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1 **Silver nanoparticles: green synthesis, self-assembly**  
2 **nanostructure and its application as SERS substrate**

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18 **Silver nanoparticles: green synthesis, self-assembly**  
19 **nanostructure and its application as SERS substrate**

20

21 **Abstract**

22 In this paper, silver nanoparticles were synthesized using citrus peel extracts such as  
23 *Citrus sinensis* (orange fruit, AgNP-Ora), *Citrus reticulata* (tangerine fruit, AgNP-Tan),  
24 and *Citrus aurantiflora* (lemon fruit, AgNP-Lem). The absorption spectra of the AgNP-  
25 Ora and AgNP-Tan colloids show localized surface plasmon band (LSPR) at 445 and  
26 423 nm, respectively. For the same synthesis method, a low intensity of the LSPR band  
27 for AgNP-Lem is observed, indicating a low yield of this reaction. However, the  
28 transmission electron microscopy images show that the colloid prepared using lemon  
29 extract also presents AgNP larger than 5 nm (5 - 55 nm), which exhibit plasmonic  
30 properties. The synthesized silver nanoparticles are spherical-like shape and are highly  
31 crystalline, and they were self-assembled on NH<sub>2</sub>-modified glass slides, obtained AgNP  
32 aggregates substrate for all three prepared silver colloids. We also explored the SERS  
33 activity of the AgNP substrates using 10<sup>-6</sup> mol L<sup>-1</sup> solutions of 4-aminebenzenethiol,  
34 rhodamine 6G, and methylene blue as Raman probe molecules. It was possible to  
35 detected with high signal-to-noise the SERS spectral pattern of all probe molecules on  
36 AgNP substrates. This simple, low cost, and greener method for synthesize silver  
37 nanoparticles may be valuable in future works about SERS sensors development and be  
38 extend to catalytic applications.

39

40 **Keywords:** Green synthesis, Ag nanoparticles, plasmonics properties, SERS activity.

## 41 **1. Introduction**

42 Silver nanoparticles have had a substantial impact across a diverse range of  
43 fields such as catalysis,<sup>1,2</sup> sensing,<sup>3,4</sup> metallic inks,<sup>5,6</sup> and medicine.<sup>7,8</sup> Silver has  
44 desirable physical properties, good relative abundance and low-cost, which make it an  
45 attractive material for several applications.<sup>9</sup> Also, it well known that silver nanoparticles  
46 have different optical and electromagnetic properties from its bulk material owing to  
47 quantum sizes and surface effects.<sup>9,10</sup> Recently, a large amount of works have been  
48 developed to study the optical properties of noble metal nanoparticles, including silver  
49 nanoparticles, which could support localized surface plasmon resonance (LSPR).<sup>11-15</sup>  
50 LSPR are electromagnetic modes associated with the excitation of collective  
51 oscillations of the electronic charge density in metals. The oscillation frequency is  
52 determined by some factors, such as density of electrons, electron mass, size and shape  
53 of the charge distribution in the nanostructure. Several unique properties can be  
54 achieved when adjusting the structure size, morphology, and composition of the metal  
55 nanoparticles, allowing to optimize the material for specific applications.<sup>15-17</sup> For  
56 example, LSPR excitations are the most important factor for generating strong optical  
57 fields in these nanostructures. Consequently, the materials that support LSPR can be  
58 used in surface-enhanced Raman scattering (SERS) studies and applications.

59 One of the most widely publicized metallic nanoparticles synthesis has been  
60 about silver.<sup>9</sup> Related to this, manufacturing and application of silver nanoparticles-  
61 based materials has become a very active field in materials science. Silver nanoparticles  
62 can be prepared by several methods including chemical reduction, electrochemical  
63 techniques, and photochemical reduction.<sup>18-20</sup> However, toxic compounds such as  
64 sodium borohydride are usually involved, some synthetic process are time consuming  
65 and/or present low-yield in silver nanoparticles formation.<sup>9,10</sup> Recently, several studies

66 has been focused on green nanoparticles synthesis approaches, including silver  
67 nanoparticles, to avoid hazardous materials.<sup>21-25</sup> Also, these eco-friendly synthetic  
68 approaches are based on use of plant extracts, which make these processes simple, low-  
69 cost effective, and efficiency produce a wide variety of shapes with sizes ranging from 1  
70 to 100 nm. Basavegowda and Lee<sup>26</sup> described the use of *Satsuma mandarin* peel extract  
71 to the synthesis of silver nanoparticles. Dubey *et al.*<sup>27</sup> reported the use of fruit extract of  
72 *Tanacetrum vulgare* to synthesize silver and gold nanoparticles. Also, the use of plant  
73 extracts can be extend to synthesized others metals, such as copper nanoparticles as  
74 showed recently by Brumbaugh *et al.*<sup>28</sup>

75 By far, beside gold nanoparticles, silver nanoparticles are the most important  
76 material widely used as SERS substrate.<sup>29</sup> Silver nanoparticles have been extensively  
77 used owing to their unique optical properties, i.e., strong LSPR, that can be tuned in the  
78 visible range when varying size, morphology, composition, and aggregates formation.<sup>30-</sup>  
79 <sup>33</sup> Recently, a great deal of research effort has been devoted to transfer SERS into  
80 practical analytical applications. However, fabrication of SERS nanostructures with  
81 both high sensitivity and high reproducibility remains difficult.<sup>29,34</sup> Also, methods to  
82 produce low-cost substrates for practical application of SERS in routine laboratory  
83 analysis and on-site analysis in the field are even limited. In this context, silver  
84 nanoparticles synthesized using plant extracts have the potential to be a good alternative  
85 to add efficiency and low-cost in the same SERS substrate preparation.

86 Although a lot of research dealing with metallic silver nanoparticles synthesis  
87 using fruit and plant extracts have been published, to the best of our knowledge, the use  
88 of this nanoparticles in plasmonics and/or SERS studies has not been investigated so  
89 far. Therefore, the objective of this study was for the first time go beyond of the  
90 synthesis process. In other words, to synthesize silver nanoparticles using citric peel

91 extracts, to explore its plasmonics properties, and to investigate its SERS activity. For  
92 this purpose, citrus peel extracts from orange, tangerine, and lemon were used to  
93 synthesize silver nanoparticles without adding external surfactant and/or capping agent.  
94 These synthesized silver nanoparticles were self-assembled on glass slides and their  
95 SERS responses were explored using 4-aminobenzenethiol (4-ABT), rhodamine 6G  
96 (Rd6G), and methylene blue (MB) as Raman probe molecules. These model molecules  
97 were chosen because they are widely used as Raman probes in SERS studies.

98

## 99 **2. Materials and methods**

### 100 **2.1. Chemicals**

101 Silver nitrate (ACS grade), 3-aminopropyl-trimethoxysilane (APTMS), 4-  
102 aminobenzenethiol (4-ABT), Rhodamine 6G (Rd6G), and methylene blue (MB) were  
103 purchased from Sigma Aldrich and used without purification. Ethylic alcohol (ACS  
104 grade), hydrogen peroxide, hydrochloric acid and sulfuric acid were purchased from  
105 Synth.

106

### 107 **2.2. Preparation of citrus peel extracts and silver nanoparticles synthesis**

108 The same procedure was used for the three different citrus peel extracts. Fresh  
109 orange (*Citrus sinensis*), tangerine (*Citrus reticulata*), and lemon (*Citrus aurantiflora*)  
110 fruits were purchased from a local supermarket in Campinas in São Paulo state, Brazil.  
111 These citric fruits were washed thoroughly with distilled water and their peels were  
112 incised into small pieces. The peels pieces (30 g) were weighed and transferred to three  
113 different 500 mL flasks containing 100 mL of deionized water, mixed, and boiled for 10  
114 min. The citrus peel extracts obtained were filtered through Whatman No. 1 filter

115 paper and the filtered were collected in three different 250 mL Erlenmeyer flasks. The  
116 pH values of the peels extracts were measured using an Analyser digital pH-meter.

117 Silver nanoparticles synthesis: the same procedure was used for the three  
118 different citrus peel extracts. Briefly, 10 mL of citrus extract was mixed with 100 mL of  
119 aqueous solution of  $\text{AgNO}_3$   $10^{-3}$  mol  $\text{L}^{-1}$  and stirred at 30 °C for 60 min. The as-  
120 synthesized silver nanoparticles were named AgNP-Ora (from orange extract), AgNP-  
121 Tan (from tangerine extract), and AgNP-Lem (from lemon extract).

122

### 123 **2.3. Silver nanoparticles deposition onto glass slides**

124 The functionalization of glass slides (1 cm x 1 cm) with APTMS for preparation  
125 of SERS substrates is described in previous work by our group.<sup>35</sup> Volumes of 100  $\mu\text{L}$  of  
126 each kind of AgNP colloids were dropped onto  $\text{NH}_2$ -modified glass slides.  
127 Subsequently, the glass slides were placed into an oven to dry at 40 °C for 15 min. After  
128 this first deposition (1D substrates), the AgNP substrates were immersed in APTMS  
129 sol-gel for 20 min. After that, the substrates 1D were washed with deionized water and  
130 dried with  $\text{N}_2$  flow. Subsequently, volumes of 100  $\mu\text{L}$  of each AgNP colloids were  
131 dropped onto substrates 1D and they were placed into a oven to dry at 40 °C for 15 min.  
132 Finally, the substrates with two AgNP depositions (2D substrates) were washed with  
133 deionized water and dried with  $\text{N}_2$  flow.

134

### 135 **2.3. Materials characterization**

136 UV-Vis spectra of the AgNP colloids were obtained using a spectrometer  
137 Agilent Cary probe 50. Fourier transform infrared (FTIR) spectra were acquired with a  
138 NICOLET 6700 FTIR. Samples were prepared for FTIR analysis by freeze-drying the  
139 aqueous citrus peel extracts, grinding the dried samples with KBr, and forming KBr

140 pellets. The AgNP 1D and 2D substrates scanning electron microscopy (SEM) images  
141 were obtained using Field Emission Gun microscope model Inspect, with acceleration  
142 voltage of 15 kV. Before analysis the samples were covered with a carbon thin film.  
143 Also, an energy-dispersive X-ray spectroscopy (EDS) elemental analysis were carried  
144 out for the AgNP substrates. High-resolution transmission electron microscopy  
145 (HRTEM) images were obtained using a JEOL JEM-3010 microscope (300 kV, 1.7 Å  
146 point resolution). The sample was prepared by drop-drying the colloidal AgNP solution  
147 on a holey carbon coated Cu grid.

148

#### 149 **2.4. SERS measurements**

150 All measurements were accomplished using a confocal Jobin-Yvon T64000  
151 Raman spectrometer system, equipped with a liquid N<sub>2</sub> cooled CCD detector and a  
152 microscope was used. The excitation source was a laser at 633 nm. The laser power at  
153 the sample surface was about 2.5 mW. The laser was focused with a 100x focal-lens  
154 objective to a spot of about 1 μm. For all measurements, the laser exposure time was  
155 dependent on the Raman probe molecule (10 s for 4-ABT and for Rd6G, and 5 s for  
156 MB). Aliquots of 25 μL of 10<sup>-6</sup> mol L<sup>-1</sup> 4-ABT, Rd6G or MB ethanolic solutions were  
157 dropped onto 2D AgNP substrates. After the solvent evaporation the substrates were  
158 ready to be analyzed.

159

### 160 **3. Results and discussion**

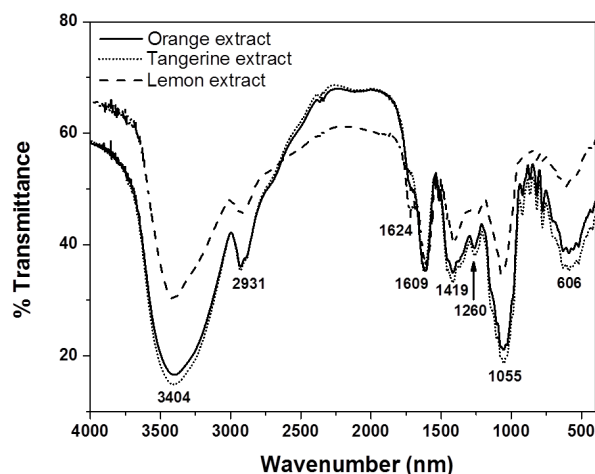
#### 161 **3.1. Synthesis and characterization of AgNP using citrus peel extracts**

162 The green methods of silver nanoparticles synthesis using plants extracts were  
163 reported to be eco-friendly when compared to traditional chemical methods, since these  
164 greener procedures are free of added chemical compounds such as surfactants and



165 capping agents. Then, the first step here is understanding what are functional groups  
166 involved in the silver nanoparticles reduction and the nanoparticle chemical stability.  
167 The FTIR spectral pattern of the three lyophilized citrus peel extracts are similar to each  
168 other (Fig. 1). Also, the general features of the FTIR spectra are common to spectra of  
169 plant extracts reported elsewhere.<sup>36-38</sup> The broad band at  $3404\text{ cm}^{-1}$  is assigned to  $\text{-O-H}$   
170 stretch from hydroxyl groups expected to be present in the peel extracts, such as sugars  
171 and polyphenols. Although the extracts have been lyophilized, this band may also be a  
172 contribution from incorporated water molecules. There are several possible assigns for  
173 the band located at  $1055$ ,  $2931$  and its shoulder at  $2869\text{ cm}^{-1}$  due to variety of  $\text{N-}$  and  
174  $\text{O-}$ containing functional groups. Also, these bands may be a contribution of  $\text{C-C}$   
175 stretch and  $\text{C-H}$  stretch from hydrocarbon chains. The band at  $1609\text{ cm}^{-1}$  is assigned to  
176 amide I band from protein carbonyl stretch. Bands located at around  $1419\text{ cm}^{-1}$  are  
177 attributed to  $\text{C-O-H}$  bending vibrations. As observed, the FTIR spectra of the citrus  
178 peel extracts indicate the presence of several functional groups, which can be associated  
179 with the presence of bioactive compounds such as flavonoids, citric acid, carotenoids,  
180 and aromatic compounds. Many of those compounds exhibit antioxidant properties, and  
181 consequently, can play important role as reducing and/or capping agents in  
182 nanoparticles synthesis.<sup>39,40</sup>

183



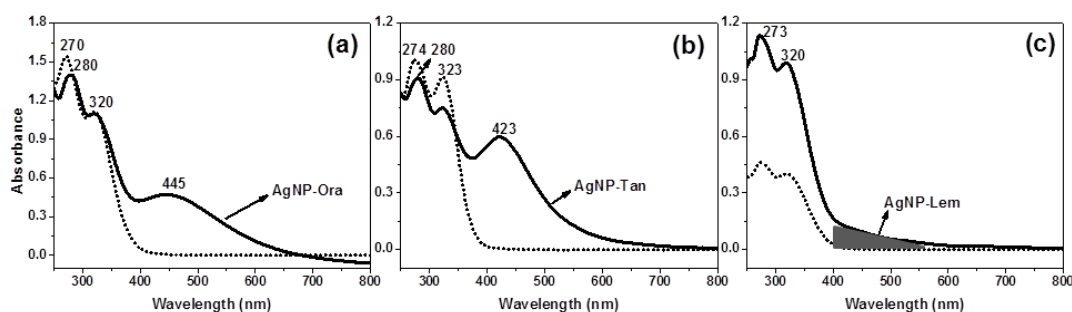
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185 **Figure 1.** FTIR spectra of the lyophilized citrus peel extracts.

186

187 During the reaction process, the color of the solutions turned from light yellow  
188 to brown (AgNP-Ora), dark-red (AgNP-Tan), and yellow (AgNP-Lem) as shown in  
189 Supplementary material (Fig. SM1). UV-vis absorption spectra of the peel extracts and  
190 of the AgNP aqueous solutions were used to understanding the differences observed in  
191 the reaction process (Fig. 2). All UV-vis absorption spectra of the citrus peel extracts  
192 exhibit a similar pattern, showing absorption bands at 320 and at around 270 nm. For  
193 AgNP-Ora and AgNP-Tan is observed the absorption band at 445 and 423 nm,  
194 respectively, which is consistent with the localized surface plasmon band (LSPR)  
195 related to the formation of silver nanoparticles (Fig. 2(a) and (b)). However, it is  
196 observed a lack of LSPR band in the AgNP-Lem UV-vis spectrum (Fig. 2(c)), which  
197 suggests the formation of silver clusters and/or ultrasmall silver nanoparticles.  
198 Ultrasmall nanoparticles and atomically precise clusters lose their visible extinction  
199 features as a function of the size, being characterized for absorbing into the UV  
200 range.<sup>41,42</sup> The formation of ultrasmall silver nanoparticles and/or clusters consisting of  
201 small number of atoms, and their chemical stability in aqueous solution, was reported  
202 and discussed by Linnert *et al.*<sup>42</sup> The silver species such as  $\text{Ag}_4^+$  and  $\text{Ag}_n$  clusters ( $n <$

203 10 atoms) are commonly formed in many procedures of silver nanoparticles colloidal  
204 synthesis and the bands located at around 280 and 323 nm are associated with their  
205 formation (Fig. 2). Also, these bands may be a contribution of the peel extracts  
206 absorption. The ultrasmall AgNP, also named quasi-metallic particles, shows absorption  
207 at around 360 nm.<sup>42</sup> The silver particles which possess metallic properties presents  
208 absorption from 380 nm to visible range.



209

210 **Figure 2.** UV-vis spectra of the AgNP colloids (solid line) and of their respective citrus  
211 peel extract (point line) orange (a), tangerine (b), and lemon (c).

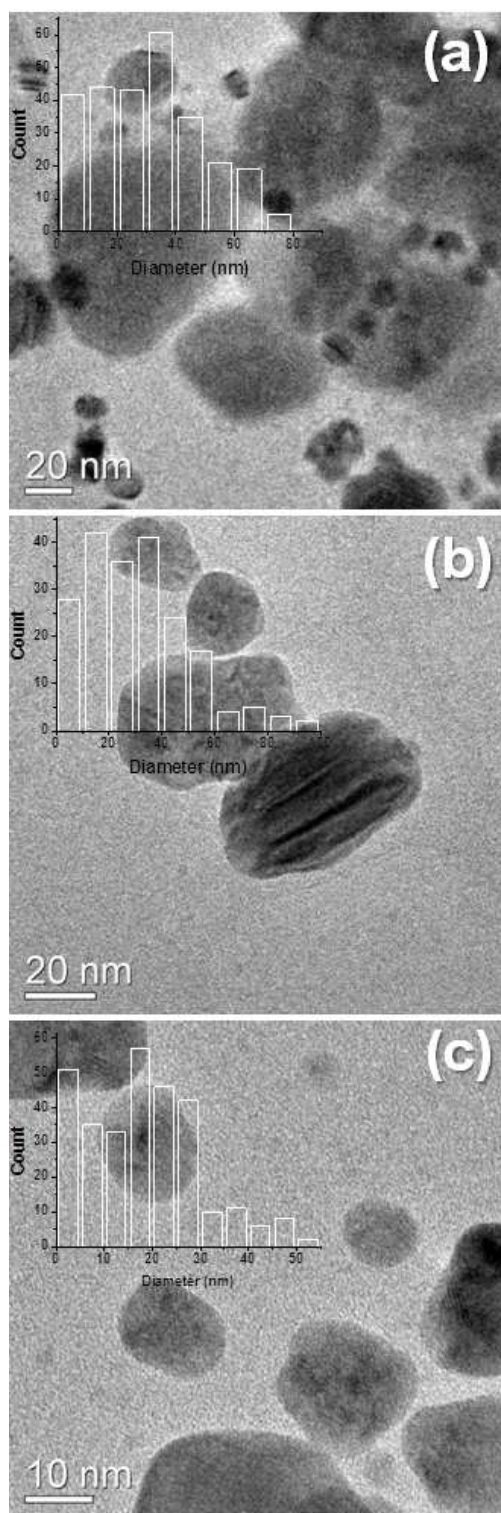
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213 According to the FTIR data, the citrus peel extracts present the same chemical  
214 composition. However, the UV-vis result indicates that 60 min of the reaction time  
215 should not be enough to growing the silver nanoparticles when using the lemon peel  
216 extract. This can be explained according to pH values of the citrus peel extracts, which  
217 were 4.0 for lemon extract and 6.0 for orange and tangerine extracts. However, in the  
218 end of the reaction the pH values were 5.0 for AgNP-Lem, and 7.0 for AgNP-Ora and  
219 for AgNP-Tan. In other words, the lemon peel extract is more acidic than the orange  
220 and tangerine extracts. This result suggests that the amount of acidic molecules such as  
221 citric acid and ascorbic acid is highest for the lemon extract. This kind of molecules  
222 present a strong reducing power, and also, play roles of capping agents in nanoparticles  
223 synthesis.<sup>43-45</sup> Consequently, a high concentration of these molecules can easily to

224 reduce  $\text{Ag}^+_{(\text{aq})}$  ions in solution, forming small silver particles, which are rapidly capped  
225 for those molecules. Although the lack of a defined LSPR band in the UV-vis  
226 absorption spectrum of AgNP-Lem, it can be observed in Fig. 2(c), the gray area of the  
227 spectrum, that the absorption between 400 and 550 nm is not zero. This result indicates  
228 the formation of AgNP between 5 - 100 nm range as showed by HRTEM analysis (Fig.  
229 3). However, the concentration of these silver nanoparticles should be lower than others  
230 two citrus peel extracts, since a defined LSPR band was not observed in that UV-vis  
231 spectrum.

232         Representatives HRTEM images of the silver nanoparticles colloids show the  
233 formation of nanoparticles with average sizes of  $31.0 \pm 18.3$  nm for AgNP-Ora,  $29.8 \pm$   
234  $18.7$  nm for AgNP-Tan, and  $18.5 \pm 11.6$  nm for AgNP-lem (Fig. 3). All the samples  
235 present AgNP with size below 5 nm, which are defined as ultrasmall nanoparticles<sup>46</sup> as  
236 evidenced before by UV-vis analysis. Although the lack of a defined LSPR band at UV-  
237 vis spectrum of the silver colloid prepared using lemon peel extract, the HRTEM  
238 analysis shows the presence of the formation of AgNP-Lem until 55 nm. The majority  
239 of the AgNP formed are spherical-like shape and are highly crystalline (Fig. SM2).

240



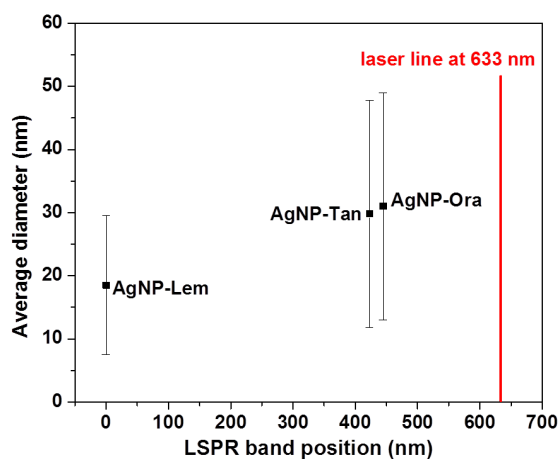
241

242 **Figure 3.** HRTEM images of the AgNP-Ora (a), AgNP-Tan (b), and AgNP-Lem (c).

243

244 Up to now, we have concluded that the same procedure to synthesize silver  
245 nanoparticles using citrus peel extracts resulted in the formation of AgNP with different  
246 size distributions due to the complexity of the chemical composition of the extracts.  
247 However, the colloids prepared contain high concentration of silver nanoparticles with  
248 preferential spherical symmetry. Also, the AgNP exhibit LSPR bands in the visible  
249 range, which is dependent on its size. These characteristics qualify the as-prepared  
250 AgNP to be used as SERS substrate. For example, using the laser line at 633 nm as  
251 source in Raman/SERS measurements the AgNP-Ora and AgNP-Tan seems to be the  
252 better samples than AgNP-Lem. As can be seen in Fig. 4, the last sample does not  
253 exhibit a defined LSPR band in the visible range, indicating an unfavorable condition to  
254 be used in SERS, where the laser line and surface plasmon coupling is important.<sup>47</sup>  
255 However, the others two samples exhibit LSPR bands near the line at 633 nm, which is  
256 a more appropriate condition for SERS. Although the AgNP prepared using lemon  
257 extract does not exhibit one defined LSPR band in visible range, the HRTEM analysis  
258 shows that there are silver nanoparticles with an adequate size to be tested in SERS.  
259 Then, the methodology of self-assembling AgNP was adopted to prepare the  
260 nanoparticles substrates for SERS as shown in Fig. 5.

261



262

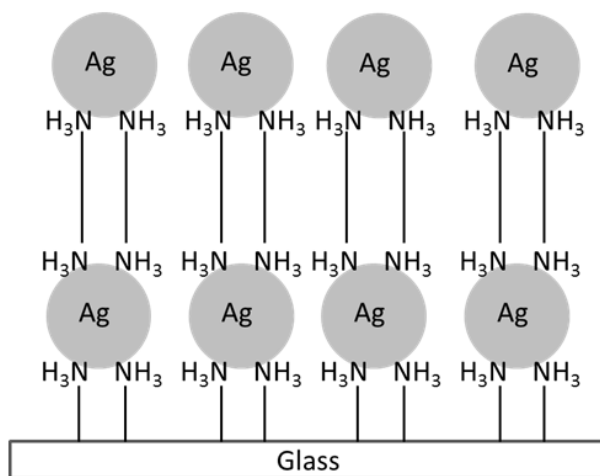
263 **Figure 4.** LSPR band position of the AgNP-Ora, AgNP-Tan, and AgNP-Lem as  
264 function of the average size and its position compared to the laser line at 633 nm.

265

### 266 3.2. Preparation of self-assembled AgNP substrates

267 The strategy of self-assembling plasmonic nanostructures has been employed  
268 with great success by many researches aiming to prepare efficient materials for specific  
269 applications.<sup>48</sup> For example, the organization of AgNP using APTMS sol gel molecules  
270 as linker offers a favorable condition for coupling surfaces plasmons of adjacent  
271 nanoparticles and enhancing the electric field in interparticle gaps (in the so-called "hot  
272 spots"). Due to hot spots generation and consequently amplification of Raman signals  
273 this approach has been employed for preparing SERS substrates.<sup>49,50</sup> In Fig. 5 is shown  
274 a representative scheme showing the strategy employed in the present work to prepare  
275 the self-assembled AgNP structures.

276

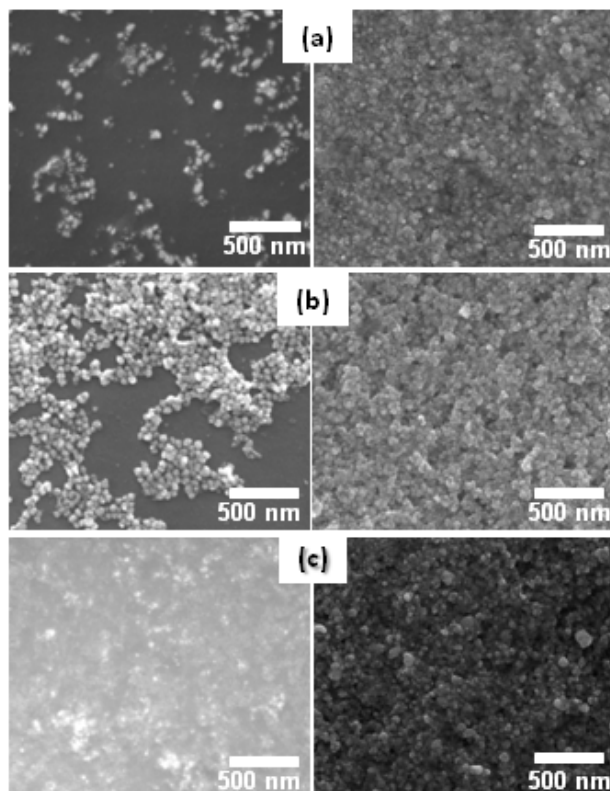


277

278 **Figure 5.** Representation of the bilayer AgNP structure on NH<sub>2</sub>-modified glass slide.

279 The APTMS sol gel molecules are the linker between the AgNP layers. Drawing out of  
280 scale, only for illustration.

281 Fig. 6 shows the SEM images of the AgNP substrates prepared with one  
282 deposition and two depositions of AgNP. As can be seen in Fig. 6, one deposition is not  
283 enough to cover the glass surface of AgNP as well as leads to formation of small AgNP  
284 aggregates, which are dispersed on glass surface. The AgNP-Lem 1D substrate presents  
285 few small aggregates comparing with other two 1D substrate due to its low  
286 concentration and/or small size of the AgNP in this colloid (Fig. 6(c)). As can be  
287 observed in Fig. 6 (images on the right side), after two depositions of AgNP, the  
288 aggregates are larger than before, forming rich areas of AgNP, even for the AgNP-Lem  
289 substrate. This result shows that the self-assembly strategy is efficient in the preparation  
290 of the AgNP substrate, promoting interparticle nanogaps generation and stability due  
291 the formation of chemical bonds between the AgNP and the available  $-NH_2$  groups  
292 (Fig.5).



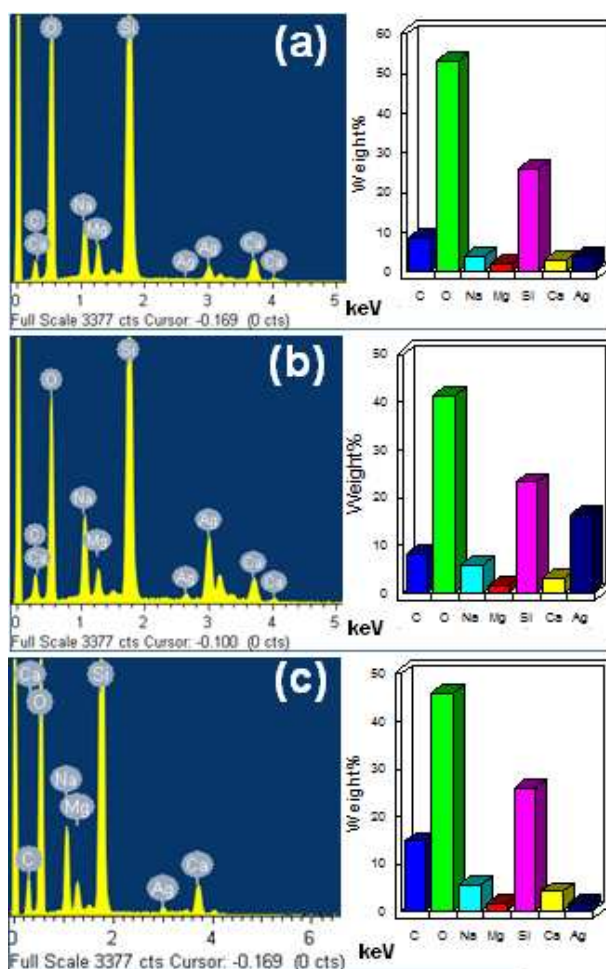
293



294 **Figure 6.** SEM images of the 1D substrates (left side) and 2D substrates (right side) for  
295 AgNP-Ora (a), AgNP-Tan (b), and AgNP-Lem (c).

296

297 EDS elemental analysis data show the same elements composition for all three  
298 AgNP 1D substrates (Fig. 7). The two peaks around 3.0 keV are characteristic of  
299 metallic Ag, which are assigned to Ag-L<sub>α</sub> and Ag-L<sub>β</sub> lines. The difference of the  
300 percentage of silver is related to the position where the EDS analysis was recorded on  
301 the AgNP aggregates. Also, the high percentage of silver observed for the AgNP-Tan  
302 1D substrate is in agreement with the SEM data, which showed the formation of larger  
303 AgNP aggregates than the other two AgNP substrates. Carbon and oxygen are from the  
304 organic compounds present in citrus peel extracts. Also, the oxygen signal may be a  
305 contribution from the glass, used as support, as well as the other observed glass  
306 components such as sodium, magnesium, silicon, and calcium.



307

308 **Figure 7.** EDS data of the 1D substrates: AgNP-Ora (a), AgNP-Tan (b), and AgNP-  
 309 Lem (c).

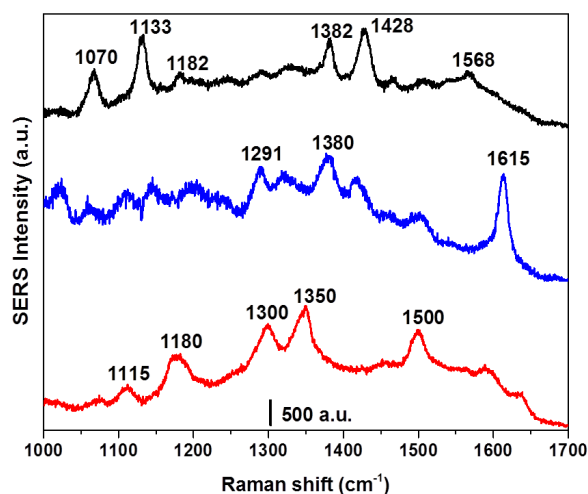
310

### 311 3.3. SERS activity of the AgNP 2D substrates

312 To evaluate the SERS activity of the AgNP 2D substrates 4-aminobenzenethiol,  
 313 rhodamine 6G, and methylene blue were chosen as Raman probe molecules. As can be  
 314 seen in Fig. 8, it is possible to detect the three molecules ( $10^{-6}$  mol L $^{-1}$ ) on AgNP-Ora  
 315 substrates. It is not our intention here to discuss in detail all assignments for the bands in  
 316 all SERS spectra. The main bands for 4-ABT (at 1070 cm $^{-1}$  assigned to  $\nu_{CS+CC}$ ), MB (at  
 317 1615 cm $^{-1}$  assigned to  $\nu_{C=C}$ ), and Rd6G (at 1500 cm $^{-1}$  assigned to  $\nu_{C=C}$ ) are in  
 318 agreement with previous works reported in the literature.<sup>51-53</sup> The most important issue

319 in this work is to show that the as-prepared AgNP, using citrus peel extracts, can be  
320 employed as efficiency in SERS, similar to the silver nanoparticles prepared by other  
321 synthetic approaches. However, the method reported here is simply, eco-friendly and  
322 cheap.

323



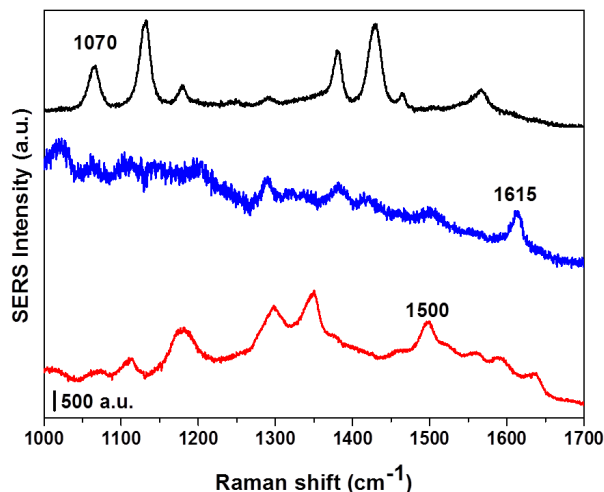
324

325 **Figure 8.** SERS spectra of  $10^{-6}$  mol L<sup>-1</sup> 4-ABT (black), MB (blue), and Rd6G (red) on  
326 AgNP-Ora substrates.

327

328 As can be visualized in Figs. 9 and 10, the substrates prepared using the AgNP-  
329 Tan or AgNP-Lem also exhibited SERS activity, being possible to detect the three  
330 molecules probe. For the AgNP-Tan substrate the signal-to-noise Raman signal from 4-  
331 ABT is higher than the Raman signal for the two dyes. A similar result is observed  
332 when analyzing the SERS spectra in Fig. 10 for AgNP-Lem, where can be observed a  
333 better quality in the SERS spectrum for 4-ABT. The main difference is the Rd6G SERS  
334 spectrum, where is not observed the band around 1500 cm<sup>-1</sup>, as observed in Figs. 8 and  
335 9. However, it can be observed a band at 1544 cm<sup>-1</sup>, which also can be attributed to this  
336 molecule. This fluctuation of the position of some bands is commonly observed in

337 SERS studies, and it can be related to specific orientation of the probe molecule  
338 adsorbed on substrate surface as well as single molecule behavior.<sup>51-53</sup>  
339

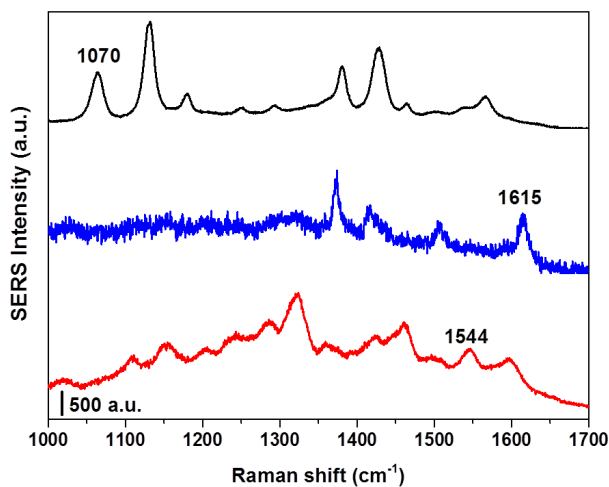


340

341 **Figure 9.** SERS spectra of  $10^{-6}$  mol L<sup>-1</sup> 4-ABT (black), MB (blue), and Rd6G (red) on  
342 AgNP-Tan substrates.

343

344



345

346 **Figure 10.** SERS spectra of  $10^{-6}$  mol L<sup>-1</sup> 4-ABT (black), MB (blue), and Rd6G (red) on  
347 AgNP-Lem substrates.

348

349 One of the most interesting result here is the SERS activity of the AgNP-Lem  
350 substrate for the three Raman probe molecules. As discussed above, the as-prepared  
351 silver nanoparticles using the lemon extracts is lack in LSPR due the low yield of the  
352 reaction. However, the HRTEM images show the presence of silver nanoparticles larger  
353 than 5 nm, which posses plasmonic properties. This explains the SERS activity of the  
354 AgNP-Lem substrate. Also, other important aspect here is the strategy of using the self-  
355 assembly approach to prepare the AgNP substrates. When the particles are in close  
356 packed assembly, then the electromagnetic field influences its neighbor. Consequently,  
357 the oscillating electrons in one particle feel the electric field due to the oscillation of the  
358 second particle, which can lead to plasmon coupling and generation of collective  
359 plasmon oscillations of the aggregate structures.<sup>48,54</sup> The plasmon coupling is an  
360 important contribute to amplification of the optical and surface-enhanced Raman  
361 scattering signals. This explain the SERS activity of the all AgNP aggregates substrates  
362 and also, being extremely decisive for the AgNP-Lem substrate SERS activity.

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#### 364 4. Conclusions

365 In this work, we present a green synthesis approach to prepare silver  
366 nanoparticles using citrus peel extracts and their self-assembly on glass slides to be used  
367 as SERS substrate. The as-synthesized AgNP are stable and exhibit LSPR band at  
368 visible range. The nanoparticles were self-assembled onto amine-modified glass slides,  
369 forming AgNP aggregates substrates. Self-assembled substrates of AgNP-Ora, AgNP-  
370 Tan, and AgNP-Lem have been further tested for SERS of 4-ABT, MB, and Rd6G at  
371  $10^{-6}$  mol L<sup>-1</sup> concentration. It was possible to detected with high signal-to-noise the  
372 SERS spectral pattern of all probe molecules for all the three AgNP substrates.  
373 Furthermore, the Raman results show that the SERS intensities for all AgNP substrates

374 are high when using 4-ABT probe molecule than the SERS intensities of the dyes probe.  
375 This simple, low cost, and greener method for synthesise silver nanoparticles may be  
376 valuable in SERS sensors development and can be extended for others fields such as  
377 catalytic applications.

378

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399 **References**

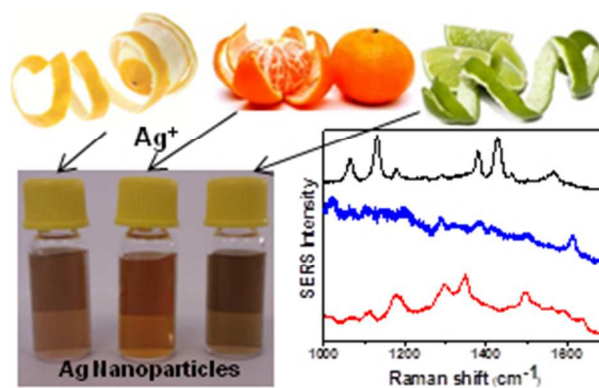
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Plasmonic silver nanoparticles synthesized using citrus peel extracts exhibit SERS activity for different Raman probe molecules.  
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