RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This Accepted Manuscript will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/advances

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

COMMUNICATION

A single-step emulsion approach to prepare fluorescent nanoscale coordination polymers for bioimaging

Zhiyong Sun,^a Yangxue Li,^b Xingang Guan,^b Tingting Sun,^b Li Chen, ^{*a} Zhigang Xie,^{*b} and Xiabin Jing^b

s Received (in XXX, XXX) Xth XXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

A crystalline nanoscale coordination polymer (NCP) based on fluorene and Zr⁴⁺ was synthesized through simply microemulsion method. The emulsion method is facile and ¹⁰ convenient for NCPs producing, the nanoparticles are fluorescent and thermal-stable, most important, they are in nanoscale thus make them applicable for bioimaging.

Luminescence nanomaterials have drawn a great of interest in bioscience such as bioimaging, drug carriers, and disease 15 detection.¹ luminescence bioimaging, In luminescent nanomaterials are used to label a molecule of interest and to render luminescent signals. Many new luminescent nanomaterials, have been synthesized for bioimaging application including semiconductor nanocrystals,² upconversion ²⁰ nanophosphors,³ quantum dots⁴ and other nanoparticles.⁵⁻⁸ Among them, coordination polymers (CPs), also called metalorganic frameworks, are attractive not only for their porous structures and high surface areas, but also for the convenience of them to be designed and incorporated with function groups which

- ²⁵ made CPs have potential applications in catalysis,⁹⁻¹¹ gas storage,¹²⁻¹³ separation¹⁴ and bioscience.¹⁵⁻¹⁶ However, CPs particles are usually in micrometer scale, which limit their application. Recently, Lin's group reported the ability to scale down the size of CPs to nanoscale,¹⁷ thus made the luminescent ³⁰ CPs an excellent material for biological applications.¹⁸⁻²¹
 - Conventionally, CPs are synthesized via time-consuming hydrothermal or solvothermal methods, these methods require several hours or days for crystallization and nanoparticles formation. Other approaches involve alternative energy sources
- ³⁵ such as microwave irradiation or ultrasound usually decreased the crystallization time, but required special power-consuming apparatus. It is highly desirable to develop new method to prepare the CPs in simple and mild condition. Very

^{*a*} Department of Chemistry, Northeast Normal University, 5268 Renmin 40 Street, Changchun 130024, P. R. China

Email: chenl686@nenu.edu.cn ^bState Key Laboratory of Polymer Physics and Chemistry, Changchun Institute of Applied Chemistry, Chinese Academy of Sciences, 5625 Renmin Street, Changchun 130022, P. R. China.

45 E-mail: xiez@ciac.ac.cn; Tel: +86-431-85262779 †Electronic Supplementary Information (ESI) available: Experimental procedures, characterization data, See DOI: 10.1039/b000000x/



Scheme 1 Preparation of ligand and NCPs

⁵⁰ recently, several groups have reported some new methods to make CPs, like spray-dry strategy²² and microfluidic approach.²³ Microemulsion method (MEM) is usually used for the preparation of polymeric nanoparticles. It is easy and convenient, and the nanoparticles prepared by MEM usually have narrow size ⁵⁵ distribution.²⁴ Very recently, Eddaoudi et al. reported a singlestep emulsion-based technique to assembly of CPs into 3D hollow superstructures.²⁵

Polyfluorenes are of interest and currently being investigated to use in light-emitting diodes, field-effect tranistors and plastic solar cells.²⁶⁻²⁸ Fluorene-based materials have been studied in bioscience because of their stability and safety to living cells.²⁹⁻³¹ Some reports on the conjugated polymers based on fluorene for biomarkers and detection of proteins and DNA were published in last several years.³²⁻³⁵



Fig. 1 SEM (a, b) and TEM (c) of the NCPs. FT-IR Spectroscopy (d) of the ligand (red) and the particles (black).

- Herein, we synthesized novel nanoscale coordination polymers (NCPs) with microemulsion method using Zr⁴⁺ and fluorene derivatives. The NCPs were characterized by thermogravimetric analysis (TGA), powder X-ray diffraction (PXRD), Fourier
- ¹⁰ transform infrared spectroscopy (FT-IR), scanning electron microscopy (SEM) and transmission electron microscopy (TEM). Then the bioimaging experiment was carried out for they are fluorescent and in nanoscale which is easy to endolysis by living cells.
- ¹⁵ Scheme 1 shows the preparation of the ligand and NCPs. The ligand (2, 7-(4-carboxyl-benzene)-9, 9'-dioctyl-9H-fluorene) was prepared through Suzuki coupling reaction and subsequent hydrolysis reaction. The structures were confirmed by ¹H-NMR (Fig. S1, ESI). Then the ligand and ZrCl₄ were dissolved in DMF
- ²⁰ with a small amount of trifluoroacetic acid (TFA) and hydrochloric acid (HCl) to modulate the crystallinity of the nanoparticles, then the mixture was dropped into a heated silicon oil bath to get the NCPs. Silicon oil is immiscible with DMF and used to form emulsion drops, in which the particles were
- ²⁵ prepared after aggregation, nucleus and crystallization³⁶ (See the ESI for detail).

The SEM (a, b) and TEM (c) of the NCPs were presented in Fig. 1. The images show the particles were in good dispersion, and in nanoscale with diameter of about 50 nm, which is easy to ³⁰ endocytosis by living cells for bioimaging application. We can

see that the particles are in nearly the same sphere morphous from the SEM pictures.

Fig. 1d shows the FT-IR of the ligand and the nanoparticles. The peaks at 2900 cm^{-1} are due to C-H stretching vibration of the

 $_{35}$ C₈H₁₇ chain of the fluorene for both the ligand and the particles. Compared with the ligand, the missing of the peak at 1691 cm⁻¹ of the NCPs is responsible for the coordination bond formation in coordination polymers. Instead, the appearance of two new peaks at 1602 cm⁻¹



Fig. 2 (a) PXRD of as synthesized NCPs (red) and UiO-66 (black). (b) TGA. (c) Simulated structure of NCPs, the green ⁴⁵ parts represent the ligand, colors of atoms O (red), Zr (light blue); MTT (d) of Hela cells after NCPs endocytosis.

and 1420 cm⁻¹ are responsible for antisymmetric and symmetric carboxylate stretching vibrations, the value between 1602 cm⁻¹ ⁵⁰ and 1420 cm⁻¹ is less than 200 cm⁻¹, this indicates the ligand is connected to Zr⁴⁺ through bidentate coordination state.³⁷⁻⁴⁰

Since the NCPs are crystalline, as indicated by the PXRD shown in Fig. 2a, we decided to study the structure of our NCPs. From Fig. 2a, we can see the PXRD pattern of our NCPs fit well but a 55 little blue shifted compared to that of UiO-66, so we concluded our NCPs would share the same structure with UiO-66,⁴¹ which is consistent with the analysis of FT-IR. And we simulated the structure of our NCPs as seen in Fig. 2c. The possible reason of the slight difference is that the unit cell volume of our NCPs is

- ⁶⁰ larger than UiO-66, the diffraction angles shift to the small angles. We assumed there are two reasons for this: (1) The ligand is different; our ligand is much longer compared to terephthalic acid, which is the ligand of UiO-66, and the octane chains of our ligand could increase the cell volume. (2) We added TFA and ⁶⁵ HCl as modulation to control the crystallization, and the TFA
- could coordinate to Zr^{4+} ,⁴² then left after activated , thus would affect the PXRD pattern.

We also studied the TGA of the nanoparticles, the result is shown in Fig. 2b. The weight loss of the material is steady ⁷⁰ decreased from room temperature to about 300 °C due to the solvent loss. The weight loss from 300 to 500 °C is because of the ligand decomposing. And from 500 to 630 °C, the rate of weight loss decreased and the finally get to a steady state. This is because that the final substance is the very stable zirconium ⁷⁵ oxide. The TGA result shows our NCPs are as stable as UiO-66.⁴³⁻⁴⁴ Moreover, the amount of the Zr⁴⁺ calculated from the result of TGA is 22.56 wt %, which fits well with the result of ICP-MS, 23.08 %.

The particles remain fluorescence when dispersed in water (as so shown in Fig. S2 and figure S3, ESI) and common organic solvents. They are possible used for bioimaging for the fluorescence and nanoscale size. Determination of the biocompatiblitity of NCPs is very important before using them in

40

living cell.⁴⁵⁻⁴⁶ As shown in Fig 2d, no obvious cytotoxicity to human breast cancer cells (HeLa) is observed via cell viability examination. Even with a concentration of NCPs up to 100 µg mL⁻¹, the cell viability is still greater than 90 % after 24 h,



Fig. 3 Microscopic images of the cells. the scale bars are 500 µm and 100 µm for (a) and (b), respectively.

- 10 demonstrating their excellent biocompatibility. Then the bioimaging experiment was further exploited to demonstrate their biomedical applications using HeLa cells. Fig. 3a and 3b confirmed our nanoparticles are in the cytoplasm of the cells, which indicated by Lin's and Petoud's groups.^{17,47} The signal of
- 15 the nanoparticles is high under microscopic with irritation. To further confirm the NCPs remain intact within HeLa cells, we carried out the control experiment with the ligand as the bioimaging agent, the pictures are presented in Fig S4 (ESI). Because the ligand is not soluble in water environment so it
- 20 aggregates into large particles, which is different with the NCPs. We can also see that the cells were in good shape and condition under different resolutions.

In summary, we have synthesized a nanoscale coordination 25 polymer material through a convenient microemulsion method. The nanoparticles were characterized through FIRT and PXRD, which indicated the NCPs formed through coordination bond and were crystalline. SEM and TEM pictures confirmed the size of the particles were in good distribution and in nanoscale which

- 30 made them possible for endocytosis by living cells. The fluorescence of the NCPs in Hela cells after endocytosis for bioimaging confirmed our idea for the possible application on bioscience of the NCPs.
- 35 This work was supported by the National Natural Science Foundation of China (Project No 91227118 and 21104075).

Notes and references

1. Lee, K. J.; Oh, W. K.; Song, J.; Kim, S.; Lee, J. and Jang, J., Chem 40 Commun (Camb), 2010, 46, 5229.

2. Satapathi, S.; Pal, A. K.; Li, L.; Samuelson, L. A.; Bello, D. and Kumar, J., RSC Advances, 2014, 4, 1116.

- 3. Ren, W.; Tian, G.; Jian, S.; Gu, Z.; Zhou, L.; Yan, L.; Jin, S.; Yin, W. and Zhao, Y., RSC Advances, 2012, 2, 7037.
- 45 4. Guo, W.; Chen, N.; Dong, C.; Tu, Y.; Chang, J. and Zhang, B., RSC Advances, 2013, 3, 9470.

5. Mitra, S.; Chandra, S.; Pathan, S. H.; Sikdar, N.; Pramanik, P. and Goswami, A., RSC Advances, 2013, 3, 3189.

6. Ribeiro, T.; Raja, S.; Rodrigues, A. S.; Fernandes, F.; Farinha, J. P. S. 105 36. Faustini, M.; Kim, J.; Jeong, G. Y.; Kim, J. Y.; Moon, H. R.; Ahn, 50 and Baleizão, C., RSC Advances, 2013, 3, 9171.

- 7. Sun, H.; Yang, L.; Yang, H.; Liu, S.; Xu, W.; Liu, X.; Tu, Z.; Su, H.; Zhao, Q. and Huang, W., RSC Advances, 2013, 3, 8766.
- 8. Zhang, Z.; Hao, J.; Zhang, J.; Zhang, B. and Tang, J., RSC Advances, 2012, 2, 8599.
- 55 9. Liu, Y.; Xuan, W. and Cui, Y., Adv Mater, 2010, 22, 4112.
- 10. Prier, C. K.; Rankic, D. A. and MacMillan, D. W., Chem Rev, 2013, 113. 5322.
- 11. Shi, J., Chem Rev, 2013, 113, 2139.
- 12. Liu, C.; Li, F.; Ma, L. P. and Cheng, H. M., Adv Mater, 2010, 22, 60 E28.
- 13. Suh, M. P.; Park, H. J.; Prasad, T. K. and Lim, D. W., Chem Rev, 2012, 112, 782.
- 14. Li, J. R.; Sculley, J. and Zhou, H. C., Chem Rev, 2012, 112, 869.
- 15. Yang, Y.; Zhao, Q.; Feng, W. and Li, F., Chem Rev, 2013, 113, 192.
- 65 16. Horcajada, P.; Gref, R.; Baati, T.; Allan, P. K.; Maurin, G.; Couvreur, P.; Ferey, G.; Morris, R. E. and Serre, C., Chem Rev, 2012, 112, 1232.
- 17. Liu, D.; Huxford, R. C. and Lin, W., Angew Chem Int Ed Engl, 2011, 50. 3696.
- 18. Della Rocca, J. and Lin, W., European Journal of Inorganic 70 Chemistry, 2010, 2010, 3725.
- 19. Rieter, W. J.; Pott, K. M.; Taylor, K. M. and Lin, W., J Am Chem Soc, 2008, 130, 11584.
- 20. Huxford, R. C.; Della Rocca, J. and Lin, W., Current Opinion in Chemical Biology, 2010, 14, 262.
- 75 21. Rieter, W. J.; Taylor, K. M.; An, H.; Lin, W. and Lin, W., J Am Chem Soc, 2006, 128, 9024.

22. Arnau Carne'-Sa'nchez, I. I., Mary Cano-Sarabia and Daniel Maspoch, Nat Chem, 2013, 5.

- 23. Faustini, M.; Kim, J.; Jeong, G. Y.; Kim, J. Y.; Moon, H. R.; Ahn, 80 W. S. and Kim, D. P., J Am Chem Soc, 2013, 135, 14619.
- 24. Qian, Q.; Huang, X.; Zhang, X.; Xie, Z. and Wang, Y., Angew Chem Int Ed Engl, 2013, 52, 10625.
- 25. Pang, M.; Cairns, A. J.; Liu, Y.; Belmabkhout, Y.; Zeng, H. C. and Eddaoudi, M., J Am Chem Soc, 2013, 135, 10234.
- 85 26. Kappaun, S.; Scheiber, H.; Trattnig, R.; Zojer, E.; List, E. J. and Slugovc, C., Chem Commun (Camb), 2008, 5170.
- 27. Watanabe, K.; Koyama, Y.; Suzuki, N.; Fujiki, M. and Nakano, T., Polymer Chemistry, 2014, 5, 712.
- 28. Behrendt, J. M.; Wang, Y.; Willcock, H.; Wall, L.; McCairn, M. C.; 90 O'Reilly, R. K. and Turner, M. L., Polymer Chemistry, 2013, 4, 1333.

29. Kahveci, Z.; Martinez-Tome, M. J.; Mallavia, R. and Mateo, C. R., Biomacromolecules, 2013, 14, 1990.

30. Chien, R.-H.; Lai, C.-T. and Hong, J.-L., The Journal of Physical Chemistry C, 2011, 115, 20732.

- 95 31. Bilalis, P.; Katsigiannopoulos, D.; Avgeropoulos, A. and Sakellariou, G., RSC Advances, 2014, 4, 2911.
- 32. Petkau, K.; Kaeser, A.; Fischer, I.; Brunsveld, L. and Schenning, A. P., J Am Chem Soc, 2011, 133, 17063.
- 33. Song, J.; Lv, F.; Yang, G.; Liu, L.; Yang, Q. and Wang, S., Chem 100 Commun (Camb), 2012, 48, 7465.
- 34. Zhu, C.; Yang, Q.; Liu, L. and Wang, S., Angew Chem Int Ed Engl, 2011, 50, 9607.
- 35. Zhu, C.; Yang, Q.; Liu, L. and Wang, S., Journal of Materials Chemistry, 2011, 21, 7905.
- W. S. and Kim, D. P., J Am Chem Soc, 2013, 135, 14619-26.

37. Zelenak, V.; Vargova, Z. and Gyoryova, K., *Spectrochim Acta A Mol Biomol Spectrosc*, 2007, **66**, 262-72.

 Nguyen Thi, T. V.; Luu, C. L.; Hoang, T. C.; Nguyen, T.; Bui, T. H.; Duy Nguyen, P. H. and Pham Thi, T. P., *Advances in Natural Sciences:* 5 *Nanoscience and Nanotechnology*, 2013, 4, 035016.

39. Ebrahim, A. M. and Bandosz, T. J., ACS Appl Mater Interfaces, 2013, 5, 10565.

40. Ebrahim, A. M.; Levasseur, B. and Bandosz, T. J., *Langmuir*, 2013, **29**, 168.

10 41. Garibay, S. J. and Cohen, S. M., *Chem Commun (Camb)*, 2010, 46, 7700.

42. Vermoortele, F.; Bueken, B.; Le Bars, G.; Van de Voorde, B.; Vandichel, M.; Houthoofd, K.; Vimont, A.; Daturi, M.; Waroquier, M.; Van Speybroeck, V.; Kirschhock, C. and De Vos, D. E., *J Am Chem Soc*, 15 2013, **135**, 11465.

43. Kandiah, M.; Nilsen, M. H.; Usseglio, S.; Jakobsen, S.; Olsbye, U.; Tilset, M.; Larabi, C.; Quadrelli, E. A.; Bonino, F. and Lillerud, K. P., *Chemistry of Materials*, 2010, **22**, 6632.

44. Wiersum, A. D.; Soubeyrand-Lenoir, E.; Yang, Q.; Moulin, B.;

²⁰ Guillerm, V.; Yahia, M. B.; Bourrelly, S.; Vimont, A.; Miller, S.; Vagner, C.; Daturi, M.; Clet, G.; Serre, C.; Maurin, G. and Llewellyn, P. L., *Chem Asian J*, 2011, **6**, 3270.

45. Zhang, X.; Zhang, X.; Wang, S.; Liu, M.; Tao, L. and Wei, Y., Nanoscale, 2013, 5, 147.

25 46. Zhang, X.; Wang, S.; Xu, L.; Feng, L.; Ji, Y.; Tao, L.; Li, S. and Wei, Y., *Nanoscale*, 2012, 4, 5581.

47. Foucault-Collet, A.; Gogick, K. A.; White, K. A.; Villette, S.; Pallier, A.; Collet, G.; Kieda, C.; Li, T.; Geib, S. J.; Rosi, N. L. and Petoud, S., *Proc Natl Acad Sci U S A*, 2013, **110**, 17199.

30

RSC Advances

A facile and convenient microemulsion method is demonstrated to perpare fluorescent nanoscale coordination polymers. And the nanoscale coordination polymers exhibted brightly blue fluorescence and good biocompatiability, thus endowed them the ability for bioimaging.

