Capacity

As = Absorbance of sample

 $A_b = Absorbance of blank$ 

### 80 DPPH radical scavenging assay

Radical scavenging activity of compounds against stable DPPH (2,2-diphenyl-2-picrylhydrayl hydrate) was determined spectrophotometrically. When DPPH reacts with an antioxidant compound, which can donate hydrogen, DPPH reduced. The ss changes in colour (from deep-violet to light yellow) were measured at 520 nm on a UV-vis spectrophotometer. A solution of DPPH was mixed with the synthesized compounds; 4a-f gives rise to the reduced form with the loss of violet color.

### $\mathbf{DPPH} \cdot + \mathbf{RH} = \mathbf{DPPH} + \mathbf{R} \cdot$

Where RH is the reduced form and  $R^{\cdot}$  is free radical

The DPPH radical scavenging activities of all the synthesized compounds, 4a-f and ascorbic acid were determined according to the reported method.<sup>31</sup> Initially, 1 mg/mL of the test material solutions and the standard (ascorbic acid), at a concentration of <sup>95</sup> 10, 50 and 100 µg/ml, respectively, were added to 1 mL of 0.2 mM freshly prepared DPPH ethanol solution. The reaction mixture were shaken vigorously and allowed to stand for 30 min at room temperature, under dark conditions. The absorbance of each reaction mixture was measured at 520 nm. The percentage <sup>100</sup> of DPPH radical scavenging activity (%) of the sample was calculated as follows:

### % Inhibition) = $[(A_b-A_s)/A_b] \times 100$

Where  $A_b$ = absorption of blank sample  $A_s$ = absorption of sample

### 3.3 Fluorescence studies

### UV-vis Spectra

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The UV absorption spectrum of compound, 4a in various <sup>110</sup> solvents such as DMF, ethanol and DMSO solution were recorded in the range of 200–800 nm. The excitation peaks appears at 314 nm and 365 nm respectively (**Fig. 5a**).

### Fluorescence spectra

<sup>115</sup> The fluorescence emission spectra of the synthesized compounds, **4a-c** were recorded in DMSO with excitation

wavelength at 365 nm. The fluorescence emission spectrum of the compounds, **4a-c** were depicted in **Fig. 5b**.

Table 1Percentage inhibition of H2O2 assay of 3-acetyl-2-5methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1H)-ones, 4a-f

Conc.	%Inhibition							
μL	4a	4b	4c	4d	<b>4e</b>	4f		
10	41.63	40.62	40.81	11.07	22.61	10.54		
20	50.85	45.68	48.14	20.84	31.74	18.19		
30	58.38	53.64	54.76	32.42	48.89	32.16		
50	84.46	83.82	74.05	52.68	72.14	52.05		
100	88.70	87.64	83.64	76.21	82.38	74.89		
IC <sub>50</sub>	1.84	2.07	2.07	2.46	2.90	2.38		

 
 Table 2 Percentage inhibition of DPPH assay of 3-acetyl-2methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1H)-ones, 4a-f

Conc.	%Inhibition						
μL	4a	4b	4c	4d	4e	4f	4a
10	24.96	21.62	22.13	18.70	17.28	19.15	16.78
50	55.73	51.89	50.20	54.00	48.55	52.26	41.51
100	92.77	88.15	85.73	86.92	67.35	85.09	64.42
IC <sub>50</sub>	1.76	1.88	1.91	1.90	2.22	1.93	2.38

<sup>10</sup> **Table-3** Reaction parameters of 3-acetyl-2-methyl-1phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-ones, **4a-f** 

		Time		Yield (%)			
Entry	Product	СН	MW	CЦ	MW	M.P(°C)	
		(h)	(min)	СП	IVI VV		
1	4a	2.45	1.20	79	82	209	
2	4b	2.40	1.10	78	84	202	
3	4c	2.50	1.40	86	95	180 <sup>29</sup>	
4	4d	2.45	1.20	83	88	187	
5	4e	2.45	1	85	89	194	
6	4f	3.05	1.10	82	85	245 <sup>29</sup>	
<sup>a</sup> Conventional heating- stirring, 85 °C, gla. AcOH, conc. H <sub>2</sub> SO <sub>4;</sub>							

<sup>b</sup>Microwave irradiated, MK-10.

### Conclusions

- <sup>15</sup> In present study, we report an efficient one pot MK-10 catalysed synthesis of fused isochromenopyrrole-5(1*H*)-ones gave an enhanced yield with lesser time than the conventional heating. Out of the synthesized compounds **4c & 4b** exhibits strong fluorescence emission. The synthesized compounds were
- <sup>20</sup> successfully screened for the free radical scavenging activity by  $H_2O_2$  and DPPH assay method. Compounds **4a**, **4b** & **4c** showed a high percentage of inhibition similar to that of standard Ascorbic acid.

### Notes and references

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90

### FIGURE CAPTIONS

- Fig.1: Comparison of percentage inhibition of H<sub>2</sub>O<sub>2</sub> assay of 3-acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-ones, **4a-f**
- **Fig.2:** Comparison of percentage inhibition of DPPH assay of 3-acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1*H*)-ones, **4a-f**
- Fig.3: [a] Comparison of UV–Vis spectrum (DMF, ethanol and DMSO) of 4a. [b] Fluorescence emission spectra of compounds, 4a-c

Scheme 1: Synthesis of 3-acetyl-2-methyl-1-phenylisochromeno [4, 3-b] pyrrol-5(1H)-ones, 4a-f



Fig.1









Fig.2



Fig.3





Scheme 1