RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This Accepted Manuscript will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/advances

Spectral properties of 4-(4-hydroxy-1-naphthylazo)benzenesulfonic acid and its application for colorimetric determination of trace Fe³⁺

Lei Hu, Li Nie, Guangnian Xu, Han Shi, Xiaoqing Xu, Xiangzhong Zhang, Zhengquan Yan*

Anhui Provincial Laboratory of Biomimetic Sensor and Detecting Technology & Solar Photovoltaic Materials Research Center, West Anhui University, Lu'an 237012, China

Tel: +86-0564-3305801

E-mail: yanzhq2008@163.com

A multifunctional dye, 4-(4-hydroxy-1-naphthylazo)benzenesulfonic acid (HNABA) was identified and synthesized. The dye, combining hydroxyl group, azo group and carboxyl group, possessed excellent optical absorption properties changing with pH, solvents and coexisted metal ions. Especially, its spectral property kept tremendously stable under acid or neural conditions and was effectively applied for colorimetric determination of Fe³⁺ from brick-red to light red in aqueous under the physiological pH condition (pH 7.0). Under the optimum conditions, the detection possessed a linear range of $9.5 \sim 400 \times 10^{-8}$ mol·L⁻¹ with a correlation coefficient (R) of 0.9938 and a limit detection (3σ , n=20) of 4.2×10^{-9} mol·L⁻¹. The relative standard deviation (R.S.D.) was lower than 3.5% (n=5). The proposed method was successfully to determinate trace Fe³⁺ in three real environmental water samples. The action mechanism between HNABA and Fe³⁺ ion was discussed in detail.



Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

ARTICLE TYPE

Spectral properties of 4-(4-hydroxy-1-naphthylazo)benzenesulfonic acid and its application for colorimetric determination of trace Fe³⁺

Lei Hu, Li Nie, Guangnian Xu, Han Shi, Xiaoqing Xu, Xiangzhong Zhang, Zhengquan Yan^{*}

Received (in XXX, XXX) XthXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX 5 DOI: 10.1039/b000000x

A multifunctional dye, 4-(4-hydroxy-1-naphthylazo)benzenesulfonic acid (**HNABA**) was identified and synthesized. The dye, combining hydroxyl group, azo group and carboxyl group, possessed excellent optical absorption properties changing with pH, solvents and coexisted metal ions. Especially, its spectral property kept tremendously stable under acid or neural conditions and was effectively applied for colorimetric determination of Fe³⁺ from brick-red to light red in aqueous under the physiological pH condition (pH 7.0). Under the optimum conditions, the detection possessed a linear range of

9.5~400×10⁻⁸ mol·L⁻¹ with a correlation coefficient (R) of 0.9938 and a limit detection (3σ , n=20) of 4.2 ×10⁻⁹ mol·L⁻¹. The relative standard deviation (R.S.D.) was lower than 3.5% (n=5). The proposed method was successfully to determinate trace Fe³⁺ in three real environmental water samples. The action ¹⁵ mechanism between **HNABA** and Fe³⁺ ion was discussed in detail.

1. Introduction

To detect metal ions has continually attracted tremendous attention in recent years due to its importance in environment, biology and chemistry domains. Iron, the third most abundant

- ²⁰ element in the earth's crust, is not only a very important macroelement in the environment, but also an essential mineral nutrient for human health.^{1, 2} People who lack of iron will be suffering from iron deficiency anemia and other serious impact on human health, while excess iron would cause an increased risk of cancer,
- ²⁵ heart disease and other illnesses such as haemochromatosis.³⁻⁸ Therefore, there is an urgent need to develop chemical sensors that are capable of detecting and monitoring iron levels in environmental samples. Considerable efforts have been made to develop various detection methods, such as fluorescent ³⁰ spectrometry,⁹⁻¹⁴ electro-chromic techniques^{15, 16} flow injection spectro-photometry,^{17, 18} chromatography,^{19, 20} mass spectrometry,²¹ nuclear magnetic resonance,²² and inductively coupled plasma mass spectrometry.²³ However, these
- instrumentally intensive methods often require utilizing 35 sophisticated instrumentation or complicated pretreatment procedures, and are not suitable for on-line or in-field monitoring. By virtue of its simplicity, rapidity, non-destructive characteristics and the distinguishable ability by naked eyes instead of the complex instruments especially,²⁴⁻²⁹ colorimetric
- ⁴⁰ sensors for Fe³⁺ have attracted considerable attention in recent years.³⁰⁻³³ Typically, Yun, *et al.*³¹ presented an easy naked-eye detection method of Fe³⁺ with a detection limit of 0.024 μ g·mL⁻¹, based on 1-nitroso-2-naphthol, an excellent color-forming chelating agent. Adebayo, *et al.*³⁴ found a novel 8-
- ⁴⁵ hydroxyquinoline-based colorimetric sensor for the simple and rapid determination of Fe³⁺ using the reaction of Fe³⁺ with the sensor to form a metaloxine complex in chloroform solution.

Wallace, *et al.*³⁰ developed a system that was able to detect low levels of Fe^{3+} using a squaraine dye to model on siderophores. All these confirm that organic colorimetric concerns are a promising

⁵⁰ these confirm that organic colorimetric sensors are a promising, easy and practical strategy for detecting Fe³⁺. However, the sensitivity of common colorimetric sensors is still lower than that of instrumentally intensive methods mentioned above. To develop novel and efficient colorimetric sensing materials ⁵⁵ remains a big challenge for a long time in the future.

Herein, to improve the sensitivity and selectivity of Fe³⁺ detection in aqueous, a multifunctional dye, 4-(4-hydroxy-1-naphthylazo)benzenesulfonic acid (**HNABA**), was identified and applied for Fe³⁺ detection practically after its optical properties ⁶⁰ were studied in detail. The multifunctional dye was expected to possess high selectivity and sensitivity to Fe³⁺ using both hydroxyquinoline group³¹ and -N=N- group^{35, 36} as chelating groups and to further increase the solubility in aqueous solution combining -SO₃H, -OH and -N=N- group. The action ⁶⁵ mechanism between **HNABSC** and Fe³⁺ was discussed by means of Job's plots and theoretical calculation.

2. Experiments

2.1 Reagents and apparatus

All the chemicals in the experiment were of AR grade and used ⁷⁰ as received from Sinopharm Chemical Reagent Co. Ltd. Water used throughout was doubly deionized.

A 1.0 mmol·L⁻¹ Fe³⁺ standard solution for testing was prepared in doubly deionized water at room temperature and diluted to appropriate concentration daily. **HNABA** was synthesized 75 according to our previous work^{37, 38} and a 5.0×10^{-3} mol·L⁻¹ **HNABA** stock solution was prepared in doubly deionized water at room temperature and stored at 4°C. Phosphate buffers (PB) were prepared by mixing a 0.01 mol·L⁻¹ H₃PO₄ solution, a 0.01 $mol \cdot L^{-1} K_2 HPO_4$ solution, a 0.01 $mol \cdot L^{-1} KH_2 PO_4$ solution or a 0.01 $mol \cdot L^{-1} KOH$ solution in a proper ratio to acquire the desired pH (pH = 3.0, 4.0, 5.0, 6.0, 7.0, 7.5, 8.0, 9.0, 10.0).

- FTIR spectra of **HNABA** with KBr disc were acquired using a ⁵ Nicolet NEXUS 870 FTIR spectrophotometer at room temperature from 4000-500 cm⁻¹. ¹H NMR spectra were recorded using a Bruker AMX-500 spectrometer operating at 400 MHz, with tetramethyl-silane (TMS) used as the reference and D₂O as solvent. Elemental analysis was conducted using an Elemental
- ¹⁰ Vario EL-III apparatus. UV-vis spectra were recorded on a Lambda 35 UV/Vis spectrometer using a 1-cm square quartz cell. pH was measured by a PHS-25pH meter.

2.2 Preparations of HNABA

- According to the literatures,^{37, 38} *p*-amino benzenesulfonic acid 15 (0.87 g, 5 mmol) was dissolved in an ice-water solution of 15% sodium nitrite (0.38 g, 5.5 mmol). After cooling to 0 °C, the solution was added to concentrated hydrochloric acid (1.2 mL) and stirred for 30 min. The excess nitrous acid was destroyed with about 5 mg urea. The mixture was then added dropwise to
- ²⁰ 10 mL buffered aqueous solution (KH₂PO₄/ Na₂HPO₄, pH=6) containing naphthol (0.73 g, 5 mmol) and stirred for another 2 h at 0~5 °C. The resultant precipitate was filtered and purified by recrystallizing three times from ethanol to provide dark red crystal **HNABA** in the yield of 92.1 %.
- ²⁵ IR (KBr), v (cm⁻¹): 3433~2500 (OH, SO₃H), 1594, 1518 (Ar), 1376 (Ar-N(R₁R)), 1169 (S-O). ¹H-NMR (D₂O, 400 Hz) δ (ppm): 8.74 (d, J=8.5 Hz, 1 H, Ar-H), 8.61 (d, J=8.4 Hz, 1 H, Ar-H), 8.40 (d, J=8.4 Hz, 1 H, Ar-H), 8.12 (d, J=7.8 Hz, 1 H, Ar-H), 8.06 (d, J=7.8 Hz, 1 H, Ar-H), 7.48 (t, J=6.4 Hz, 1 H, Ar-H), 7.51
- $_{30}$ (t, J=6.7 Hz, 1 H, Ar-H), 7.15 (t, J=6.5 Hz, 1 H, Ar-H), 5.15 (s, 1 H, -OH). Anal. Calcd for $C_{16}H_{12}N_2SO_4$: H, 3.68; C, 58.53; N, 8.51; S, 9.77. Found: H, 3.73; C, 58.49; N, 8.50; S, 9.79.

2.3 Fe³⁺ detection procedure

- For the Fe^{3+} determination, 1.0 mL PB (pH 7.0), 1.0 mL 5.0×10^{-4} ³⁵ mol·L⁻¹ of **HNABA** and 1.0 mL of the appropriate Fe³⁺ solution or sample were transferred into a 10 mL volumetric flask. The mixture was stirred thoroughly and finally diluted to 10 mL with doubly deionized water. After 20 min, the absorption spectra were measured from 200 nm to 600 nm and the band-slit was set
- ⁴⁰ as 2.0 nm. The absorption intensity difference at 478 nm was used for quantitative analysis. The decreased absorption intensity of **HNABA** was represented as $\Delta A = A_0 A$, where A_0 and A were the absorption intensities of the systems in the absence and presence of Fe³⁺, respectively.

45 3. Results and discussion

3.1 The UV-vis absorption spectrum of HNABA

It is well known that **HNABA** is a strong polar compound, whose solubility is quite low in nonpolar solvents. To illustrate the effect of solvents on the absorption spectrum of **HNABA**, the UV-vis ⁵⁰ spectra in different polar solvents, *i.e.*, N,N-dimethyl formamide (DMF), tetrahydrofuran (THF), acetone, ethanol and water were recorded as shown in **Fig. 1**.



Fig. 1 The UV–Vis spectra of HNABA in nonpolar solvents, DMF, THF, Acetone, water and Ethanol

In Fig. 1, we can find that HNABA has a strong and sharp 60 absorption peak at ca. 478 nm in polar protic water and ethanol, attributed to the whole molecular π -conjugated system, with a molar absorptivity (ε) of 2.54 ×10⁴ L·mol⁻¹ cm⁻¹ and 1.95 ×10⁴ L·mol⁻¹·cm⁻¹, respectively, meaning that **HNABA** exists in the form of monomolecule, *i.e.*, polar protic water and ethanol are 65 both good solvents for HNABA. While in polar aprotic acetone, THF and DMF, the absorption intensity at ca. 478 nm decreases and a new blue-shift absorption peak emerges at ca. 420 nm. The absorption gets wide and blue-shift might be attributed to the Haggregation of HNABA in the poor solvents as acetone, THF and 70 DMF.^{39, 40} The conclusion could be further confirmed by the fact that the absorptions at ca. 240, 266, 290, 330 nm appear clearly in polar protic water and ethanol while decrease or disappear almost in polar aprotic acetone. THF and DMF, attributing to $\pi \rightarrow \pi^*$. $n \rightarrow \pi^*$ transitions of C=C, N=N and O=C bonds in non-75 conjugated benzenesulfonic acid and naphthol moieties.



Fig. 2 Effect of pH on the UV–Vis spectra of **HNABA** (From up to down: 3.0, 4.0, 5.0, 6.0, 7.0, 7.5, 8.0, 9.0, 10.0)

For an acidic **HNABA** molecule, pH of the system will have a ⁸⁰ more significant influence on its existing formation and so make its absorption spectrum changed. Here the effect of pH on the absorption spectrum was investigated in polar protic water with a pH range of 3.0~10.0. As shown in **Fig. 2**, it is interesting to find that the absorption at 478 nm keeps quite stable as long as pH is less than 7.0. When pH is more than 8.0, the absorption intensity at 478 nm decreases gradually and red shifts. The reason may be that SO₃H and OH groups in **HNABA** molecules all change into SO₃⁻ and O⁻ ions under basic conditions, which would enlarge ⁵ the whole molecular π -conjugated system, but decrease the molecular dipole moment. The same phenomenon is also found in the absorbance at *ca.* 240, 266, 290, 330 nm as shown in Fig. 2, meaning that **HNABA** could possess the best optical absorption property under a wide pH range, *i.e.*, physiological conditions

¹⁰ (pH *ca*. 7.0) will be selected in the next experiments.

3.2 Special response to Fe³⁺

To demonstrate the selectivity of **HNABA** sensing to Fe³⁺, we had investigated its colorimetric response to some other environmentally relevant metal ions, *i.e.*, Ag⁺, Al³⁺, Ba²⁺, Cd²⁺, ¹⁵ Co³⁺, Cu²⁺, Zn²⁺, Mn²⁺, K⁺, Mg²⁺, Na⁺, Ni²⁺, Pb²⁺, Hg²⁺, Cr³⁺ and Fe²⁺ at high concentrations in aqueous solutions at pH 7.0. After the addition of 100 equiv (4.0×10⁻⁴ mol·L⁻¹) different metal ions above and Fe³⁺ (4.0×10⁻⁶ mol·L⁻¹), the relative changes in absorption intensities of the sensing system were recorded as ²⁰ shown in **Fig. 3**, respectively. It can be seen from Fig. 3 that the

- ²⁰ shown in **Fig. 5**, respectively. It can be seen from Fig. 5 that the absorption intensities of the **HNABA** sensing system in the presence of Ag⁺, Al³⁺, Ba²⁺, Cd²⁺, Co³⁺, Cu²⁺, Zn²⁺, Mn²⁺, K⁺, Mg²⁺, Na⁺, Ni²⁺, Pb²⁺, Hg²⁺, Cr³⁺ and Fe²⁺ show negligible change even with 100-fold higher concentrations than that of Fe³⁺
- ²⁵ and the alterations of ΔA are all less than 5% (detection error). The results indicate that **HNABA** possesses excellent selectivity to Fe³⁺ even in the presence of other coexisting metal ions under very high concentrations.



Fig. 3 Effect of different metal ions on the UV–Vis spectra of HNABA

To illustrate the response speed of **HNABA** to Fe³⁺ and the stability of the proposed system, the effect of incubation time on the absorption intensity was also investigated. The results show that a maximum and constant ΔA is reached after all reagents are ³⁵ added and incubated for *ca*. 20 min at room temperature. ΔA

- remains constant for more than 1 h, implying that the **HNABA** sensor to Fe^{3+} is stable and reliable.
- The influence of ionic strength on ΔA of the system at 478 nm was also investigated by varying NaCl concentrations from ${}_{40} 1.0 \times 10^{-2} \text{ mol} \cdot \text{L}^{-1}$ to $1.0 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$. It is worthwhile noting that all the ion strengths tested have no obvious effect on ΔA , hinting that the target sensing system is quite stable and may be applied in various kinds of surroundings.

3.3 Analytical parameters and samples detection

⁴⁵ **Fig. 4** shows the colour change and the absorption spectra of **HNABA** at different concentrations of Fe³⁺ between 9.5~400 ×10⁻⁸ mol·L⁻¹. From Fig. 4a, it is easy to find that the colour of the sensing system changes from brick-red to light red, which can be detected by naked eye. Also, the calibration graph, the ⁵⁰ detection limit and precision for Fe³⁺ detection are obtained under the optimal conditions from Fig. 4b and 4c. A linear relationship between ΔA and Fe³⁺ concentration is exhibited in the range of 9.5~400 ×10⁻⁸ mol·L⁻¹ with a correlation coefficient of 0.9938. The regression equation is $A = -0.04834 \times 10^{-4} + 1.37 \times 10^{-3}$ c. ⁵⁵ Based on the definition of detection limit, three times of average deviation of UV absorption at 478 nm in 20 blank samples without Fe³⁺ divided by the slope absolute value of the standard curve in Fig. 4c here, the limit of detection (LOD) for Fe³⁺ is up to 4.2×10^{-9} mol·L⁻¹.



⁶⁰ Fig. 4 The colour change (4a) and the absorption spectra (4b) of HNABA in different Fe³⁺ concentrations (From up to down: 0, 9.5, 20, 60, 100, 150, 200, 250, 300, 350, 400 ×10⁻⁸ mol·L⁻¹); and the linear relationship between the ΔA of HNABA at 478 nm and c_{Fe}^{3+} (4c)

⁶⁵ To confirm its feasibility, the proposed method was applied to determine Fe³⁺ in 3 environmental water samples from the Pi River, underground water and tap water in campus, respectively (**Table 1**). All the samples were obtained by filtering several times and concentrated by 100 times before testing. For recovery studies, some known concentrations of Fe³⁺ were added to the environmental water samples and the total Fe³⁺ concentrations were determined following the method proposed above. The s recoveries of different known amounts of Fe³⁺ spiked were obtained from 97.8 % to 102.2% with a satisfying analytical precision (R.S.D. \leq 3.5 %).

 Table 1 Determination results for environmental water samples (n=5)^a

	Samples ^b	C_{Fe}^{3+} in	Spiked	Found	Recovery	R.S.D.
		sample ^b (nM)	(nM)	(nM)	(%)	(%)
	1 (the Pi River)	788.5	500.0	1277.7	97.8	2.2
2	(underground) 3 (pure water)	985.2 0.00	500.0 500.0	1496.0 492.8	102.2 98.6	3.5 1.3

10 a. PB, pH 7.0.

b. The environmental water Fe³⁺ concentration determined using **HNABA** with the proposed method. The real values are the table values $\times 10^{-2}$ nmol·L⁻¹ for the detected water samples were concentrated 100 times.

15 3.4 Action mechanism between HNABA and Fe³⁺

To investigate the nature of the bonding between **HNABA** and Fe^{3+} , the binding stoichiometry of **HNABA** with Fe^{3+} was determined by using Job's plot. For the Job plot analyses, a series of solutions with varying mole fraction of Fe^{3+} were prepared by ²⁰ maintaining the total **HNABA** and Fe^{3+} concentration constant $(6.0 \times 10^{-5} \text{ mol} \cdot L^{-1})$. The absorption intensity at 478 nm was measured for each solution. As shown in **Fig. 5**, a 1:1 stoichiometry for the complex between **HNABA** and Fe^{3+} can be drawn from Job's plots,⁴¹ which confirms that Fe^{3+} might

25 coordinate with nitrogen atoms in -N=N bonds or with oxygen atoms in naphthol rings.



Fig. 5 Job's plot of **HNABA**-Fe³⁺ system. [**HNABA**] + [Fe³⁺] = 8.0×10^{-5} mol·L⁻¹ in aqueous at pH 7.0

- ³⁰ Theoretical calculations have been carried out to further understand the nature of the bonding between **HNABA** and Fe³⁺. The structures of **HNABA** before and after coordinating with Fe³⁺ were shown in **Fig. 6**, which were optimized using the the B3LYP/6-31G level of theory and method implemented in the
- ³⁵ Gaussian 03 suite of program.⁴² From the results, we can find easily that the terminal phenyl rings distorted greatly, once the binding of Fe³⁺ with nitrogen atoms in –N=N bonds in **HNABA**,

which resulted in the original conjugated system destroyed and so the absorbance at 478 nm deduced and even quenched.



⁴⁰ Fig. 6 Optimized geometries of HNABA before and after acted with Fe³⁺ ion. O, N, C, H, S and Fe atom are represented as red, blue, gray, whitegray, yellow and blue-purple, respectively

4. Conclusions

In conclusion, a multifunctional dye, 4-(4-hydroxy-1-naphthylazo) 45 benzenesulfonic acid (HNABA) possessed a strong absorption (ε = $2.54 \times 10^4 \text{ L} \cdot \text{mol}^{-1} \cdot \text{cm}^{-1}$) at *ca*. 478 nm in polar protic water and kept tremendously stable under acid or neural conditions. Based on the results, the dve was successfully developed for trace Fe³⁺ detection with high selectivity and sensitivity under the ⁵⁰ physiological pH condition (pH 7.0). The optimal test conditions were obtained (After 20 min incubation time at room temperature under pH =7.0, c_{HNABA} 5.0×10⁻⁵ mol·L⁻¹ in water) by investigating the influences of solvent, pH, ion intensity and incubation time on detection sensitivity. The linear range to s5 detect Fe^{3+} in aqueous environment was 9.5~400 ×10⁻⁸ mol·L⁻¹ with a detection limit of 4.2 $\times 10^{-9}$ mol·L⁻¹ and a correlation coefficient of 0.9938. The action mechanism of HNABA and Fe³⁺ ion was confirmed by means of Job's plots, experimental and theoretical deduction as well.

60 Acknowledgements

The authors gratefully acknowledge the financial supports from the National Natural Science Fund of China (No: 21277103) and Anhui Provincial Natural Science Fund (No: 1308085ME57).

Notes and references

- 65 Anhui Provincial Laboratory of Biomimetic Sensor and Detecting Technology & Solar Photovoltaic Materials Research Center, West Anhui University, Lu'an 237012, China; Tel: 86 5643305801; E-mail: yanzhq2008@163.com
- † Electronic Supplementary Information (ESI) available: [details of any 70 supplementary information available should be included here]. See DOI: 10.1039/b000000x/
 - R. K. Shervedani, A. Hatefi-Mehrjardi and A. Asadi-Farsani, *Anal Chim Acta*, 2007, 601, 164-171.
- 75 2. M. Shamsipur, M. Sadeghi, A. Garau and V. Lippolis, *Anal Chim Acta*, 2013, 761, 169-177.
 - 3. J. Mao, Q. He and W. S. Liu, *Talanta*, 2010, 80, 2093-2098.
 - 4. P. Blatny, F. Kvasnicka and E. Kenndler, *J Chromatogr A*, 1997, 757, 297-302.
- 80 5. M. J. C. Marenco, C. Fowley, B. W. Hyland, D. Galindo-Riano, S. K. Sahoo and J. F. Callan, *J Fluoresc*, 2012, 22, 795-798.
 - 6. T. A. Ali, G. G. Mohamed, M. M. I. El-Dessouky, S. M. Abou El Ella and R. T. F. Mohamed, *Int J Electrochem Sc*, 2013, 8, 1469-1486.
- 7. N. R. Chereddy, S. Thennarasu and A. B. Mandal, *Dalton T*, 2012, 41, 11753-11759.

- 8. B. D. Wang, J. Hai, Z. C. Liu, Q. Wang, Z. Y. Yang and S. H. Sun, *Angew Chem Int Edit*, 2010, 49, 4576-4579.
- 9. Y. Y. Du, M. Chen, Y. X. Zhang, F. Luo, C. Y. He, M. J. Li and X. Chen, *Talanta*, 2013, 106, 261-265.
- 5 10. C. H. Ma, L. P. Lin, Y. Y. Du, L. B. Chen, F. Luo and X. Chen, *Anal Methods-Uk*, 2013, 5, 1843-1847.
- 11. K. G. Qu, J. S. Wang, J. S. Ren and X. G. Qu, *Chem-Eur J*, 2013, 19, 7243-7249.
- 12. S. H. Wang, L. Y. Du, A. M. Zhang and B. Li, *Anal Lett*, 1997, 30, 2099-2107.
- X. F. Wu, B. W. Xu, H. Tong and L. X. Wang, *Macromolecules*, 2010, 43, 8917-8923.
- 14. X. S. Zhu and R. R. Jiang, J Fluoresc, 2011, 21, 385-391.
- M. Becuwe, P. Rouge, C. Gervais, M. Courty, A. Dassonville-Klimpt,
 P. Sonnet and E. Baudrin, *J Colloid Interf Sci*, 2012, 388, 130-136.
- 16. M. M. Zareh, I. F. A. Ismail and M. H. Abd El-Aziz, *Electroanal*, 2010, 22, 1369-1375.
- 17. B. Haghighi and A. Safavi, Anal Chim Acta, 1997, 354, 43-50.
- 18. W. Ruengsitagoon, Talanta, 2008, 74, 1236-1241.
- 20 19. M. Sugiyama, Y. Naraki and T. Hori, J Liq Chromatogr R T, 2009, 32, 788-800.
 - 20. H. Matsumiya, N. Iki and S. Miyano, Talanta, 2004, 62, 337-342.
 - 21. I. A. Korobeinikova, G. B. Pronchev and A. N. Ermakov, *J Anal Chem*, 2011, 66, 740-744.
- 25 22. H. Fujii, J Am Chem Soc, 2002, 124, 5936-5937.
- 23. S. B. Khan, M. M. Rahman, H. M. Marwani, A. M. Asiri, K. A. Alamry and M. A. Rub, *Appl Surf Sci*, 2013, 282, 46-51.
- 24. L. Hu, Z. Q. Yan and H. Y. Xu, Rsc Adv, 2013, 3, 7667-7676.
- 25. L. Hu, Y. F. Zhang, L. Nie, C. G. Xie and Z. Q. Yan, *Spectrochim* 30 *Acta A*, 2013, 104, 87-91.
- 26. Z. Q. Yan, S. Y. Guang, H. Y. Xu and X. Y. Liu, *Analyst*, 2011, 136, 1916-1921.
- 27. Z. Q. Yan, L. Hu, L. Nie and H. Lv, *Spectrochim Acta A*, 2011, 79, 661-665.
- 35 28. E. W. Baumann, Analyst, 1992, 117, 913-916.
- 29. M. A. Kabil and S. E. Ghazy, Anal Sci, 1995, 11, 817-822.
- K. J. Wallace, M. Gray, Z. L. Zhong, V. M. Lynch and E. V. Anslyn, *Dalton T*, 2005, 2436-2441.
- 31. J. Yun and H. Choi, Talanta, 2000, 52, 893-902.
- 40 32. S. Kawakubo, K. Shimada, Y. Suzuki and K. Hattori, *Anal Sci*, 2011, 27, 341-344.
 - 33. S. P. Wu, Y. P. Chen and Y. M. Sung, Analyst, 2011, 136, 1887-1891.
- 34. B. K. Adebayo, S. Ayejuyo, H. K. Okoro and B. J. Ximba, *Afr J Biotechnol*, 2011, 10, 16051-16057.
- 45 35. S. Zareba and H. Hopkala, J Pharmaceut Biomed, 1996, 14, 1351-1354.
 - 36. A. K. Sharma and I. Singh, Food Anal Method, 2009, 2, 221-225.
- 37. Z. Q. Yan, Y. F. Chen, S. Y. Guang, H. Y. Xu and L. F. Li, *Polym Sci Ser B*, 2011, 53, 535-539.
- 50 38. Z. Q. Yan, S. Y. Guang, H. Y. Xu and X. Y. Liu, *Dyes Pigments*, 2013, 99, 720-726.
 - 39. Z. Q. Yan, H. Y. Xu, S. Y. Guang, X. Zhao, W. L. Fan and X. Y. Liu, *Adv Funct Mater*, 2012, 22, 345-352.
- 40. Z. Q. Yan, S. Y. Guang, H. Y. Xu, X. Y. Su, X. L. Ji and X. Y. Liu, ⁵⁵ *Rsc Adv*, 2013, 3, 8021-8027.

- 41. H. J. Kim, J. E. Park, M. G. Choi, S. Ahn and S. K. Chang, *Dyes Pigments*, 2010, 84, 54-58.
- 42. M. J. T. Frisch, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, J. A., Jr.; Vreven, T.; Kudin, K. N.;
- 60 Burant, J. C.;, Gaussian 03, revision A.1, Gaussian, Inc.: Pittsburgh, PA, 2004.