

# Polymer Chemistry

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

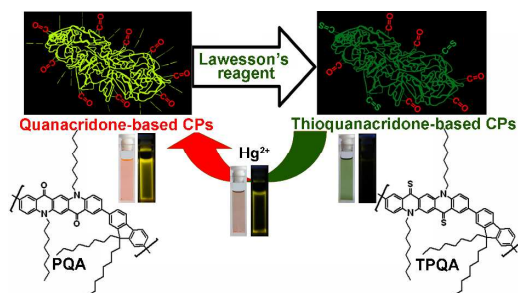
You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

*Graphics for Contents***Fluorescent conjugated polymer based on thiocarbonyl quinacridone for sensing mercury ion and bioimaging**

*Yi Qu,<sup>a,b</sup> Xinran Zhang,<sup>a</sup> Yongquan Wu,<sup>b</sup> Fuyou Li<sup>b,\*</sup> and Jianli Hua<sup>a,\*</sup>*

Highly sensitive FRET-based thiocarbonyl quinacridone fluorescent conjugated polymers for sensing and bioimaging of mercury ion have been developed.



# Fluorescent conjugated polymer based on thiocarbonyl quinacridone for sensing mercury ion and bioimaging

*Yi Qu,<sup>a,b</sup>Xinran Zhang,<sup>a</sup> Yongquan Wu,<sup>b</sup> Fuyou Li<sup>b\*</sup> and Jianli Hua<sup>a\*</sup>*

<sup>a</sup>Key Laboratory for Advanced Materials and Institute of Fine Chemicals, East China University of Science and Technology, 130 Meilong Road, Shanghai 200237, P. R. China

<sup>b</sup>Department of Chemistry & Laboratory of Advanced Materials, Fudan University, 220 Handan Road, Shanghai, 200433, P. R. China

Emails: jlhua@ecust.edu.cn (Prof. Jianli Hua); fyli@fudan.edu.cn (Prof. Fuyou Li)

†Electronic supplementary information (ESI) available: The <sup>1</sup>H NMR, GPC data, IR spectra of four quinacridone-based polymers. The selectivity experimental figures of **PTQA2** and **PTQA-NPs**. See DOI:

## Abstract

Because of the extreme toxicity of mercury ions ( $\text{Hg}^{2+}$ ), a great deal of effort has been invested in developing probes that use colorimetric and fluorometric methods to detect them. Nowadays, most of the current fluorescent probes still work in the organic solvents or a mixture of organic solvents and water. Conjugated polymers (CPs) can serve as excellent fluorophores because of their strong emission and the controllable emission wavelength. In this work, a kind of thiocarbonyl quinacridone-based CPs nanoparticle (**PTQA-NPs**) was synthesized for selectively detecting  $\text{Hg}^{2+}$  in pure water. Additionally, the fabrication of nanoparticle provides a very sensitive correction for environmental effects, the minimum detectable concentration of  $\text{Hg}^{2+}$  for this nanoparticle was as low as 1 ppb. Furthermore, we show the capability of this polymer to monitoring  $\text{Hg}^{2+}$  in HeLa cell lines by confocal laser scanning microscopy (CLSM).

**Keywords:** fluorescent probe; mercury ion; quinacridone; conjugated polymer nanoparticle; cell imaging

## Introduction

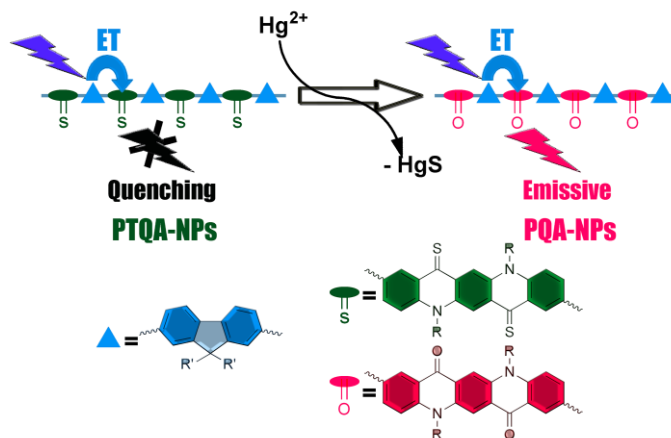
The mercury ion ( $\text{Hg}^{2+}$ ) is one of the most toxic metal ions because it can invade the nervous system, endocrine system and immune system of the organism.<sup>1-4</sup> Mercury can impair brain's cortex that caused central nerve system injury and dyskinesia. Embryo anamorphosis and mentally handicapped neonatorum also induced by infraction of mercury ion across the placenta.<sup>5-7</sup> Importantly, the concentration of the mercury pollutant is stored in animal tissues and not excreted that may be dramatically larger in the tissues of some animals at the top of a food chain.

A great deal of effort has been invested in developing easy-to-use colorimetric and fluorescent probes.<sup>8-14</sup> To achieve high selectivity, fluorescent chemodosimeters based on specific chemical reactions have been developed.<sup>15-18</sup> It is well-known that thioureas,<sup>19,20</sup> thioamides<sup>21</sup> and thiones<sup>22</sup> are all effective in reacting with mercury ion through the formation of the insoluble  $\text{HgS}$  deposits between the active  $\text{C}=\text{S}$  group and mercury ion. However, common commercial dyes in these researches showed poor stability under excitation in the presence of  $\text{Hg}^{2+}$  and oxygen. Quinacridone (QA) and its derivatives are widely used organic pigments with exceptional color and weather fastness.<sup>23</sup> Good light and temperature stability and high fluorescent quantum yield have permitted the fabrication of high performance organic fluorescent sensors based on QA derivatives. Two carbonyl groups on the skeleton of quinacridone can transfer to the thiolation analogue. Thiocarbonylquinacridone-based small molecular chemodosimeter for  $\text{Hg}^{2+}$  shows good selectivity and high detected limit (4.7 nM) that lower than the blood level (5.8 ppb) set by the U. S. EPA.<sup>24</sup> Furthermore, desulfurization of thiocarbonylquinacridone gives significant color change from dark green to bright red. As a

result, the derivative can be used as colorimetric probe for  $\text{Hg}^{2+}$ , which was seldom reported by previous studies.

It was well known that colorimetric and fluorescent probes based on water soluble CPs have attracted increasing attention and there are still challenges in this area.<sup>25-29</sup> Polyelectrolyte with amount of quaternary ammonium compounds is an useful strategy.<sup>30</sup> However, it is difficult to be modified on the insoluble pigment, such as quinacridone, which always need some more synthetic steps and lower yields. In another way, polymeric nanoparticles provide a widely universal fabrication method for hydrophobic and amphiphilic polymers.<sup>31-36</sup> In this work, we present the poly(fluorene-co-thiocarbonylquinacridone)-based colorimetric and fluorescent probe **PTQA-NPs** (Scheme 1) for sensing  $\text{Hg}^{2+}$  in both  $\text{CHCl}_3$  solution (polymer solution) and PBS buffers (polymer nanoparticles).

This sensory system provides some advantages: (1) chemodosimeter was designed in this system which can reduce the interference from complex biocircumstance; (2) facile and simple synthesis. Suzuki couple reaction was chosen for synthesizing conjugated polymer and Lawesson's reagent was used for efficient sulphuration of carbonyl group; (3) obviously fluorescent enhancement. This sensory system performed "turn on" response in blue (assigned to fluorene moiety) and orange (assigned to quinacridone moiety) channels. (4) high sensitivity. **PTQA-NPs** can detect  $\text{Hg}^{2+}$  in water at nM level.



**Scheme 1.** Schematic illustration of the sensing process of *PTQA-NPs* to  $\text{Hg}^{2+}$  with changes in both colorimetric and fluorescent approaches.

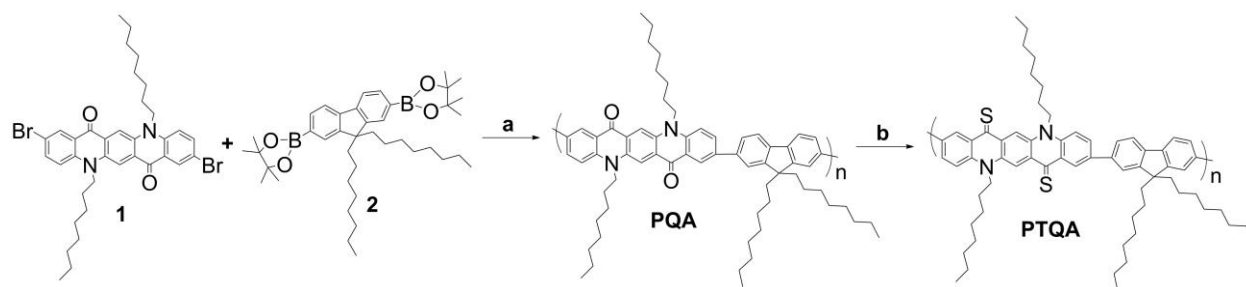
## Results and discussion

### Design and principle of PTQA-NP for sensing $\text{Hg}^{2+}$

This work was designed on the basis of energy transfer (ET) process which (As shown in Scheme 1). The main chain of CPs alternated between fluorene and quinacridone moieties and ET process occurred in the main chain under excitation. Fluorene moiety acts as light harvesting group that can achieve ET process to quinacridone (emitter) or thiocarbonyl quinacridone (quencher). Moreover, fluorene can work as internal reference that exhibited blue emission around 480 nm when  $\text{Hg}^{2+}$  was added in the aqueous. Quinacridone showed strong emission in the orange-red to NIR channel (560 ~ 750 nm). Different amount of thiocarbonyl groups were induced as ET quencher by reacting polyquinacridone with different ratio of Lawesson's reagent. These thiocarbonyl groups on quinacridone skeleton were used for sensing  $\text{Hg}^{2+}$  and the ratio of C=S/C=O could modulate the origin fluorescent intensity of CPs. During the desulfurized process, the sulfur atom captured  $\text{Hg}^{2+}$  ion and generated the insoluble substance  $\text{HgS}(\downarrow)$ .

Meanwhile, the thiocarbonyl group was recovered to carbonyl group. As a result, the  $\text{Hg}^{2+}$  was cleared and fluorescence lighted up.

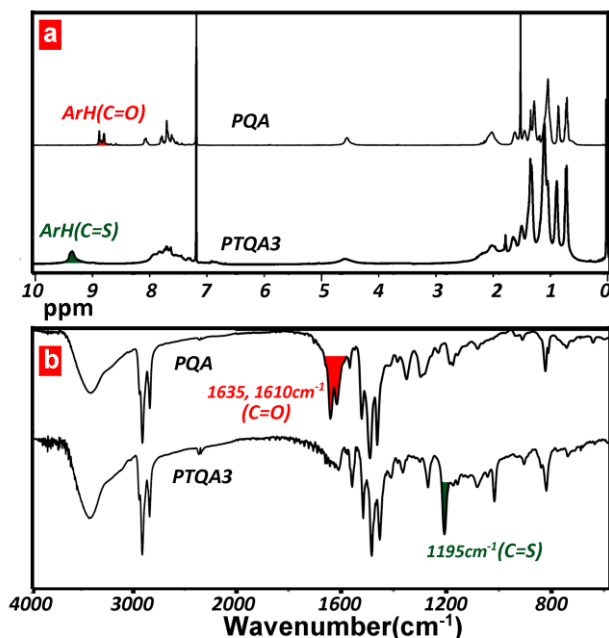
### Synthesis



**Figure 1.** The synthetic routine of **PTQA** polymers. (a)  $\text{Pd}(\text{PPh}_3)_4$ ,  $\text{K}_2\text{CO}_3$ ,  $\text{THF}/\text{H}_2\text{O}$ ; (b) Lawesson's reagent, toluene.

The synthetic route to **PQA** and **PTQA1~PTQA3** is illustrated in Figure 1. Monomer **1** is synthesized according to the previously published method.<sup>24</sup> Polymerization of **1** and 2,2'-(9,9-dioctyl-9H-fluorene-2,7-diyl)-bis(4,4,5,5-tetramethyl-1,3,2-dioxaborolane) (**2**) was carried out in the presence of palladium catalyst via the Suzuki cross-coupling reaction, providing the precursor polymer **PQA**. **PTQA1~PTQA3** with different thiolation rate could be obtained by a reaction of **PQA** with 0.4, 0.6 and 1.0 equiv. of Lawesson's reagent according to the literature with modification.<sup>37</sup>





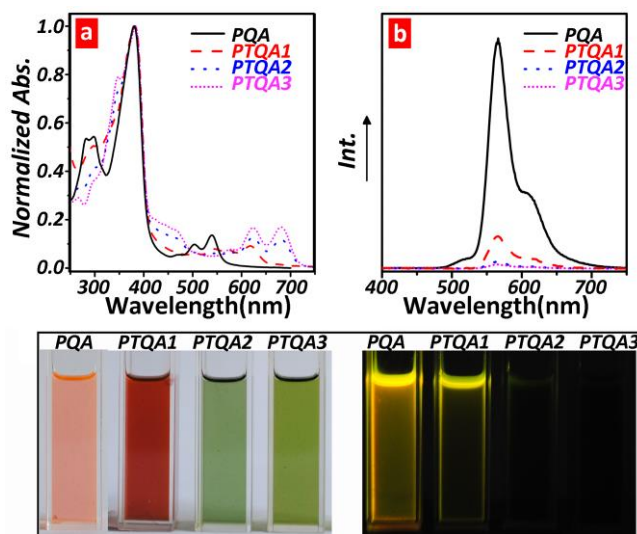
**Figure 2.** <sup>1</sup>H NMR and FT-IR spectra of **PQA** and **PTQA3**.

The <sup>1</sup>H NMR spectra of **PQA** and **PTQA1~PTQA3** were also used for determining the expected structure (see Figure 2a and Figure S1). As shown in Figure 2a, the <sup>1</sup>H NMR spectrum of **PQA** exhibited the peaks of phenylene group adjacent to C=O at 8.7–8.9 ppm and others of quinacridone group and fluorene group at 7.5–8.0 ppm. **PTQA1~PTQA3** with different amount of thiocarbonyl groups showed the signals decreased at 8.7–8.9 and increased peak around 9.4 (assigned to the protons adjacent to C=S moieties). We attempted to calculate the thiolated ratio from the integrations of both above peaks in the <sup>1</sup>H NMR spectra. Table S1 gives out a set of data of thiolated ratios for these three polymers: 26.3% (**PTQA1**), 66.7% (**PTQA2**), and 100% (**PTQA3**). The IR spectra of polymers were used for confirming the structure of **PQA** and its thiolate derivatives **PTQA1~PTQA3**. As shown in Figure 2b, The IR spectrum of **PQA** showed the sharp peaks at 1635 cm<sup>-1</sup> and 1610 cm<sup>-1</sup> due to C=O stretching vibration from quinacridone skeleton. On the other hand, the stretching vibration peaks of C=S at 1195 cm<sup>-1</sup> emerged in

**PTQA3**. These peaks can be found in **PTQA1** and **PTQA2**, indicating these polymers contained both C=O and C=S moieties (See Figure S2).

The GPC analysis revealed the weight average molecular weight ( $M_w$ ) and polydispersity of **PQA** and **PTQA1~PTQA3** (Table S2). The molecular weights of these polymers are relatively low, which may be because the copolymerization reactivity of the quinacridone and fluorene monomers is not very high. A similar phenomenon can be seen in the previous report on polycondensation of the monomers.<sup>38</sup> Overall, the molecular weights are high enough for the polymer sensors to have good sensing properties.

Figure 3 showed the normalized UV-vis absorption (a) and photoluminescence spectra (b) of **PQA** and **PTQA1~PTQA3** in chloroform. **PQA** showed the characteristic absorption band at around 380, 505 and 545 nm in chloroform solution, which corresponded to the  $\pi$ - $\pi^*$  transitions of the fluorene segments and the quinacridone units, respectively. **PTQA3** showed the same absorption of fluorene at 383 nm but the absorption peak belonged to quinacridone was disappeared. The new absorption bands emerged at 623 and 680 nm were confirmed to absorption of dithiocarbonyl quinacridone chromophore in our previous report.<sup>22</sup> Similar absorption spectra were also found in the chloroform solutions of **PTQA1** and **PTQA2**, both of which showed the characteristic absorbance of quinacridone and thiocarbonyl quinacridone chromophores.



**Figure 3.** UV-vis absorption (a) and fluorescence spectra (b) of **PQA** and **PTQA1 ~ PTQA3** in  $\text{CHCl}_3$ . Inset: color (a) and emission (b) pictures of **PQA** and **PTQA1~PTQA3**. [Polymer] = 10  $\mu\text{g/mL}$ ,  $\lambda_{\text{ex}} = 380 \text{ nm}$ .

The emission intensities of **PQA**, **PTQA1~PTQA3** in chloroform were strongly dependent on the concentration of thiocarbonyl quinacridone units (Figure 3b). In a solution of **PQA**, a stronger orange emission at 566 nm can be observed, whose quantum yield was 0.905 by using rhodamine B as the reference. Furthermore, the quantum yields of **PTQA1~PTQA3** decreased to 0.122, 0.025 and 0.017, respectively. This phenomenon can be understood by intrachain exciton migration in chloroform solution and thiocarbonyl unit acted as exciton trap in the conjugated system.<sup>39,40</sup> It is worth pointing out that the obviously color changes and stronger emission quenching of quinacridone-containing polymers with different thiocarbonyl concentration in the solvent can be observed by the naked eye (photos in Figure 3). All of the photophysical data were listed in Table 1.

**Table 1.** Photophysical properties of **PQA, PTQA1~PTQA3**

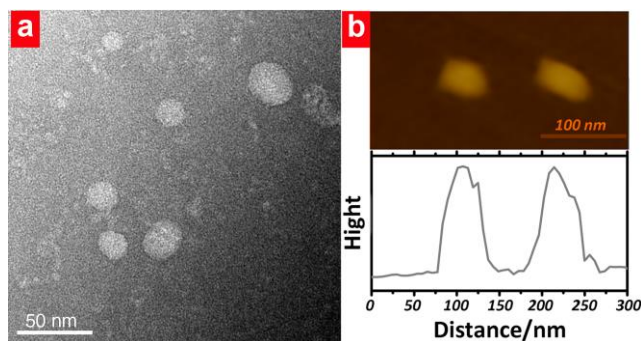
No.	$\lambda_{\text{abs.}}/\text{nm}$	$\lambda_{\text{em.}}/\text{nm}$	$\Phi$
<b>PQA</b>	380, 505, 545	566	0.905
<b>PTQA1</b>	382, 505, 545, 575, 623	566	0.122
<b>PTQA2</b>	382, 545, 575, 623, 680	563	0.025
<b>PTQA3</b>	383, 575, 623, 680	563	0.017

### Nanoparticle morphology

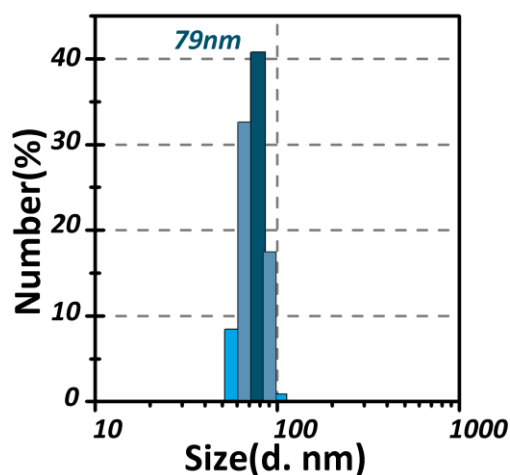
With the efficient quenching of fluorescence and lower thiolated ratio, **PTQA2** was chosen to fabricate **PTQA-NPs**. The THF solution of **PTQA2** was dispersed in water by ultrasonication and indeed conveniently formed nanoparticles. The route for preparing **PTQA-NPs** is described in supporting information. The topographic properties of these homogeneous nanoparticles were studied by transmission electron microscopy (TEM) and atomic force microscopy (AFM) and Dynamic light scattering (DLS).

The morphology of **PTQA-NPs** nanoparticles in dry state was investigated by TEM and AFM after depositing onto copper grid and mica, respectively. As shown in Figure 4a, ellipsoidal nanoparticles with an average diameter of 30 nm were observed by TEM. The AFM image in Figure 4b also shows the ellipsoidal morphology of **PTQA-NPs** nanoparticles with an average diameter of 50 nm. It was anticipated that the **PTQA-NPs** in aqueous solution with sub-50 nm diameter should be beneficial to perform detecting experiment in aqueous and cell uptake experiment. DLS measurement was further carried out to determine the hydrodynamic particle size of the whole nanoparticle range. The DLS measurements revealed that all the **PTQA-NPs** had a relatively narrow size distribution with a mean size of around  $79 \pm 6$  nm (Figure 5) in aqueous solution. The colloid particle diameter data obtained from TEM and AFM were

measured in the dry state while DLS result was the hydrodynamic diameter of the micelle measured in the solution state. Therefore, the particle sizes tested by TEM and AFM are smaller than that by DLS.



**Figure 4.** (a) TEM image of PTQA-NPs, carbon-coated copper grids, 1% phosphotungstic acid (PTA) negative stain; (b) AFM image with cross-sectional analysis of PTQA-NPs.



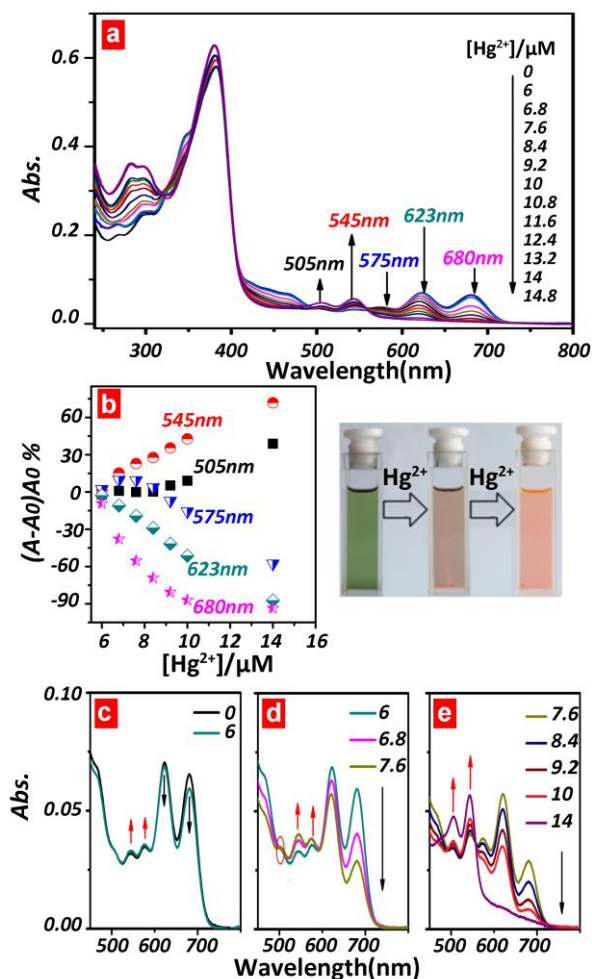
**Figure 5.** Histogram of the particle size distribution of PTQA-NPs.

### Sensing Properties

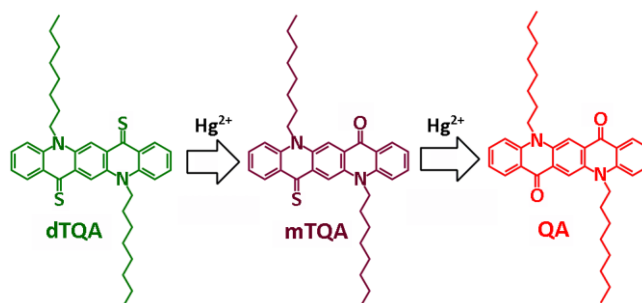
The number of fluorescence-quenching groups plays an important role on the sensor capability. On the one hand, if there are not enough groups, the fluorescence cannot be fully quenched, resulting in an unsuccessful fluorescence-enhanced sensor; on the other hand, if there are too many groups, the sensor will lose its sensitivity because it can't release the fluorescence-

enhanced signal in time when the concentration of analyte is low. Although the thiolation rate of **PTQA3** is higher than that of **PTQA2**, probe sensitivity of **PTQA3** can be affected since the fluorescence signal is weak at the beginning of adding  $\text{Hg}^{2+}$  ions. In addition, due to the similar fluorescence intensities of **PTQA2** and **PTQA3**, choosing **PTQA2** for sensor performance testing can enhance the sensitivity of fluorescent probe. Then, we studied the sensing properties of **PTQA2** and its nanoparticle (**PTQA-NPs**). The colorimetric sensing properties of **PTQA2** to different metal ions were characterized in the solution of chloroform ( $\text{CHCl}_3$ , 10  $\mu\text{g/mL}$ ). Upon addition of  $\text{Hg}^{2+}$ , shown in Figure 6a, the absorbance at 623 nm and 680 nm were decreased and new absorption peak emergent at 505 nm and 545 nm. Additionally, the shoulder peak at 575 nm was assigned to monothiocarbonyl quinacridone which could be observed in small molecules.<sup>22</sup> The absorption changes of these five peaks were showed in Figure 6b. A small parabolic curve described the absorbance changes at 575 nm indicates generation and disappearance of monothiocarbonyl quinacridone moiety. The magnifying figures showed the characteristic absorbance of thiocarbonyl quinacridone varies with the concentration of  $\text{Hg}^{2+}$  (Figure 6c ~ 6d). As shown in Figures 6c and d, when the concentration of  $\text{Hg}^{2+}$  increased from 0 to 7.6  $\mu\text{M}$ , the absorbance was fade-out at 623 nm and 680 nm and fade-in at 545 nm and 575 nm. In this course, dithiocarbonyl component of **PTQA2** transferred to monothiocarbonyl and dicarbonyl ones. With the addition of  $\text{Hg}^{2+}$  from 7.6 to 14  $\mu\text{M}$ , the peak at 505 nm was emergent and enhanced that indicated generation of quinacridone (Figure 6e). Absorbance at 575 nm decreased in this stage means the monothiocarbonyl component was converted into quinacridone. The peak at 545 nm was still enhanced in this stage and we surmises that it is composed of absorptions of monothiocarbonyl quinacridone and quinacridone. For the same reason, the band at 623 nm is composed of both absorptions of dithiocarbonyl and monothiocarbonyl components that made it

decreased slower than the one at 680 nm. Further investigation of these absorption bands indicated that the dithiocarbonyl quinacridone was first converted to monothiocarbonyl quinacridone and then disulfurized to quinacridone (Scheme 2). Additionally, when  $\text{Hg}^{2+}$  was added to  $\text{CHCl}_3$  solution of **PTQA2**, a dramatic color change from green to brown to orange was observed, indicating that thiocarbonyl quinacridone-based sensor can be used as colorimetric probe for naked eye recognition.

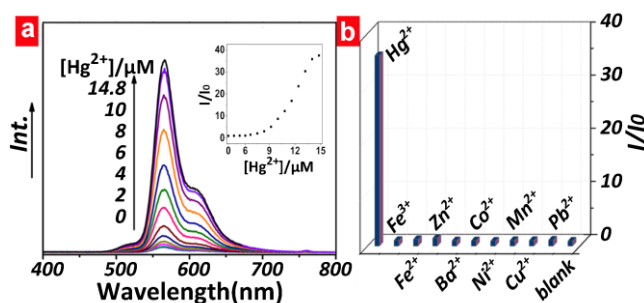


**Figure 6.** (a~d) UV-vis spectra of **PTQA2** (10  $\mu\text{g/mL}$ ) upon the titration of  $\text{Hg}^{2+}$  in  $\text{CHCl}_3$ ; (e) absorbance changes of **PTQA2** with  $\text{Hg}^{2+}$  at different wavelength.



Scheme 2. Sensing mechanism of thiocarbonyl quinacridone-based chemosensors.

Figure 7a showed the fluorescence titration spectra of **PTQA2** with  $\text{Hg}^{2+}$ . When  $\text{Hg}^{2+}$  ion was added to the solution of **PTQA2**, about 36-fold emission enhancement at 566 nm was observed. Moreover, the selective studies of **PTQA2** were recorded in Figure 7b and Figure S3a. 14  $\mu\text{M}$  of  $\text{Hg}^{2+}$  and 40  $\mu\text{M}$  of other metal ions were added to  $\text{CHCl}_3$  solutions of **PTQA2** and only  $\text{Hg}^{2+}$  causes about 35 folds emission enhancement at 566 nm.

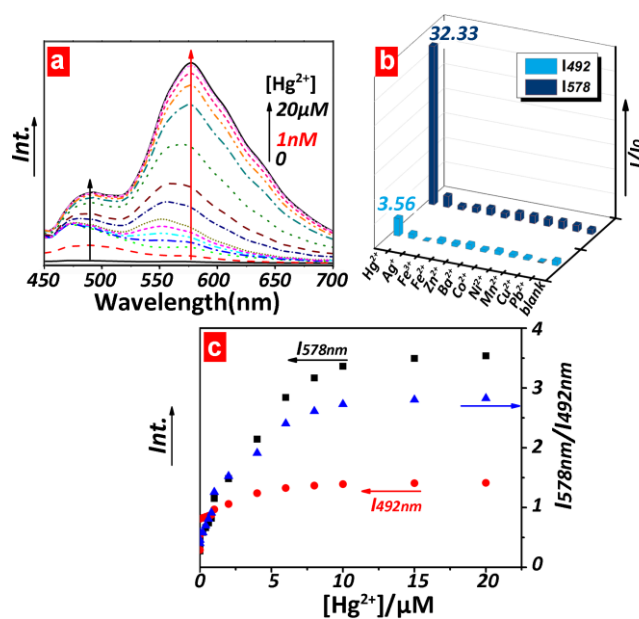


**Figure 7.** (a) Fluorescent spectra of **PTQA2** (10 $\mu\text{g/mL}$ ) upon titration of  $\text{Hg}^{2+}$  in  $\text{CHCl}_3$  solution. inset: Plot of fluorescence intensity change of **PTQA2** ( $\lambda_{\text{ex}} = 380 \text{ nm}$ ,  $\lambda_{\text{em}} = 566 \text{ nm}$ ); (b) Selective properties of **PTQA2** towards different metal ions in  $\text{CHCl}_3$  solution ( $[\text{Hg}^{2+}] = 14 \mu\text{M}$  and other metal ions  $[\text{M}^{n+}] = 40 \mu\text{M}$ ).

Moreover, the sensing property of **PTQA-NPs** was also studied in the PBS solution (see Figure 8). Differently, a weak emission at 492 nm belonged to fluorene was first observed in the solution of **PTQA-NPs** when nM level of  $\text{Hg}^{2+}$  was added in the system. Considering of the



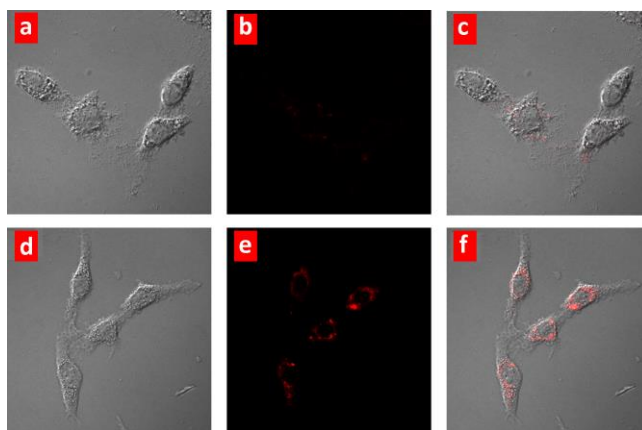
aggregation morphology of this nanoparticle, we presumed that the energy transfer process from fluorene unit to thiocarbonyl quinacridone moiety could be disturbed by a little amount of  $\text{Hg}^{2+}$ . And then, **PTQA-NPs** emitted the fluorescence of fluorene. Upon addition of  $\text{Hg}^{2+}$ , a new emission at 578 nm was emerged and enhanced for the reproduction of quinacridone groups. The increasing emission at 492 nm indicated that the inefficient energy transfer still occurred in the nanoparticles. Good selectivity was also founded in the PBS solution of **PTQA-NPs** (Figure 8b and S3b). With addition of 14  $\mu\text{M}$  of  $\text{Hg}^{2+}$ , the emission intensities enhanced 3.56-fold and 32.33-fold at both 492 nm and 578 nm, respectively. To compare with other metal ions, only  $\text{Hg}^{2+}$  caused distinct fluorescent output. As shown in Figure 8c, the emission intensity of **PTQA-NPs** efficiently increased after 5  $\mu\text{M}$  of  $\text{Hg}^{2+}$  was added, which means the high sensitivity of the probe.



**Figure 8.** (a) Fluorescent spectra of **PTQA-NPs** (10 $\mu\text{g/mL}$ ) upon titration of  $\text{Hg}^{2+}$  in PBS solution; (b) Selective properties of **PTQA-NPs** towards different metal ions in PBS solution ( $[\text{Hg}^{2+}] = 14 \mu\text{M}$  and other metal ions  $[\text{M}^{\text{n}+}] = 40 \mu\text{M}$ ); (c) Plot of fluorescence intensity change of **PTQA-NPs**.

### Monitoring Hg<sup>2+</sup> in Living Cells

After in vitro experiments, we demonstrated the applicability of **PTQA-NPs** in monitoring intracellular Hg<sup>2+</sup>. The bioimaging experiments were carried out by confocal laser scanning microscopy (CLSM). As shown in Figure 9, HeLa cells incubated with **PTQA-NPs** (10 µg/mL) for 1 h at 37 °C showed no emission. When the cells were supplemented with **PTQA-NPs** in the growth medium for 1 h at 37 °C and then incubated with 10 µM Hg<sup>2+</sup> for 20 min at 37 °C, a strong enhancement in the red emission was observed in the intracellular region. Bright-field measurements confirmed that the HeLa cells with or without treatment with Hg<sup>2+</sup> remained viable throughout the imaging experiments. Furthermore, overlay of luminescence and bright field images demonstrated that the red luminescence was evident in the cytoplasm over the nucleus and membrane, which was also confirmed by xz cross-sectional image (Figure S4).



**Figure 9.** CLSM images in HeLa cells. (Top, a-c) image of HeLa cells incubated with 10 µg/mL **PTQA-NPs** for 60 min at 37 °C. (a) bright field image; (b) red channel image, and (c) overlay of (a), (b). (Bottom, d-f) image of HeLa cells stained with 10 µg/mL **PTQA-NPs** for 60 min at 37 °C, then further treated with 10 µM Hg<sup>2+</sup> at 37 °C for 20 min. (d) brightfield image; (e) red channel image and (f) overlay of (d), (e). Emission was collected by red channel from 560-660 nm,  $\lambda_{\text{ex}} = 488$  nm.

## Conclusion

In summary, we have demonstrated a series of highly sensitive FRET-based thiocarbonyl quinacridone conjugated polymers for sensing and bioimaging of  $\text{Hg}^{2+}$  ion. The thiocarbonyl quinacridone chromophore, a useful  $\text{Hg}^{2+}$  sensor, was successfully conjugated with fluorene as an energy donor. For **PTQA2** in chloroform, the addition of mercury ion results in green to brown to orange absorption color change and more than 30-fold fluorescence enhancement at visible region wavelengths. Moreover, this conjugated polymer system could be used for detection of  $\text{Hg}^{2+}$  in aqueous solution with high sensitivity, which can detect 1 ppb  $\text{Hg}^{2+}$  in the PBS solution of **PTQA-NPs**. This concentration was lower than the blood level (5.8 ppb) set by the U. S. EPA. Our data confirm that thiocarbonyl quinacridone-based conjugated polymer system is a viable fluorescent probe for  $\text{Hg}^{2+}$  in biosystem.

## Acknowledgements:

This work was supported by NSFC/China (21372082, 2116110444 and 21172073), and the National Basic Research 973 Program (2013CB733700 and 2013CB834701).

## References:

1. J. Kaiser, *Science*, 1998, **279**, 1850–1851.
2. D. Zeller and S. Booth, *Science*, 2005, **310**, 777–779.
3. S. E. Bryan, C. Lambert, K. J. Hardy and S. Simons, *Science*, 1974, **186**, 832–833.
4. A. Curley, V. A. Sedlak, E. F. Girling, R. E. Hawk, W. F. Barthel, P. E. Pierce and W. H. Likosky, *Science*, 1971, **172**, 65–67.
5. J. W. Spann, R. G. Heath, J. F. Kreitzer and L. N. Locke, *Science*, 1972, **175**, 328–331.

6. A. H. Stern, *Science*, 2004, **303**, 763–766.
7. V. K. Bains, K. Loomba, A. Loomba and R. Bains, *Br Dent J*, 2008, **205**, 373–378.
8. Q. Xu, S. Lee, Y. Cho, M. H. Kim, J. Bouffard and J. Yoon, *J. Am. Chem. Soc.*, 2013, **135**, 17751–17754.
9. J. Y. Jung, M. Kang, J. Chun, J. Lee, J. Kim, Y. Kim, S.-J Kim, C. Lee and J. Yoon, *Chem. Commun.*, 2013, **49**, 176–178.
10. S. Xu, B. Hu, S. E. Flower, Y.-B. Jiang, J. S. Fossey, W.-P. Deng and T. D. James, *Chem. Commun.*, 2013, **49**, 8314–8316.
11. K. Lawrence, S. E. Flower, G. Kociok-Kohn, C. G. Frost and T. D. James, *Anal. Methods-UK*, 2012, **4**, 2215–2217.
12. X. Zhou, H. Li, H. Xiao, L. Li, Q. Zhao, T. Yang, J. Zuo and W. Huang, *Dalton Trans.*, 2013, **42**, 5718–5723.
13. H. Son, J. H. Lee, Y.-R Kim, I. S. Lee, S. Han, X. Liu, J. Jaworski and J. H. Jung, *Analyst*, 2012, **137**, 3914–3916.
14. Q. Liu, J. Peng, L. Sun, and F. Li, *ACS Nano*, 2011, **5**, 8040–8048.
15. Y. Yang, Q. Zhao, W. Feng and F. Li, *Chem. Rev.*, 2013, **113**, 192–270.
16. J. Zhang, Y. Zhou, J. Yoon and J. Kim, *Chem. Soc. Rev.*, 2011, **40**, 3416–3429.
17. H. Kim, Z. Guo, W. Zhu, J. Yoon and H. Tian, *Chem. Soc. Rev.*, 2011, **40**, 79–93.
18. X. Chen, T. Pradhan, F. Wang, J. Kim and J. Yoon, *Chem. Rev.*, 2012, **112**, 1910–1956.
19. Y. Liu, M. Chen, T. Cao, Y. Sun, C. Li, Q. Liu, T. Yang, L. Yao, W. Feng and F. Li, *J. Am. Chem. Soc.*, 2013, **135**, 9869–9876.

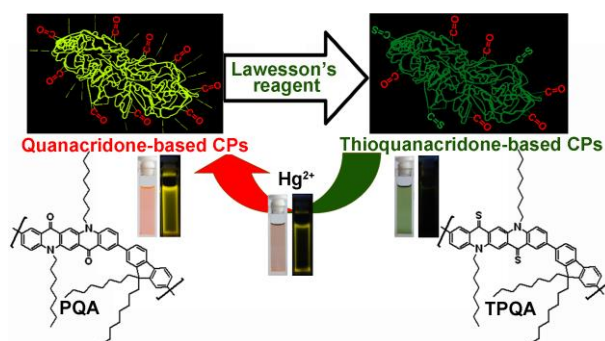
20. W. H. Ma, Q. Xu, J. J. Du, B. Song, X. J. Peng, Z. Wang, G. D. Li and X. F. Wang, *Spectrochim. Acta, Part A*, 2010, **76**, 248–252.
21. K. C. Song, J. S. Kim, S. M. Park, K. C. Chung, S. Ahn and S. K. Chang, *Org. Lett.*, 2006, **8**, 3413–3416.
22. G. Zhang, D. Zhang, S. Yin, X. Yang, Z. Shuai and D. Zhu, *Chem. Commun.*, 2005, **41**, 2161–2163.
23. H. Li, C. Gu, L. Jiang, L. Wei, W. Hu and H. Fu, *J. Mater. Chem. C*, 2013, **1**, 2021–2027.
24. Y. Qu, J. Yang, J. Hua and L. Zou, *Sensor Actuat. B-Chem.*, 2012, **161**, 661–668.
25. C. A. Traina, R. C. Bakus II and G. C. Bazan, *J. Am. Chem. Soc.*, 2011, **133**, 12600–12607.
26. X. Zhou, H. Li, H. Xiao, L. Li, Q. Zhao, T. Yang, J. Zuo and W. Huang, *Dalton Trans.*, 2013, **42**, 5718–5723.
27. J. Song, J. Zhang, F. Lv, Y. Cheng, B. Wang, L. Feng, L. Liu and S. Wang, *Angew. Chem. Int. Ed.*, 2013, **52**, 13020–13023.
28. J. Liu, Y. Zhong, P. Lu, Y. Hong, J. W. Y. Lam, M. Faisal, Y. Yu, K. S. Wong and B. Z. Tang, *Polym. Chem.*, 2010, **1**, 426–429.
29. R. Hu, J. W. Y. Lam, J. Liu, H. H. Y. Sung, I. D. Williams, Z. Yue, K. S. Wong, M. M. F. Yuen and B. Z. Tang, *Polym. Chem.*, 2012, **3**, 1481–1489.
30. C. Zhu, Q. Yang, L. Liu and S. Wang, *Chem. Commun.*, 2011, **47**, 5524–5526.
31. L. Feng, C. Zhu, H. Yuan, L. Liu, F. Lv and S. Wang, *Chem. Soc. Rev.*, 2013, **42**, 6620–6633.

32. Y. Yuan, R. T. K. Kwok, G. Feng, J. Liang, J. Geng, B. Z. Tang and B. Liu, *Chem. Commun.*, 2014, DOI: 10.1039/C3CC47585A.
33. J. Geng, K. Li, D. Ding, X. Zhang, W. Qin, J. Liu, B. Z. Tang and B. Liu, *Small*, 2012, **8**, 3655–3663.
34. E. Zhao, H. Li, J. Ling, H. Wu, J. Wang, S. Zhang, J. W. Y. Lam, J. Z. Sun, A. Qin and B. Z. Tang, *Polym. Chem.*, 2013, DOI: 10.1039/C3PY01387A.
35. G. Wang, H. Yin, J. C. Y. Ng, L. Cai, J. Li, B. Z. Tang and B. Liu, *Polym. Chem.*, 2013, **4**, 5297–5304.
36. Z. Yang, Y. Yuan, R. Jiang, N. Fu, X. Lu, C. Tian, W. Hu, Q. Fan and W. Huang, *Polym. Chem.*, 2014, DOI: 10.1039/C3PY01197F.
37. A. Levai and J. Jekő, *J. Heterocyclic Chem.*, 2005, **42**, 739–742.
38. J. Liu, B. Gao, Y. Cheng, Z. Xie, Y. Geng, L. Wang, X. Jing and F. Wang, *Macromolecules*, 2008, **41**, 1162–1167.
39. S. W. Thomas, G. D. Joly and T. M. Swager, *Chem. Rev.*, 2007, **107**, 1339–1386.
40. S. Rochat and T. M. Swager, *ACS Appl. Mater. Interfaces*, 2013, **5**, 4488–4502.

## For Table of Contents use only

Fluorescent conjugated polymer based on thiocarbonyl quinacridone for sensing mercury ion and bioimaging

Yi Qu,<sup>a,b</sup> Xinran Zhang,<sup>a</sup> Yongquan Wu,<sup>b</sup> Fuyou Li<sup>b\*</sup> and Jianli Hua<sup>a\*</sup>



Highly sensitive FRET-based thiocarbonyl quinacridone fluorescent conjugated polymers for sensing and bioimaging of mercury ion have been developed.