

NJC

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxxx

ARTICLE TYPE

Mesoporous Silica - Chondroitin Sulphate Hybrid Nanoparticles for Targeted and Bio-responsive Drug Delivery

Krishna Radhakrishnan^a, Jasaswini Tripathy^{ab}, Akshay Datey^{bc}, Dipshikha chakravorty^{bc}, Ashok M. Raichur^{*ac}

Received (in XXX, XXX) Xth XXXXXXXXX 20XX, Accepted Xth XXXXXXXXX 20XX
DOI: 10.1039/b000000x

This work proposes the fabrication of a novel targeted drug delivery system based on mesoporous silica-biopolymer hybrids that can release drugs in response to biological stimuli present in cancer cells. The proposed system utilizes mesoporous silica nanoparticles as a carrier to host the drug molecules. A biopolymer cap is attached onto these particles which serves the multiple functions of drug retention, targeting and bio-responsive drug release. The biopolymer chondroitin sulphate, used here is a glycosaminoglycan that can specifically bind to receptors over-expressed on cancer cells. This molecule also possess the property to disintegrate on exposure to enzymes over-expressed in cancer cells. When these particles interact with cancer cells, the chondroitin sulphate present on the surface recognizes and attaches to the CD44 receptors facilitating the uptake of these particles. The phagocytised particles are then exposed to the degradative enzymes such as hyaluronidase present inside the cancer cells which degrades the cap resulting in drug release. By utilizing a cervical cancer cell line we have demonstrated the targetability and intracellular delivery of hydrophobic drugs encapsulated in these particles. It was observed that the system was capable of enhancing the anticancer activity of hydrophobic drug curcumin. Overall, we believe that this system might prove to be a valuable candidate for targeted and bioresponsive drug delivery.

Introduction

Bio-responsive drug delivery involves the utilization of a biologically relevant molecule or phenomenon to elicit drug release from a suitable drug carrier. These autonomous systems eliminate the need to externally manipulate the location and time of drug release. The stimuli utilized in these systems include biomolecules such as glucose^{1,2}, enzymes³⁻⁵, ATP^{6,7}, pH variations⁸⁻¹⁰ and redox potential variations^{11,12}.

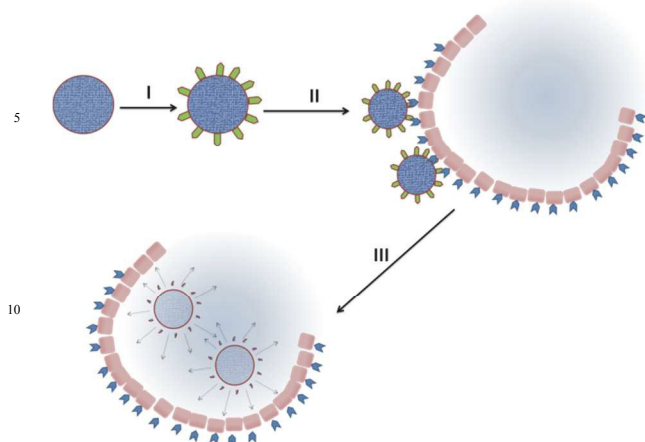
Recently several drug delivery systems have been designed that have the ability to modulate drug release in response to these various stimuli. These systems include polymeric nanoparticles, lipid based nanoparticles, hydrogel particles and mesoporous silica nanoparticle (MSN) – hybrids.^{8,13} Of these MSN-hybrids offer several advantages such as (i) ability to retain drug molecules for prolonged time periods (ii) High drug loading efficiency due to high porosity and high specific surface area (iii) possibility for surface modification to provide additional functionalities and (iv) biochemical stability. Moreover the mesopores of the MSN can be blocked with suitable capping agents to block the outward leakage of encapsulated drug molecules. Based on the properties of these capping agents, the systems may be tuned to respond to a wide array of physical, chemical or biological stimuli.^{14,15}

Although a few MSN hybrid systems have been reported with the ability to respond to diverse bio-stimuli such as variations in

redox potentials;¹⁶⁻¹⁸ pH variations;¹⁹⁻²¹ and enzymes,²²⁻²⁴ there is a lot more to be explored in this area. One important development in this field has been the introduction of capping agents that serve the dual purpose of targeting and stimuli response. An example for this kind of systems would be hyaluronic acid capped MSNs that exhibit targetability and bio-responsive drug delivery.¹⁸

In the present study, we report the fabrication of another bio-responsive MSN-hybrid system with the ability to target cancer cells. MSN - hybrid drug delivery system have been fabricated by capping the mesopores of drug loaded MSNs with chondroitin sulphate (CHD). These particles display the ability to deliver hydrophobic drugs specifically into cancer cells and by utilizing the internal stimuli present there, induce bio-responsive drug release. CHD is a sulfated glycosaminoglycan widely distributed in the body. It is an FDA approved drug clinically used for treatment of arthritis and osteoporosis. In the present system we have used CHD as a biocompatible capping agent that prevents the outward diffusion of the drug molecules from the MSN mesopores which in turn minimizes premature leakage ahead of reaching the target site. CHD also acts as a stimuli responsive component of the system as it can undergo degradation in the presence of enzymes such as lysosomal hyaluronidase present inside cancer cells.^{25,26} This molecule is also known to interact with cell surface receptors such as CD44 over-expressed on the surface of cancer cells.²⁷ Although CHD has been known to

possess such remarkable properties, no significant attempts have been made to utilize it as a targeted and bioresponsive platform



Scheme I General scheme illustrating drug loaded MSN-CHD nanoparticles being taken up by cancer cells and releasing the drug inside the cancer cells. (I) CHD capping on drug loaded MSN. (II) The CHD capping interacts with the CD 44 receptors overexpressed on the cancer cells resulting in efficient particle uptake. (III) Once inside the cell, the CHD cap undergoes enzymatic degradation utilizing the local biological stimuli resulting in drug release.

for drug delivery. This is the first report that utilizes CHD for targeted and stimuli responsive drug delivery.

The synthesis, receptor mediated uptake and intracellular drug release of these particles have been illustrated in scheme I. When the CHD capped MSN particles interact with cancer cells, the CHD present on the surface recognizes and attaches to the CD44 receptors facilitating the uptake of these particles. The phagocytised particles are then exposed to the degradative enzymes such as hyaluronidase present inside the cancer cells which degrades the cap resulting in drug release.

Materials and Methods

N-cetyltrimethyl-ammonium bromide (CTAB), NaOH, Tetraethoxysilicate (TEOS), aminopropyltrimethoxysilane (APTMS), Phosphate Buffer Saline (PBS) 10X, CHD, 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC), Hyaluronidase, MTT ((3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide), DMEM, FCS (Fetal Calf serum) and antibiotics were purchased from Sigma Aldrich, USA.

Synthesis of MSN - NH₂

Mesoporous silica nanoparticles were synthesized by modifying a previously reported procedure. Briefly, 1g of CTAB in water was kept in 2 M NaOH solution and heated up to 80 °C. 5g of TEOS was added to this mixture and stirred for 2 h. The resulting white precipitate was isolated by removing the supernatant by centrifugation at 12000 rpm. This precipitate was then washed with methanol 4 times to remove un-reacted reactants. Finally the particles were calcined at 540 °C to remove the surfactant template.

For the synthesis of amino-propyl functionalized MSNs, APTMS was used as a linker. Briefly, 200 mg of MSN was dried at 110 °C under vacuum for 15 h to remove adsorbed moisture.

The MSN particles were then dispersed in dry toluene (to prevent homopolycondensation of APTMS) by ultrasonication. 1mL APTMS was then added to this mixture and refluxed for 20 h under argon atmosphere. The resulting MSN-NH₂ was further purified by ultracentrifugation and washing cycles with toluene to eliminate un-reacted siloxane moieties. MSN-NH₂ obtained was later dried at 110 °C under vacuum to remove the solvent.

Attachment of CHD to MSNs

Initially the carboxyl groups of CHD was activated using a carbodiimide, EDC. In a typical experiment 3 mg/mL CHD solution in MES buffer (pH 5.5) was mixed with EDC (0.15 M) and stirred at room temperature for 2 hours. This activated the carboxyl groups of CHD and resulted in the formation of an O-Acyl-isourea intermediate. 50 mg of dried MSN-NH₂ powder was then added to this solution and kept under constant stirring for 4 hours at ambient temperature. The activated carboxyl groups of CHD reacted readily with the amine functional groups of MSN-NH₂ resulting in the formation of amide bonds. The resulting MSN-CHD nanoparticles were purified by ultracentrifugation and washing cycles with double-distilled water to eliminate un-reacted reactants. The purified MSN-CHD was dried overnight at 33°C under vacuum and stored at 4°C till further use.

Material characterization

The morphological analysis of MSN was carried out using transmission electron microscopy (TEM). Tecnai F-30 (FEI, Eindhoven, The Netherlands), equipped with a Schottky Field Emission source was used to obtain the TEM images. The samples for TEM analysis was prepared by placing a drop of the nanoparticle solution onto a copper grid.

Fourier transform infrared spectroscopy (FTIR) spectra were obtained at various stages of functionalisation on MSN, using a Thermo-Nicolet 6700 spectrometer, Thermo Scientific. The FTIR spectra of the samples pelletized using potassium bromide was recorded in the range 400 cm⁻¹ to 4000 cm⁻¹ with a resolution of 4 cm⁻¹. TGA was further utilised to characterize the various functionalisation steps on the MSNs. STA 409PC, Netzsch with a heating rate of 10 °C min⁻¹ was utilized to obtain the TGA spectra of the MSN samples at various stages of functionalisation. The comparative weight loss of the sample was measured as a function of temperature which was varied between 30°C and 800 °C under Ar atmosphere to record the TGA spectra. While the weight loss occurring before 110 °C is attributable to the loss of physically adsorbed moisture, the weight loss in the 120 °C to 750°C regime is due to the various functional groups, organic moieties attached on to the surface of MSNs. The ζ-potential variations of MSN particles was measured using a Zetasizer Nanoseries NanoZS90 equipment (Malvern Southborough, MA). The measurements were recorded by suspending the required particles in pH adjusted water.

Drug loading and release experiments

The drug loading was carried out prior to the capping process. In order to realize drug loading, the MSN-NH₂ nanoparticles were suspended in a solution containing the required drug molecules (either 0.8mg/mL curcumin in ethanol or 0.6mg/mL fluorouracil in H₂O). This suspension was kept for 12 hours under stirring to

aid efficient diffusion of the drug molecules into the pores of the MSN. The drug loaded nanoparticles were then centrifuged to remove un-adsorbed drug molecules. These particles were then added to a solution containing EDC activated CHD to seal the mesopores.

The drug uptake and release was calculated by measuring the absorbance of the drug molecules in the loading or release media. Absorbance measurements were carried out using a Perkin Elmer Lambda 35 spectrometer (Perkin Elmer, USA) with a wavelength range of 190 to 1100 nm. In order to estimate the amount of unloaded drugs remaining in the drug loading solution, the absorbance of the supernatant subsequent to centrifugation of the loading solution - MSN mixture was recorded. This data in conjunction with the data from the standard curve plotted for various concentrations of the drug, gave the amount of drug present in the supernatant after drug loading.

In a typical drug release experiment, 3 mg of the drug loaded nanoparticles were incubated in 2 mL of the respective release media. In order to measure the amount of drug released, the particles were centrifuged and samples were taken from the supernatant. An equal volume of the release media was replaced to compensate for the loss due to sampling. The release curve was obtained by plotting the amount of drug released as a function of time. Total amount of drug released from MSN-CHD particles kept under probe sonication (to completely remove the CHD cap) for 120 minutes was taken as the 100% drug release.

Cell viability assessment using MTT assay

HeLa cells (human cervical cancer cells) and COLO 205 (cervical cancer) were cultured in Dulbecco's Modified Medium (DMEM) and RPMI medium respectively, supplemented with 10% FBS and antibiotics. The cells were cultured in a sterile incubator at 37 °C, 5% CO₂. When the cells reached 90% confluence, they were harvested and seeded onto a 96 well plate at a density of 5 X 10⁴ cells/mL. Once these cells attached and flattened onto the well plate, different concentrations of the various nanoparticles were added and incubated for 24 hours. After the required incubation time, 20 μl of MTT dye, a soluble yellow tetrazolium salt (5mg/ml) was added to each well and kept for another 4 h at 37°C. This resulted in the formation of formazan crystals which were then solubilized. The absorbance of the resulting solution was measured and the cell viability was calculated by taking the untreated cells as the control.

Confocal microscopy imaging

The CLSM images were obtained using a Carl Zeiss LSM Confocal scanning system equipped with a 100x oil immersion objective with a numerical aperture of 1.4. Briefly COLO 205 cells were seeded on cover-slip at a cell density of 10⁵/cm² and incubated with curcumin loaded MSN-PRMs. After the required time period, the cells were washed with PBS and fixed using 4% para-formaldehyde. This was followed by DAPI staining and washing with PBS. Finally the cells were visualized under the CLSM microscope after adding the mounting media.

Statistical analysis

The Graphpad Prism software (version 5) was used for all statistical analysis. The significance of the ζ-potential variations was calculated by paired t test and ANOVA. The anticancer

activity of free curcumin and curcumin in MSN-CHD (at 40 μg/mL) was compared using a Student's *t* test. A *P* value of less than 0.05 was taken as significant.

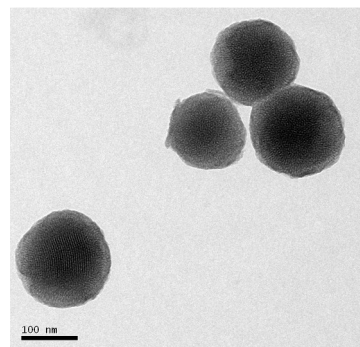


Fig.1 TEM micrographs of the synthesized MSNs. Dilute solutions containing MSNs were coated onto a carbon grid and dried to remove the aqueous solvents.

Result and Discussion

The MSNs were prepared by means of a surfactant templated approach. CTAB which is a cationic surfactant which form micelle structures in solution. These micelles acts as templates onto which the silica species get adsorbed and later undergoes hydrolysis and condensation to form silica. The mesoporous structure of the MSNs are obtained by calcination at 540 °C which removes the surfactant template. Thus prepared MSNs with diameter of around 170 nm was observed using TEM microscopy as shown in figure 1. The N₂ adsorption - desorption isotherms indicated a specific surface area of 781 m²g⁻¹ and average pore diameter of 2.5 nm. The typical type IV curve observed is indicative of the mesoporous structure of these particles (Supporting information). The MSNs were then functionalized with amine groups by a post synthetic grafting approach involving APTMS. Subsequent to the amine functionalisation, CHD molecules were attached via carbodiimide mediated crosslinking. This resulted in the formation of amide linkage between the amine groups of MSN and carboxyl groups of CHD. The efficiency of the capping was expected to be further enhanced by the electrostatic interaction between the negatively charged CHD and positively charged MSN-NH₂.

Table 1. Zeta Potential variations of MSNs suspended in water recorded at various stages of MSN-CHD fabrication.

Particle	Zeta Potential (mV) ^a
MSN	-27.40
MSN-NH ₂	5.03
MSN-CHD	-34.16

^a Values represent average of three measurements. The zeta values have changed significantly after each functionalisation step according to a one way ANOVA test performed. ($p < 0.001$)

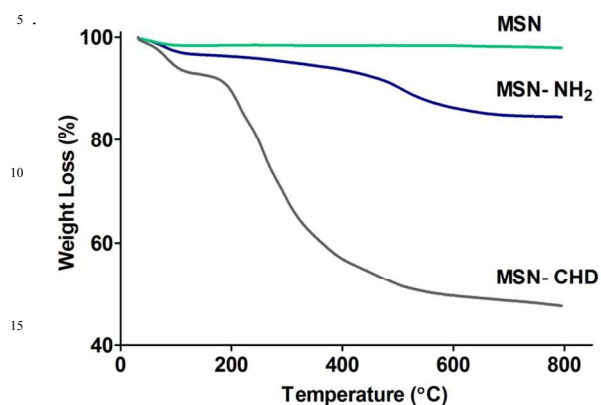


Fig. 2. TGA thermograms of the particles taken at various stages of functionalization. Samples were heated at a heating rate of $10\text{ }^{\circ}\text{C min}^{-1}$ and the corresponding weight loss was measured. The graph displayed a progressive increase in weight loss corresponding to each functionalization step. MSNs maximum weight loss after the grafting of the bulky CHD groups.

After removing the unreacted CHD molecules, the particles were characterized for successful cap attachment. Zeta potential measurements (Table 1.) taken immediately after the cap attachment showed that the positive zeta potential of MSN-NH₂ nanoparticles have shifted from +5.03 mV to -34.16 mV due to attachment of CHD.

The formation of chemical bonds and attachment of organic groups to the surface was characterized using TGA and IR spectroscopy. The TGA analysis carried out at different stages of functionalisation showed that there is progressive increase in weight loss after each functionalisation step (Figure 2). This is a strong indicator for attachment of the amine groups and CHD onto the MSN surface. This was further confirmed using IR spectroscopy. The FTIR spectra of MSN samples (Figure 3) at all stages of functionalisation showed the characteristic peak of Si-O-Si at 1087 cm^{-1} . Upon amine functionalisation a strong peak appeared at 1580 cm^{-1} corresponding to NH bending vibrations of the amine functional group attached to the surface. The CHD cap attachment resulted in the emergence of a new peak at 1650 cm^{-1} which is attributable to the C=O stretching vibration of the carboxyl groups contributed by CHD.

CHD is a glycosaminoglycan made up of repeating units of D-

glucuronic acid and N-acetyl-D galactosamine that are sulfated at different positions. Cells degrade CHD by using enzymes predominantly present in the lysosomes. The major enzyme involved in this process is an endo- β -N-acetylhexosaminidase called hyaluronidase which is known to cleave linkages between N acetylgalactosamine or N-acetyl-galactosamine sulfate, and glucuronic acid in CHD. In order to study the bio-responsive drug release from MSN-CHD nanoparticles we have utilized this enzyme as the bio-stimuli model. Another reason for choosing hyaluronidase as the stimuli is the fact that it is widely over-expressed in the microenvironment of several cancers.²⁸

The drug loaded MSN-CHD nanoparticles were incubated in MES buffer at pH 5.5 in the presence of various concentrations of hyaluronidase to simulate cancer cell microenvironment and the drug release was observed using absorbance spectroscopy. As a control, drug loaded MSN-CHD hybrids were incubated in PBS to simulate the physiological pH conditions in the absence of the bio-st

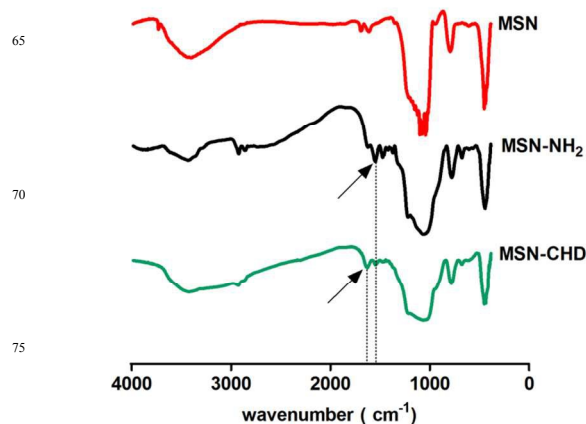


Fig. 3 FTIR spectra of MSN, MSN-NH₂ and MSN-CHD confirmed the attachment of the various functional molecules onto the surface of MSNs

As shown in Figure 4., the particle exhibited minimal drug leakage in the absence of the bio-stimuli. In contrast, the particles incubated in the presence of the enzyme exhibited enhanced drug release rate. This indicates that the system can specifically induce drug release in response to biological stimuli associated with cancer cells.

The drug release from the particles reached nearly 84.2 % within 25 hours when incubated in media containing 500 U/mL while the particles incubated in 250 U/mL showed a lower drug release of only 56.45 % within the same time period. This indicates that the drug release from the particles can be modulated by the variations in the concentration of the hyaluronidase enzyme. One possible explanation for this behavior is that at lower concentrations of the enzyme the CHD cap degradation and removal may be relatively low and this partially degraded cap could be hindering the release of the drug molecules. Since the concentration of hyaluronidase I is significantly higher in certain cancer cells when compared to normal cells, this property could be useful in targeting such cancer cells.²⁸

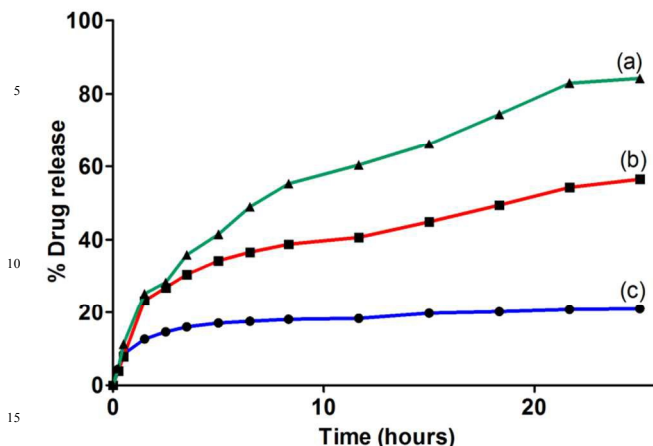


Fig. 4. Release profile from MSN -CHD in the presence of (a) 500 U/mL Hyaluronidase (b) 250 U/mL Hyaluronidase and (c) in the absence of enzyme. The release media was either prepared in MES buffer pH 5.5 (in the case of enzyme containing media) or in PBS buffer pH 7.4 (as in the case of control media). The concentration of drugs in the release media was evaluated using absorbance spectroscopy. The release curves indicate a concentration dependent variation in drug release rates in the presence of the triggering enzyme Hyaluronidase.

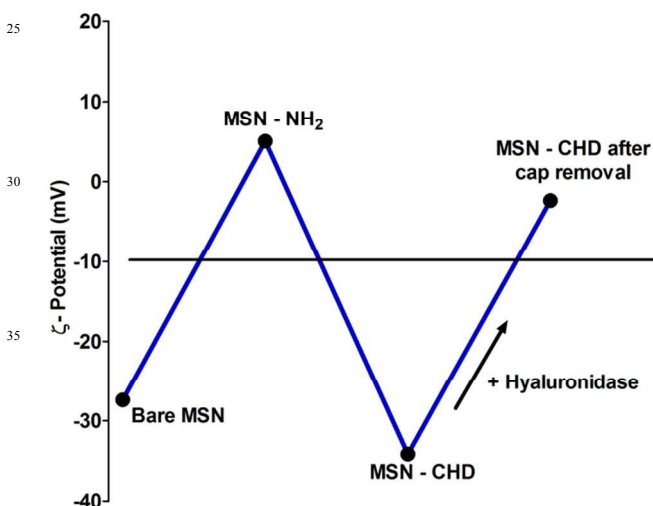


Fig. 5 Graph showing variation in zeta potential at various stages of MSN functionalisation and CHD cap removal. The particles were suspended in water before recording the zeta potential measurements. The zeta potential fluctuated during fabrication and cap removal indicating variations in the surface chemistry of the particles at various stages.

The cap removal process in the presence of hyaluronidase enzyme was further evaluated using zeta potential measurements. It was observed that the zeta potential of MSN -CHD which was at - 34.16 mV changed to a value of - 2.34 mV when the particles were incubated in the presence of hyaluronidase I. This indicates that the CHD cap has been degraded into fragments by hyaluronidase enzyme and this has resulted in the reduction of the negative charge on the surface of MSNs.

Next we evaluated the performance of the system in a biological scenario by using cancer cells. For this we loaded a hydrophobic anticancer drug curcumin into the pores of the particles and then observed the interactions of these particles with

cancer cells. Curcumin is a photochemical drug that exhibits green fluorescence when excited with 420 nm wavelength light. This property of curcumin has been utilized to study the cell interactions of the drug loaded MSN-CHD using confocal laser scanning microscopy (CLSM). The curcumin loaded MSN-CHD particles were incubated with HeLa cells and CLSM images were taken. As evidenced from the CLSM images in Figure 6., the particles were efficiently taken up by HeLa cells. This efficiency is attributable to the high density of CD 44 receptors present on these cancer cells. CHD molecules have been reported to bind with CD 44 receptors and this could have enhanced the uptake of the nanoparticles into the cell. In order to further verify this hypothesis we blocked the CD 44 receptors on the cancer cells prior to incubation with MSN-CHD. This was carried out by using free hyaluronic acid which is another major ligand for this receptor. As seen in the CLSM images shown in Figure 6, the uptake of curcumin loaded MSN-CHD was considerably reduced when the CD 44 receptors present on the cancer cell surface was blocked. This is a very good indicator that the cellular uptake of the MSN-CHD nanoparticles had taken place via the CD 44 receptors present on the cancer cells. Furthermore, CD 44 negative L929 mouse fibroblast cells were then used as a negative control to study the specificity of MSN-CHD nanoparticles towards CD 44 positive cancer cells. HeLa and L929 cells were incubated with the same concentration of

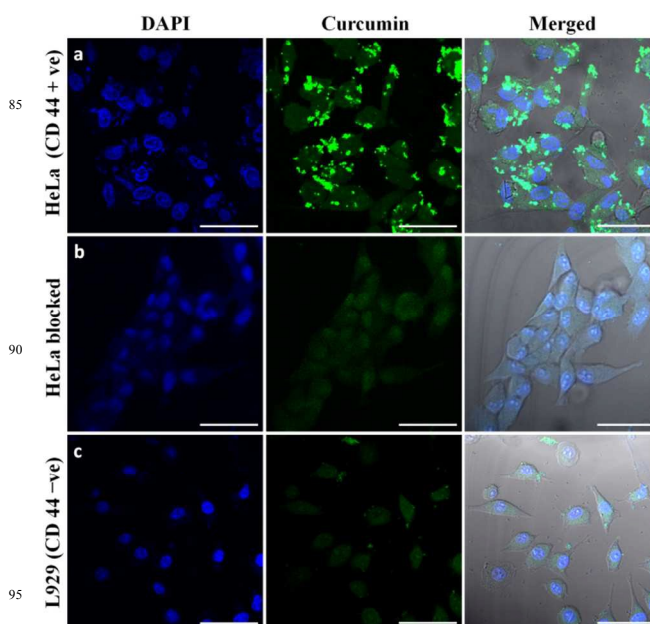


Fig. 6 CLSM images of (a) HeLa cells (b) HeLa cells whose CD44 receptors have been blocked with free HA and (c) L929 cells incubated with curcumin loaded MSN-CHD nanoparticles for 2 hours. (Scale bar = 50 μ m). The CLSM images indicate that the MSN-CHD particles are taken up by cancer cells which have free unblocked HA receptors.

curcumin loaded MSN-CHD nanoparticles and CLSM images were taken. The CLSM images as shown in Figure 6. indicate that the particles have preferentially accumulated inside the HeLa cells with respect to L929. While a significant green fluorescence was observed from untreated HeLa cells, only negligible fluorescence was coming from L929. This supports the fact that

the MSN-CHD particles have the ability to accumulate selectively in CD 44 positive cancer cells.

Since the loaded curcumin is a hydrophobic anticancer drug, we next studied the anticancer activity of curcumin loaded MSNs against HeLa cells. Due to its hydrophobic nature, curcumin is known to show very limited cytotoxicity in aqueous solutions. In fact, we observed that when free curcumin suspended in water was incubated with cancer cells, no significant fluorescence was observed from the cells (Figure 7), proving that free curcumin in water is not taken up by cancer cells.

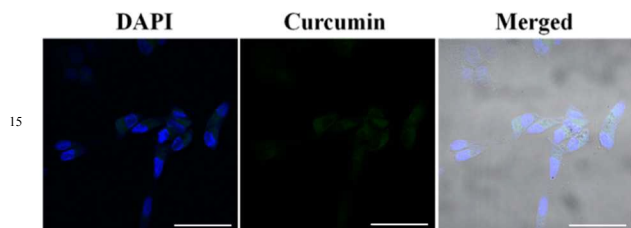


Fig. 7 CLSM images of HeLa cells incubated with free curcumin suspended in water. Due to the hydrophobic nature and poor aqueous solubility of free curcumin, it can be seen that it is not efficiently taken up by cancer cells. (Scale bar = 50 μm).

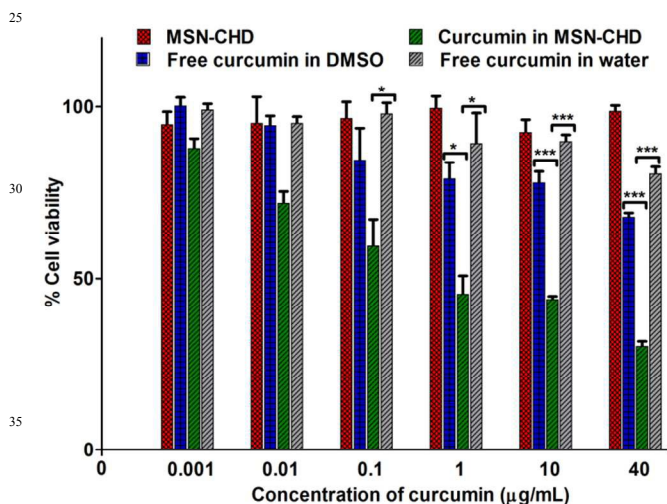


Fig. 7 Results of MTT assay showing the anticancer activity of curcumin encapsulated inside MSN-CHD when compared to free curcumin in DMSO or in water. The error bar represents standard deviation of three measurements. The readings that are significantly different according to a one way ANOVA test have been marked in the graph (***) $p < 0.0001$, * $p < 0.05$.

One way to overcome this limitation is to encapsulate curcumin inside suitable carriers that can be suspended in aqueous solutions. Additionally it would be beneficial if these carriers can deliver the drug directly into the interior of the cancer cells. We hypothesized that the bio-responsive MSN-CHD hybrids could act as a suitable carrier for this kind of intracellular drug delivery. To demonstrate this we studied the anticancer activity of free curcumin and curcumin encapsulated in MSN-CHD against HeLa cells. As a control we have evaluated the viability of HeLa cells incubated with unloaded MSN-CHD. In

order to validate any cytotoxicity contribution from the carrier, the unloaded MSN-CHD concentrations were adjusted to the corresponding concentrations of the loaded MSN-CHD studied.

The results of the MTT assay carried out reveals that curcumin encapsulated inside MSN-CHD shows more potent anticancer activity against HeLa cells when compared to free curcumin. In fact only 59.4% of the cells survived when treated with 0.1 $\mu\text{g/mL}$ curcumin encapsulated inside MSN-PRM as opposed to the 97.94% cells surviving after treatment with free curcumin in water ($p < 0.05$). More significant difference in anticancer activity was observed at higher concentrations of curcumin with the percentage of cell death being 69.87% for curcumin in MSN-PRM (40 $\mu\text{g/mL}$) and 19.6% for free curcumin in water (40 $\mu\text{g/mL}$) ($p < 0.0001$). Interestingly, the anticancer activity of curcumin in MSN-CHD was significantly higher than curcumin dissolved in DMSO as well. The difference was significant at higher concentrations with the cell death being around 32.18% at 40 $\mu\text{g/mL}$ of curcumin ($p < 0.0001$). This may be due to the enhanced ability of curcumin in MSN-CHD to accumulate inside HeLa cells as opposed to free curcumin. We also observed that the unloaded MSN-CHD particles exhibited no significant cytotoxicity within the range of concentrations tested.

Conclusion

In summary we have reported the fabrication of a novel bioresponsive and targeted drug delivery system which utilizes Mesoporous silica nanoparticles as the drug carrier and Chondroitin Sulphate as the degradable cap. The cap in addition to being bio-degradable, provides targetability to the particles towards CD 44 overexpressing cells. In the proof of concept studies, we have demonstrated that these particles can release drug molecules in the presence of the enzyme stimuli hyaluronidase and that the particles can deliver hydrophobic anticancer drugs specifically to CD 44 overexpressing cancer cells. The particles were also seen to enhance the anticancer activity of the hydrophobic drug curcumin. These findings clearly indicate that the Mesoporous silica nanoparticle-Chondroitin sulphate system is a highly promising candidate for bioresponsive and targeted drug delivery.

Notes and references

^a Department of Materials Engineering, Indian Institute of Science, Bangalore, 560012, India, Fax: +91 80 23600472, Tel: +91 80 22933238 Email: amr@materials.iisc.ernet.in

^b Department of Microbiology and Cell Biology, Indian Institute of Science, Bangalore, 560012, India,

^c Bioengineering Program, Indian Institute of Science, Bangalore, 560012, India

^d School of Applied sciences (Chemistry), KIIT University, Bhubaneswar 751024, Odisha, India

1. K. Ishihara and K. Matsui, *J. Polym. Sci. Part C Polym. Lett.*, 1986, **24**, 413–417.
2. Y. Zhao, B. G. Trewyn, I. I. Slowing, and V. S.-Y. Lin, *J. Am. Chem. Soc.*, 2009, **131**, 8398–8400.
3. R. V. Ulijn, N. Bibi, V. Jayawarna, P. D. Thornton, S. J. Todd, R. J. Mart, A. M. Smith, and J. E. Gough, *Mater. Today*, 2007, **10**, 40–48.
4. K. Radhakrishnan and A. M. Raichur, *Chem. Commun.*, 2012, **48**, 2307–2309.
5. H. Li, S. S. Yu, M. Miteva, C. E. Nelson, T. Werfel, T. D. Giorgio, and C. L. Duvall, *Adv. Funct. Mater.*, 2013, **23**, 3040–3052.
6. S. Biswas, K. Kinbara, T. Niwa, H. Taguchi, N. Ishii, S. Watanabe, K. Miyata, K. Kataoka, and T. Aida, *Nat. Chem.*, 2013, **5**, 613–620.
7. V. C. Özalp and T. Schäfer, *Chem. – Eur. J.*, 2011, **17**, 9893–9896.
8. A. E. Felber, M.-H. Dufresne, and J.-C. Leroux, *Adv. Drug Deliv. Rev.*, 2012, **64**, 979–992.
9. Y. Qiu and K. Park, *Adv. Drug Deliv. Rev.*, 2012, **64**, Supplement, 49–60.
10. Q. Yin, J. Shen, Z. Zhang, H. Yu, and Y. Li, *Adv. Drug Deliv. Rev.*, 2013.
11. N. Graf and S. J. Lippard, *Adv. Drug Deliv. Rev.*, 2012, **64**, 993–1004.
12. Y.-J. Pan, Y.-Y. Chen, D.-R. Wang, C. Wei, J. Guo, D.-R. Lu, C.-C. Chu, and C.-C. Wang, *Biomaterials*, 2012, **33**, 6570–6579.
13. R. de la Rica, D. Aili, and M. M. Stevens, *Adv. Drug Deliv. Rev.*, 2012, **64**, 967–978.
14. I. I. Slowing, J. L. Vivero-Escoto, C.-W. Wu, and V. S.-Y. Lin, *Adv. Drug Deliv. Rev.*, 2008, **60**, 1278–1288.
15. B. G. Trewyn, S. Giri, I. I. Slowing, and V. S.-Y. Lin, *Chem. Commun.*, 2007, **31**, 3236–3245.
16. Z. Luo, K. Cai, Y. Hu, L. Zhao, P. Liu, L. Duan, and W. Yang, *Angew. Chem. Int. Ed.*, 2011, **50**, 640–643.
17. Q. Zhang, F. Liu, K. T. Nguyen, X. Ma, X. Wang, B. Xing, and Y. Zhao, *Adv. Funct. Mater.*, 2012, **22**, 5144–5156.
18. B. Chang, D. Chen, Y. Wang, Y. Chen, Y. Jiao, X. Sha, and W. Yang, *Chem. Mater.*, 2013, **25**, 574–585.
19. D. Tarn, M. Xue, and J. I. Zink, *Inorg. Chem.*, 2013, **52**, 2044–2049.
20. Q. He, Y. Gao, L. Zhang, Z. Zhang, F. Gao, X. Ji, Y. Li, and J. Shi, *Biomaterials*, 2011, **32**, 7711–7720.
21. C.-H. Lee, S.-H. Cheng, I.-P. Huang, J. S. Souris, C.-S. Yang, C.-Y. Mou, and L.-W. Lo, *Angew. Chem. Int. Ed.*, 2010, **49**, 8214–8219.
22. C. Coll, L. Mondragón, R. Martínez-Mañez, F. Sancenón, M. D. Marcos, J. Soto, P. Amorós, and E. Pérez-Payá, *Angew. Chem. Int. Ed.*, 2011, **50**, 2138–2140.
23. A. Bernardos, E. Aznar, M. D. Marcos, R. Martínez-Mañez, F. Sancenón, J. Soto, J. M. Barat, and P. Amorós, *Angew. Chem. Int. Ed.*, 2009, **48**, 5884–5887.
24. A. Schlossbauer, J. Kecht, and T. Bein, *Angew. Chem. Int. Ed.*, 2009, **48**, 3092–3095.
25. T. Honda, T. Kaneiwa, S. Mizumoto, K. Sugahara, and S. Yamada, *Biomolecules*, 2012, **2**, 549–563.
26. R. Stern and M. J. Jędrzejewski, *Chem. Rev.*, 2006, **106**, 818–839.
27. E. Harris and P. Weigel, *Biochem. -- Fac. Publ.*, 2009.
28. Z. Chen, Z. Li, Y. Lin, M. Yin, J. Ren, and X. Qu, *Chem. Weinh. Bergstr. Ger.*, 2013, **19**, 1778–1783.