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COMMUNICATION

Traumatic brain injury induces elevation of Co in the human brain

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Traumatic brain injury (TBI) is the most common cause of death and disability in young adults, yet the molecular mechanisms that follow TBI are poorly understood. We previously reported a perturbation in iron (Fe) levels following TBI. Here we report that the distribution of cobalt (Co) is modulated in post-mortem human brain following injury. We also investigated how the distribution of other biologically relevant elements changes in TBI. Cobalt is increased due to TBI while copper (Cu), magnesium (Mg), manganese (Mn), phosphorus (P), potassium (K), rubidium (Rb), selenium (Se) and zinc (Zn) remain unchanged. The elevated Co has important implications for positron emission tomography neuroimaging. This is the first demonstration of the accumulation of Co in injured tissue explaining the previous utility of ⁵⁵Co-PET imaging in TBI.

Traumatic brain injury (TBI) is a major health and socioeconomic problem worldwide which according to the World Health Organization projection will become the major cause of death and disability by 2020. Annually there are an estimated 10 million people affected by TBI.¹ Repeated or moderate to severe TBI is suspected to be a risk factor for the development of Alzheimer's disease² and sporadic Parkinson's disease.³ In humans the molecular mechanisms leading to neurodegeneration and poor neurological outcome remain unclear.

Transition elements, although in trace amounts, are vital for biological function and for all facets of life. Approximately half of all enzymes are metalloenzymes.⁴ Zinc has been implicated as having a positive impact on the outcome of TBI in mouse models,^{5,6} but there is a lack of information on how metals change as a result of TBI in humans, save for a recent report of iron (Fe) accumulation post-injury.⁷ In this study we analyze the level of cobalt (Co), copper (Cu), iron (Fe), magnesium (Mg), manganese (Mn), phosphorus (P), potassium (K), rubidium (Rb), selenium (Se) and zinc (Zn) in human brain tissue of patients who died from severe traumatic brain injury.

All procedures were conducted in accordance with the Australian National Health and Medical Research Council's National Statement on Ethical Conduct in Human Research (2007), the Victorian Human Tissue Act (1982), the National Code of Ethical Autopsy Practice (2002) and the Victorian Government policies and practices in relation to post-mortem.

Trauma brain samples from 27 individuals who died after closed head injury were obtained from the Australian Neurotrauma Tissue and Fluid Bank. Cases were aged between 17 and 78 years (mean = 48 years) and the causes of injury include motor vehicle accident, motorbike accident, nursing home accident, household accident, stair accident and falls. The post-mortem intervals varied between 33 and 129 hours (mean = 81 hours). Patients were divided in 3 groups to compare the acute and delayed times after injury: 10 cases (8 males and 2 females) had survival time of less than 17 minutes, designated 'acute' group, when death occurred upon arrivals of the paramedics (cases 1-10); 8 cases (7 males and 1 female) were selected with a survival time between 30 minutes and 3 hours (mean = 1 hour) and designated as 'early' group (cases 11-18); and 9 cases (7 males and 2 females) had survival time between 6 and 122 hours (mean = 43 hours) and designated 'late' group (cases 19-27). The late cohort also contains tissue from the same area on the non-injured (late-NI) side of the brain. The brain region analyzed was located in proximity of the injured tissue and was identified macroscopically by a neuropathologist (Dr McLean). Control brain samples of 10 individuals, aged between 16 and 78 (mean = 56 years), without brain injury or other neuropathology were obtained from the National Neural Tissue Resource Centre of Australia (cases 28-37). Clinical information and epidemiological details of all patients are described in Table 1.

Approximately 0.25g of frozen tissue was allowed to thaw from -80 °C on ice and then homogenized using a BioMasher (Omni International). Tissue was placed in the BioMasher, the plunger was inserted and then the apparatus was centrifuged at 10,000 rpm with a benchtop centrifuge. After centrifugation Tris buffer saline (TBS, 50 mM Tris pH 8.0, 150 mM NaCl) containing EDTA free protease

Table 1 Details of the 27 trauma and 10 control cases. Cases 1-10: cases with a survival time between 0 and 17 minutes; Cases 11-18: cases with a survival time between 30 minutes and 3 hours; cases 19-27: Cases with a survival time between 6 and 261 hours; Cases 28-37: control cases. All brains were obtained at autopsy. PMI = post mortem interval (time between death and brain retrieval); M = male; F = female.

Case	Age	Sex	Cause of Injury	PMI	Cause of death	Survival time
1 ^a	51	M	Motor vehicle accident	60 hrs	Brain + multiple injuries	< 17 mins
2 ^a	63	M	Household accident	70 hrs	Brain injury	< 17 mins
3 ^a	27	M	Suicide	84 hrs	Brain + multiple injuries	< 17 mins
4 ^a	41	M	Suicide	96 hrs	Brain + multiple injuries	< 17 mins
5 ^a	57	F	Motor vehicle accident	87 hrs	Brain + multiple injuries	< 17 mins
6 ^a	49	M	Motor vehicle accident	107 hrs	Brain + multiple injuries	< 17 mins
7 ^a	45	M	Motor vehicle accident	43 hrs	Brain + multiple injuries	< 17 mins
8 ^a	21	M	Motor vehicle accident	100 hrs	Brain injury	< 17 mins
9 ^a	41.3	M	Aviation accident	114 hrs	Brain + multiple injuries	< 17 mins
10 ^a	57.6	F	Motor vehicle accident	97 hrs	Brain injury	< 17 mins
11 ^b	16.8	M	Motor vehicle accident	85 hrs	Brain + multiple injuries	< 3 hrs
12 ^b	78.7	M	Household accident	45 hrs	Brain injury	< 3 hrs
13 ^b	18.3	M	Motor vehicle accident	79 hrs	Brain + multiple injuries	< 3 hrs
14 ^b	34.7	M	Motorbike accident	66 hrs	Brain + multiple injuries	< 3 hrs
15 ^b	22.9	F	Motor vehicle accident	108 hrs	Brain + multiple injuries	< 3 hrs
16 ^b	52.8	M	Motorbike accident	65 hrs	Brain + multiple injuries	< 3 hrs
17 ^b	19.6	M	Suicide	33 hrs	Brain + multiple injuries	< 3 hrs
18 ^b	59.8	M	Motor vehicle accident	71 hrs	Brain + multiple injuries	< 3 hrs
19 ^c	46.0	M	Fall	129 hrs	Brain injury	6 hrs
20 ^c	56.3	M	Motor vehicle accident	65 hrs	Brain injury	8 hrs
21 ^c	64.6	M	Fall	61 hrs	Brain injury	8 hrs
22 ^c	75.9	M	Staircase fall	89 hrs	Brain injury	10 hrs
23 ^c	59.6	F	Motor vehicle accident	80 hrs	Brain injury	35 hrs
24 ^c	61.7	M	Fall	40 hrs	Brain injury	93 hrs
25 ^c	38.9	F	Staircase fall	101 hrs	Brain injury	122 hrs
26 ^c	70.9	M	Motor vehicle accident	114 hrs	Brain injury	76 hrs
27 ^c	73.7	M	Fall	91 hrs	Brain injury	29 hrs
28 ^d	16	M	-	-	Suicide by hanging	-
29 ^d	48.7	M	-	50 hrs	Cardiac failure	-
30 ^d	51.6	M	-	64 hrs	Asthma	-
31 ^d	52.3	M	-	52 hrs	Cardiomyopathy	-
32 ^d	59.6	M	-	43 hrs	Pulmonary embolism	-
33 ^d	64.1	M	-	24 hrs	Ischaemic heart disease	-
34 ^d	66.9	M	-	10 hrs	Pneumonia	-
35 ^d	64.4	M	-	24 hrs	Pulmonary embolism	-
36 ^d	77.5	M	-	53 hrs	Myocardial infarction	-
37 ^d	60	F	-	48 hrs	Myocardial infarction	-

^a Acute group; ^b Early group; ^c Late group; ^d Control group

inhibitors (Roche) was added at a ratio of 1:4 (w/v). The sample was centrifuged at 175,000 *g* for 30 minutes at 4 °C. The TBS supernatant was collected and stored at -80 °C before analysis. The resulting pellet was washed with a volume of TBS equal to the amount used for homogenization and centrifuged for 15 minutes at 175,000 *g* at 4 °C.

The TBS wash supernatant was removed and the pellet was resuspended in 7 M urea 2 M thiourea 4% CHAPS 30 mM bicine pH 8.5 and centrifuged as before. The resulting pellet was then incubated with 70% formic acid for 16 hours at room temperature before being centrifuged at 16,000 *g* for 30 minutes.

Table 2 Distribution of trace elements in different cellular fractions and total levels from human brain. Concentration based on wet weight of tissue, mean \pm standard deviation, range in parentheses. $n = 34$. * excludes values from the 'acute' cohort ($n = 25$); ** excludes values from the 'late' cohort ($n = 25$); reported in Ayton *et al.*⁷

	Co (ng g ⁻¹)	Cu (μ g g ⁻¹)	Fe (μ g g ⁻¹)	Mg (μ g g ⁻¹)	Mn (ng g ⁻¹)	P (mg g ⁻¹)	K (mg g ⁻¹)	Rb (μ g g ⁻¹)	Se (ng g ⁻¹)	Zn (μ g g ⁻¹)
Soluble	1.0 \pm 0.7 (0.3-2.7)**	2.2 \pm 0.9 (0.8-4.4)	8.7 \pm 4.2 (2.5-25.3)*	58.7 \pm 28 (21-189)	51 \pm 24 (19-145)	0.5 \pm 0.3 (0.2-1.6)	1.5 \pm 0.7 (0.4-4.4)	2.2 \pm 1.0 (0.6-5.3)	57 \pm 26 (19-152)	2.8 \pm 1.1 (1.3-7.6)
Membrane	1.4 \pm 0.6 (0.4-3.3)	1.3 \pm 0.4 (0.6-2.3)	36.9 \pm 10 (19-63)	34.5 \pm 10 (19-73)	118 \pm 34 (56-206)	1.0 \pm 0.4 (0.3-1.9)	0.4 \pm 0.1 (0.2-1.0)	0.6 \pm 0.2 (0.3-1.4)	79 \pm 35 (0.5-150)	5.1 \pm 2.6 (1.9-18)
Formic Acid	3.5 \pm 1.3 (2-9)	1.3 \pm 0.3 (0.8-2.1)	32.3 \pm 12 (10-55)	42.0 \pm 14 (17-78)	119 \pm 31 (66-210)	1.2 \pm 0.3 (0.7-2.1)	0.3 \pm 0.1 (0.1-0.6)	0.4 \pm 0.2 (0.2-0.9)	134 \pm 35 (70-260)	6.3 \pm 1.6 (4-11)
Total^a	5.8 \pm 2 (4+10)*	4.8 \pm 1.2 (2.5-7.5)	76.0 \pm 17 (39-107)**	136.2 \pm 36 (63-255)	291 \pm 60 (198-425)	2.7 \pm 0.6 (1.7-4.2)	2.2 \pm 0.7 (0.8-5.0)	3.2 \pm 1.0 (1.2-6.0)	270 \pm 60 (123-400)	14.3 \pm 4.4 (8.6-29.5)

^a Total = soluble + membrane + formic acid.

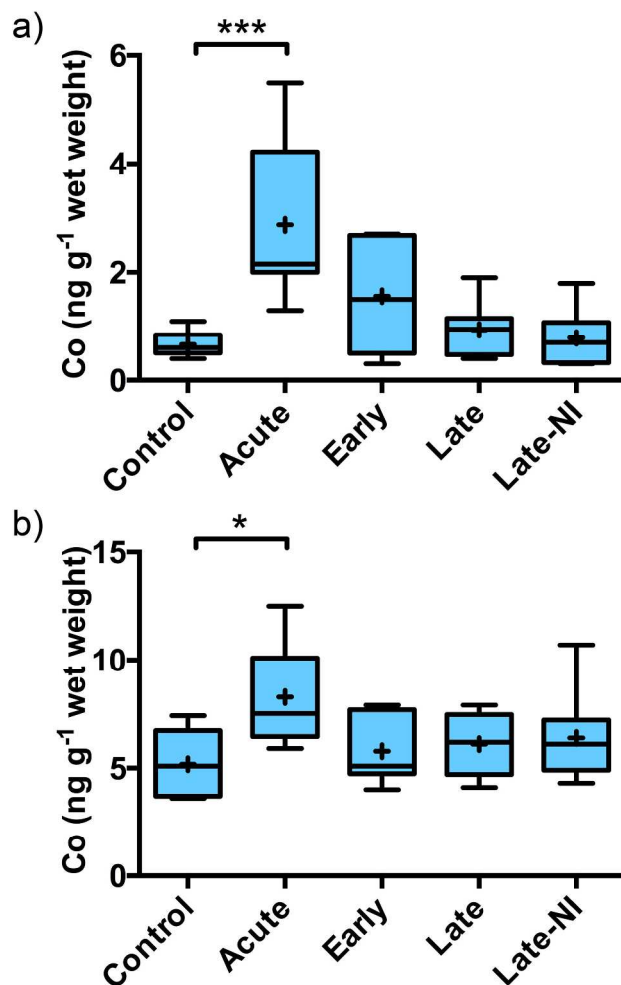


Fig. 1 Elevated levels of Co in the human brain following TBI as measured by ICP-MS. a) Co is significantly elevated in the acute TBI cases in the TBS soluble extracted material compared to all other groups (*** Tukey post-hoc test $p < 0.001$) and b) in the total Co levels calculated as the sum of soluble, membrane and formic acid extracted Co is also significantly elevated but to a lesser extent (* Tukey post-hoc test $p < 0.05$). Boxes represent interquartile range; error bars represent minimum and maximum values; + represents mean and line represents median.

Inductively coupled plasma mass spectrometry (ICP-MS) was used to determine the quantity of Co, Cu, Fe, Mg, Mn, P, K, Rb, Se and Zn in the TBS soluble, membrane and formic acid extracted homogenates. Supernatants were diluted 1:15 with 1% nitric acid (Suprapur, Merck).

Measurements were made using an Agilent 7700 series ICP-MS instrument under routine multi-element operating conditions using a helium collision gas cell. The instrument was calibrated using 0, 5, 10, 50 and 100 ppb of certified multi-element ICP-MS standard calibration solutions (ICP-MS-CAL2-1, ICP-MS-CAL-3 and ICP-MS-CAL-4, Accustandard) for a range of elements. We used a certified internal standard solution containing 200 ppb of yttrium (⁸⁹Y) as an internal control (ICP-MS-IS-MIX1-1, Accustandard). The sample was introduced via the automated liquid sampler (Agilent) using a peristaltic pump at a flow rate of 0.4 mL min⁻¹.

Statistical analysis was performed using SigmaStat (SysStat, San Jose, CA), Prism 5.0 (GraphPad, La Jolla, CA) and SPSS software (SPSS Inc., Chicago, IL). Kolmogorov-Smirnov test (with Lilliefors' correction) was used to test data for normality within each group and values were transformed by natural logarithm calculation if required. One-way ANOVA was followed by multiple comparisons using the Tukey's *post-hoc* test to identify significant difference between trauma and control groups. Statistical significance was considered at the 5% level ($p < 0.05$).

Quantitative measurement of Co, Cu, Fe, Mg, Mn, P, K, Rb, Se and Zn was performed on the three extracted pools of material: soluble, membrane and formic acid (Table 2). The total level of each element measured were consistent with the total level per gram of wet tissue for Co, Cu, Fe, Mg, Mn, P, K, Rb, Se and Zn levels previously reported for normal human brain tissue.⁸⁻¹⁰ Cobalt and Fe (previously reported by Ayton *et al.*⁷) were the only elements that changed significantly in response to TBI. Co was significantly elevated in the acute TBI patients (Fig. 1a). The elevation of Co was restricted to the soluble fraction with no evidence of a change in Co levels in the membrane or FA fractions (Table 2). Although the elevation in Co remained significantly elevated in the total (Fig. 1b, [total] = [soluble + membrane + FA]) the signal to noise was much greater due to the addition of the membrane and FA fractions. The elevation of Co returned to baseline 3-hours post-injury.

Finally, the fractionation protocol allowed a more detail analysis about how the elements are distributed in the tissue (Fig. 2). Analyzing the proportion of an element in each fraction showed that elements K and Rb are greater than 70% associated with the soluble extracted material consistent with these elements being free ions.¹¹ Alternatively, elements largely associated to membrane bound proteins, such as Fe, had greater than 80% of the element associated with the membranous pellet (membrane plus FA fraction) consistent with Fe being a cofactor

for membrane associated proteins involved in the electron transport chain, for example.

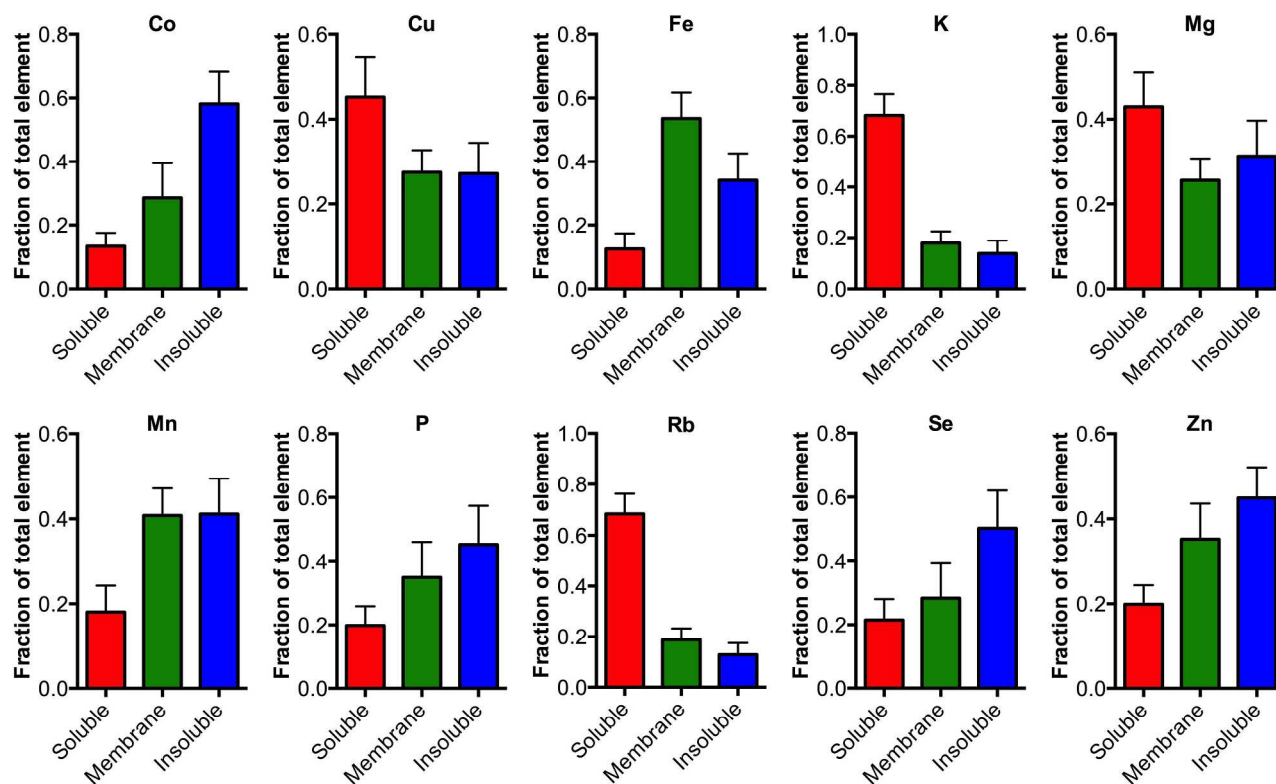


Fig. 2 Distribution of elements in the cellular fractions. The elements Co, Cu, Fe, Mg, Mn, P, K, Rb, Se and Zn were measured in the soluble, membrane and formic acid fractions using ICP-MS, no significant changes were observed between the tissue from TBI and controls (one-way ANOVA) except for Co as noted in Figure 1. The fraction of each element in the respective pool is graphed (mean \pm 1 standard deviation). As expected elements that are truly free ions such as Rb and K are mostly distributed in the soluble phase.

Calcium (Ca) overload and inflammatory process are both attributed to the resulting neuronal death. Co is used as a surrogate marker of Ca accumulation in degenerating neurons¹². Radioisotopes of Co have been successfully used to detect ischemic damage, extravasation or inflammation in several neurodegenerative pathologies.¹³⁻¹⁵ This is the first study to demonstrate that TBI induces the specific uptake of *in situ* Co into brain tissue. Cobalt is predominantly found as cobalamin (vitamin B₁₂), which is used as a cofactor for methyl transfer reactions that are vital for DNA synthesis and fatty acid synthesis. Alternatively, Co is also utilized in a vitamin B₁₂ independent fashion in the enzyme methionine aminopeptidase 2,¹⁶ which removes the N-terminal methionine of newly synthesized proteins and plays a vital role in the angiogenesis of blood vessels and as such is a target of anticancer angiogenesis compounds. Long-term hypoxia in mice has been shown to both elevate brain Co levels and increase activity of vitamin B₁₂.¹⁷ Previous positron emission tomography studies have used ⁵⁵CoCl₂ to detect areas of ischemic damage, suggesting that the observed increase in Co in acute patients may also be due to an influx of non-vitamin B₁₂ Co. Additionally, the cellular fractionation of the tissue into soluble and membrane pools demonstrate that the increased Co is due to an actual accumulation of Co, likely from the blood. There is an emerging role for cobalamin in regulating the production of growth factors and cytokines in the CNS¹⁸ and this may be related to the increased uptake of this element in

response to TBI. Vitamin B₁₂ and its cobalamin products,¹⁹ specifically the thiolato- derivatives²⁰ have also been shown to possess potent antioxidant properties *in vitro*, which may explain the rapid influx of Co to brain regions under increased oxidative stress, as is the case following acute TBI.²¹

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Notes and references

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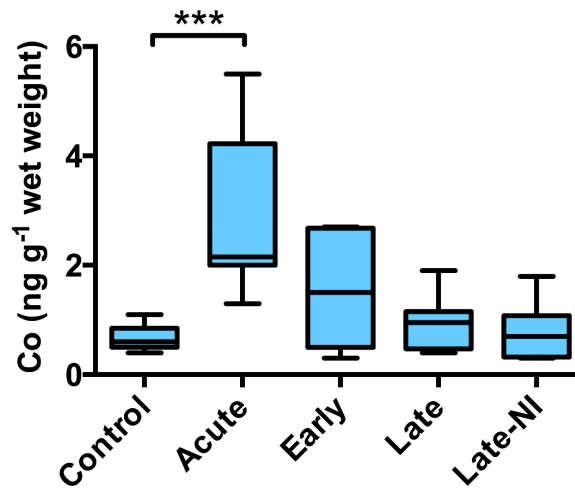
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4 **Graphical abstract:**
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Following acute traumatic brain injury (less than three hours post-event), cobalt levels in the brain are significantly elevated. This elevation in brain cobalt levels may have important implications for positron emission tomography neuroimaging for assessing traumatic brain injury severity.