# **RSC Advances**



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This Accepted Manuscript will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

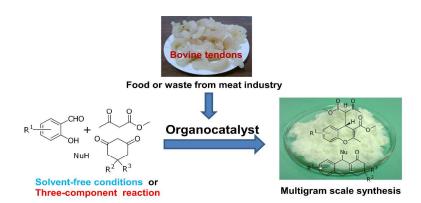
Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/advances

#### A Table of contents entry:

A new concept of catalysts which are prepared from renewable materials is demonstrated for the synthesis of coumarins and chromenes.



#### Cite this: DOI: 10.1039/c0xx00000x

#### www.rsc.org/xxxxxx

# **ARTICLE TYPE**

### Organocatalyst from renewable materials for the synthesis of coumarins and chromenes: three-component reaction and multigram scale synthesis<sup>†</sup>

55

Rapeepat Sangsuwan,<sup>a</sup> Sasithorn Sangher,<sup>b</sup> Thammarat Aree,<sup>c</sup> Chulabhorn Mahidol,<sup>a,b</sup> Somsak <sup>5</sup> Ruchirawat,<sup>a,b,d</sup> and Prasat Kittakoop\*<sup>a,b,d</sup>

Received (in XXX, XXX) XthXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

A new concept of catalysts which are prepared from renewable materials is demonstrated. It is known that amino acids (e.g., proline and hydroxyproline) are robust organocatalysts for several reactions. Bovine

- <sup>10</sup> tendons which are proteins rich in hydroxyproline and proline were used as a source of amino acids. An acid hydrolysate of tendons (a TH catalyst) could catalyze two reactions: (i) the synthesis of coumarins and chromenes under solvent-free conditions and (ii) the synthesis of densely functionalized 4Hchromenes *via* a three-component reaction. Moreover, an economical and easily accessible TH catalyst is applicable in a multigram scale synthesis of coumarins and chromenes, as well as in the three-component
- <sup>15</sup> reaction for chromene synthesis. A catalytic activity of hydroxyproline for the synthesis of 4*H*-chromenes *via* the three-component reaction was also discovered. The present work demonstrates not only the green catalysts from renewable materials, but also an environmentally benign preparation of coumarins and chromenes.

#### Introduction

A benzopyran chemical class, the fusion of a benzene ring and a pyran ring with various levels of saturation and oxidation, appears in many natural products.<sup>1</sup> 1-Benzopyran skeletons including coumarin 1 and 4*H*-chromene 2 (Fig.1) are important scaffolds in many drugs and bioactive natural products.

Fig. 1 Structure of coumarin 1 and 4H-chromene 2

- <sup>30</sup> Coumarins (2*H*-chromen-2-one derivatives) possess many biological activities including anticoagulant, anticancer, enzyme inhibition, vasorelaxant, antimicrobial, antioxidant, and anti-inflammatory, and anti-HIV activities.<sup>2-12</sup> In addition to the biological properties, they have been used in food additives, <sup>35</sup> cosmetics, optical brightener, and fluorescent and laser dyes.<sup>13</sup>
- 4*H*-Chromene moiety is present in many bioactive compounds

<sup>a</sup>Chulabhorn Graduate Institute, Chemical Biology Program, Kamphaeng Phet 6 Road, Laksi, Bangkok 10210, Thailand. Fax: +66-2-5538545; Tel:

 <sup>45</sup> <sup>d</sup>Center of Excellence on Environmental Health and Toxicology (EHT), CHE, Ministry of Education, Thailand.
 †Electronic Supplementary Information (ESI) available. See DOI: 10.1039/b000000x/ <sup>50</sup> and drug leads.<sup>14-18</sup> The examples of drugs currently used are warfarin **3a** and cromoglicic acid **3b** as anticoagulants and anti-asthmatic agents, respectively (Fig. 2). Because of their important use in pharmaceuticals, there are many synthetic methods towards the synthesis of these privileged structures.

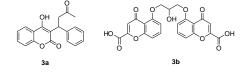


Fig. 2 Structure of warfarin 3a cromoglicic acid 3b

- Several synthetic routes have been reported for the synthesis of coumarins.<sup>19-21</sup> Whereas, chromene derivatives were synthesized by various methods, for example, DBU-catalyzed reaction between salicylic aldehydes and ethyl 2-methylbuta-2,3-dienoate;<sup>22</sup> tandem benzylation and cyclization by FeCl<sub>3</sub>;<sup>23</sup> Cu(I) <sup>65</sup> catalyzed domino reactions;<sup>24</sup> and triazine functionalized ordered mesoporous organosilica as an organocatalyst.<sup>25</sup> However, these procedures are not green methods since metals and hazardous solvents are used in the synthesis. Recently, green methods were developted for the synthesis of chromene and coumarin, for <sup>70</sup> example, four-component catalyst-free reaction in water<sup>26</sup> and biocatalytic domino reaction with the enzyme alkaline protease.<sup>27</sup> However, the development of new methods and catalysts, which provide economical and environmentally friendly routes, is still
- 75 Among various organocatalysts, amino acids play an important

required for both coumarin and chromene syntheses.

<sup>40 +66-86-9755777;</sup> E-mail: prasat@cri.or.th

<sup>&</sup>lt;sup>b</sup>Chulabhorn Research Institute, Kamphaeng Phet 6 Road, Laksi, Bangkok 10210, Thailand.

<sup>&</sup>lt;sup>c</sup>Department of Chemistry, Faculty of Science, Chulalongkorn University, Bangkok 10330, Thailand.

role in asymmetric organocatalysis. L-Proline and derivatives have been extensively used as efficient organocatalysts,<sup>28,29</sup> and they are employed in several asymmetric organic syntheses.<sup>30</sup> L-Proline and other amino acids have been demonstrated to be

- 5 powerful catalysts for various reactions, i.e., aldol reactions, Mannich reactions, Michael reaction, and a-functionalizations of carbonyl compounds.<sup>31</sup> Utilization of renewable materials for fine and industrial chemicals is one of the important topics in green chemistry,<sup>32-36</sup> because renewable resources provide not only the
- 10 reduction of environmental impacts, but also economical feasibility and sustainable productions. Normally, tendons are composed of various collagen fibers, which are proteins rich in hydroxyproline and proline;<sup>37-39</sup> both amino acids are known as robust organocatalysts.<sup>28-31,40</sup> In the present work, we demonstrate
- 15 a new concept of catalysts which are prepared from renewable materials. We used bovine tendons as a source of amino acids, and found that a tendon hydrolysate (TH) served as an excellent organocatalyst, catalyzing the synthesis of coumarins and 4Hchromenes under solvent-free conditions. The TH catalyst also
- 20 efficiently catalyzed the three-component reactions for the construction of densely functionalized 4H-chromenes, recently reported by Gu and coworkers.<sup>41</sup> Moreover, the cheap and easily accessible TH catalyst was applicable in a multigram scale synthesis of coumarins and chromenes, as well as in a multigram 25 scale chromene synthesis via the three-component reactions.

#### **Results and discussion**

The TH catalyst was simply prepared from the acid hydrolysis 30 of bovine tendons. The hydrolysate obtained from acid hydrolysis was neutralized by base, and then extracted with MeOH to give the TH catalyst. The reaction between salicylaldehyde (4a) methyl acetoacetate (5) catalyzed by the TH catalyst under solvent-free condition (at 55 °C) was initially investigated; this

- 35 reaction gave coumarin 6a and two diastereomers of chromene 7a as products (Fig. 3). We also found that methyl acetoacetate provided better yields of products than that of ethyl acetoacetate. To ensure that the reaction was not catalyzed by the residue of base or acid which was probably present in the prepared catalyst,
- 40 portions of the TH catalyst were dissolved in water, and pH of solutions were found to be at 6.5-7.2, confirming that the reaction was not due to the activity of base or acid.

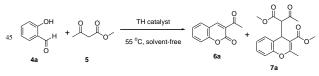


Fig. 3 Reaction of salicylaldehyde (4a) and methyl acetoacetate (5) catalyzed by the TH catalyst.

50

Next, we investigated the influence of the molar ratio of substrates (salicylaldehyde 4a to methyl acetoacetate 5) on the product ratios of coumarin (6a) to chromene (7a). As shown in Table 1, increasing the molar ratio of methyl acetoacetate (5)

55 gave a slighly increase in chromene (7a) production. We next investigated the effect of temperature for the reaction, and the reactions were conducted at room temperature (26-28 °C), 55 °C, and 80 °C. It was found that the times used for the reaction at

room temperature, 55 °C, and 80 °C were 96 h, 29 h, and 19 h, 60 respectively (the yields were >96% as indicated by <sup>1</sup>H NMR spectrum). Although increasing temperature could shorten the time for the reactions, performing the reaction at 55 °C was more reasonable, in term of the reduction of energy consumption, than that at 80 °C. We therefore decided to carry out the reaction at 55 65 °C for further experiments.

Table 1 Effects of the molar ratio of 4a : 5 on the yield of 6a and 7a<sup>a</sup>.

Entry	4a : 5	Time (h)	6a:7a <sup>b</sup>
1	1:1	65	15:8
2	1:3	65	3:4
3	1:5	65	7:11
4	1:7	65	1:2

<sup>a</sup> Reactions were performed using salicylaldehyde (50 mg, 0.41 mmol) and 20% (by weight) of the TH catalyst.

<sup>b</sup> The ratio of **6a**:**7a** was determined by <sup>1</sup>H NMR spectrum.

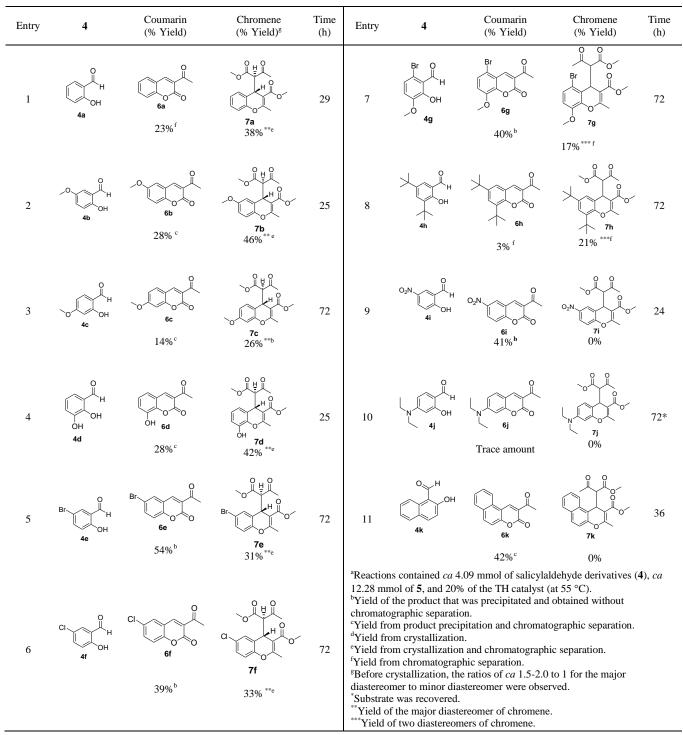
Next, we investigated the reaction of methyl acetoacetate (5) 70 and a variety of salicylaldehyde derivatives (4), using the molar ratio 1:3 of salicylaldehyde derivatives (4) to methyl acetoacetate (5) with 20 % of the TH catalyst (at 55 °C). As shown in Table 2, both coumarins and chromenes were obtained in low to moderate yields (3-54%). Unsubstituted salicylaldehyde and those bearing 75 electron-donating groups (Table 2, entries 1-4 and 8) favored the formation of chromenes (7a, 7b, 7c, 7d, and 7h) with respect to coumarins (6a, 6b, 6c, 6d, and 6h). In contrast, salicylaldehyde bearing an electron-withdrawing group (Table 2, entries 5 and 6) provided coumarins (6e and 6f) more than chromenes (7e and 7f). 80 5-Nitrosalicylaldehyde and 2-hydroxy-1-naphthaldehyde did not provide chromenes (7i and 7k), therefore only coumarins (6i and 6k) were obtained (Table 2, entries 9 and 11). This could be because the nitro group increases the acidity of the hydroxyl group, which is likely to be deprotonated, and thus reducing the 85 electrophilicity of the aldehyde group. It was found that 4-(diethylamino)salicylaldehyde (4j) did not give both coumarin

(6j) and chromene (7j) (Table 2, entry 10). This could be because a diethylamino group of 4j probably makes an aldehyde group less reactive via para-donating electron from a nitrogen atom. As mentioned earlier, this reaction gave two diastereomers of

chromene products with the ratio of ca 1.5-2.0 to 1 for the major diastereomer to minor diastereomer. <sup>1</sup>H NMR spectrum (ESI, Fig. 1S) of the two diastereomers (0.429 g) of chromene 7b indicated the ratio of 2 to 1 (a major isomer to a minor isomer). However, 95 after crystallization from EtOH/CH<sub>2</sub>Cl<sub>2</sub>, a large amount (0.397 g) of the major diastereomer of chromene 7b was obtained, leaving only 0.029 g of the filtrate containing the two chromene diastereomers at the ratio of 5:3 (ESI, Fig. 2S). This result suggested that only the major diastereomer could be 100 crystallizable, and that the minor diastereomer was converted to the major diastereomer during crystallization. As shown in Table 2 (entries 1-6), large amounts of the major diastereomer of chromenes 7a-7f (26-46% yield) were obtained after crystallization. It should be noted that chromenes 7g and 7h could 105 not be crystallized, and unfortunately they could not be separated by chromatographic techniques; therefore, percentage yields of chromenes 7g and 7h were of a mixture of the two diastereomers (Table 2, entries 7 and 8).

The structure of a major diastereomer of chromene **7a** was elucidated by analysis of spectroscopic data; extensive analysis of 2D NMR data established a planar structure of **7a**. Fortunately, appropriate crystals of the major diastereomer of chromenes **7a**, 5 **7b**, **7e**, and **7f** were obtained, and they were subjected to a single crystal X-ray analysis, which conclusively disclosed the relative

configuration of *S*\* and *R*\* for the positions 4 and 2', respectively (Fig. 4). We proposed that the isomerization between the two chromene diastereomers proceeds *via* an enol intermediate (ESI, <sup>10</sup> Fig. 3S). It is worth mentioning that this is the first report on the preparation of a single 4*S*,2'*R*-diastereomer of chromenes (e.g. **7a-7f**), particularly under environmentally benign conditions.



15 Table 2 Synthesis of coumarins and chromenes from salicylaldehyde derivatives (4) and methyl acetoacetate (5) using the TH catalyst<sup>a</sup>.

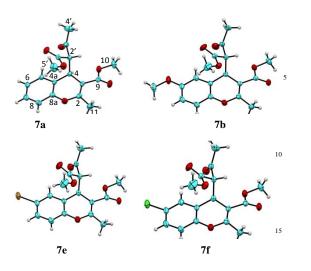


Fig. 4 ORTEP plots (30% probability level) of chromenes **7a**, **7b**, **7e**, and **7f** (color codes: C = cyan, O = red, Br = orange, Cl = green, H= white).

Tendons are proteins rich in hydroxyproline and proline amino acids.<sup>37-39</sup> The fact that both proline and hydroxyproline are robust organocatalysts,<sup>28-31,40</sup> we therefore propose that the two amino acids in the TH catalyst are possibly responsible for the <sup>25</sup> catalytic activity. Analysis of an amino acid content in the TH catalyst revealed that, among 20 natural amino acids, the percentage of proline and hydroxyproline were 15.89% and 13.12%, respectively (ESI, Table 2S), which were relatively high as compared to other amino acids, except glycine (19.23%). To <sup>30</sup> prove our hypothesis, we then used L-proline (20% mol) as a

catalyst for the reaction entries 1, 2, and 5 (Table 2). Indeed, we found that L-proline gave the same coumarins and chromenes as

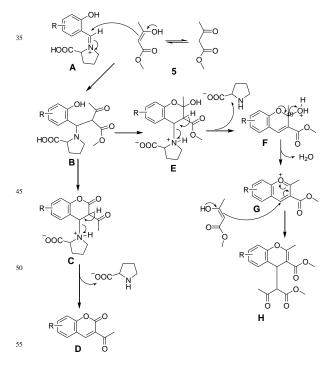


Fig. 5 A proposed mechanism for the formation of coumarin and chromene.

that obtained from the TH catalyst; respective yields of 60 coumarins **6a**, **6b**, and **6e** were 29%, 28%, and 50%, while those of chromenes **7a**, **7b**, and **7e** were 40%, 18%, and 24%, respectively. However, hydroxyproline (20% mol) did not catalyze such reaction, possibly due to the insolubility of hydroxyproline in solvent-free conditions (its precipitate was 65 observed). This result conclusively indicated that hydroxyproline

did not involve in the reaction under solvent-free conditions. We then performed the reaction in EtOH:H<sub>2</sub>O (9:1), in which hydroxyproline could be completely dissolved; the reaction proceeded under this condition, giving respective yields of 23%,

<sup>70</sup> 14%, and 24% for coumarins **6a**, **6b**, and **6e** and 9%, 17%, and 8% for chromenes **7a**, **7b**, and **7e**. Therefore, hydroxyproline also had a catalytic activity for the formation of chromenes and coumarins under a relatively polar condition that could dissolve hydroxyproline, but not under solvent-free conditions as shown in

<sup>75</sup> Table 2. A proposed mechanism for the formation of coumarin and chromene catalyzed by amino acids is shown in Fig. 5. The reaction starts with a condensation of an aldehyde and a catalyst (represented as L-proline), giving rise to an iminium intermediate **A**, which subsequently reacts with methyl acetoacetate (**5**), to

<sup>80</sup> give the intermediate **B**. Lactonization of **B** leads to the formation of the intermediate **C**. A release of a catalyst from the intermediate **C** gives rise to a coumarin **D**. Alternatively, a phenolic group of the intermediate **B** intramolecularly attacks a carbonyl to give the intermediate **E**; a release of a catalyst from <sup>85</sup> the intermediate **E** yields the intermediate **F**. Loss of water assisted by lone pair electrons on oxygen gives the oxonium intermediate **G**, which in turn reacts with methyl acetoacetate (**5**) to give a chromene **H** (Fig. 5).

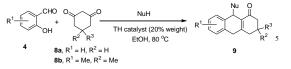
Since the TH catalyst is inexpensive, we performed multigram <sup>90</sup> scale synthesis of coumarin **6a** and chromene **7a** with three separated (solvent-free) conditions (Table 3). With *ca* 10 g of a substrate **4a**, respective yields of 28-41% and 33-37% for coumarin **6a** and chromene **7a** were obtained (Table 3), indicating that a practical multigram scale synthesis of coumarins <sup>95</sup> and chromenes is feasible with the TH catalyst. The TH catalyst is a green catalyst derived from renewable materials (bovine tendons), and the solvent-free conditions employed for this multigram scale synthesis is also environmentally friendly.

100	Table 3 Multigram	scale synthesis of	f coumarin <b>6a</b>	and chromene 7a.
-----	-------------------	--------------------	----------------------	------------------

				mene / u		
Entry	Weight	Ratio of	Temperature	Time	Yield of 6a	Yield of 7a
	of <b>4a</b> (g)	4a:5	(°C)	(h)	(g) (%)	(g) (%)
1	10.0	1:3	55	72	4.3 (28%)	8.6 (33%)
2	10.6	1:7	55	72	6.8 (41%)	10.4 (37%)
3	10.4	1:7	80	15	4.8 (30%)	9.9 (36%)

Next we used the TH catalyst for the synthesis of densely functionalized 4*H*-chromenes *via* a three-component reaction. Gu et al. reported the elegant green chemistry route for the synthesis <sup>105</sup> of densely functionalized 4*H*-chromenes *via* a three-component reaction of salicylaldehydes, 1,3-cyclohexanediones, and nucleophiles, employing L-proline as catalyst, and some products could be easily isolated by filtration, avoiding the use of chromatographic separations.<sup>41</sup> We followed Gu method for the <sup>110</sup> synthesis of 4*H*-chromenes, using our TH catalyst. As shown in Table 4, the reaction of salicylaldehyde derivatives (**4**), dimedone

 Table 4 Three-component reaction of salicylaldehyde derivatives (4),
 dimedone derivatives (8), and nucleophiles (NuH) by the TH catalyst.



Entry	NuH	Product	Time (h)	Yield (%)
1	N.N. N.N. N.H.	NN 9a	24	67
2	SH	S S O S O S O S O S O S O S O S O S O S	24	79
3			24	56
4	⊂, Lz		24	95
5	0	9d OF OH Haco O	24	76
6	Hz	9e Br	24	43
7		9f H <sub>3</sub> CO	16	66
8	н Х Соон	9g	16	73
9	OCH3	9h OCH3 Br	16	73
10			4	70

derivatives (8), and nucleophiles (NuH) gave chromenes (9a-9j) 10 with yields of 43-95%. It should be noted that chromene products (9a-9j) were obtained by filtration and washing with ethanol (without chromatographic separations). Chromenes 9a-9c were previously synthesized by Gu and coworkers with respective yields of 98, 88, and 87%,<sup>41</sup> however, these chromenes (9a-9c) 15 obtained from the present work (by the TH catalyst) were 67, 79, 56%, respectively (Table 4, entries 1-3). It should be noted that Gu and coworkers cooled the reaction mixture to 0 °C before filtration and washing with ethanol,<sup>41</sup> however, in the present work, this process was performed at room temperature (26-28 <sup>20</sup> °C). This may be the reason that the yields of 9a-9c were lower than Gu method.<sup>41</sup> Compounds 9d–9j were new chromenes (Table 4, entries 4-10) obtained from the TH catalyst with yields  $\geq$  70%, except **9f** (43%) and **9g** (66%). Although chromene products were simply obtained by filtration and washing with 25 ethanol, we observed that the derivatives that are relatively polar (e.g., 9c and 9f, Table 4, entries 3 and 6) had substantial yield losses from ethanol washing. Wheras non-polar products (e.g.,

9d) did not have much vield losses from ethanol washing, and

thus providing good yield (95%). Apart from the chromene synthesis reported by Gu and 30 coworkers,<sup>41</sup> there were other works recently reported for the synthesis of densely functionalized 4H-chromenes using ZnO nanoparticles,<sup>42</sup> tetrabutylammonium fluoride,<sup>43</sup> L-proline,<sup>44</sup> and iron (III) chloride and triphenylphosphine<sup>45</sup> as catalysts. The TH 35 catalyst is much cheaper than those catalysts previously employed for such chromene synthesis,<sup>41-45</sup> and we therefore used the TH catalyst for multigram scale synthesis of chromenes. As shown in Fig. 6, a scale of ca 10 g salicylaldehyde derivatives was used with 20% of the TH catalyst. After stirring the reaction 40 mixture at 80 °C for 15-18 h, chromene products (9k-9p) were simply obtained by filtration and washing with ethanol, giving yields of 56, 91, 74, 85, 96, and 97%, respectively (Fig. 6). This result indicates that the economical and green TH catalyst is applicable in the multigram scale synthesis of chromenes; the 45 products were easily obtained without chromatographic separations.

Since hydroxyproline was also present in the TH catalyst (ESI, Table 2S), we then performed the three-component reaction for the synthesis of 4*H*-chromenes **9n** and **9p** using hydroxyproline <sup>50</sup> as a catalyst. Surprisingly, quantitative yields of the chromenes **9n** (99.5%) and **9p** (99.8%) were obtained from the reaction; this result indicated that hydroxyproline in the TH catalyst should also involve in the formation of chromenes. This is the first report on a catalytic activity of hydroxyproline for the synthesis of 4*H*-<sup>55</sup> chromenes *via* the three-component reaction.

Overall, the present study highlights a feasible eco-friendly multigram scale synthesis of coumarins and chromenes with a green catalyst from renewable materials, bovine tendons. It should be noted that bovine tendons are used as food ingredients <sup>60</sup> in some countries in Asia, but they are waste from meat industry in Western countries. The multigram scale synthesis of coumarins and/or chromenes reported here could be performed under solvent-free conditions (Table 3) or without chromatographic separations of the products (Fig. 6). Therefore, both the catalyst <sup>65</sup> and the synthetic method are green and friendly to the environment.

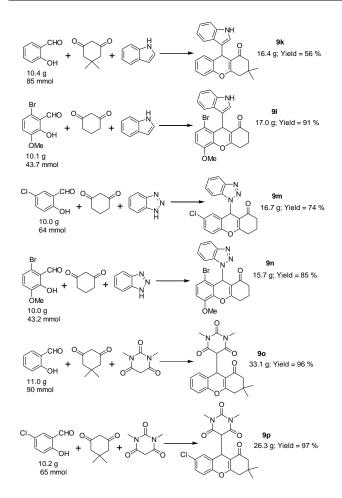


Fig. 6 Multigram scale synthesis of chromenes 9k-9p.

#### Conclusions

- <sup>5</sup> The economical, easily accessible, and environmentally friendly TH catalyst was prepared from the acid hydrolysis of bovine tendons which are renewable materials. The TH catalyst catalyzed the synthesis of coumarins and chromenes (e.g., **7a-7h**) under the solvent-free conditions, and it also catalyzed the
- <sup>10</sup> synthesis of densely functionalized 4*H*-chromenes *via* the threecomponent reaction. Moreover, the TH catalyst is applicable in the multigram scale synthesis of coumarins and chromenes, as well as in the three-component reaction for the construction of the 4*H*-chromenes. A multigram scale preparation of chromenes with
- <sup>15</sup> a single 4*S*,2'*R*-diastereomer was demonstrated for the first time. Proline, not hydroxyproline, in the TH catalyst possibly catalyzes the formation of coumarins and chromenes under solvent-free conditions. In addition to proline, hydroxyproline was also found to catalyze the three-component reaction for the synthesis of 4*H*-
- 20 chromenes. To the best of our knowledge, this is the first report on the green catalyst prepared from renewable materials.

#### **Experimental section**

#### 25 Instruments

Melting points were measured on Buchi 535 Melting Point

Apparatus and reported without correction. UV–Vis spectra were obtained from Shimadzu UV-1700 PharmaSpec Spectrophotometer. FTIR data were obtained using a universal <sup>30</sup> attenuated total reflectance (UATR) attachment on a Perkin– Elmer Spectrum One spectrometer. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker AM 300 NMR instrument (operating at 300 MHz for <sup>1</sup>H and 75 MHz for <sup>13</sup>C) and a Bruker AVANCE 400 NMR spectrometer (operating at 400 MHz for <sup>1</sup>H and <sup>35</sup> 100 MHz for <sup>13</sup>C). APCI TOF MS spectra were obtained from a Bruker MicroTOF<sub>LC</sub> spectrometer.

#### Preparation of the TH catalyst

Two batches of the TH catalyst were prepared, and the TH <sup>40</sup> catalysts from both batches had the same catalytic properties. Amino acid content in the TH catalyst is in Table 2S (ESI).

*The first batch.* Bovine tendon (135 g) was hydrolyzed in 110 mL of 6 M HCl. A mixture was stirred and refluxed for 12 h; pH of the mixture was adjusted to 7 with saturated NaOH. The <sup>45</sup> mixture was dried by a rotary evaporator, vaccum-dried, and then dissolved in methanol. The methanol soluble part was collected and dried to yield 7 g of the TH catalyst.

*The second batch.* Bovine tendon (256 g) was hydrolyzed in 90 mL of 6 M HCl. The mixture was stirred and refluxed for 18 h, <sup>50</sup> and its pH was adjusted to 7 with saturated NaOH. The mixture was dried by a rotary evaporator, vaccum-dried, and extracted with 100 mL of MeOH (eleven times, a total volume of 1100 mL). The MeOH extracts were combined and dried, giving 24 g of the TH catalyst.

## General procedure for the synthesis of coumarins and chromenes using the TH catalyst

A mixture of a salicylaldehyde derivative (500 mg, 1 equiv.) and methyl acetoacetate (3 equiv.) was added the TH catalyst (20 % by weight of a salicylaldehyde derivative). The reaction was stirred at 55 °C, and the progress of a reaction was monitored by TLC. After the reaction was complete (the time for each reaction is indicated in Table 2), if coumarin product precipitated, it was collected by filtration. However, if the coumarin product did not for precipitate, the reaction mixture was left at room temperature, in order to crystallize chromene product. In case that both chromene and coumarin did not crystallize or precipitate, the reaction mixture was subjected to chromatographic separations (i.e., Sephadex LH-20 column chromatography (CC), silica gel CC, 70 and preparative TLC). In some cases, chromene or coumarin in the fractions, obtained from abrometographic separations users

the fractions obtained from chromatographic separations were recrystallized from EtOH/CH<sub>2</sub>Cl<sub>2</sub>.

Multigram scale synthesis of coumarin **6a** and chromene **7a** was performed with *ca* 10 g of salicylaldehyde **4a**, using 20% of <sup>75</sup> the TH catalyst. The ratio of substrates **4a** and **5**, time, and temperature for the reaction were indicated in Table 3. After the reaction was complete, it was added an equal volume of CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O; compounds **6a** and **7a** were in the CH<sub>2</sub>Cl<sub>2</sub> layer, while the TH catalyst was in the H<sub>2</sub>O layer. The CH<sub>2</sub>Cl<sub>2</sub> layer was left <sup>80</sup> at room temperature, and chromene **7a** crystallized from the mixture, followed by coumarin **6a**. Finally, a mother liquid was separated by Sephadex LH-20 CC (eluted with MeOH); fractions were left at room temperature, and **6a** or **7a** individually crystallized from the fractions.

#### Synthesis of coumarin 6a and chromene 7a

Both 6a and 7a were prepared according to the general procedure. The crude reaction mixture was left at room temperature, and the major diastereomer of chromene 7a s crystallized from the mixture. After removing crystals of 7a, the

- reaction mixture was then washed with water to remove a catalyst, dried under vacuum, and further purified by Sephadex LH-20 CC (eluted with 100 % MeOH), followed by silica gel preparative TLC (developed with 20 % EtOAc in hexane) to
- <sup>10</sup> obtain coumarin **6a** and a mixture of the two diastereomers of chromene **7a**. The mixture of the two diastereomers of chromene **7a** was dissolved in a mixture of EtOH/CH<sub>2</sub>Cl<sub>2</sub> (2:1) and left at room temperature, and the major diastereomer of chromene **7a** again crystallized from a solution.
- <sup>15</sup> (*S*\*)-Methyl 4-((*R*\*)-1-methoxy-1,3-dioxobutan-2-yl)-2methyl-4*H*-chromene-3-carboxylate (7a). Colorless crystals; mp 119-122 °C; UV (MeOH)  $\lambda_{max}$  (log ε) 268 (3.68); IR (UATR)  $\nu_{max}$ : 3002, 2952, 2840, 1713, 1640, 1584, 1488, 1459, 1434, 1381, 1356, 1291, 1218, 1188, 1154, 1106, 1064, 990, 947,
- <sup>20</sup> 822, 758 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 2.21 (s, 3H), 2.44 (s, 3H), 3.44 (s, 3H), 3.65 (d, *J* = 4.5 Hz, 1H), 3.76 (s, 3H), 4.76 (d, *J* = 4.5 Hz, 1H), 6.98 (dd, *J* = 1.0 Hz, 8.1 Hz, 1H), 7.05 (td, *J* = 1.2 Hz, 7.5 Hz, 1H), 7.19 (td, *J* = 1.1 Hz, 8.0 Hz, 1H) 7.29 (dd, *J* = 1.5 Hz, 7.7 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.7,
- $_{25}$  29.5, 34.6, 51.5, 51.9, 66.1, 103.4, 115.8, 121.4, 124.3, 128.2, 129.1, 151.2, 163.9, 167.1, 168.9, 201.6; APCI-TOF MS: calcd. for  $C_{17}H_{18}NaO_6,\ \textit{m/z}$  341.0996 (M+Na)<sup>+</sup>, found 341.1000.

#### Synthesis of coumarin 6b and chromene 7b

- <sup>30</sup> Compounds **6b** and **7b** were prepared according to the general procedure. Coumarin **6b** precipitated from the reaction mixture. After removing coumarin **6b**, the reaction mixture was washed with water to remove a catalyst, dried under vacuum, and further purified by Sephadex LH-20 CC (eluted with 100 % MeOH) to
- <sup>35</sup> obtain additional amounts of coumarin **6b** and the mixture of two diastereomers of chromene **7b**. The mixture of two diastereomers of chromene **7b** was dissolved in a mixture of EtOH/CH<sub>2</sub>Cl<sub>2</sub> (2:1) and left at room temperature, and the major diastereomer of chromene **7b** crystallized from a solution.
- <sup>40</sup> (*S*\*)-Methyl 6-methoxy-4-((*R*\*)-1-methoxy-1,3-dioxobutan-2yl)-2-methyl-4*H*-chromene-3-carboxylate (7b). Yellow crystals; mp 97-98 °C; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 279 (3.83); IR (UATR)  $\nu_{max}$ : 2997, 2952, 2838, 1713, 1636, 1601, 1496, 1433, 1380, 1351, 1283, 1244, 1204, 1155, 1106, 1068, 1034, 992, 947,
- <sup>45</sup> 870, 814, 774, 721, 696 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ: 2.16 (s, 3H), 2.38 (s, 3H), 3.41 (s, 3H), 3.62 (d, J = 6 Hz, 1H), 3.71 (s, 6H), 4.69 (d, J = 3 Hz, 1H), 6.68 (dd, J = 3 Hz, 9 Hz, 1H), 6.80 (d, J = 3 Hz, 1H), 6.86 (d, J = 9 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ: 19.3, 29.0, 34.4, 51.0, 51.6, 55.0, 65.7,
- <sup>50</sup> 102.1, 112.6, 114.0, 116.1, 121.7, 144.9, 155.7, 163.8, 166.7, 168.6, 201.3; ESI-TOF MS: calcd. for  $C_{18}H_{20}NaO_7$ , m/z 371.1101 (M+Na)<sup>+</sup>, found 371.1106.

#### Synthesis of coumarin 6c and chromene 7c

<sup>55</sup> Compounds **6c** and **7c** were prepared according to the general procedure. The major diastereomer of chromene **7c** precipitated from the reaction mixture. After removing precipitate of **7c**, the resulting mixture was left at room temperature, and coumarin **6c** 

- was precipitated. After removing coumarin **6c**, the reaction <sup>60</sup> mixture was purified by Sephadex LH-20 CC, eluted with 100 % MeOH, to obtain fractions that contained coumarin **6c**, and these fractions were combined. A combined fraction containing **6c** was dried by a rotary evaporator, and dissolved in EtOH/CH<sub>2</sub>Cl<sub>2</sub>; coumarin **6c** crystallized from EtOH/CH<sub>2</sub>Cl<sub>2</sub>.
- <sup>65</sup> (*S*\*)-Methyl 7-methoxy-4-((*R*\*)-1-methoxy-1,3-dioxobutan-2yl)-2-methyl-4*H*-chromene-3-carboxylate (7c). White solid; UV (MeOH)  $\lambda_{max}$  (log ε) 271 (3.75), 222 (4.21); IR (UATR)  $\nu_{max}$ : 3003, 2952, 2841, 1705, 1638, 1583, 1508, 1437, 1385, 1358, 1337, 1288, 1260, 1243, 1206, 1189, 1157, 1140, 1069, 1035,
- <sup>70</sup> 1005, 952, 849, 805, 778, 715 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 2.21 (s, 3H), 2.43 (s, 3H), 3.47 (s, 3H), 3.64 (d, *J* = 3 Hz, 1H), 3.76-3.77 (s, 6H), 4.70 (d, *J* = 3 Hz, 1H), 6.54 (d, *J* = 3 Hz, 1H), 6.80 (d, *J* = 3 Hz, 1H), 6.63 (d, *J* = 9 Hz, 1H), 7.19 (d, *J* = 6 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.8, 29.6, 34.2,
- $_{75}$  51.6, 52.1, 55.4, 66.1, 101.1, 103.8, 110.8, 113.3, 129.8, 152.1, 159.6, 164.0, 167.3, 169.2, 202.0; ESI-TOF MS: calcd. for  $C_{18}H_{20}NaO_7,\ m/z\ 371.1101\ (M+Na)^+,\ found\ 371.1109.$

#### Synthesis of coumarin 6d and chromene 7d

- <sup>80</sup> Compounds **6d** and **7d** were prepared according to the general procedure. Coumarin **6d** precipitated from the reaction mixture. After removing coumarin **6d**, the reaction mixture was left at room temperature, and the major diastereomer of chromene **7d** crystallized from the mixture. After removing crystals of **7d**, the
- 85 reaction mixture was washed with water to remove a catalyst, dried under vacuum, and further purified by Sephadex LH-20 CC (eluted with 100 % MeOH), followed by silica gel CC (eluted with 20 % EtOAc in petroleum ether) to obtain additional amounts of coumarin 6d and a mixture of two chromene of diastereomers 7d. The mixture of the two chromene mixture of the two chromenes.
- <sup>90</sup> diastereomers **7d**. The mixture of the two chromene diastereomers **7d** was dissolved in EtOH/CH<sub>2</sub>Cl<sub>2</sub> (2:1) and left at room temperature; the major diastereomer of chromene **7d** crystallized from EtOH/CH<sub>2</sub>Cl<sub>2</sub>.
- (*S*\*)-Methyl 8-hydroxy-4-((*R*\*)-1-methoxy-1,3-dioxobutan-2-95 yl)-2-methyl-4*H*-chromene-3-carboxylate (7d). Colorless crystals; mp 124-126 °C; UV (MeOH)  $\lambda_{max}$  (log ε) 286 (3.62), 267 (3.65), 205 (4.20); IR (UATR)  $\nu_{max}$ : 3420, 3002, 2953, 2845, 1712, 1641, 1617, 1598, 1480, 1435, 1382, 1356, 1211, 1160, 1084, 995, 843, 781, 734 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 100 2.21 (s, 3H), 2.49 (s, 3H), 3.47 (s, 3H), 3.64 (d, *J* = 4.9 Hz, 1H), 3.78 (s, 3H), 4.76 (d, *J* = 4.8 Hz, 1H), 5.44 (s, 1H), 6.79-6.86 (m, 2H), 6.92-6.98 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.6, 29.5, 34.7, 51.6, 52.1, 66.0, 104.2, 114.6, 119.9, 122.1, 124.5, 139.1, 143.5, 163.3, 167.1, 168.9, 201.7; ESI-TOF MS: calcd. for 105 C<sub>17</sub>H<sub>18</sub>NaO<sub>7</sub>, *m*/z 357.0944 (M+Na)<sup>+</sup>, found 357.0952.

#### Synthesis of coumarin 6e and chromene 7e

Compounds **6e** and **7e** were prepared according to the general procedure. Coumarin **6e** precipitated from the reaction mixture. <sup>110</sup> After removing coumarin **6e**, the reaction mixture was left at room temperature, and the major diastereomer of chromene **7e** crystallized from the mixture. After removing crystals of **7e**, the reaction mixture was then washed with water to remove a catalyst, dried under vacuum, and further purified by silica gel <sup>115</sup> preparative TLC (developed with 25 % EtOAc in petroleum ether) to obtain the mixture of two chromene diastereomers **7e**.

The mixture of the two chromene diastereomers 7e was dissolved in EtOH/CH<sub>2</sub>Cl<sub>2</sub> and left at room temperature; the major diastereomer of chromene 7e crystallized from EtOH/CH<sub>2</sub>Cl<sub>2</sub>. (*S*\*)-Methyl 6-bromo-4-((*R*\*)-1-methoxy-1,3-dioxobutan-2-

- (3 )-Methyl **3 b b b b b b c** -
- <sup>10</sup> 3H), 3.48 (s, 3H), 3.64 (d, J = 3 Hz, 1H), 3.77 (s, 3H), 4.72 (d, J = 3 Hz, 1H), 6.87 (d, J = 9 Hz, 1H), 6.29 (m, 1H), 7.46 (s, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.7, 29.5, 34.3, 51.7, 52.2, 65.9, 103.3, 116.7, 117.6, 123.6, 131.2, 131.9, 150.5, 163.9, 166.9, 168.8, 201.24; ESI-TOF MS: calcd. for C<sub>17</sub>H<sub>17</sub>BrNaO<sub>6</sub>, <sup>15</sup> m/z 419.0101 (M+Na)<sup>+</sup>, found 419.0093.

#### Synthesis of coumarin 6f and chromene 7f

Compounds **6f** and **7f** were prepared according to the general procedure. Coumarin **6f** precipitated from the reaction mixture.

- <sup>20</sup> After removing coumarin **6f**, the reaction mixture was left at room temperature, and the major diastereomer of chromene **7f** crystallized from the mixture. After removing crystals of **7f**, the reaction mixture was washed with water to remove a catalyst, dried under vacuum, and purified by silica gel preparative TLC
- <sup>25</sup> (developed with 25 % EtOAc in petroleum ether) to obtain a mixture of the two diastereomers of **7f**. The mixture of two chromene diastereomers **7f** was dissolved in EtOH/CH<sub>2</sub>Cl<sub>2</sub> and left at room temperature; the major diastereomer of chromene **7f** crystallized from EtOH/CH<sub>2</sub>Cl<sub>2</sub>.
- <sup>30</sup> (*S*\*)-methyl 6-chloro-4-((*R*\*)-1-methoxy-1,3-dioxobutan-2-yl)-2-methyl-4*H*-chromene-3-carboxylate (7f).Colorless crystals; mp 111-113 °C; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 271 (3.48); IR (UATR)  $\nu_{max}$ : 3003, 2953, 2841, 1716, 1694, 1643, 1582, 1483, 1435, 1381, 1351, 1281, 1223, 1190, 1158, 1116, 1069, 992, 950,
- <sup>35</sup> 887, 820, 775, 664 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 2.21 (s, 3H), 2.43 (s, 3H), 3.48 (s, 3H), 3.64 (d, J = 4.4 Hz, 1H), 3.77 (s, 3H), 4.72 (d, J = 4.2 Hz, 1H), 6.92 (d, J = 8.7 Hz, 1H), 7.16 (dd, J = 2.5 Hz, 8.7 Hz, 1H), 7.32 (d, J = 2.4 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.6, 29.5, 34.4, 51.6, 52.1, 65.9, 103.2,
- <sup>40</sup> 117.1, 123.1, 128.2, 129.0, 129.2, 150.0, 163.9, 166.9, 168.8, 201.3; ESI-TOF MS: calcd. for C<sub>17</sub>H<sub>17</sub>ClNaO<sub>6</sub>, *m/z* 375.0606 (M+Na)<sup>+</sup>, found 375.0603.

#### Synthesis of coumarin 6g and chromene 7g

- <sup>45</sup> Compounds **6g** and **7g** were prepared according to the general procedure. Coumarin **6g** precipitated from the reaction mixture. After removing coumarin **6f**, the reaction mixture was washed with water to remove a catalyst, dried under vacuum, and purified by preparative TLC (50 % EtOAc in petroleum ether) to obtain <sup>50</sup> the mixture of two diastereomers of chromene **7g**.
- **3-Acetyl-5-bromo-8-methoxy-2H-chromen-2-one (6g).** Yellow solid; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 320 (3.80), 259 (3.62), 213 (4.08); IR (UATR)  $\nu_{max}$ : 3084, 2942, 1733, 1686, 1592, 1563, 1466, 1437, 1358, 1328, 1264, 1227, 1202, 1152, 1095, 953, 924, 832,
- <sup>55</sup> 765 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ: 2.74 (s, 3H) , 3.97 (s, 3H) , 7.04 (d, J = 9 Hz, 1H), 7.47 (d, J = 6 Hz, 1H), 8.73 (s, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ: 30.5, 56.5, 113.9, 116.1, 118.5, 125.3, 128.2, 145.7, 146.5, 146.6, 158.0, 195.0; ESI-TOF

MS: calcd. for  $C_{12}H_9BrNaO_4$ , m/z 318.9576 (M+Na)<sup>+</sup>, found <sup>60</sup> 318.9587.

Methyl 5-bromo-8-methoxy-4-(1-methoxy-1,3-dioxobutan-2-yl)-2-methyl-4*H*-chromene-3-carboxylate (7g). Yellow viscous oil; UV (MeOH)  $\lambda_{max}$  (log ε) 292 (3.82), 265 (4.01), 202 (4.61); IR (UATR)  $v_{max}$ : 2997, 2951, 2841, 2172, 1717, 1644, 1602,

- <sup>65</sup> 1578, 1477, 1435, 1382, 1357, 1312, 1237, 1210, 1155, 1096, 1062, 883, 800 cm<sup>-1</sup>; The major diastereomer: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 2.36 (s, 3H), 2.51 (s, 3H), 3.54 (s, 3H), 3.73 (s, 3H), 3.86 (s, 3H), 3.95 (d, *J* = 3 Hz, 1H), 4.81 (d, *J* = 3 Hz, 1H), 6.71-6.75 (m, 1H), 7.27-7.31 (m, 1H); <sup>13</sup>C NMR
- <sup>70</sup> (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.4, 30.0, 36.2, 51.3, 52.0, 56.2, 62.3, 101.5, 111.9, 112.4, 122.4, 127.4, 127.8, 147.3, 163.7, 167.1, 167.8, 201.1; The minor diastereomer: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 2.11 (s, 3H), 2.47 (s, 3H), 3.66 (s, 3H), 3.79 (s, 3H), 3.87 (s, 3H), 3.94-3.97 (m, 1H), 4.85 (d, *J* = 3 Hz, 1H), 6.71-6.75
- <sup>75</sup> (m, 1H), 7.27-7.31(m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ: 19.1, 30.1, 36.8, 51.7, 52.3, 56.3, 62.4, 103.8, 111.4, 112.4, 122.4, 127.4, 127.8, 147.2, 162.9, 167.1, 168.2, 201.2; ESI-TOF MS: calcd. for C<sub>18</sub>H<sub>19</sub>BrNaO<sub>7</sub>, *m*/*z* 449.0206 (M+Na)<sup>+</sup>, found 449.0215.

#### Synthesis of coumarin 6h and chromene 7h

Compounds **6h** and **7h** were prepared according to the general procedure. The reaction mixture was washed with water to remove a catalyst, dried under vacuum, and purified by Sephadex <sup>85</sup> LH-20 CC (eluted with 100 % MeOH) to obtain the mixture of two chromene diastereomers **7h** and the fractions that contained coumarin **6h**. The fractions containing **6h** were combined and

- coumarin **6h**. The fractions containing **6h** were combined and purified again by preparative TLC (40 % hexane in  $CH_2Cl_2$ ) to give coumarin **6h**.
- <sup>90</sup> Methyl 6,8-di-tert-butyl-4-(1-methoxy-1,3-dioxobutan-2-yl)-2methyl-4H-chromene-3-carboxylate (7h). Yellow viscous oil; UV (MeOH) λ<sub>max</sub> (log ε) 276 (3.83); IR (UATR) ν<sub>max</sub>: 2954, 2870, 1716, 1643, 1598, 1435, 1380, 1361, 1274, 1246, 1213, 1199, 1170, 1074, 994, 883, 832, 774 cm<sup>-1</sup>; The major
- <sup>95</sup> diastereomer: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.28-1.29 (s, 9H), 1.42 (s, 9H), 2.21 (s, 3H), 2.50 (s, 3H), 3.45 (s, 3H), 3.55 (d, J = 6.1 Hz, 1H), 3.75 (s, 3H), 4.72 (m, 1H) 7.15 (m, 1H), 7.22 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 19.5, 29.4, 30.0, 30.4 (3C), 31.4 (3C), 34.8, 36.1, 51.4, 52.0, 66.4, 103.8, 121.6, 122.4, 123.9,
- <sup>100</sup> 127.3, 136.1, 146.3, 163.9, 167.1, 169.0, 201.7; The minor diastereomer: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ: 1.28-1.29 (s, 9H), 1.42 (s, 9H), 1.78 (s, 3H), 2.49 (s, 3H), 3.64 (d, *J* = 6.5 Hz, 1H) 3.71 (s, 3H), 3.77 (s, 3H), 4.72 (m, 1H) 7.15 (m, 1H), 7.22 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ: 19.3, 29.5, 29.7, 30.0 (3C), <sup>105</sup> 31.4 (3C), 34.5, 36.7, 51.4, 52.2, 64.9, 104.2, 121.8, 122.5, 123.7,
- 127.2, 136.4, 146.6, 163.4, 167.1, 168.6, 202.3; ESI-TOF MS: calcd. for  $C_{25}H_{34}NaO_6$ , m/z 453.2248 (M+Na)<sup>+</sup>, found 453.2261.

#### Synthesis of coumarin 6i and chromene 7i

<sup>110</sup> Compound **6i** was prepared according to the general procedure. The reaction mixture was added ethanol to precipitate coumarin **6i**, and **6i** was then collected by filtration. Chromene **7i** was not obtained from this reaction.

#### $\scriptstyle 115$ Synthesis of coumarin 6j and chromene 7j

The synthesis was performed according to the general

procedure. However, the reaction did not give coumarin 6j and chromene 7j.

#### Synthesis of coumarin 6k and chromene 7k

<sup>5</sup> The synthesis was performed according to the general procedure. Coumarin **6k** precipitated from the reaction mixture. After removing coumarin **6k**, the resulting reaction mixture was then purified by silica gel preparative TLC (100 % CH<sub>2</sub>Cl<sub>2</sub>) to obtain additional amounts of coumarin **6k**. Chromene **7k** was not <sup>10</sup> formed from this reaction.

# General procedure of three-component reactions of salicylaldehyde derivatives (4), 1,3-cyclohexanedione derivatives (8), and nucleophiles (NuH) by the TH catalyst

- A mixture of salicylaldehyde derivatives (4) (1.0-1.7 mmol; 1 equiv.), 1,3-cyclohexanedione derivatives (8) (1 equiv.), NuH (1 equiv.) and the TH catalyst (20% by weight) in ethanol (3 mL) was stirred at 80  $^{\circ}$ C. The time for each reaction was indicated in Table 4. The reaction mixture was cooled to room temperature,
- <sup>20</sup> and it was filtered and washed with ethanol to yield chromenes. Spectroscopic data of chromenes **9a–9c**,<sup>41</sup> **9d**,<sup>43</sup> and **9e**<sup>46</sup> were identical to those reported in the liturature.

Multigram scale synthesis of chromenes was performed in the same manner as that mentioned above. Amounts of ca 10 g of

- <sup>25</sup> salicylaldehyde derivatives (4), with 1 equivalent of 1,3cyclohexanedione derivatives (8) and NuH, were used in the experiment; the reaction time was 15-18 h (Fig. 6). Spectroscopic data of chromene **9k** were in good agreement with those in the literature.  $^{42,44,45}$
- **7-Bromo-9-(1***H***-indol-3-yl)-2,3,4,9-tetrahydro-1***H***-xanthen-1one (9f). white solid; UV (MeOH) \lambda\_{max} (log ε) 280 (4.34) 221 (4.78); IR (UATR) \nu\_{max}: 3747, 3403, 3331, 3056, 2949, 2303, 1638, 1575, 1474, 1456, 1422, 1375, 1338, 1233, 1181, 1135,**
- <sup>35</sup> 1097, 1068, 997, 912, 816, 795, 739 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.62-2.06 (m, 2H), 2.32-2.36 (m, 2H), 2.57-2.79 (m, 2H), 5.28 (s, 1H), 6.96-7.01 (m, 2H), 7.07-7.14 (m, 2H), 7.22-7.30 (m, 3H), 7.36 (d, J = 8 Hz, 1H), 8.15 (s, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 20.3, 27.7, 29.6, 37.0, 111.4, 113.6, 117.2,
- <sup>40</sup> 118.0, 118.8, 119.5, 119.7, 121.7, 122.7, 125.5, 127.4, 130.5, 132.8, 136.5, 148.6, 165.7, 197.1; ESI-TOF MS: calcd. for  $C_{21}H_{16}BrNNaO_2$ , *m/z* 416.0257 (M+Na)<sup>+</sup>, found 416.0256.

#### 9-(1H-Indol-3-yl)-6-methoxy-3,3-dimethyl-2,3,4,9-tetrahydro-

<sup>45</sup> **1***H*-**xanthen-1-one (9g).** Orange solid; UV (MeOH)  $\lambda_{max}$  (log ε) 283 (4.09), 223 (4.65), 203 (4.59); IR (UATR)  $v_{max}$ : 3408, 3336, 3050, 2958, 2869, 1637, 1582, 1542, 1505, 1456, 1421, 1374, 1336, 1284, 1264, 1190, 1167, 1145, 1112, 1096, 1035, 1013, 957, 838, 782, 738, 704 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ: 0.05 (a, 2H), 1.10 (a, 2H), 2.17 (d, L= 16 Hz, 1H), 2.25 (d, L= 16).

- <sup>50</sup> 0.95 (s, 3H), 1.10 (s, 3H), 2.17 (d, J = 16 Hz, 1H), 2.25 (d, J = 16 Hz, 1H), 2.51 (d, J = 18Hz, 1H), 2.59 (d, J = 17 Hz, 1H), 3.76 (s, 3H), 5.25 (s, 1H), 6.56 (dd, J = 9, 3 Hz, 1H), 6.63 (d, J = 3 Hz, 1H), 6.98 (td, J = 8, 1 Hz, 1H), 7.05 (m, 2H), 7.15 (d, J = 2Hz, 1H), 7.25 (d, J = 8 Hz, 1H), 7.37 (d, J = 8 Hz, 1H), 8.02 (brs, 1H);  $^{13}$ C NUE (75 MUE (75 C))  $^{23}$  C 2 0, 41 5 50)
- $_{55} \ ^{13}\text{C NMR} \ (75 \ \text{MHz}, \ \text{CDCl}_3) \ \delta: \ 27.6, \ 28.9, \ 29.0, \ 32.0, \ 41.5, \ 50.9, \\ 55.4, \ 101.2, \ 111.2, \ 111.5, \ 113.0, \ 117.3, \ 119.0, \ 119.2, \ 120.5, \\ 121.5, \ 122.3, \ 125.6, \ 130.6, \ 136.5, \ 150.0, \ 158.8, \ 164.1, \ 197.4;$

ESI-TOF MS: calcd. for  $C_{24}H_{23}NNaO_3$ , *m*/z 396.1570 (M+Na)<sup>+</sup>, found 396.1574.

#### 3-(3,3-Dimethyl-1-oxo-2,3,4,9-tetrahydro-1*H*-xanthen-9-yl)-

**1***H*-indole-2-carboxylic acid (9h). White solid; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\varepsilon$ ) 296 (3.77), 227 (4.01); IR (UATR)  $\nu_{\text{max}}$ : 3261, 2937, 2877, 1676, 1644, 1579, 1547, 1485, 1446, 1417, 1376, 1350, 65 1319, 1289, 1257, 1227, 1201, 1183, 1143, 1199, 1034, 1015, 913, 875, 758, 736, 710, 682 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, DMSO*d*<sub>6</sub>)  $\delta$ : 0.87 (s, 3H), 1.05 (s, 3H), 1.98 (d, *J* = 16 Hz, 1H), 2.24 (d, *J* = 16 Hz, 1H), 2.60 (d, *J* = 18 Hz, 1H), 2.68 (d, *J* = 18 Hz, 1H), 6.25 (s, 1H), 6.87 (t, *J* = 7 Hz, 1H), 6.99 (td, *J*= 8, 3 Hz, 1H), 11.50 (s, 1H), 13.18 (brs, 1H); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>)  $\delta$ : 26.8, 27.3, 28.8, 31.5, 40.5, 50.3, 111.5 (2C), 112.7, 116.2, 119.2, 119.9, 123.7, 124.0, 124.7, 124.9, 125.2, 127.7, 129.7, 136.1, 148.6, 163.6, 164.1,196.0; ESI-TOF MS: calcd. for C<sub>24</sub>H<sub>22</sub>NO<sub>4</sub>, 75 *m*/z 388.1543 (M+H)<sup>+</sup>, found 388.1554.

#### 7-Bromo-9-((4-methoxybenzyl)thio)-2,3,4,9-tetrahydro-1H-

**xanthen-1-one** (**9i**). White crystals; mp 115-117 °C; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 279 (3.57), 205 (4.13); IR (UATR)  $\nu_{max}$ : <sup>80</sup> 2951, 2830, 1641,1608, 1574, 1509, 1474, 1411, 1374, 1301, 1233, 1170, 1133, 1033, 995, 908, 814, 734, 670 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): 1.81-2.02 (m, 2H), 2.27-2.49 (m, 4H), 3.57 (d, *J* = 14 Hz, 1H), 3.66 (d, *J* = 14 Hz, 1H), 3.79 (s, 3H), 4.94 (s, 1H), 6.80-6.89 (m, 3H), 7.17-7.20 (m, 3H), 7.28 (dd, *J* = 9,2 Hz, <sup>85</sup> 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): 19.9, 27.6, 34.7, 34.8, 36.8, 55.3, 111.8, 113.8 (2C), 117.5, 117.9, 124.8, 129.9 (2C), 130.2, 131.1, 132.5, 149.5, 158.6, 166.9, 196.2; ESI-TOF MS: calcd. for C<sub>21</sub>H<sub>19</sub>BrNaO<sub>3</sub>S, *m/z* 453.0131 (M+Na)<sup>+</sup>, found 453.0133.

<sup>90</sup> **5**-(**3**,**3**-Dimethyl-7-nitro-1-oxo-2,**3**,**4**,**9**-tetrahydro-1*H*-xanthen-**9**-yl)-1,**3**-dimethylpyrimidine-2,**4**,**6**(1*H*,**3***H*,**5***H*)-trione (**9j**). White solid; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 318 (3.94), 231 (4.23), 203 (4.51); IR (UATR)  $\nu_{max}$ : 3744, 3070, 2960, 2869, 1747, 1675, 1651, 1584, 1526, 1446, 1422, 1381, 1343, 1287, 1235, 1209, 95 1194, 1147, 1127, 1089, 1031, 905, 840, 801, 749, 735, 702 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.13 (s, 3H), 1.15 (s, 3H) 2.29 (d, *J* = 16 Hz, 1H), 2.37 (d, *J* = 16 Hz, 1H), 2.55 (s, 2H), 3.21 (s, 3H), 3.26 (s, 3H), 3.90 (d, *J* = 2 Hz, 1H), 4.97 (s, 1H), 7.17 (dd, *J* = 8,2 Hz, 1H), 8.1-8.2 (m, 2H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 100 26.9, 28.5, 28.6, 29.4, 32.1, 34.6, 41.3, 50.6, 55.4, 108.9, 117.7, 123.2, 124.4, 124.6, 144.5, 151.0, 154.6, 166.6, 166.7, 167.2, 197.5; ESI-TOF MS: calcd. for C<sub>21</sub>H<sub>21</sub>N<sub>3</sub>NaO<sub>7</sub>, *m*/z 450.1272 (M+Na)<sup>+</sup>, found 450.1262.

#### 105 8-Bromo-9-(1*H*-indol-3-yl)-5-methoxy-2,3,4,9-tetrahydro-

**xanthen-1-one (91).** White solid; UV (EtOH)  $\lambda_{max}$  (log ε) 291 (4.36), 273 (4.40), 221 (4.96), 213 (4.96); IR (UATR)  $\nu_{max}$ : 3338, 2319, 1644, 1573, 1471, 1428, 1372, 1311, 1217, 1182, 1135, 1092, 1062, 869, 800, 745 cm<sup>-1</sup>; <sup>1</sup>NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ: 1.66-1.97 (m, 2H), 2.17-2.36 (m, 2H), 2.60-2.76 (m, 2H), 3.87 (s, 3H), 5.30 (s, 1H), 6.86-7.01 (m, 3H), 7.08 (d, *J* = 2.4 Hz, 1H), 7.28 (d, *J* = 8.6 Hz, 2H), 7.35 (d, *J* = 7.9 Hz, 1H), 10.90 (s, 1H); <sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ: 19.9, 26.7, 29.5, 36.3, 55.9, 111.6, 111.9, 113.7, 114.3, 116.4, 118.3, 118.6, 120.5, 124.7, 115 125.2 (2C), 128.2, 136.0, 140.2, 147.1, 165.5, 195.9; ESI-TOF

MS: calcd. for  $C_{22}H_{18}BrNNaO_3$ , m/z 446.0362 (M+Na)<sup>+</sup>, found 446.0363.

#### 9-Benzotriazol-1-yl-7-chloro-2,3,4,9-tetrahydro-xanthen-1-

- <sup>5</sup> **one (9m).** Yellow solid; UV (EtOH)  $\lambda_{max}$  (log ε) 265 (4.45) ; IR (UATR)  $v_{max}$ : 3747, 2942, 1732, 1648, 1583, 1480, 1421, 1383, 1240, 1182, 1079, 1001, 823, 747 cm<sup>-1</sup>; <sup>1</sup>NMR (300 MHz, CDCl<sub>3</sub>) δ: 2.03-2.17 (m, 2H), 2.39-2.44 (m, 2H), 2.73-2.97 (m, 2H), 7.02 (s, 1H), 7.18-7.31 (m, 3H), 7.36 (d, J = 7.5 Hz, 1H),
- <sup>10</sup> 7.51 (t, J = 7.5 Hz, 1H), 7.74 (d, J = 8.4 Hz, 1H), 8.01 (d, J = 8.4 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 20.1, 27.8, 36.4, 48.2, 109.3, 109.6, 118.7, 120.0, 120.9, 123.9, 127.6, 128.8, 130.3, 130.5, 132.3, 145.7, 148.6, 169.2, 196.0; ESI-TOF MS: calcd. for C<sub>19</sub>H<sub>14</sub>ClN<sub>3</sub>NaO<sub>2</sub>, *m/z* 374.0667 (M+Na)<sup>+</sup>, found 374.0658.
- 15

#### 9-Benzotriazol-1-yl-8-bromo-5-methoxy-2,3,4,9-tetrahydro-

- **xanthen-1-one (9n).** White solid; UV (EtOH)  $\lambda_{\text{max}}$  (log  $\varepsilon$ ) 293 (3.96), 264 (4.26), 207 (4.61) ; IR (UATR)  $v_{\text{max}}$ : 2962, 2940, 2838, 1659, 1609, 1578, 1476, 1443, 1386, 1332, 1312, 1278, 20 1263, 1231, 1205, 1187, 1158, 1149, 1139, 1125, 1101, 1092,
- 1064, 1002, 941, 917, 873, 818, 802, 776, 764, 735, 743, 702, 657 cm<sup>-1</sup>; <sup>1</sup>NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.95-2.10 (m, 2H), 2.36-2.40 (m, 2H), 2.80-2.97 (m, 2H), 3.96 (s, 3H), 6.85 (d, *J* = 8.8 Hz, 1H), 7.11 (s, 1H), 7.28 (d, *J* = 8.9 Hz, 1H), 7.32 (td, *J*<sub>t</sub> = 8.1 Hz,
- <sup>25</sup>  $J_d$  = 0.7 Hz, 1H), 7.55 (td, J = 7.9 Hz,  $J_d$  = 0.7 Hz, 1H), 7.95 (d, J = 8.4 Hz, 1H), 8.15 (d, J = 8.4 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ: 20.0, 27.6, 36.4, 48.1, 56.4, 111.2, 111.7, 112.9, 114.1, 119.3, 120.4, 123.6, 127.2, 128.4, 133.5, 142.2, 144.8, 147.7, 168.8, 196.2; ESI-TOF MS: calcd. for C<sub>20</sub>H<sub>16</sub>BrN<sub>3</sub>NaO<sub>3</sub>, *m/z* <sup>30</sup> 448.0267 (M+Na)<sup>+</sup>, found 448.0267.

#### $\hbox{5-(3,3-Dimethyl-1-oxo-2,3,4,9-tetrahydro-1} H-xan then -9-yl)-$

**1,3-dimethyl-pyrimidine-2,4,6-trione** (90). White solid; UV (EtOH)  $\lambda_{\text{max}}$  (log  $\varepsilon$ ) 267 (3.30), 222 (3.38), 204 (3.50); IR <sup>35</sup> (UATR)  $v_{\text{max}}$ : 3749, 2957, 2886, 1746, 1675, 1643, 1582, 1457, 1421, 1387, 1319, 1289, 1230, 1185, 1114, 1034, 775, 756 cm<sup>-1</sup>; <sup>1</sup>NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.13 (s, 3H), 1.18 (s, 3H), 2.30 (d, *J* = 17.3 Hz, 1H), 2.36 (d, *J* = 16.3 Hz, 1H), 2.48 (d, *J* = 17.7 Hz, 1H), 2.56 (d, *J* = 17.6 Hz, 1H), 3.07 (s, 3H), 3.21 (s, 3H), 3.85 (d, <sup>40</sup> *J* = 2.7 Hz, 1H), 4.87 (s, 1H), 7.03 (d, *J* = 8.22 Hz, 1H) 7.08-7.09 (m, 2H), 7.22-7.29 (m, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$ : 27.2,

(iii, 2H), 7.22-7.29 (iii, 1H); C NMR (75 MHz, CDC1<sub>3</sub>) 6, 27.2, 28.1, 28.2, 29.2, 32.0, 36.3, 41.4, 50.5, 54.9, 108.8, 116.6, 120.5, 124.9, 127.9, 129.0, 150.4, 151.1, 166.9, 167.2, 168.0, 192.7; ESI-TOF MS: calcd. for  $C_{21}H_{22}N_2NaO_5$ , m/z 405.1421 (M+Na)<sup>+</sup>, 45 found 405.1430.

# 5-(7-Chloro-3,3-dimethyl-1-oxo-2,3,4,9-tetrahydro-1*H*-xanthen-9-yl)-1,3-dimethyl-pyrimidine-2,4,6-trione (9p).

- White solid; UV (EtOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 265 (4.24), 225 (4.31), 206 <sup>50</sup> (4.42); IR (UATR)  $\nu_{max}$ : 2959, 2876, 1734, 1677, 1645, 1449, 1420, 1382, 1287 1234, 1187, 1119, 1033, 827, 757 cm<sup>-1</sup>; <sup>1</sup>NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.12 (s, 3H), 1.14 (s, 3H), 2.28 (d, *J* = 15.9 Hz, 1H), 2.35 (d, *J* = 16.1 Hz, 1H), 2.47 (d, *J* = 18.5 Hz, 1H), 2.54 (d, *J* = 17.8 Hz, 1H), 3.16 (s, 3H), 3.24 (s, 3H), 3.85 (d, *J* =
- <sup>55</sup> 2.5 Hz, 1H), 4.84 (s, 1H), 6.98 (d, J = 8.7 Hz, 1H), 7.16 (d, J = 2.3 Hz, 1H), 7.21 (dd,  $J_a = 8.7$  Hz,  $J_b = 2.4$  Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ: 27.1, 28.3, 28.4, 29.3, 32.0, 35.5, 41.4, 50.5, 55.1, 108.5, 118.0, 122.9, 127.9, 129.0, 130.0, 148.9, 151.1,

166.8, 167.0, 167.8, 197.4; ESI-TOF MS: calcd. for  $_{60}$  C<sub>21</sub>H<sub>21</sub>ClN<sub>2</sub>NaO<sub>5</sub>, *m*/*z* 439.1031 (M+Na)<sup>+</sup>, found 439.1036.

#### Single crystal X-ray analysis

X-ray diffraction data for chromenes **7a**, **7b**, **7e**, and **7f** were collected at 296(2) K on a Bruker X8 APEX II KAPPA CCD <sup>65</sup> diffractometer using graphite monochromatized Mo-K $\alpha$  radiation ( $\lambda = 0.71073$  Å). The structures were solved using SHELXS-97 and refined using full-matrix least squares on  $F^2$  with SHELXL-97.<sup>47</sup> Final *R*-values and selected refinement details of **7a**, **7b**, **7e**, and **7f** are given in Table 1S (ESI). Data of **7a**, **7b**, **7e**, and **7f** are cCDC numbers for **7a**, **7b**, **7e**, and **7f** are CCDC numbers for **7a**, **7b**, **7e**, and **7f** are CCDC 952576, CCDC 952577, CCDC 952578, and CCDC 952579. Copies of these data can be obtained, free of charge, *via* www.ccdc.cam.ac.uk/conts/retrieving.html (or 12 Union Road, <sup>75</sup> Cambridge CB2 1EZ, UK, fax: +44 1223 336033, e-mail: deposit@ccdc.cam.ac.uk).

#### Acknowledgments

P. K. is supported by The Thailand Research Fund and the Center of Excellence on Environmental Health and Toxicology,
Science & Technology Postgraduate Education and Research Development Office (PERDO), Ministry of Education. T. A. acknowledges financial support by the National Research University Project (FW657B) from the Office of the Higher Education Commission, Ministry of Education. R. S. thanks the
<sup>85</sup> Ministry of Science and Technology for a student grant.

#### Notes and references

- E. Schweizer and D. Meeder-Nycz, Chemistry of Heterocyclic Compounds: Chromenes, Chromanones, and Chromones, Volume 31, 1977, 11-139.
- A. M. Holbrook, J. A. Pereira, R. Labiris, H. McDonald, J. D. Douketis, M. Crowther and P. S. Wells, *Arch. Intern. Med.*, 2005, 165, 1095.
- M. E. Riveiro, A. Moglioni, R. Vazquez, N. Gomez, G. Facorro, L.
   Piehl, E. R. de Celis, C. Shayo and C. Davio, *Bioorg. Med. Chem.*, 2008, 16, 2665-2675.
  - A. Chilin, R. Battistutta, A. Bortolato, G. Cozza, S. Zanatta, G. Poletto, M. Mazzorana, G. Zagotto, E. Uriarte, A. Guiotto, L. A. Pinna, F. Meggio and S. Moro, *J. Med. Chem.*, 2008, **51**, 752-759.
- 100 5. M. Campos-Toimil, F. Orallo, L. Santana and E. Uriarte, *Bioorg. Med. Chem.*, 2002, **12**, 783-786.
  - D. A. Ostrov, J. A. Hernández Prada, P. E. Corsino, K. A. Finton, N. Le and T. C. Rowe, *Antimicrob. Agents Chemother.*, 2007, 51, 3688-3698.
- <sup>105</sup> 7. N. A. Gormley, G. Orphanides, A. Meyer, P. M. Cullis and A. Maxwell, *Biochemistry*, 1996, **35**, 5083-5092.
  - C. A. Kontogiorgis, K. Savvoglou and D. J. Hadjipavlou-Litina, J. Enzyme Inhib. Med. Chem., 2006, 21, 21-29.
- G. Melagraki, A. Afantitis, O. Igglessi-Markopoulou, A. Detsi, M.
   Koufaki, C. Kontogiorgis and D. J. Hadjipavlou-Litina, *Eur. J. Med. Chem.*, 2009, 44, 3020-3026.
  - R. W. Fuller, H. R. Bokesch, K. R. Gustafson, T. C. McKee, J. H. Cardellina, J. B. McMahon, G. M. Cragg, D. D. Soejarto and M. R. Boyd, *Bioorg. Med. Chem. Lett.*, 1994, 4, 1961-1964.

75

- D. Viña, M. J. Matos, M. Yáñez, L. Santana and E. Uriarte, *Med. Chem. Comm.*, 2012, 3, 213-218.
- S. Vazquez-Rodriguez, R. Figueroa-Guíñez, M. J. Matos, L. Santana, E. Uriarte, M. Lapier, J. D. Maya and C. Olea-Azar, *Med. Chem. Comm.*, 2013, 4, 993-1000.
- 13. S. R. Trenor, A. R. Shultz, B. J. Love and T. E. Long, *Chem. Rev.*, 2004, **104**, 3059-3078.
- W. Kemnitzer, J. Drewe, S. Jiang, H. Zhang, C. Crogan-Grundy, D. Labreque, M. Bubenick, G. Attardo, R. Denis, S. Lamothe, H.
- 10 Gourdeau, B. Tseng, S. Kasibhatla and S. X. Cai, *J. Med. Chem.*, 2008, **51**, 417-423.
- W. Kemnitzer, S. Jiang, Y. Wang, S. Kasibhatla, C. Crogan-Grundy, M. Bubenik, D. Labrecque, R. Denis, S. Lamothe, G. Attardo, H. Gourdeau, B. Tseng, J. Drewe and S. X. Cai, *Bioorg. Med. Chem. Lett.*, 2008, 18, 603-607.
- D. Baiz, T. A. Pinder, S. Hassan, Y. Karpova, F. Salsbury, M. E. Welker and G. Kulik, *J. Med. Chem.*, 2012, 55, 8038-8046.
- K. S. Shih, J. H. Wang, Y. W. Wu, C. M. Teng, C. C. Chen and C. R. Yang, *PloS one*, 2012, 7, e42389.
- 20 18. S. A. Patil, J. Wang, X. S. Li, J. Chen, T. S. Jones, A. Hosni-Ahmed, R. Patil, W. L. Seibel, W. Li and D. D. Miller, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 4458-4461.
- K. Cattopadhyay, A. Recio, 3rd and J. A. Tunge, Org. Biomol. Chem., 2012, 10, 6826-6829.
- 25 20. Y. Jiang, W. Chen and W. Lu, RSC Adv., 2012, 2, 1540-1546.
- Y. J. Jang, S. E. Syu, Y. J. Chen, M. C. Yang and W. Lin, Org. Biomol. Chem., 2012, 10, 843-847.
- L. Z. Dai, Y. L. Shi, G. L. Zhao and M. Shi, *Chem. Eur. J.*, 2007, 13, 3701-3706.
- 30 23. J. Fan and Z. Wang, Chem. Commun., 2008, 5381-5383.
- 24. C. C. Malakar, D. Schmidt, J. Conrad and U. Beifuss, *Org. Lett.*, 2011, **13**, 1972-1975.
- J. Mondal, A. Modak, M. Nandi, H. Uyama and A. Bhaumik, *RSC Adv.*, 2012, 2, 11306-11317.
- 35 26. K. Kumaravel and G. Vasuki, *Green Chem.*, 2009, **11**, 1945-1947.
- C.-H. Wang, Z. Guan and Y.-H. He, *Green Chem.*, 2011, 13, 2048-2054.
- B. List, R. A. Lerner and C. F. Barbas, J. Am. Chem. Soc., 2000, 122, 2395-2396.
- 40 29. K. A. Ahrendt, C. J. Borths and D. W. C. MacMillan, J. Am. Chem. Soc., 2000, **122**, 4243-4244.
  - 30. D. W. C. MacMillan, *Nature*, 2008, **455**, 304-308.
- 31. L. W. Xu and Y. Lu, Org. Biomol. Chem., 2008, 6, 2047-2053.
- 32. M. Philippe, B. Didillon and L. Gilbert, *Green Chem.*, 2012, **14**, 952-956.
- T. Saito, R. H. Brown, M. A. Hunt, D. L. Pickel, J. M. Pickel, J. M. Messman, F. S. Baker, M. Keller and A. K. Naskar, *Green Chem.*, 2012, 14, 3295-3303.
- R. Malacea, C. Fischmeister, C. Bruneau, J. L. Dubois, J. L.
  Couturier and P. H. Dixneuf, *Green Chem.*, 2009, 11, 152-155.
- U. Biermann, U. Bornscheuer, M. A. Meier, J. O. Metzger and H. J. Schafer, *Angew. Chem. Int. Ed. Engl.*, 2011, **50**, 3854-3871.
- 36. M. Firdaus and M. A. R. Meier, Green Chem., 2013, 15, 370-380.
- 37. S. Jimenez, M. Harsch and J. Rosenbloom, *Biochem. Biophys. Res.* 5 *Commun.*, 1973, **52**, 106-114.
- 38. C. S. Ku, M. Sathishkumar and S. P. Mun, Chemosphere, 2007,

**67**, 1618-1627.

- M. J. Barnes, B. J. Constable, L. F. Morton and P. M. Royce, Biochem. J., 1974, 139, 461-468.
- 60 40. D. Font, C. Jimeno and M. A. Pericas, Org. Lett., 2006, 8, 4653-4655.
  - 41. M. Li, B. Zhang and Y. Gu, Green Chem., 2012, 14, 2421-2428.
  - 42. P. P. Ghosh and A. R. Das, J. Org. Chem., 2013, 78, 6170-6181.
  - 43. S. Gao, C. H. Tsai and C. F. Yao, *Synlett*, 2009, 949-954.
- 65 44. N. C. Ganguly, S. Roy, P. Mondal and R. Saha, *Tetrahedron Lett.*, 2012, **53**, 7067-7071.
  - 45. M. Li and Y. Gu, Adv. Synth. Catal., 2012, 354, 2484-2494.
  - N. Sato, M. Jitsuoka, T. Shibata, T. Hirohashi, K. Nonoshita, M. Moriya, Y. Haga, A. Sakuraba, M. Ando, T. Ohe, H. Iwaasa, A. Gomori, A. Ishihara, A. Kanatani and T. Fukami, *J. Med. Chem.*, 2008, **51**, 4765-4770.
  - 47. G. M. Sheldrick, Acta Cryst., 2008, A64, 112-122.