Organic & Biomolecular **Chemistry**

Accepted Manuscript

This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](http://www.rsc.org/Publishing/Journals/guidelines/AuthorGuidelines/JournalPolicy/accepted_manuscripts.asp).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](http://www.rsc.org/help/termsconditions.asp) and the Ethical quidelines still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

www.rsc.org/obc

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

ARTICLE TYPE

Multi-substituted 8-Aminoimidazo[1,2-*a***]pyrazines by Groebke-Blackburn-Bienaymé Reaction and Their Hsp90 Inhibitory Activity**

Jing Ren,1,# Min Yang, 2,# Hongchun Liu,³ Danyan Cao,¹ Danqi Chen,¹ Jian Li,² Le Tang,¹ Jianhua He,² Yue-Lei Chen, 1,* Meiyu Geng,³ Bing Xiong, 1,* and Jingkang Shen1,*

⁵*Received (in XXX, XXX) Xth XXXXXXXXX 20XX, Accepted Xth XXXXXXXXX 20XX* **DOI: 10.1039/b000000x**

Using 2,3-diamino pyrazine substrate and yttrium triflate catalyst, various 2-alkyl and aryl substituted 3,8 diaminoimidazo[1,2-*a***]pyrazines could be efficiently prepared**

- ¹⁰**through Groebke-Blackburn-Bienaymé MCR. In particular, a novel 2-piperonyl 3,8-diaminoimidazo[1,2-***a***]pyrazine structure could be prepared exclusively with this new method and was found with moderate Hsp90 inhibitory activity. Crystal complex with N-terminus ATP domain of Hsp90 and**
- ¹⁵**one of the new Hsp90 inhibitors was also obtained to elucidate the activity origin of 2-piperonyl 3,8-diaminoimidazo[1,2** *a***]pyrazines.**

Introduction

8-Aminoimidazo[1,2-*a*]pyrazines **I** are highly useful for ²⁰medicinal chemistry research, for it's obvious similarity with adenine **II**, which is ubiquitous in bioactive structures. Consequently, installation of different substituents on 8 aminoimidazo[1,2-*a*]pyrazine ring system become an intriguing topic, and this usually could be achieved via certain ring

- 25 formation reactions from substituted fragments.¹ Preparation of multi-substituted 8-aminoimidazo[1,2-*a*]pyrazine ring system is more demanding and less reported. It is known that multicomponent reactions (MCRs) are advantageous in the syntheses of heavily substituted heterocycles. However, syntheses
- ³⁰of multi-substituted 8-aminoimidazo[1,2-*a*]pyrazines through MCR pathway were rarely discussed and seemed to be less efficient: For example, David et al.² attempted the preparation of 2-substituted 3,8-diaminoimidazo[1,2-*a*]pyrazines through Groebke-Blackburn-Bienaymé MCR³ and obtained poor yields.
- 35 Interestingly, one recent article by Pirali *et al.*⁴ described more efficient syntheses of 2-substituted 3,8-diaminoimidazo[1,2 *a*]pyrazines **VI** starting from chloropyrazine **1**, isocyanides **III** and aldehydes **IV** via two step sequences including Groebke-Blackburn-Bienaymé MCR and the following aminolysis of
- ⁴⁰intermediate **V**. Nevertheless, it was found that only aryl carboxaldehydes were productive with the reported condition, and this meant that the R^2 on 2-substituted 3,8diaminoimidazo[1,2-*a*]pyrazine products **VI** could only be aryl groups (Scheme 1).

Scheme 1. A comparison of imidazo[1,2-*a*]pyrazines **I** and adenine **II** and one of the known synthetic method 4 to 8-aminoimidazo[1,2-*a*]pyrazines **VI**. Reaction conditions: a) compound **1** (1 equiv), R^2CHO (1.2 equiv), MeCN, reflux, 2 h; b) TMSCl (1.2 equiv), MeCN/DCM, 30 min; c) $R¹NC$ ⁵⁰(1.2 equiv), reflux, overnight; d) aq. NH4OH/dioxane, 100 ◦C, overnight.

During our study of Groebke-Blackburn-Bienaymé MCR, ⁵ a serendipitous experiment indicated that 2,3-diaminopyrazine was a superior substrate for this MCR leading to 2-aryl, 2-benzyl, and 2-alkyl 3,8-diaminoimidazo[1,2-*a*]pyrazines. Using this efficient ⁵⁵methodology, a panel of 2-piperonyl substituted 3,8 diaminoimidazo[1,2-*a*]pyrazines could be prepared for the first time. Some of 2-piperonyl substituted 3,8-diaminoimidazo[1,2 *a*]pyrazines were found with moderate Heat Shock Protein 90 (Hsp90) inhibitory activity, and could serve as leads for further ⁶⁰optimization. These works will be described herein.

Results and discussion

diaminoimidazo[1,2-*a*]pyrazines.

Initially, using yttrium triflate catalyst, it was found that the *N*-2,4-dimethoxybenzyl (*N*-DMB) protected diamino pyrazine **2** reacted with isocyanide **3** and phenyl acetaldehyde **4** to give the 65 desired product **5** with good yield, after removing the *N*protection with acid (Table 1, entry 1). This was unique since in comparison experiments, TMSCl⁴ failed to give the conversion and TfOH only gave low yield (Table 1, entry 2-3). Changing from diamino pyrazine **2** back to chloropyrazine **1** again failed to ⁷⁰give the reaction with TMSCl, and this was consistent with the findings by Pirali *et al.*⁴ (Table 1, entry 4). With TfOH or rare earth-metal triflate catalysts, some product **5** could be observed after aminolysis, however the yield was low (Table 1, entry 5-7). As phenyl acetaldehyde **4** is a special type of alkyl aldehyde, 75 above discovery served as a successful example of Groebke-Blackburn-Bienaymé MCR leading to 2-alkyl 3,8-

[a] Reaction conditions: For entry 1 and 3: a) compound **2** (1.0 equiv), compound **3** (1.2 equiv), compound **4** (1.2 equiv), catalyst (10 mol%), ⁵MeOH, 70 °C, 5 h; b) TFA, DCM, 50 °C, 1 h. For entry 2 and 4: a) compound **4** (1.2 equiv), MeCN, 100 °C, 2 h; b) TMSCl (1.2 equiv), MeCN/DCM, rt, 30 min; c) compound **3** (1.2 equiv), 100 °C, 12 h. For entry 5, 6, and 7: a) compound **1** (1.0 equiv), compound **3** (1.2 equiv), compound **4** (1.2 equiv), catalyst (10 mol%), MeOH, 100 °C, 12 h; b) aq. 10 NH₄OH/dioxane, sealed tube, 120 ℃, 24 h. [b] Yield was determined after flash chromatography.

To further explore the scope of the yttrium triflate catalyzed synthesis of 2-substituted 3,8-diaminoimidazo[1,2-*a*]pyrazines, ¹⁵different substrates, including pyrazines **6-8**, isocyanides **9-11**, as well as aldehydes **4** and **12-16**, were subjected to the new reaction condition described in Table 1, entry 1 (Scheme 2). For common electron rich substituted phenyl acetaldehydes, the yields were moderate to good with different isocyanides. Since the yields

- ²⁰were still lower than that with phenyl acetaldehyde **4** (Table 1, entry 1), it might be speculated that the electron donating groups on the phenyl ring of the aldehyde component reduced the yield. Retaining phenyl acetaldehyde **4**, chloro-substituted pyrazine **6** gave product **20** again with good yield. Above examples (for
- ²⁵products **17**-**21**) were all based on *N*-DMB protected pyrazines **6**, and several points were noteworthy: 1) Compared to chloropyrazine **1** starting material, *vic*-diamino substitution on diamino pyrazine **2** could significantly improve the MCR yield with various isocyanide and aldehyde components; 2) As
- ³⁰illustrated in the general reaction schemes in Scheme 2 and Table 1, deprotection of *N* 8 -DMB on 3,8-diaminoimidazo[1,2 *a*]pyrazine intermediate (formed from pyrazine **2**) gave better yield than aminolysis of 8-Cl on 3,8-diaminoimidazo[1,2*a*]pyrazine intermediate (formed from chloropyrazine **1**), 35 rendering the syntheses of 2-substituted 3,8-diaminoimidazo[1,2-
- *a*]pyrazines more efficient.

 Next, morpholine substituted pyrazine **7** reacted with acetal aldehyde **15** and different isocyanides to give products **22** and **23** with good yields (Scheme 2). This was not trivial, since simple ⁴⁰aldehydes such as **15** used to be poor partner for this type of reaction. Meanwhile, pyrazines **7** and **8** could react with aryl carboxaldehydes **14** and **16** as well to give the desired 2 substituted 3,8-diaminoimidazo[1,2-*a*]pyrazines **24-26** (Scheme

2). The reactions with morpholine substituted pyrazine **7** and *N*-⁴⁵cyclohexyl pyrazine **8** were highly practical for the following reasons: 1) The diamino substituted pyrazine substrates improved the MCR yield substantially, with both alkyl aldehyde and aryl carboxaldehyde; 2) It eliminated the necessity of *N*-deprotection, and yielded more interesting *N*-alkylated 2-substituted 3,8- ⁵⁰diaminoimidazo[1,2-*a*]pyrazines; 3) Morpholine or alkylated amino group was widely used in medicinal chemistry.

For Groebke-Blackburn-Bienaymé MCR, there might be several reasons accounting for the yield improvement using diamino substituted pyrazine substrates and yttrium triflate ⁵⁵catalyst: 1) *vic*-Diamino substitution could enhance the nucleophilicity of pyrazine substrates and result in improved condensation reaction; 2) *vic*-Diamino substitution could provide a better chelation environment for yttrium metal, ⁶ which was an essential catalyst for Groebke-Blackburn-Bienaymé MCR. This ⁶⁰might also explain why TMSCl failed to work with diamino substituted pyrazine substrates.

General scheme of the reaction: [a]

Scheme 2. New condition for Groebke-Blackburn-Bienaymé MCR led to various multisubstituted 3,8-diaminoimidazo[1,2-*a*]pyrazines. [a] ⁵Reaction conditions: For products **17-21**: a) pyrazines (1.0 equiv), isocyanides (1.2 equiv), aldehydes (1.2 equiv), $Yb(OTf)$ ₃ (10 mol%), MeOH, 70 °C, 5 h; b) TFA, DCM, 50 °C, 1 h. For products **22-26**: pyrazines (1.0 equiv), isocyanides (1.2 equiv), aldehydes (1.2 equiv), Yb(OTf)₃ (10 mol %), MeOH, 70 °C, 5 h. [b] Yield was determined after 10 flash chromatography.

On the basis of above discovery, we looked for the applications in medicinal chemistry. It is known that Hsp90 belongs to a large family of heat shock proteins, which are evolutionarily conserved and required for essential housekeeping functions such as protein 15 folding, assembly, and transportation.⁷ Discovering new Hsp90 inhibitors is an important approach combating cancer. ⁸ We noticed that some ATP-competitive Hsp90 inhibitors contains 8,9-disubstituted adenine motif, 9 which might be replaced with 2,3-disubstituted 8-aminoimidazo[1,2-*a*]pyrazine scaffold, ²⁰according to the structure similarity between adenine and 8-

aminoimidazo[1,2-*a*]pyrazine. Further computational study using scaffold hopping concept also suggested that our new product **19** or similarly substituted 8-aminoimidazo[1,2-*a*]pyrazines might possess Hsp90 inhibitory activity due to the remarkable 25 resemblance to PU-H71, 10 an Hsp90 inhibitor in Phase I clinical trial (Scheme 3).

Scheme 3. The structure similarity between compound **19** and a known Hsp90 inhibitor PU-H71.

30

To test the hypothesis, a focused library of 8 aminoimidazo[1,2-*a*]pyrazines bearing 2-piperonyl and 3-alkyl amino substitutions was prepared using the new methodology (Scheme 4). With diamino pyrazine **2** and piperonals **27** and **28**, ³⁵moderate to good yields were documented for products **29**-**32**. When chloro substituted diamino pyrazine **6** was used, the yields

were again good for products **34** and **35**, and this was consistent with the observations in Scheme 2, conversions from **6**, **3**, and **4** to **22**.

The inhibitory activity of the library against Hsp90 was measured and analyzed, too (Scheme 4): 1) The halogenated 2 piperonyl substitution was found essential for Hsp90 binding, as suggested by the activity comparison of compounds **19**, **29**, and **30**: Compared to compound **19**, the bromine atom on compound ⁴⁵**29** dramatically increased the activity to 5.34 µM, while the iodine on compound **30** further improves the value to 2.95 µM. 2) The substitution on C3 of 8-aminoimidazo[1,2-*a*]pyrazine scaffold affected the Hsp90 affinity, too, as demonstrated by the activities of compounds **29**, **31**, **32**, and **33**. The 2-*tert*-butyl ⁵⁰amine group on compound **31** or the 2-cyclohexyl amino group on compound **32** may not be long enough to reach the van der Waals surface of Hsp90 protein, and resulted in decreased activtities. 3) The 6-chloro substitution on 8-aminoimidazo[1,2 *a*]pyrazine was found destructive to Hsp90 inhibitory activity, ⁵⁵and this was demonstrated by the data of compounds **34** and **35**. Generally, the moderate inhibitory activity of 2-piperonyl and 3 alkyl amino 8-aminoimidazo[1,2-*a*]pyrazines suggested that this new scaffold could serve as a promising starting point for further structural optimization.

60 General scheme of the reaction: [a]

65

Scheme 4. Synthesis and activity study of a Hsp90 targeted focused library. [a] Reaction conditions: a) pyrazines (1.0 equiv), isocyanides (1.2 equiv), aldehydes (1.2 equiv), Yb(OTf)₃ (10 mol%), MeOH, 70 °C, 5 h; ⁷⁰b) TFA, DCM, 50 °C, 1 h. [b] Yield was determined after flash chromatography. [c] See supporting information for details.

For better understanding the interactions between 2,3 disubstituted 8-aminoimidazo[1,2-*a*]pyrazine and Hsp90 binding site, compound **29** was cocrystallized with N-terminus ATP ⁵domain of Hsp90 and the co-crystal structure was successfully solved (pdb entry: $4R3M$).¹¹ As shown in Figure 1, in Hsp90 Nterminus ATP domain, the 8-aminoimidazo[1,2-*a*]pyrazine ring was situated at the same position as the adenine ring of PU-H71(Figure 1B). The 8-aminoimidazo[1,2-*a*]pyrazine ring system 10 formed several indirect hydrogen bonds with the binding site of

- Hsp90 through four conserved water molecules (red spheres in Figure 1), and two direct hydrogen bonds with ASP93 and THR184 residues. The 2-piperonyl substitution showed typical π - π stacking interaction with PHE138 residue, and the bromine on
- 15 2-piperonyl substitution was close to GLY135 residue on the Hsp90 backbone. A halogen bond with the oxygen atom of GLY135 residue was thus speculated for the short Br-O distance of 3.4 Å. In addition, compared to the PU-H71 structure, discrepancy between the orientations of 3-benzyl amino group on
- ²⁰compound **29** and 9-alkyl amino group on PU-H71 could be observed. This might account partially for the activity difference between these two compounds.
	- A:

Figure 1. A: Co-crystal structure of Hsp90 with compound **29 (**pdb entry: 4R3M**)**. B: superimposition of crystal structures **29-** $Hsp90^{11}$ and PU-H71-Hsp90 (pdb entry: 2FWZ). Protein was 30 shown in cartoon style, and the crystal water molecules were shown in red spheres. The carbon atoms of compound **29** were in green, and that of PU-H71 in blue.

Conclusions

In conclusion, with 2,3-diamino pyrazine substrate and yttrium ³⁵triflate catalyst, various 2-alkyl and aryl substituted 3,8 diaminoimidazo[1,2-*a*]pyrazines could be efficiently prepared through Groebke-Blackburn-Bienaymé MCR. Elimination of the chloride aminolysis step made this method more favorable compared with previous synthetic approaches. We further ⁴⁰demonstrate that 2-piperonyl 3,8-diaminoimidazo[1,2-*a*]pyrazine structures, prepared exclusively with our new synthetic method, were potential Hsp90 inhibitors. Crystal complex with Nterminus ATP domain of Hsp90 and one of the new Hsp90 inhibitors was also obtained to elucidate the activity origin of 2- 45 piperonyl 3,8-diaminoimidazo $[1,2-a]$ pyrazines. The new scaffold, moderate preliminary biological activity, and crystal structure details suggested together that 2-piperonyl 3,8 diaminoimidazo[1,2-*a*]pyrazines were promising leads for Hsp90 inhibitor discovery.

⁵⁰**Acknowledgements**

This work was supported by the National Natural Science Foundation of China (81102306, 81273368, 81402850) and National Science & Technology Major Project "Key New Drug Creation and Manufacturing Program", China (2012ZX09301001, ⁵⁵2014ZX09507-002).

Notes and references

2 ⁶⁰*Shanghai Institute of Applied Physics, Chinese Academy of Sciences, 239 Zhang Heng Road, Shanghai 201203, P. R. China.*

³Division of Antitumor Pharmacology, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 555 Zuchongzhi Road, Shanghai 201203, P. R. China.

65 † Electronic Supplementary Information (ESI) available: Experimental details, crystallographic data, and copies of NMR spectra. See DOI: 10.1039/b000000x/

⁷⁰1. *(a)* T. L. Gilchrist, *Heterocyclic Chemistry* (3rd Edition), Prentice Hall, **1997**; *(b)* J. A. Joule, K. Mills, *Heterocyclic Chemistry* (4th Edition), Blackwell Science Ltd., **2000**; For some recent examples, see: *(c)* H. Zeng, D. B. Belanger, P. J. Curran, G. W. Shipps, Jr., H. Miao, J. B. Bracken, M. A. Siddiqui, M. Malkowski and Y. Wang, ⁷⁵*Bioorg. Med. Chem. Lett*., 2011, **21**, 5870-5875; *(d)* S. Martinez Gonzalez, A. I. Hernandez, C. Varela, M. Lorenzo, F. Ramos-Lima, E. Cendon, D. Cebrian, E. Aguirre, E. Gomez-Casero, M. I. Albarran, P. Alfonso, B. Garcia-Serelde, G. Mateos, J. Oyarzabal, O. Rabal, F. Mulero, T. Gonzalez-Granda, W. Link, J. Fominaya, M. Barbacid, J. ⁸⁰R. Bischoff, P. Pizcueta, C. Blanco-Aparicio and J. Pastor, *Bioorg. Med. Chem. Lett*., 2012, **22**, 5208-5214; *(e)* S. Martinez Gonzalez, A. I. Hernandez, C. Varela, S. Rodriguez-Aristegui, R. M. Alvarez, A. B. Garcia, M. Lorenzo, V. Rivero, J. Oyarzabal, O. Rabal, J. R. Bischoff,

¹Medicinal Chemistry Department, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 555 Zuchongzhi Road, Shanghai 201203, P. R. China. E-mail: bxiong@simm.ac.cn

M. Albarran, A. Cebria, P. Alfonso, W. Link, J. Fominaya and J. Pastor, *Bioorg. Med. Chem. Lett*., 2012, **22**, 1874-1878; *(f)* S. Martinez Gonzalez, A. I. Hernandez, C. Varela, S. Rodriguez-Aristegui, M. Lorenzo, A. Rodriguez, V. Rivero, J. I. Martin, C. G.

- ⁵Saluste, F. Ramos-Lima, E. Cendon, D. Cebrian, E. Aguirre, E. Gomez-Casero, M. Albarran, P. Alfonso, B. Garcia-Serelde, J. Oyarzabal, O. Rabal, F. Mulero, T. Gonzalez-Granda, W. Link, J. Fominaya, M. Barbacid, J. R. Bischoff, P. Pizcueta and J. Pastor, *Bioorg. Med. Chem. Lett*., 2012, **22**, 3460-3466; *(g)* J. M. Bartolome-
- 10 Nebreda, F. Delgado, M. L. Martin-Martin, C. M. Martinez-Viturro, J. Pastor, H. M. Tong, L. Iturrino, G. J. Macdonald, W. Sanderson, A. Megens, X. Langlois, M. Somers, G. Vanhoof and S. Conde-Ceide, *J. Med. Chem*., 2014, **57**, 4196-4212.
- 2. *(a)* D. B. Salunke, E. Yoo, N. M. Shukla, R. Balakrishna, S. S. 15 Malladi, K. J. Serafin, V. W. Day, X. Wang and S. A. David, *J. Med. Chem*., 2012, **55**, 8137-8151. *(b)* N. M. Shukla, D. B. Salunke, E. Yoo, C. A. Mutz, R. Balakrishna and S. A. David, *Bioorg. Med. Chem.*, 2012, **20**, 5850-5863.
- 3. *(a)* H. Bienaymé and K. Bouzid, *Angew.Chem., Int. Ed.*, 1998, **110**, ²⁰2349-2352; *(b)* C. Blackburn, B. Guan, P. Fleming, K. Shiosaki and S. Tsai, *Tetrahedron Lett*., 1998, **39**, 3635-3638; *(c)* K. Groebke, L. Weber and F. Mehlin, *Synlett*., 1998, 661-663.
- 4. M. Guasconi, X. Lu, A. Massarotti, A. Caldarelli, E. Ciraolo, G. C. Tron, E. Hirsch, G. Sorba and T. Pirali, *Org. Biomol. Chem*., 2011, **9**, ²⁵4144-4149.
- 5. *(a)* T. Meng, Z. Zhang, D. Hu, L. Lin, J. Ding, X. Wang and J. Shen, *J. Comb. Chem*., 2007, **9**, 739-741; *(b)* H. Sun, H. Zhou, O. Khorev, R. Jiang, T. Yu, X. Wang, Y. Du, Y. Ma, T. Meng and J. Shen, *J. Org. Chem*., 2012, **77**, 10745-10751; *(c)* H. Zhou, W. Wang, O. ³⁰Khorev, Y. Zhang, Z. Miao, T. Meng and J. Shen, *Eur. J. Org. Chem*., 2012, 5585-5594.
- 6. S. Kobayashi, in *Lanthanides: Chemistry and Use in Organic Synthesis*, ed. S. Kobayashi, Springer Berlin Heidelberg, 1999, 63- 118.
- ³⁵7. *(a)* M. Feder, G. Hofmann, *Annu Rev Physiol*., 1999, **61**, 243-282; *(b)* D. Picard, *Cell Mol Life Sci*., 2002, **59**, 1640-1648; *(c)* C. Garrido, S. Gurbuxani, L. Ravagnan, G. Kroemer, *Biochem. Biophys. Res. Commun.,* 2001, **286**, 433-442.
- 8. *(a)* M. Hwang, L. Moretti, B. Lu, *Curr Med Chem*., 2009, **16**, 3081- ⁴⁰3092; *(b)* W. Xu, L. Neckers, *Clin Cancer Res* 2007, **13**, 1625-1629; *(c)* J. Ren, J. Li, Y. Wang, W. Chen, A. Shen, H. Liu, D. Chen, D.
- Cao, Y. Li, N. Zhang, Y. Xu, M. Geng, J. He, B. Xiong, J. Shen, *Bioorg. Med. Chem. Lett*., 2014, **24**, 2525-2529; *(d)* R. Bhat, S. R. Tummalapalli and D. P. Rotella, *J. Med. Chem.* 2014, DOI: ⁴⁵10.1021/jm500823a.
- 9. *(a)* L. Llauger, H. He, J. Kim, J. Aguirre, N. Rosen, U. Peters, P. Davies, G. Chiosis, *J. Med. Chem.,* 2005, **48**, 2892-2905; *(b)* S.-H. Kim, A. Bajji, R. Tangallapally, B. Markovitz, R. Trovato, M. Shenderovich, V. Baichwal, P. Bartel, D. Cimbora, R. McKinnon, R.
- ⁵⁰Robinson, D. Papac, D. Wettstein, R. Carlson, K. M. Yager, *J. Med. Chem.,* 2012, **55**, 7480-7501.
	- 10. H. He, D. Zatorska, J. Kim, J. Aguirre, L. Llauger, Y. She, N. Wu, R. M. Immormino, D. T. Gewirth, G. Chiosis, *J. Med. Chem*., 2005, **49**, 381-390.
- ⁵⁵11. See supporting information for details.