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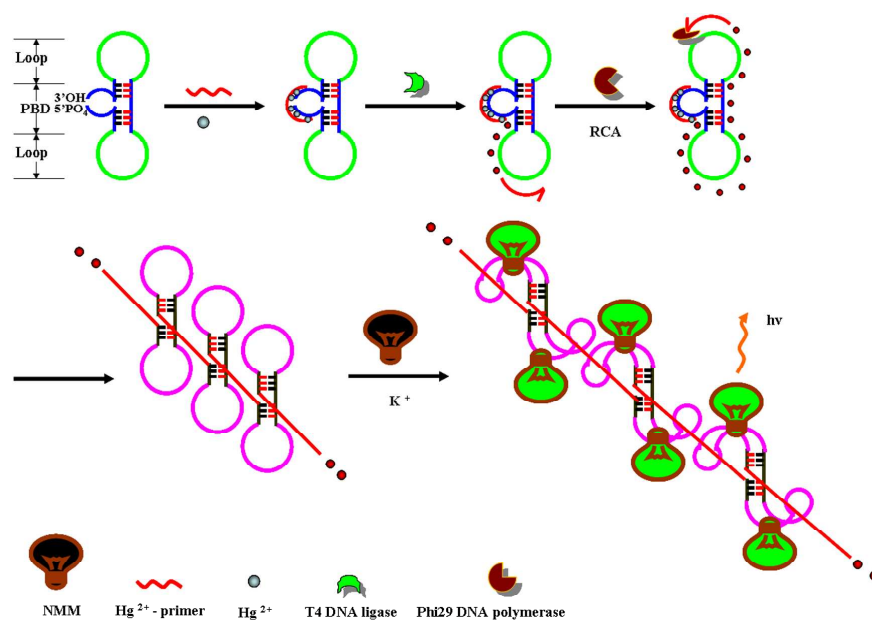
# Target-Responsive dumbbell probe-mediated rolling circle amplification strategy for highly sensitive $\text{Hg}^{2+}$ detection

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**Scheme 1** Schematic representation of the process of the label-free fluorescent sensing system based on target-responsive dumbbell probe-mediated rolling circle amplification (D-RCA) for detection of mercuric ion.



1 **Target-Responsive dumbbell probe-mediated rolling circle**  
2 **amplification strategy for highly sensitive Hg<sup>2+</sup> detection**

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24 **Abstract:**

25 A novel label-free amplified fluorescent sensing scheme based on  
26 target-responsive dumbbell probe-mediated rolling circle amplification (D-RCA) has  
27 been developed for sensitive and selective detection of mercuric ion. In this strategy,  
28 we reported an ingeniously designed dumbbell-shaped DNA probe (D-DNA) that  
29 integrates target-binding, amplification and signaling within one multifunctional  
30 design. An  $\text{Hg}^{2+}$ -primer DNA ( $\text{Hg}^{2+}$ -p-DNA) was designed to complementary to the  
31 region of D-DNA but with T–T mismatches. The mismatched  $\text{Hg}^{2+}$ -primer cannot  
32 initiate the RCA reaction in the absence of  $\text{Hg}^{2+}$ . Stable T– $\text{Hg}^{2+}$ –T can be formed in  
33 the presence of target  $\text{Hg}^{2+}$ , thus induces the elongation and amplification reaction by  
34 a RCA mechanism, resulting in numerous cascade dumbbell probes with duplex  
35 G-rich quadruplex structure oligomer. Upon the addition of N-methyl mesoporphyrin  
36 IX (NMM), the signal reporter, the strong interaction between the G-quadruplex and  
37 NMM brings about a great fluorescence enhancement. In this way, we successfully  
38 converted each  $\text{Hg}^{2+}$ -triggered D-RCA reaction event into the detectable fluorescent  
39 signals, which were significantly amplified by RCA in an isothermal fashion. This  
40 approach can detect 80 fM mercuric ions, much lower than those of previously  
41 reported biosensors, and exhibits high discrimination ability. More significantly, the  
42 dynamic range of D-RCA is extremely large, covering 5 orders of magnitude. We also  
43 demonstrate  $\text{Hg}^{2+}$  quantification with this highly sensitive and selective D-RCA  
44 strategy in real samples.

45 *Keyword:* Hg<sup>2+</sup> detection; G-quadruplex; label-free; dumbbell probe-mediated rolling  
46 circle amplification.

## 47 **1. Introduction**

48 Mercuric ion (Hg<sup>2+</sup>), one of the most toxic heavy metal ions and a severe  
49 environmental pollutant that is not biodegradable, has serious deleterious effects on  
50 human health, especially in the central nervous system, even at low concentration<sup>1,2</sup>.  
51 So far, several modern analytical techniques for mercury detection have been  
52 established, such as atomic absorption/emission spectroscopy<sup>3</sup>, selective cold vapor  
53 atomic fluorescence spectrometry<sup>4, 5</sup> and inductively coupled plasma mass  
54 spectrometry (ICP-MS)<sup>6</sup>. However, most of them require expensive and sophisticated  
55 instrumentation and/or complicated sample preparation processes. Alternatively,  
56 chemical sensors based on small organic fluorophores for Hg<sup>2+</sup> are easy and rapid to  
57 operate.<sup>7-10</sup> But many of these systems are limited in practical use, due to their poor  
58 sensitivity<sup>11, 12</sup>, strong dependence on organic solvents<sup>13, 14</sup> and cross-sensitivities  
59 toward other metal ions<sup>15</sup>.

60 The recent discovery of Hg<sup>2+</sup>-mediated thymine–thymine (T–T) DNA  
61 base-pairing provides an efficient platform for constructing highly selective sensing  
62 systems for highly selective Hg<sup>2+</sup> detection by using T-containing oligonucleotides<sup>15-17</sup>.  
63 Numerous Hg<sup>2+</sup> detection methods including fluorescent<sup>18, 19</sup>, colorimetric, and  
64 electrochemical biosensor have been developed<sup>20-24</sup>. Among them, the development of  
65 homogeneous fluorescence sensors is one of the most attractive and interesting areas  
66 due to its simplicity and rapidness<sup>25-28</sup>. Recently, several fluorescent sensing systems  
67 based on molecular beacon<sup>29</sup>, linear quencher–fluorophore probes,<sup>30, 31</sup> graphene

68 oxide<sup>32</sup>, gold nanorods<sup>33</sup>, conjugates polymers and carbon nanomaterial<sup>34, 35</sup> have  
69 been reported for Hg<sup>2+</sup> assay. These methods mainly relied on the fluorescence  
70 resonance energy transfer (FRET) process. Although these methods possess high  
71 sensitivity and selectivity, these methods require at least one end tag modification of  
72 DNA probe. The requirement of fluorescently labeled probes increases the  
73 experimental cost and the design complexity, making the assays time-consuming,  
74 laborious and cost-intensive.<sup>36, 37</sup> To circumvent the aforementioned limitations, a  
75 kind of DNA intercalators (SYBR Green I, TOTO-3 and YOYO-1), specific  
76 fluorometric dyes, were used to develop label-free Hg<sup>2+</sup> sensor<sup>38</sup>. Recently, Xing  
77 developed a new label-free Hg<sup>2+</sup> ions assay with polymerase assisted fluorescence  
78 amplification, resulting in a low detection limit of 40 pM and high specificity<sup>39</sup>.  
79 However, there is a dispute about the toxicity of some intercalation dyes. In theory,  
80 intercalation dyes are not safe enough and may be a threat to the health of the  
81 operators<sup>29</sup>. Recently, Feng *et al* developed a novel label-free fluorescent sensor for  
82 Hg<sup>2+</sup> based on target-induced structure-switching of G-quadruplex.<sup>40</sup> Although the  
83 label-free fluorescence method does not require any chemical modification for DNA,  
84 it shows poor detection limits with 25 nM, which can hardly meet the demanded  
85 sensitivity which is below 10 nM (the maximum contamination level defined by the  
86 US Environmental Protection Agency (EPA)). Consequently, the development of a  
87 novel, label-free and signal-amplified approach for further improving sensitivity and  
88 selectivity is highly desirable.

89 More recently, an isothermal amplification method designated as rolling circle

90 amplification (RCA), has gained considerable attention as a novel tool to amplify the  
91 recognition events detections. Due to the simplicity, robustness and high signal  
92 amplification of RCA, numerous novel methods based on RCA has been popularly  
93 employed in DNA,<sup>41</sup> RNA,<sup>42</sup> and protein<sup>43</sup> detection. Xing firstly reported a novel  
94 electrochemiluminescent sensor for Hg<sup>2+</sup> detection by using padlock probe-based  
95 RCA, resulting in a detection limit of 100 pM.<sup>44</sup> Although the assay exhibits  
96 acceptable sensitivity and high specificity, the approaches need the complex labeling  
97 procedure and chemical modification for DNA, making the assays time-consuming,  
98 laborious and cost-intensive. Consequently, the development of a novel, label-free and  
99 signal-amplified approach for further improving sensitivity and selectivity is highly  
100 desirable.

101 In this study, we developed a novel label-free fluorescent amplification sensing  
102 scheme based on target-triggered dumbbell probe-mediated rolling circle  
103 amplification (D-RCA)-responsive G-quadruplex formation for highly sensitive and  
104 selective detection of Hg<sup>2+</sup>. A dumbbell-shaped DNA probe (D-DNA) that integrates  
105 target-binding, amplification and signaling within one multifunctional design was  
106 ingeniously designed. Hg<sup>2+</sup>-primer DNA (Hg<sup>2+</sup>-p-DNA) was designed to  
107 complementary to the region of D-DNA but with T-T mismatches. It can initiate  
108 rolling circle amplification (D-RCA) in the presence of Hg<sup>2+</sup> target, resulting in  
109 numerous cascade dumbbell probes with duplex G-rich quadruplex structure oligomer.  
110 Upon the addition of N-methyl mesoporphyrin IX (NMM), the strong interaction  
111 between the G-quadruplex and NMM brings about a great fluorescence enhancement



112 for  $\text{Hg}^{2+}$  detection. The scheme takes advantage of the highly specific T- $\text{Hg}^{2+}$ -T  
113 complex-dependence of the DNA hybridization, the powerful signal amplification  
114 capability of RCA and the significant increase of the fluorescence signal from the  
115 interaction between N-methyl mesoporphyrin IX (NMM) and G-quadruplexes. NMM  
116 is a commercially available unsymmetrical anionic porphyrin characterized by a  
117 pronounced structural selectivity for G-quadruplex forms. It is weakly fluorescent, but  
118 exhibits a prominent enhancement in its fluorescence upon binding to G-quadruplex  
119 DNA. We aim to improve the sensitivity of  $\text{Hg}^{2+}$  detection via RCA and the  
120 selectivity by using the dumbbell probe to reduce non-specific amplification and the  
121 highly specific T- $\text{Hg}^{2+}$ -T complex-dependence of the DNA hybridization. In addition,  
122 we achieve the label-free by the strong interaction between the G-quadruplex and  
123 NMM.

### 124 **Scheme 1**

125

## 126 **2. EXPERIMENTAL SECTION**

### 127 **2.1 Materials and Reagents**

128 Phi29 DNA polymerase, T4 DNA ligase, and dNTP were obtained from  
129 Fermentas (Lithuania). The oligonucleotides with the following sequences were  
130 obtained from Shanghai Sangon Biological Engineering Technology & Services Co.,  
131 Ltd. (China): the dumbbell probe (D-DNA): 5'-TCTTTCTTCCGA CATCAACCCA  
132 AAACCCAAAA CCCAAAAC CCAAG ATGTTCG CACGCTAAA CCC AAA ACCC  
133 AA AACCCAAAAC CCAATA GCGT GGTGTTTCCT-3'.  $\text{Hg}^{2+}$ - specific primer

134 DNA ( $\text{Hg}^{2+}$ -p-DNA): 5'-CTTGTTTGTT GGTTTCTC-3'. The  $\text{Hg}^{2+}$  stock solution  
135 ( $1.0 \times 10^{-3}$  M) was prepared in ultrapure water with 2 drops of concentrated nitric acid.  
136 Other chemicals (analytical grade) were obtained from standard reagent suppliers.  
137 Water ( $\geq 18.2$ M) was used and sterilized throughout the experiments. N-methyl  
138 mesoporphyrin IX (NMM) was purchased from J&K Scientific Ltd. (Beijing, China).

## 139 **2.2 Apparatus**

140 All the fluorescence measurements were performed on a Hitachi F-7000  
141 spectrofluorimeter (Hitachi, Japan). The excitation wavelength was 399 nm, and the  
142 spectra are recorded between 600 and 620 nm. The fluorescence emission intensity  
143 was measured at 608 nm.

144

## 145 **2.3 Ligation reactions**

146 The ligation reaction was carried out with 20  $\mu\text{l}$  of a reaction mixture [40 mM  
147 Tris-HCl, 10 mM  $\text{MgCl}_2$ , 10 mM dithiothreitol (DTT) (pH 7.8), 500 mM ATP, 75 nM  
148 the dumbbell probe (D-DNA) and 25 nM  $\text{Hg}^{2+}$ -p-DNA]. Before adding T4 DNA  
149 ligase, the oligonucleotide mixture was firstly denatured at 95 °C for 3min, and  
150 cooled slowly to room temperature over a 10-min period. Then, the reaction mixture  
151 was incubated to hybridize at 37 °C for 30 min in the presence of 5  $\mu\text{L}$   $\text{Hg}^{2+}$  solution  
152 of different concentration. Then, 1  $\mu\text{L}$  (5 U) T4 DNA ligase was added to the mixture  
153 and incubated at 37 °C for 2 h. The full hybridization equilibrium between the  
154 D-DNA and p-DNA is a key factor for ensuring efficient ligation reaction. Therefore,  
155 the effect of molar ratio of the D-DNA to p-DNA was evaluated in **Fig. S1**(see

156 supporting information).

## 157 **2.5 RCA reactions**

158 For RCA, the aforementioned ligation product was mixed with 4  $\mu\text{L}$   $10 \times$   
159 reaction buffer [330 mM Tris-acetate, 100 mM  $\text{Mg}(\text{Ac})_2$ , 660 mM Potassium Acetate  
160 (KAc), 1% Tween 20 and 10 mM DTT (pH 7.9) ], 2  $\mu\text{L}$  (10 u/ $\mu\text{L}$ ) phi29 DNA  
161 polymerase and 8  $\mu\text{L}$  10 mM dNTPs. The reaction mixture was incubated at 37  $^\circ\text{C}$  for  
162 120 min.

## 163 **2.6 Measurement of fluorescent spectra**

164 The RCA amplification product was mixed with 5  $\mu\text{L}$  250  $\mu\text{M}$  N-methyl  
165 mesoporphyrin IX (NMM) and 5  $\mu\text{L}$  470 mM KCl, final volume of 50  $\mu\text{L}$ . The  
166 reaction mixture was incubated at 37  $^\circ\text{C}$  for 30 min. The fluorescent spectra were  
167 measured using a spectrofluorophotometer. The excitation wavelength was 399 nm,  
168 and the spectra are recorded between 600 and 620 nm. The fluorescence emission  
169 intensity was measured at 608 nm.

170

## 171 **3. Results and discussion**

### 172 **3.1 Sensing strategy**

173 The designed strategy is conceptually depicted in Scheme 1. The dumbbell  
174 padlock probe (D-DNA) with two stem-loop structures was chosen as RCA unit for  
175 the amplified sensing system based on its high sensitivity and selectivity. The D-DNA  
176 three kinds of domains, a  $\text{Hg}^{2+}$ -specific primer-binding domain ( $\text{Hg}^{2+}$ -PBD), a stem  
177 domain and two loop domains.  $\text{Hg}^{2+}$ -specific primer ( $\text{Hg}^{2+}$ -p-DNA) can be hybridized

178 to the dumbbell padlock probe template in the presence of  $\text{Hg}^{2+}$  based on T- $\text{Hg}^{2+}$ -T  
179 construction. Phi29 DNA polymerase can initiate the RCA reaction, resulting in  
180 numerous cascade dumbbell probes with duplex G-rich oligomer. This G-rich  
181 oligomer fold into a quadruplex structure with monovalent ions. Upon the addition of  
182 NMM, the signal reporter, the strong interaction between the “activated”  
183 G-quadruplex and NMM brings about a great fluorescence enhancement. Eventually,  
184 RCA provides an amplified detection signal for the target  $\text{Hg}^{2+}$ . In contrast, in the  
185 absence of target  $\text{Hg}^{2+}$ ,  $\text{Hg}^{2+}$ -p-DNA can not be hybridized to the template D-DNA,  
186 and the RCA process could not proceed. Therefore, there was no fluorescence. In this  
187 way, we successfully converted each  $\text{Hg}^{2+}$ -triggered RCA reaction event into the  
188 detectable fluorescent signals, which were significantly amplified by RCA in an  
189 isothermal fashion.

### 190 **3.2 The verification of the sensing strategy**

191 In this novel strategy, RCA was a crucial step, which mediated the generation  
192 and amplification of the fluorescence signal. To verify the amplification of the RCA  
193 reaction, the agarose gel electrophoresis experiment was performed. The RCA  
194 products were investigated by gel electrophoresis. It is observed that the RCA  
195 products in lane 2-3 reaction show extremely low mobility in **Fig. 1A**. The anticipated  
196 high molecular weight of RCA product was confirmed in lane 2, 3. Compared with  
197 lane 2, 3, lane 1 displayed no bands in negative control experiment. These results give  
198 immediate evidence for the high molecular weight of these products, indicating  $\text{Hg}^{2+}$   
199 acted as a trigger of the RCA reaction and the signal enhancement had a positive

200 correlation with the  $\text{Hg}^{2+}$  level. In addition, the amplification of the RCA reaction was  
201 also verified using fluorescent intensity. Typical fluorescence spectra characteristics  
202 of the sensing strategy in response to  $\text{Hg}^{2+}$  are shown in **Fig. 1B**. Compared with the  
203 background fluorescence of the NMM, 10 nM  $\text{Hg}^{2+}$  resulted in a significant  
204 fluorescence enhancement while a control experiment without  $\text{Hg}^{2+}$  only exhibited a  
205 negligible fluorescence change. The result provided a convincing proof of the  
206 detection mechanism of the proposed sensing strategy shown in Scheme 1.

207 **Fig. 1.**

208 **3.3 Optimization of parameters-dependent signal amplification of the sensing**  
209 **strategy**

210 In order to achieve the system's best sensing performance, several experimental  
211 parameters affecting RCA were investigated. The full hybridization equilibrium  
212 between the  $\text{Hg}^{2+}$ -p-DNA and D-DNA is a key factor for ensuring RCA reaction.  
213 Therefore, the effect of the concentration of  $\text{Hg}^{2+}$ -p-DNA was evaluated. As shown in  
214 **Fig. S2A** (see supporting information), the fluorescence intensity increased with the  
215 increase of the concentration of  $\text{Hg}^{2+}$ -p-DNA. When the concentration of  
216  $\text{Hg}^{2+}$ -p-DNA reached 10 nM, the maximum fluorescence intensity was achieved.  
217 Thereafter, the fluorescence response exhibited a gradual decrease with a further  
218 increase of the concentration of  $\text{Hg}^{2+}$ -p-DNA. This was probably because a large  
219 excesses of  $\text{Hg}^{2+}$ -p-DNA disturbed their hybridization with the D-DNA in a  
220 head-to-tail fashion and the subsequent D-RCA reaction.<sup>45</sup> As a result, 10 nM was  
221 selected as the optimal concentration of  $\text{Hg}^{2+}$ -p-DNA due to its strongest fluorescence

222 intensity. In theory, more complementary copies (duplex G-quadruplexes DNA) of the  
223 dumbbell probes template are generated with the elongation of RCA reaction time;  
224 stronger signal amplification will be produced. So the effect of RCA reaction time on  
225 the fluorescence signal was examined, which is shown in **Fig. S2B** (see supporting  
226 information). The fluorescence intensity enhanced quickly with the increase in  
227 reaction time, and nearly reached a plateau after 120 min. This might be attributed to  
228 the fact that the RCA reaction had reached equilibrium caused by exhaustion of the  
229 RCA substrates or inactivation of phi29 DNA polymerase. Therefore, 120 min was  
230 chosen as the optimum time for the RCA reaction. This time was in agreement with  
231 the reported RCA reaction time of 1-2 h.

232 To achieve the best sensing performance, the concentration of NMM and  $K^+$  were  
233 also optimized. As shown in **Fig. S2C-D**(see supporting information). The  
234 experimental results indicated that a concentration of 100 mM of  $K^+$  and 25  $\mu$ M of  
235 NMM could provide maximum S/N ratio for the sensing system.

### 236 **3.4 The sensibility and detection range of the designed sensing system**

237 In our study, we evaluated the sensitivity of the  $Hg^{2+}$  ions fluorescence sensor  
238 under optimized conditions. **Fig. 2A** illustrates fluorescence spectra of sensing system  
239 after addition of different concentrations of  $Hg^{2+}$  ions (0 - 10 nM) under the optimal  
240 conditions. Significantly, the fluorescence intensity gradually increased along with  
241  $Hg^{2+}$  ions concentration, leading to an impressively large dynamic range that spans  
242 five orders of magnitude (0 to 10 nM). **Fig. 2B** shows the logarithmic relationship  
243 between the fluorescence response and the different concentrations of  $Hg^{2+}$  ions. The

244 linear concentration ranges were 100 fM up to 10 nM for  $\text{Hg}^{2+}$  ions over a 6-decade  
245 concentration range with a linear correlation coefficient of 0.992. The calculated limit  
246 of low detection is 80 fM for  $\text{Hg}^{2+}$  ions in terms of the  $3\sigma$  rule. The detection limit of  
247 the newly designed  $\text{Hg}^{2+}$  ions sensing system is 2-3 orders of magnitude lower than  
248 that of previously reported fluorescent methods<sup>38, 39, 46</sup>. The ultrahigh sensitivity was  
249 attributed to the following factors. First, the introduction of  $\text{Hg}^{2+}$  ions would trigger  
250 the RCA reaction, and eventually RCA product with many tandem repeated duplex  
251 G-quadruplexes was generated in large quantities by RCA, substantially amplifying  
252 each target-triggered RCA reaction event. Second, free NMM and D-DNA showed  
253 relatively low background fluorescence, which dramatically increased the  
254 fluorescence signal-to-noise (S/N) ratio.

255 **Fig. 2.**

### 256 **3.5 The selectivity of this sensing system**

257 **Fig. 3.**

258 To test the selectivity of this novel method for  $\text{Hg}^{2+}$  ions detection, control  
259 experiments were executed to evaluate whether other environmentally relevant metal  
260 ions performed the similar function as the  $\text{Hg}^{2+}$  ions in the current biosensor. As  
261 revealed in **Fig. 3**, one can find that only the  $\text{Hg}^{2+}$  ions samples show a significant  
262 higher fluorescence intensity relative to the competing metal ions samples at identical  
263 conditions. The competing metal ions exhibited almost the same fluorescence  
264 response as the blank solution without target, and did not induce any significant signal.  
265 In addition, the co-existence of other metal ions with  $\text{Hg}^{2+}$  in the sample also did not

266 affect  $\text{Hg}^{2+}$  detection (**Fig.4**). This result obviously indicated that the proposed  
267 strategy had sufficient selectivity in  $\text{Hg}^{2+}$  ions detection. The high selectivity makes it  
268 promising for practical applications. The excellent specificity was attributed to using  
269 the dumbbell probe to reduce non-specific amplification and the highly specific  
270 T- $\text{Hg}^{2+}$ -T complex-dependence of the DNA hybridization.

271

**Fig. 4.**

### 272 **3.6 Application of designed sensing system in the real samples**

273 In order to verify the usefulness of the proposed sensor for the identification and  
274 detection of  $\text{Hg}^{2+}$  in practical applications, the detection of natural  $\text{Hg}^{2+}$  in real  
275 samples (tap water, orange juice, and lake water) was demonstrated here by the use of  
276 the designed sensing system. The samples collected were first centrifuged for 10 min  
277 at 10000 rpm, then filtered through a 0.22  $\mu\text{m}$  membrane and detected according to  
278 the general procedure with three replicates. Prior to the assay, the samples were  
279 diluted 10 times with ultrapure water so that the level of  $\text{Hg}^{2+}$  was within the linear  
280 ranges. The results averaged from three determinations are summarized in Table 1 (in  
281 supporting information). The data listed in **Table 1**(see supporting) indicated that the  
282 obtained results were highly consistent with those from the atomic fluorescence  
283 spectrometry (AFS) method. Meanwhile, when the known amount of  $\text{Hg}^{2+}$  was added  
284 in these samples, the recovery of  $\text{Hg}^{2+}$  ranged from 98% to 107%. These confirm that  
285 the designed sensor has a high accuracy and sensitivity to meet the requirements of  
286 the application.

287

**Table 1**



#### 288 **4. Conclusions**

289 In summary, we have successfully demonstrated a novel label-free fluorescent  
290 sensing scheme based on target-responsive dumbbell probe-mediated rolling circle  
291 amplification (D-RCA) for sensitive and selective detection of  $\text{Hg}^{2+}$  ions monitoring.  
292 The label-free cascade fluorescence amplification strategy shows promising  
293 capabilities of specific and sensitive analysis for  $\text{Hg}^{2+}$  ions detection. The advantages  
294 of the strategy include: (i) an extremely low detection limit as low as 80 fM for  $\text{Hg}^{2+}$   
295 ions is respectively achieved. More significantly, the dynamic range of the based on  
296  $\text{Hg}^{2+}$ -mediated D-RCA is extremely large, covering 6 orders of magnitude; (ii) this  
297 label-free design does not require any chemical modification for DNA or  
298 sophisticated equipments, which offers the advantages of simplicity and cost  
299 efficiency. (iii) The strong T- $\text{Hg}^{2+}$ -T interaction and the uniquely designed dumbbell  
300 probe ensure the advantages of excellent specificity and reduce non-specific  
301 amplification. In addition, the label-free cascade fluorescence amplification strategy  
302 based on D-RCA strategy shows a high accuracy for the application. We expect that  
303 this highly sensitive and inexpensive D-RCA strategy will become a promising  
304 quantification method in both environmental and food safety fields.

305

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397 **Figure captions**

398 **Scheme 1** Schematic representation of the process of the label-free fluorescent  
399 sensing system based on target-responsive dumbbell probe-mediated rolling circle  
400 amplification (D-RCA) strategy for detection of mercuric ion.

401

402 **Fig. 1.** (A) Agarose gel (0.7%) electrophoresis experiments. The products of RCA  
403 reaction (1 h) (indicated by 1-3) were denatured at 95 °C for 5 min and quenched with  
404 ice-cooled water for 10 min. The marker was indicated by M. Line 1, the control  
405 without  $\text{Hg}^{2+}$ ; Line 2-3, the positive with  $\text{Hg}^{2+}$ . The high molecular weight RCA  
406 products are observed in lines 2-3. (B) The fluorescent intensity and fluorescence  
407 emission spectra under different conditions: NMM;  $\text{Hg}^{2+}$ -p-DNA + D-DNA +  $\text{Hg}^{2+}$   
408 (10 nM) + Phi29 DNA polymerase + dNTPs + NMM;  $\text{Hg}^{2+}$ -p-DNA + D-DNA +  
409 Phi29 DNA polymerase + dNTPs + NMM.

410

411 **Fig. 2.** (A) Fluorescence emission spectra obtained in the dumbbell probe-mediated  
412 RCA label-free cascade amplification strategy for detection of  $\text{Hg}^{2+}$  ions with varying  
413 concentrations from 0 - 10 nM. (B) Linear relationship between the fluorescent  
414 intensity and the concentration of  $\text{Hg}^{2+}$  ions. Each data point represents an average of  
415 6 measurements (each error bar indicates the standard deviation)

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417

418 **Fig. 3.** Selectivity of the label-free cascade amplification strategy for  $\text{Hg}^{2+}$  ions  
419 compared to other tested metal ions, respectively.

420

421 **Fig. 4.** Fluorescence intensity obtained in the different solutions. (a) 10 nM Hg<sup>2+</sup> in  
422 the standard solution, (a') 10 nM Hg<sup>2+</sup> in a mixture solution, (b) the standard solution  
423 as control, (b') the mixture solution. The mixture solution containing a mixture of  
424 other metal ions (Mix; Ni<sup>2+</sup>, Fe<sup>2+</sup>, Fe<sup>3+</sup>, Cd<sup>2+</sup>, Co<sup>2+</sup>, Cu<sup>2+</sup>, Mn<sup>2+</sup>, Pb<sup>2+</sup> and Zn<sup>2+</sup> (each  
425 100 nM)).

426 **Table 1.** Real Detection and Recovery Test of Hg<sup>2+</sup> in Different Real Samples.

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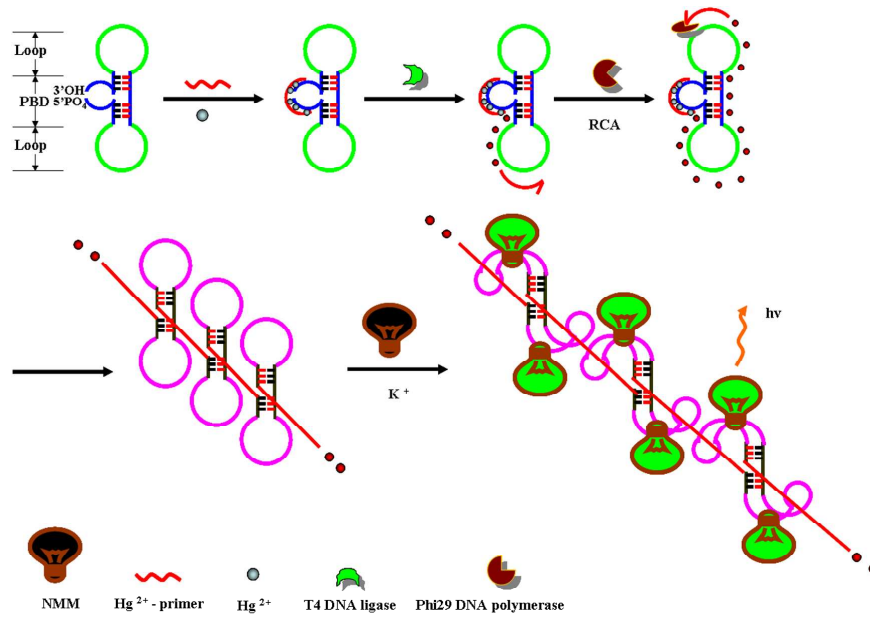
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## Scheme 1



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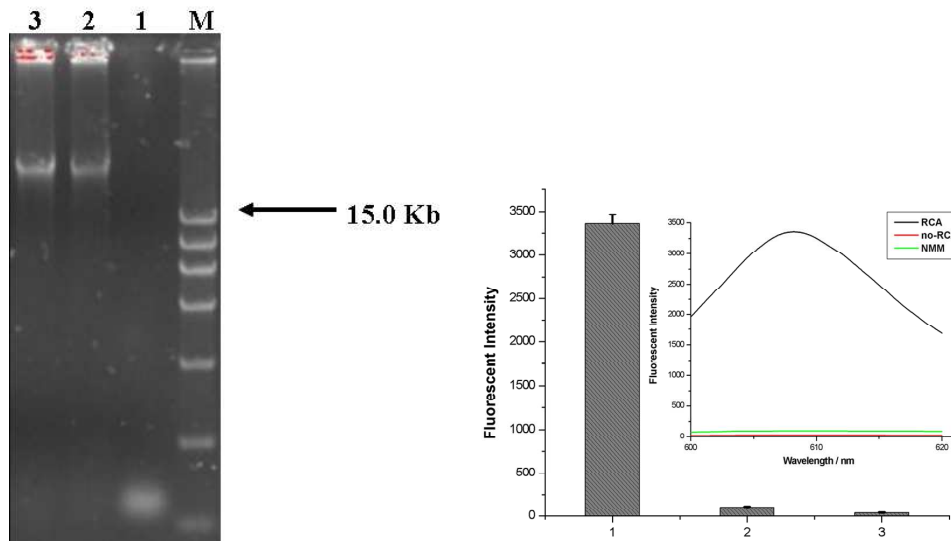
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Fig. 1.



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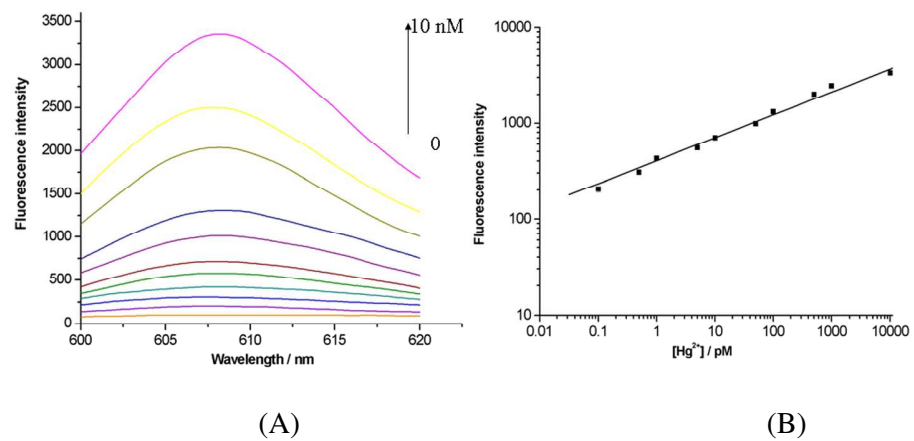
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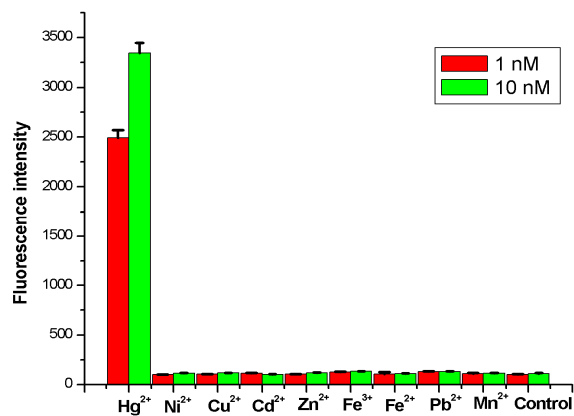
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473 **Fig. 2.**

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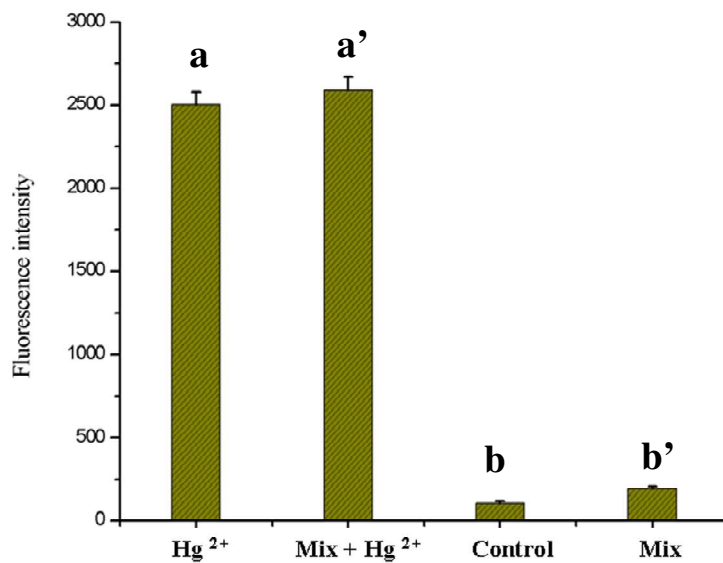
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476 **Fig. 3.**

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479 **Fig. 4.**



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**Table 1.****Table 1. Real Detection and Recovery Test of Hg<sup>2+</sup> in Different Samples**

sample	AFS method (pM) <sup>a</sup>	proposed method (pM) <sup>b</sup>	spiked Hg <sup>2+</sup> (pM)	measured Hg <sup>2+</sup> (pM)	recovery (%)
Orange juice	10.02	1.13	1.0	1.02 ± 0.03	102
Lake water	65.70	6.42	1.0	0.98 ± 0.02	98
tap water	18.5	1.92	1.0	1.07 ± 0.05	107

<sup>a</sup>The data represent natural Hg<sup>2+</sup> levels detected by standard atomic fluorescence spectrometry.

<sup>b</sup>The Hg<sup>2+</sup> concentrations were obtained after 10 times dilution with ultrapure water.

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