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3 The blue line: luminol-H<sub>2</sub>O<sub>2</sub>-Cu NCs. Luminol:  $5 \times 10^{-5}$  M in pH 11.8 (sodium

4 hydroxide solution);  $H_2O_2$ : 0.15 M; Cu NCs: 12.8 mg L<sup>-1</sup>.

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Figure 2. Effects of the reaction conditions on the luminol-H2O2-Cu NCs CL system. (A) Effects of pH :Luminol:  $1 \times 10^{-5}$  M; H<sub>2</sub>O<sub>2</sub>: 0.15 M; Cu NCs: 12.8 mg L<sup>-1</sup> (B) Effect of luminol concentration: pH: 11.8; H<sub>2</sub>O<sub>2</sub>: 0.15 M; Cu NCs: 12.8 mg L<sup>-1</sup> (C) Effect of H<sub>2</sub>O<sub>2</sub> concentration : PH:11.8; Luminol:  $5 \times 10^{-5}$  M; Cu NCs: 12.8mg L<sup>-1</sup> (D) Effect of flow rate: Luminol:  $5 \times 10^{-5}$  M; H<sub>2</sub>O<sub>2</sub>: 0.15 M; pH: 11.8. (E)Effect of Cu NCs: Luminol:  $5 \times 10^{-5}$  M; H<sub>2</sub>O<sub>2</sub>: 0.15 M; pH: 11.8.





19 Figure 3. Chemiluminescence spectra for luminol- $H_2O_2$ -Au NCs system. Luminol: 5 ×

20  $10^{-5}$  M in pH 11.8 (sodium hydroxide solution); H<sub>2</sub>O<sub>2</sub>: 0.15 M; Cu NCs: 12.8 mg

 $21 L^{-1}$ .





Figure 4. UV-visible absorption spectra of (a) Cu NCs; (b) luminol-H<sub>2</sub>O<sub>2</sub>-Cu NCs; (c)



25 luminol-H<sub>2</sub>O<sub>2</sub>- H<sub>2</sub>O.



Figure 5. Standard calibration curve for H<sub>2</sub>O<sub>2</sub> assay.



**Figure 6.** Selectivity for  $H_2O_2$  assay against other common cations.  $H_2O_2$ : 1 mM;

- 31 The concentration of each cations was 1M.
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38 Scheme 2. Possible mechanism for the luminol-H<sub>2</sub>O<sub>2</sub>-Cu NCs CL system.

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1	Luminol chemiluminescence enhanced by copper nanoclusters and its analytical
2	application
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23 Abstract

25	It was found that Cu nanoclusters could enhance the chemiluminescence (CL)
26	emission from the luminol-hydrogen peroxide system in an alkaline medium. Herein, the
27	CL spectra, UV-visible spectroscopy and radical scavengers were conducted to explore
28	the possible enhancement mechanism. The enhanced CL should attribute to the catalysis
29	of Cu nanocluster, which effectively catalyzed the decomposition of $\mathrm{H_2O_2}$ to produce
30	double hydroxyl radical. The inhibiting effects of some organic compounds were also
31	investigated. Then, the proposed method has been successfully applied to determine $H_2O_2$
32	in enviroment water samples with satisfactory accuracy and precision.
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34	Keywords
35	Cu nanoclusters; chemiluminescence; luminol; hydrogen peroxide;
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### 45 Introduction

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In recent years, chemiluminescence and related analytical techniques have attracted extensive interest and have been developed as important and powerful tools in different fields,<sup>1-9</sup>because of its inherent strengths: high sensitivity, a wide linear range, simple instrumentation, and, in many cases, lack of background scattering light interference.

However, resulting from the weak CL emission of traditional CL system, people 51 centered their interest on some new material for the purpose of enhancement of the CL 52 intensity. Catalysts, such as transition metal ions and peroxidases, have been applied for 53 that purpose.<sup>10-13</sup> Lately, much attention has been paid to the chemiluminescence of 54 nanomaterials system, providing amplified CL emission. Many researches have 55 56 demonstrated that use of nanoparticles in CL reactions has proposed new methods to enhance the inherent sensitivity and expand new applications in detection. For example, 57 Cui and co-workers have reported many prominent works about noble metal 58 nanoparticles-catalyzed CL systems, such as Au, Ag, and Pt nanoparticles, which 59 significantly enhanced many traditional CL systems.<sup>14-16</sup> Yu et al. have decorated Pt-Co 60 bimetallic alloy nanoparticles on graphene to catalyze luminol CL system for sensing 61 glucose.<sup>17</sup> In other situations, metal oxide nanoparticles, such as Fe<sub>2</sub>O<sub>3</sub>, ZnO, Co<sub>2</sub>O<sub>3</sub>, 62 CoFe<sub>2</sub>O<sub>4</sub>, CeO<sub>2</sub>, ZnS and CuO, have also used in the CL reaction.<sup>18-25</sup> However, the 63 application of metal nanocluster as catalysts for the CL system has not yet been reported, to 64 the best of our knowledge. 65



Metal nanoclusters (NCs) consisting of several to tens atoms have recently attracted

67	much attention. <sup>26-28</sup> Because their unique physical, electrical, and optical properties have
68	made metal NCs as promising candidates in the fields of catalysis, chemical sensors,
69	electronic devices, and biological imaging. <sup>29-32</sup> Until now, the application of metal NCs in
70	analytical fields is mainly focus on their fluorescence properties and very little on their
71	catalytic properties for biological or chemical sensing application. <sup>33-35</sup> Therefore, it is very
72	meaningful to investigate novel sensing platforms based on their catalytic activitys of metal
73	NCs.

74 In this paper, we report the catalytic property of copper (Cu) NCs in luminol CL system for the first time. Compared with the noble metals Au and Ag, the metal Cu is relatively 75 76 abundant, inexpensive, and readily available from commercial sources. It was found that 77 Cu NCs could enhance greatly CL from Luminol-H<sub>2</sub>O<sub>2</sub> system. A possible enhancement 78 mechanism of Cu NCs on luminol CL was exploited. The effect of Cu NCs on the luminol-79 H<sub>2</sub>O<sub>2</sub>-Cu NCs CL system was studied. Experimental results suggested that some organic compounds containing -OH, -NH<sub>2</sub>, -SH groups could inhibit the CL signal of luminol-80 H<sub>2</sub>O<sub>2</sub>-Cu NCs system. It indicated that the proposed system had great potential for the 81 82 determination of such compounds. Meanwhile, the feasibility of the present method for 83 H<sub>2</sub>O<sub>2</sub> detection was also researched. Under optimum conditions, the CL intensity was linear 84 with H<sub>2</sub>O<sub>2</sub> concentration.

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- 88 Experimental

- 90 *Reagents and materials*
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All chemicals and reagents were of analytical grade and used as received without further purification, and ultrapure water was used throughout. Bovine serum albumin (BSA) was purchased from Sangon Biotech Co., Ltd. (Shanghai, China). CuSO<sub>4</sub>·5H<sub>2</sub>O was purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). 30% (v/v) H<sub>2</sub>O<sub>2</sub>, sodium hydroxide and nitro blue tetrazolium (NBT) were purchased from Kelong Reagent Co., Chengdu, China. Thiourea, ascorbic (AA) were commercially purchased from Chongqing Chemical Regent Company (Chongqing, China).

A  $1.0 \times 10^{-2}$  M stock solution of luminol (3-aminophthalhydrazide) was prepared by dissolving luminol (Sigma) in 0.1 M sodium hydroxide solution. Working solutions of luminol were prepared by diluting the stock solution. Working solutions of H<sub>2</sub>O<sub>2</sub> were prepared fresh daily by dilution of 30% (v/v) H<sub>2</sub>O<sub>2</sub>.

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### 104 Synthesis of BSA-Cu nanoclusters

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BSA modified Cu NCs were prepared in aqueous solution following a previous method<sup>36</sup>. In a typical experiment, 1mL aqueous CuSO<sub>4</sub>·5H<sub>2</sub>O solution (20 mM) was added to the BSA solution (5 mL, 15mg/mL) under vigorous stirring for 5min at room temperature. Then, The solution PH was adjusted to 12 by adding NaOH solution and the mixture was allowed to proceed under vigorous stirring at 55°C for 8 h. The solution was

- then dialyzed in ultra-pure water for 48 h to remove unreacted  $Cu^{2+}$ . The final solution was
- 112 stored at  $4^{\circ}$ C in refrigerator when not in use.
- 113
- 114 General procedure for CL analysis
- 115

The chemiluinescence detection was conducted on a laboratory-built flow injection CL 116 117 system (Xi'an Remax Company, Xi'an, China), consisting of two peristaltic pumps to 118 deliver the reactants to the flow cell. (Scheme 1) One delivered Cu NCs and  $H_2O_2$  (or 119 samples) with two channels at a flow rate (per tube) of 1.9 mL/min. The other pump was 120 used to carry luminol solution at the same flow rate. The PTFE tubing (0.8 mm i.d.) was 121 used to connect all components in the flow system. A six-way injection valve equipped 122 with an 8 cm long sampling loop was used to inject. The CL signal produced was detected 123 by a photomultiplier tube (operated at -550 V), and was then recorded by a computer 124 equipped with a data acquisition interface. Data acquisition and treatment were performed 125 with BPCL software running under Windows XP. When the CL system was used to study 126 the effect of organic compounds and the free radical scavengers, one peristaltic pump was 127 used to deliver Cu NCs and the mixture of  $H_2O_2$  and luminol, and the other was used to 128 carry organic compounds or free radical scavenger at 1.9 mL/min, respectively.

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130 Sample preparation

For hydrogen peroxide determination, the tap water samples were chosen for
investigation in this study. The water sample was filtered through a 4.5μm micropore
membrane before experiment.

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- 135 **Results and discussion**
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- 137 Enhancement of luminol CL

The effects of Cu NCs on the luminol-H<sub>2</sub>O<sub>2</sub> chemiluminescence system were studied. 138 As show in Fig. 1, the oxidation of luminol by H<sub>2</sub>O<sub>2</sub> generates weak CL in alkaline media. 139 140 However, the CL signal intensity could be enhanced significantly up to about 70 folds as 141 soon as adding the Cu NCs. Compared with other nano-catalysts reported in the literatures (Table 1), the CL enhancement factor on luminol- H2O2 CL system of Cu nanoclusters is 142 143 much higher than that of most catalysts mentioned. Though the catalytic activities of Au 144 and Pt nanoparticles are little higher than Cu nanoclusters, they are costly. Therefore, Cu 145 nanocluster could be an outstanding catalyst on the luminol- H<sub>2</sub>O<sub>2</sub> CL system.

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### 147 *Optimization of the reaction conditions*

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The reaction conditions were optimized for the luminol-H<sub>2</sub>O<sub>2</sub>-Cu nanoclusters CL system shown in Fig. 2. The pH of luminol solution is of great importantance in the CL reaction, so the effect of pH on the CL was tested in the range of pH 11.4–12.6 (Fig. 2A). The optimized PH condition for luminol-H<sub>2</sub>O<sub>2</sub>-Cu nanoclusters CL system was pH 11.8 When the pH of luminol solution was lower than 11.8, the CL intensity increased with increasing the pH. The effect of luminol concentration on the CL was investigated in the range from  $1.0 \times 10^{-6}$  to  $2.0 \times 10^{-4}$  M (Fig. 2B), the CL intensity increased with increasing

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luminol concentration in the range of  $1.0 \times 10^{-6}$  to  $5.0 \times 10^{-5}$  M. However, when the 156 concentration of luminol was above  $5.0 \times 10^{-5}$  M, only slight changes in the light intensity 157 were observed. Therefore,  $5.0 \times 10^{-5}$  M was selected as the optimal luminol concentration 158 159 in the present study. The effect of H<sub>2</sub>O<sub>2</sub> concentration on the CL was studied in the range of 0.01-0.5 M (Fig. 2C), the CL intensity increased with increasing  $H_2O_2$  concentration in the 160 161 range of 0.01-0.15 M and decreased when the concentration of  $H_2O_2$  is larger than 0.15 M. 162 The effects of the concentration of Cu NCs and the flow rate were also discussed (Fig. 2D, 163 2E). Considering the CL intensity and the consumption of the regents, the optimized conditions for the luminol- H<sub>2</sub>O<sub>2</sub>-Cu NCs system were as follows: 5.0× 10<sup>-5</sup> M luminol in 164 NaOH solution (PH=11.8), 0.15 M H<sub>2</sub>O<sub>2</sub>, 12.8 mg/L Cu nanoclusters and 1.90 mL/min 165 166 flow rate.

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### 168 Mechanism Discussion

A F-7500 mode fluorescence spectrophotometer has been used to discuss the CL 170 mechanism of luminol- H<sub>2</sub>O<sub>2</sub>-Cu nanoclusters sysyem. The CL spectra was obtained after 171 172 turning off the light entrance slot. As shown in Fig. 3, the maximal emission peak located at 173 425nm clearly, indicating that the luminophor was still the excited-sate 3-aminophthalate anions (3-APA\*).<sup>37-38</sup> Therefore, the adding of Cu nanoclusters did not result in forming a 174 175 new luminophor for this CL system. The enhanced CL signals were thus attributed to the possible catalysis from Cu nanoclusters. In order to further confirm the possible catalysis of 176 177 Cu nanocluster, the UV-visible absorption spectra was recorded. As shown in Fig. 4, the

178	maximum absorption peaks of Cu NCs and luminol- $H_2O_2$ -Cu NCs system are observed at
179	around 325 nm and 346 nm, respectively. Therefore, the light absorption of the mixed
180	system was approximately equal to the sum of the light absorption of the two individual
181	systems, which implied that no change was taken between the species after the reaction. As
182	a result, the enhancement of CL signals had derived from the catalytic effects of Cu NCs.
183	The CL-generation mechanism for luminol oxidation in aqueous solution has been
184	extensively studied. It was reported that $H_2O_2$ decomposition on supported metal catalysts
185	such as Au NPs, Ag NPs and CuO NPs involved the formation of hydroxyl radicals OH.
186	Furthermore, Xu et al has found Cu NCs could exhibit significant peroxidase-like activity. <sup>35</sup>
187	Similarly, we suggested that the O-O bond of $H_2O_2$ might be broken up into double
188	OH· radicals by virtue of the catalysis of Cu nanocluster. Then the OH· radicals reacted with
189	luminol anion and HO2 <sup>-</sup> to form luminol radical (L· <sup>-</sup> ) and superoxide radical anion O2 <sup>-,</sup>
190	which further reacted with each other to form the excited 3-aminophthalate anion (3-APA*).

To acquire further insight into the mechanism of the CL system, the effects of various active oxygen radical scavengers on the CL were studied. (Table 2) AA is well known as an efficient ROS scavenger, and it can terminate active oxygen radicals by electron transfer. The influence of AA on the CL signal was investigated, and the results showed it could quench the CL even at a relatively low concentration. Therefore, we confirmed that the CL reaction must happen in a radical way, in which the generation of free radicals appeared to be the key factors.

For purpose of identifying the generation of  $O_2$ .<sup>-</sup> and OH· in the CL reaction, NBT was frequently used for the detection of  $O_2$ .<sup>-</sup> radicals.  $O_2$ .<sup>-</sup> can reduce NBT to its deep blue

200	diformazan form. The color changed from yellow to blue when 1mM mol/L NBT was added
201	to the CL system, and then the CL intensity decreased by a factor of $\sim$ 52.3. The result
202	confirmed that $O_2 \cdot $ was involved in the CL process. OH $\cdot$ is always supposed to be one of the
203	most potent oxidizers among the oxgen-centered free radicals. Thiourea is an effective
204	radical scavenger for OH. When 1.0 mM thiourea is added to CL system, a distinct
205	inhibition is observed by a factor of $\sim 61$ . It indicated that OH $\cdot$ is generated in the CL
206	process.
207	Based on the above results, the whole enhanced mechanism is summarized in Scheme 2.

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### 209 Inhibition effects of organic compounds

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211 Some organic compounds containing hydroxyl (OH), amino (NH<sub>2</sub>), or mercapto (SH) 212 groups were found to inhibit the CL from the luminol-H2O2 system-Au NPs/Ag NPs system. It also has been reported that the reducing groups of OH, NH<sub>2</sub>, or SH are possible 213 to compete with luminol for active oxygen intermediates, giving rise to a decrease in CL 214 intensity. 14,39 Moreover, such compounds may interact with Cu NCs to interrupt the 215 216 formation of luminol radicals and hydroxyl radicals taking place on the surface of 217 nanoclusters, causing a decrease in the CL intensity. Therefore, the effects of such organic compounds on the luminol-H2O2 system-Cu NCs were studied as list in Table 3. As 218 expected, for 10<sup>-4</sup> M tested compounds, the CL signals were obviously inhibited. In 219 220 addition, the inhibition percentage varied with the spcies and concentration of the 221 compounds. The results demonstrate that the luminol-H<sub>2</sub>O<sub>2</sub> system-Cu NCs system has the

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potential to respond such compounds. Nevertheless, low selectivity does be the main

223	disadvantage of the CL detection, but this weakness can be overcome by implementation of
224	a separation unit. As a result, it is perfect to design a CL detector in HPLC and
225	high-performance capillary electrophoresis for the simultaneous determination of numerous
226	compounds.
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228	Analytical performance
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230	Hydrogen peroxide is of vital importance for medical diagnosis, because it is involved
231	in many detection processes as an intermediate product. The possibility of using the
232	proposed method to detect hydrogen peroxide is studied (Fig. 5). Under the optimum
233	conditions described above, the linear calibration range prolonged over 3 orders of
234	magnitude from 0.1 mM to 150 mM. The regression equation is $\Delta I=54.39 + 30117.8[H_2O_2]$
235	(mol/L), r=0.9984 (n=9). The limit of detection (LOD, $3\sigma$ ) for hydrogen peroxide was 0.03
236	mM. The relative standard deviation (RSD) was 3.1% for 60 mM $H_2O_2$ (n=7).
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238	Interference study

The selectivity of the proposed method was evaluated by analyzing a standard solution of 1.0 mM  $H_2O_2$ , to which varying amounts of possible interference were added. With respect to 1.0 mM  $H_2O_2$ , the tolerable limit of each exotic species was considered as a relative error less than the 5% level. As shown in Fig. 6, most of the ions had no essential effect on the detection of 1.0 mM  $H_2O_2$ . Though Fe<sup>3+</sup> is the main interference for determination, the

244	interference could be eliminated for adding the EDTA. The experimental result suggested
245	that the addition of EDTA could realise the quantitative recovery of $H_2O_2$ from the water
246	samples as compared to that without EDTA. Therefore, the results indicated that the
247	proposed CL system is highly selective for hydrogen peroxide
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249	Analytical applications
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251	The CL method based on Cu nanocluster catalysis was applied to the determination of
252	$\mathrm{H_2O_2}$ in tap water. From Table 4, it can be seen that the recovery of $\mathrm{H_2O_2}$ in tap water
253	sample ranged from 85.0 to 110.0% through standard addition experiments, which
254	demonstrated the proposed CL system was satisfactory for H <sub>2</sub> O <sub>2</sub> analysis. Meanwhile, as
255	shown in Table 5, the concentration of the $H_2O_2$ was in excellent agreement with that
256	obtained by spectrophotometric method.
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258	Conclusion
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260	In summary, Cu NCs were found to enhance greatly the Luminol- $H_2O_2$ CL signals. The
261	enhancement of CL was suggested to attribute to the catalysis of Cu NCs on the radical
262	generation and electron-transfer processes during the luminol CL reaction. Some organic

inhibit the CL signals of the luminol-H2O2-Cu NCs system under the optimized 264 265 experimental conditions, which could be potentially used to detect these compounds.

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compounds containing OH, NH2, or SH groups interacting with Cu NCs were found to

266	Moreover, the proposed method was successfully applied for $H_2O_2$ detection in water
267	sample. This work was of great importance for the investagation of new and efficient
268	catalysts for CL system and helpful for understanding of CL mechanism correspondingly.
269	Acknowledgement
270	We thank Prof. H. Z. Zheng and Prof. Y. M. Huang for measurements.
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	on luminol-H <sub>2</sub> O <sub>2</sub> CL system	
Nano-catalyst	Enhancement factor	Literature
Ag NPs	3-10	15
Au-Ag alloy NPs	5	40
ZnO NPs	18	19
CeO <sub>2</sub> NPs	22.5	22
Co Fe <sub>2</sub> O <sub>4</sub> MNPs	50	21
Fe <sub>2</sub> O <sub>3</sub> NPs	18	18
Au NPs	100	14
Pt NPs	120	16
Cu NCs	70	This worl

357

358

**Table 2.** Effect of different radical scawenger on the CL of Luminol-H2O2 in the presence

of Cu nanocluster<sup>a</sup>

360

scavengers	Intermediates	Concentration	Percent inhibition(%) <sup>b</sup>
H <sub>2</sub> O			0
Ascorbic acid	OH∙, O <sub>2</sub> .−	0.1mM	70.1
NBT	O <sub>2</sub>	1mM	52.3
Thiourea	OH·	1mM	61.0

<sup>a</sup>Solution condition: Luminol,  $5 \times 10^{-5}$  M in pH 11.8 (sodium hydroxide solution); H<sub>2</sub>O<sub>2</sub>,

362 0.15 M; Cu NCs, 12.8 mg  $L^{-1}$ 

363 <sup>b</sup>Average value of three determination

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Omera			O(0/)
Organic compo	unus Quenching (%)	Urganic compounds	Quenching (%)
Ascorbic acid	70.1 25.4	L - alanine	33.7 24 7
L- leucine Resorcinol	23.4	L - phonylaiannie	24.7
I – aspartate	43.0	L - gryenie L - histidine	16.2
L - tryptophan	17.4	L - valine	43.7
hydroquinone	84.6	Butylated hvdroxytoluene	57.2
L – glutamic ac	id 18.3	L - cysteine	42.6
L - serine	39.7	L-Proline	42.9
Tal	ble 4 Analytical results	of H <sub>2</sub> O <sub>2</sub> in tap water (n	=3)
lan water	Detected Adde	d (mM) Found (	mM) Recov
Tap water	Detected Adde	ed ( mM ) Found (	mM) Recov
Sample1	ND <sup>a</sup>	ed ( mM ) Found ( 0.20 0.1	mM) Recov 7 85
Sample1	ND <sup>a</sup> ND <sup>a</sup>	ed (mM) Found ( 0.20 0.1 10 11	mM) Recov 7 85
Sample1 Sample2 Sample3	Detected Adde	ed (mM) Found ( 0.20 0.1 10 11 150 14	mM) Recov 7 85 110 8 98
Sample1 Sample2 Sample3 ND (not detected)	Detected Adde ND <sup>a</sup> ND <sup>a</sup> ND <sup>a</sup>	ed (mM) Found ( 0.20 0.1 10 11 150 14	mM) Recov
Sample1 Sample2 Sample3 ND (not detected)	Detected Adde ND <sup>a</sup> ND <sup>a</sup> ND <sup>a</sup>	rd (mM) Found ( 0.20 0.1 10 11 150 14 rde (n=3)	mM) Recov
Sample1 Sample2 Sample3 ND (not detected) Tabl	Detected Adde ND <sup>a</sup> ND <sup>a</sup> ND <sup>a</sup> <b>e 5</b> Determination of H	ed (mM) Found ( 0.20 0.1 10 11 150 14 $\frac{1}{2}O_2$ in tap water (n=3)	mM) Recov
ap water Sample1 Sample2 Sample3 ND (not detected) Tabl Tap water	Detected Adde ND <sup>a</sup> ND <sup>a</sup> ND <sup>a</sup> e 5 Determination of H Proposed method	ed (mM)       Found ( $0.20$ $0.1$ $10$ $11$ $150$ $143$ $2O_2$ in tap water (n=3)         Spectrophotome	mM) Recov
ap water Sample1 Sample2 Sample3 ND (not detected) Tabl Tap water	Detected Adde $ND^{a}$ $ND^{a}$ $ND^{a}$ e 5 Determination of H Proposed method $H_{2}O_{2} (mM)$	ed (mM) Found ( 0.20 0.1 10 11 150 14 $2O_2$ in tap water (n=3) Spectrophotomo H <sub>2</sub> O <sub>2</sub> (	mM ) Recov 7 85 110 8 98 etric method mM )
Sample1 Sample2 Sample3 ND (not detected) Tabl Tap water Sample1	Detected Adde $ND^{a}$ $ND^{a}$ $ND^{a}$ e 5 Determination of H Proposed method $H_2O_2 (mM)$ 2. 0±0.1	rd (mM) Found ( 0.20 0.1 10 11 150 14 150 14 150 14 1202 1202 in tap water (n=3) Spectrophotomo $H_2O_2($ $1.9\pm 0.$	mM ) Recov 7 85 110 8 98 etric method mM ) 1
Sample1 Sample2 Sample3 ND (not detected) Tabl Tap water Sample1 Sample2	Detected Adde $ND^{a}$ $ND^{a}$ $ND^{a}$ e 5 Determination of H Proposed method $H_{2}O_{2} (mM)$ 2. 0±0.1 10.0±0.2	ed (mM) Found ( 0.20 0.1 10 11 150 14 $2O_2$ in tap water (n=3) Spectrophotomo H <sub>2</sub> O <sub>2</sub> ( $1.9\pm 0.$ $10.3\pm 0$	mM ) Recov 7 85 110 8 98 etric method mM ) 1 0.1