

Metallomics

Accepted Manuscript



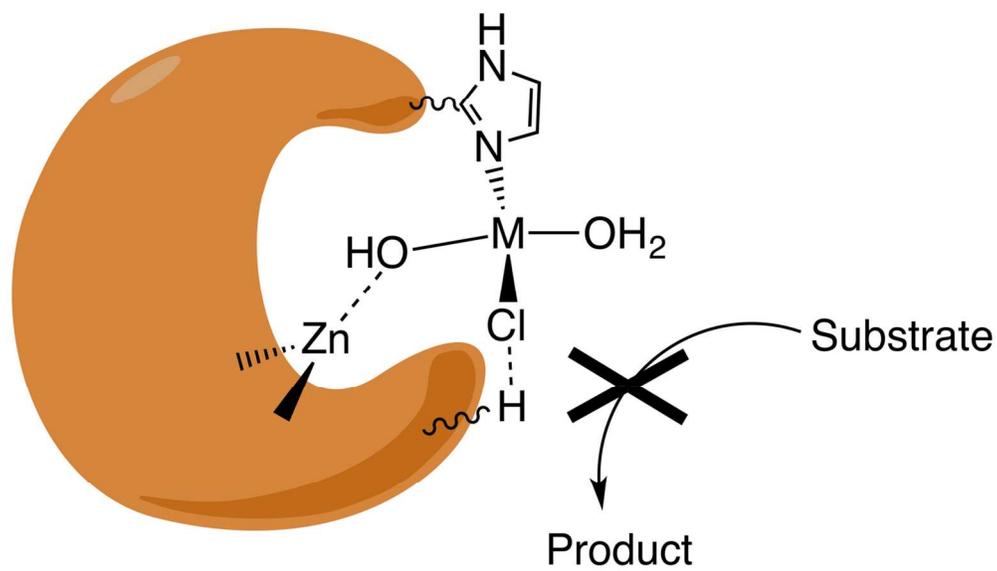
This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1
2
3 Metal complexes modulate the degradation of aggregation prone substrates by metalloproteases and
4
5
6 can be used to tackle conformational diseases.
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



66x38mm (600 x 600 DPI)

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Metal complexes and metalloproteases: targeting conformational diseases

Giuseppe Grasso,^{1*} Sylvestre Bonnet²

1 Chemistry Department, Università di Catania, Viale Andrea Doria 6, 95125, Catania, Italy

2 Leiden University, Leiden Institute of Chemistry, Gorlaeus Laboratories, P.O. Box 9502,
2300 RA Leiden

* Correspondence to:

Giuseppe Grasso, Dipartimento di Scienze Chimiche, Università di Catania, Viale Andrea Doria 6,
95125, Catania, Italy. e-mail: grassog@unict.it

Table of Contents

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abstract	3
Keywords	4
1. Metalloproteases and conformational diseases	5
2. Metal complexes as metalloproteases inhibitors	7
3. Metal complexes as metalloprotease activators	12
4. Metal complexes as artificial metalloproteases	14
4. Conclusions and perspective	17
6. Acknowledgements	18
Tables.....	19
References.....	21
Figures and Tables Legends.....	34

Abstract

In recent years many metalloproteases (MPs) have been shown to play important roles in the development of various pathological conditions. Although most of the literature is focused on matrix MPs (MMPs), many other MPs have been demonstrated to be involved in the degradation of peptides or proteins whose accumulation and dyshomeostasis are considered as being responsible for the development of conformational diseases, i.e., diseases where non-native protein conformations lead to protein aggregation. It seems clear that, at least in principle, it must be possible to control the levels of many aggregation-prone proteins not only by reducing their production, but also by enhancing their catabolism. Metal complexes that can perform this function were designed and tested according to at least two different strategies: i) intervening on the endogenous MPs by directly or indirectly modulating their activity; ii) acting as artificial MPs, replacing or synergistically functioning with endogenous MPs. These two different bioinorganic approaches are widely represented in the current literature and the aim of this review is to rationally organize and discuss both of them so to give a critical insight on these approaches and highlighting their limitations and future perspectives.

Keywords

Metal ions, neurodegenerative disease; cancer; Alzheimer's disease; metalloproteinase activity.

1. Metalloproteases and conformational diseases

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Metalloenzymes comprise a vast family of proteins which are involved in many physiological processes fundamental for the life of cells and, more generally, of living organisms. There are many ways to describe and sort metalloenzymes discovered so far. One common and rational way is to group them according to the metal ion of the catalytic site that is in charge of the biological function of the metalloenzyme.¹ Following this method, MPs can be classified into zinc-, copper-, iron-, nickel-, or manganese-containing enzymes, where the specific inorganic features of the particular metal ion give to the enzyme its specific properties. In particular, zinc always stays in the +2 oxidation state as it cannot be oxidized nor reduced in biological conditions. For this reason, a large part of metalloenzymes functioning as metalloproteases (MPs) are zinc-containing MPs (ZnMPs). ZnMPs participate in biological reactions encompassing the degradation of all major metabolites (carbohydrates, lipids, nucleic acids, and proteins/peptides), and they can be classified according to their location² or main function.³ In ZnMPs the metal ion has a catalytic role, and the mechanism for substrate degradation has been already thoroughly investigated. In the catalytic site, zinc is maintained in position by coordinating residues but it is also bound to water, thus generating a hydroxide ion that can attack the protein substrate (polarization assisted zinc water catalysis) or induce the formation of nucleophiles.⁴

Another method to sort MPs would be to identify which ones are involved with a particular disease. However, it is practically impossible to group univocally the various existing MPs and to associate them with a single disease, as most MPs seem to be involved in the development of several pathologies. In this perspective, matrix MPs (MMPs) certainly hold the sceptre of the multifunctional MP, as they play an important role in tissue remodeling, which is itself associated with various physiological and pathological processes such as angiogenesis,⁵ morphogenesis,⁶ tissue repair,⁷ cirrhosis,⁸ systemic sclerosis,⁹ and metastasis.¹⁰ Recent data suggested an active role of MMPs in the pathogenesis of aortic aneurysm, and a dysregulation of the balance between MMPs and their natural inhibitors (TIMPs) is also a characteristic of acute and chronic

1 cardiovascular diseases.¹¹ Indeed, the majority of MPs are involved in somewhat very different
2 pathologies so that the only meaningful categorization can be carried out the other way around, that
3 is, by listing all the MPs that are involved with a particular disease. In this review we will focus on
4 conformational diseases (CDs), that, by definition, are diseases caused by mutations or other
5 changes in the structure of specific proteins that lead to their aggregation and deposition.
6
7

8 Protein misfolding is an intrinsic aspect of normal folding within the complex cellular environment,
9 and its effects are minimized in living systems by the action of a range of protective mechanisms
10 that include molecular chaperones and quality control systems. MPs play a very important role in
11 regulating and controlling protein misfolding. Indeed, in physiological conditions MPs contribute to
12 lower the level of aggregation-prone proteins by degrading them if their concentration is above
13 normal levels. However, several environmental factors can contribute to protein misfolding such as
14 interaction with metal ions,^{12,13} interaction with small molecules,¹⁴ pH changes,¹⁵ etc.¹⁶ Misfolded
15 proteins have a tendency to aggregate to form a variety of species including the highly organized
16 and kinetically stable amyloid fibrils, which is considered as the main culprit for the development
17 of widespread pathological conditions such as Alzheimer's Diseases (AD), Parkinson Disease (PD),
18 or prion diseases. Amyloid fibrils and their precursors (oligomers) appear to have adverse effects on
19 cellular functions regardless of the sequence of the component peptide or protein. Table 1 lists the
20 most widespread CDs that have been associated with the aggregation of a specific protein,^{17,18}
21 together with the MPs which are known to be involved either directly in the catabolism of the
22 aggregation-prone protein, or indirectly with the disease. It is important to note that while some
23 MPs seem to be specific for a particular disease, many others are involved in different diseases.
24 Therefore, the fact that most MPs have multiple biological roles in living organisms makes the
25 design of drugs targeting specific MPs a very challenging task, all the more to tackle one particular
26 disease. For this reason, in addition to the efforts put by the scientific community into the design of
27 specific MPs modulators, new therapeutic horizons have been opened by the use of metal
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 complexes (MCs), which have catalytic functions mimicking the ones of the endogenous MPs. Both
2
3 approaches will be reviewed and discussed in the next sections.
4
5
6
7
8

9 **2. Metal complexes as metalloproteases inhibitors**

10
11 Because the activity of many MPs can be linked, directly or indirectly, to the development of
12
13 specific diseases, the investigation of biomolecular mechanisms responsible for the alteration of
14
15 enzymes activities,¹⁹⁻²² and new analytical approaches that are able to assess enzymes kinetics,²³⁻²⁶
16
17 have attracted much scientific interest in recent years. In particular, MCs are one of the most used
18
19 molecules used to inhibit MPs, and, before reviewing the molecular details of these inhibitors, some
20
21 definitions must be given. According to a recent review from Kilpin *et al.*²⁷ MCs inhibitors can
22
23 inhibit a protein through *i)* only one of their ligands; *ii)* only the metal center; *iii)* both the metal
24
25 center and one of the ligands. Most of the difficulties encountered by the scientific community
26
27 consist in obtaining potent and selective MP inhibitors that are able to inhibit one given MP, for
28
29 example by chelation of its zinc ion, but without interacting with metal ions of other MPs.
30
31 Therefore, the MC design has to take into account both the issues of drug delivery²⁸ and
32
33 coordination chemistry, i.e. the MC has to get the proper localization without undergoing collateral
34
35 reactions and then has to be properly functionalized so to interact specifically with its selected
36
37 target. The first metal complexes that were shown to inhibit MPs were “free” aluminium ions made
38
39 by dissolving AlCl₃ in aqueous solution. Calpain, cathepsin D, trypsin and α-chymotrypsin²⁹ were
40
41 shown to be inhibited by Al³⁺, which is supposed to be able to substitute Mg²⁺ or Fe³⁺ ions in
42
43 magnesium- or iron-dependent enzymes because of the similar ionic radius of these three ions. Such
44
45 substitution is believed to be the cause of enzyme inhibition, and addition of the strong metal
46
47 chelator EDTA sometimes was shown to revert inhibition.
48
49
50
51
52

53
54 Obviously, the absence of organic ligands that would make specific interactions with the binding
55
56 pocket of the MP, rules out selectivity during enzyme inhibition by “free” metal ions. For this
57
58 reason, various research groups have developed small organic molecules that show selective MP
59
60

1 inhibition by employing a zinc-binding group (ZBG) to bind the active site metal ion.³⁰ This was
2 usually carried out by using a strong chelator such as hydroxamic acid. However, the use of
3 hydroxamic acids ZBG has been shown to be limited by poor pharmacokinetics, low oral
4 bioavailability, hydrolytic liability, and inadequate selectivity for zinc, which may account for most
5 failures of those inhibitors in clinical trials.³¹ In recent years, the search for new MPIs with major
6 changes in the zinc(II) chelating groups appeared as a challenge for several research groups.^{32,33}
7 New ZBGs such as hydroxypyrimidinone³⁴ or 1-hydroxypiperazine-2,6-diones³⁵ have been
8 explored. In this perspective, it is important to realize that the zinc(II) ion is a borderline Lewis acid
9 that is able to interact with a variety of donor atoms such as sulfur, nitrogen, or oxygen. In order to
10 design an effective and selective Zn-chelating drug it is therefore important to consider the
11 thermodynamic ability of the metal chelator to coordinate the metal ions and to compete with the
12 biological ligands that bind to the cofactor in the enzymatic environment.³⁶ The wide development
13 of pharmacological inhibitors of MMPs (known as MMPIs) as potential anti-cancer agents began in
14 the 1980s, but the clinical use of these agents in oncology has been brought to a crashing halt with
15 the repeated failure of various MMPIs in multiple large-scale phase III clinical trials.³⁷ Recently,
16 instead of the hydroxamic acid moiety, carboxylates have been proposed as a chelating agent that
17 could function as a less strong and more selective MMPs inhibitor with a safer toxicity profile.³⁸
18 Other functional groups that may function as ZBGs are phosphinic acid, heterocycles,
19 pyrimidinetrione, thiol diketopiperazine, carbamoyl phosphonate, and sulfone N-
20 formylhydroxylamine.⁹⁴ The general active site of MMPs is a groove, usually presented
21 horizontally across the protein with three un-primed sub-sites to the left of the coordinated zinc and
22 three primed sub-sites to the right (S3-S2-S1-Zn-S1'-S2'-S3').³⁹ While most attempts to introduce
23 selectivity have focused on the S1' channel, other groups also investigated the possibility of
24 interaction with the S2' pocket.⁴⁰
25
26 Very recently, ruthenium, osmium,⁴¹ and platinum complexes able to inhibit MMPs through a non-
27 competitive mechanism have also been reported,⁴² for which the structure of the MCs bound to its
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 target MMP was even obtained.⁴³ The selectivity of such Pt complexes seems to be higher than that
2
3
4 of classic organic inhibitors based on Zn chelation, since they seem to interact with specific amino
5
6 acid residues of the binding pocket, which are different for each specific MMP. Figure 1 shows the
7
8 structures of catalytic domains of MMPs and a model of their interaction with a specific platinum
9
10 complex. The binding sites are in regions critical for the enzymatic activity, in particular, His224
11
12 for MMP-3 and Met404 for MMP-9 (both located in the S1' specificity loop), or Met87 for MMP-
13
14 12, which ends up very close to the active site upon formation of a salt bridge between the N-
15
16 terminal Phe84 and Asp237. It has been hypothesized by the authors that, after interaction of
17
18 [PtCl₂(smp)] at primary anchoring sites on the MMPs surface, other binding events can take place,
19
20 further stabilizing the inhibitor-enzyme interaction. For this reason, another MMP tested in this
21
22 study, MMP-2, has a very low inhibition by [PtCl₂(smp)] because it lacks potential anchoring
23
24 residues in such a favorable position to allow the concerted binding event.
25
26

27
28 Metal ions different from platinum have also been used as MMP inhibitors (see Figure 2). The
29
30 ligands bound to the metal center sometime play an important role on the inhibition properties of
31
32 the MC. For example, [Co(acacen)(NH₃)₂]Cl,⁴⁴ a complex with two substitutionally labile *trans*
33
34 ammonia ligands, is almost ten times more potent as inhibitor of MMP-9 than [Co(acacen)(Im)₂]Cl,
35
36 an analogue complex containing the substitutionally less labile imidazole ligands (Im). Binding of
37
38 coordinating residues of the protein to the Co center must play a role in this case. By covalent
39
40 attachment of the metal fragment with known MMP-inhibiting biphenylamide or biphenylsulfonate
41
42 organic fragments, organic-inorganic bifunctional compounds showed 3- to 6-fold higher activities,
43
44 with IC₅₀ values down to 0.83 μM. Later on, the binding mechanism of [Co(acacen)(NH₃)₂]Cl to
45
46 MMP-2 was investigated, and inhibition was shown to be irreversible, time-dependent, and to
47
48 involve both the structural and catalytic histidine sites. According to kinetic analysis,⁴⁵ reversible
49
50 formation of an enzyme-inhibitor complex occurs first (Figure 3a), followed by an irreversible
51
52 reaction (Figure 3b). The kinetics of irreversible metal binding increased with temperature. Thus,
53
54 metal binding to the protein involves structure loss of the protein upon binding ($\Delta S > 0$) and ligand
55
56
57
58
59
60

1
2 loss before the histidine can bind ($\Delta H > 0$). Overall, this study is one of the few quantitative analysis
3
4 of the thermodynamic and kinetics of the irreversible inhibition of an MMP by a transition metal
5
6 complex.
7

8
9 However, selectivity in the series of the tested MMPIs was not as good as anticipated, and specific
10
11 MMPIs have proven to be extremely difficult to obtain. This is due to the fact that MMPs are quite
12
13 similar structurally, particularly with respect to the catalytic site. In addition, there is evidence for
14
15 crosstalk among the different protease classes that can possibly lead to compensatory mechanisms
16
17 if one class is inhibited. Of course, even a highly specific inhibitor will be of no benefit if
18
19 modulation of its target interferes with anti-angiogenic programs or immune cell-mediated tumor
20
21 destruction. For this reason, the roles of MMPs in each cancer and disease setting must be very well
22
23 understood. For example, inhibition of MMP-9 may be useful in preventing the priming of sites for
24
25 metastatic formation, but once metastasis has occurred and has engaged an angiogenic program,
26
27 MMP-9 inhibition could promote angiogenesis and lead to larger and more aggressive metastatic
28
29 lesions.⁴⁶ Therefore, although there is ample evidence from animal models that MMPIs could
30
31 effectively modulate disease progression, successful results regarding the use of MMPIs in human
32
33 diseases are very scarce.⁴⁷ The reader interested with more detailed information on MMPI-based
34
35 drugs that have been engaged clinical phase trials is advised to read the nice review on the subject
36
37 by B. Fingleton (Ref. 47).
38
39

40
41
42 On the other hand, the development of inhibitors of other MPs different from MMPs have been
43
44 more successful, and many drugs based on this approach are currently used in humans. For
45
46 example, ACE inhibitors have been for a long time successfully applied in the treatment of
47
48 hypertension and congestive heart failure⁴⁸ and, more recently, new MPIs which contemporarily
49
50 inhibit the catalytic function of more than one MPs have been developed.⁴⁹ The benefit of this new
51
52 class of dual or triple inhibitors (ACE/NEP; NEP/ECE and ACE/NEP/ECE) has been demonstrated
53
54 but unfortunately only a few reached the clinical development stage.⁵⁰ In order to achieve the best
55
56 results in term of specificity and inhibition activity, a thorough understanding of the relationship
57
58
59
60

1 between the structure and the properties of the designed MPIs is necessary. Moreover, the
2 investigation of the inhibitor-MP complex is very useful in order to obtain crucial information on
3 the structure-function relationship of the MPs themselves. For example, the detailed analysis of the
4 crystal structure of the phosphoramidon-NEP complex shown in Figure 4 (PDB code 1DMT)
5 allowed for explaining the enzyme's broad substrate specificity.⁴⁹ The main structural features
6 observed in most selective NEP or ACE inhibitors are a ZBG (often a SH group), a pseudo-
7 dipeptide unit, and a C-terminal carboxylate group. The ZBG has a strong impact on many factors
8 such as inhibitor potency and selectivity. This is not only due to the affinity of the ZBG for the
9 metal ion in the active site, but also to its hydrogen-bonding interactions with the protein and the
10 van der Waals contacts.⁵¹ Therefore, it is possible to optimize and design such interactions in order
11 to find the best ZBG for a given metalloenzyme. In addition, specificity toward a MP can be
12 obtained by adding a bulky portion in the P1' position of the MPI, so that, for example, NEP
13 inhibition is stronger than ACE inhibition. Indeed, the S1' subsite of NEP is able to accommodate a
14 larger group than the corresponding S1' subsite in ACE, determining a different affinity toward the
15 MPI.⁵²

16 In order to identify new chelating scaffolds for MPIs, another approach has also been applied that is
17 based on the screening of low molecular weight compounds against drug targets.³² Fragments are
18 then linked or "grown" to generate potent leads, which can be further optimized to obtain improved
19 solubility and pharmacokinetics via traditional medicinal chemistry. The main advantage of such
20 approach is that active sites are more efficiently probed by small fragments that are not limited by
21 steric constraints or hydrogen-bonding mismatches. For example, by following such method it has
22 been demonstrated that substitution at either the 5- or 7-positions of the 8-hydroxyquinoline
23 inhibitor produces potent leads against MMP-2, while derivatives at the 2- and 4-positions do not.³²
24 Finally, NEP inhibition has been also proposed as a possible treatment for type 2 diabetes,¹¹⁰ but
25 warning about the risk of reducing NEP activity is also raised by *in vivo* experiments that have
26 clearly demonstrated that a reduction in NEP activity contributes to the development of AD.⁵³ For
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 this reason, activation instead of inhibition of NEP and other MPs has been also proposed as a
2 possible therapeutic route to tackle a variety of CDs. Indeed, the reason why chelators have failed in
3 the clinic is that therapeutic use of a metal chelator may result in indiscriminate binding of essential
4 metal ions, leading to potentially harmful effects such as the scavenging of iron leading to anemia.
5
6 Therefore, in the next section the use of MCs as possible positive modulators of MPs is also
7 reviewed.
8
9
10
11
12
13
14
15
16
17

18 **3. Metal complexes as metalloprotease activators**

19
20 In Table 1 the various MPs involved with different CDs are listed. Although in the case of most
21 CDs such as cancer, atherosclerosis, etc. a meaningful therapeutic approach is represented by the
22 development of MPIs, in other widespread CDs such as AD, PD and Scrapie/Creutzfeldt-Jakob
23 disease a positive modulation of some of the involved MPs could represent instead a valid
24 therapeutic approach. Indeed, CDs are characterized by the accumulation and fibrillation of a
25 misfolded protein, and increasing the activity of some of the MPs that are partly or totally
26 responsible for the catabolism of this aggregation-prone protein may lower, in principle, the
27 concentration of this protein.⁵⁴ However, as outlined above for NEP, modulating the activity of
28 MPs involved in different biomolecular functions can also be detrimental to health. Another
29 example is indeed represented by ACE, the inhibition of which is commonly targeted in elderly
30 populations because it plays a central role in the regulation of blood pressure and hypertension.
31 However, as ACE is able to degrade A β , an up-regulation of this enzyme has been proposed as a
32 possible therapeutic route for AD, while its inhibition enhances A β deposition in the brain and is
33 therefore potentially dangerous.⁵⁵
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51

52 In this scenario, MCs were not only proposed as MPs inhibitors (see previous section), but also as
53 MPs activators. In the latter case, the up-regulation of MPs usually occurs through a multistep
54 biomolecular process, rather than by direct interaction of the MC with the enzyme. The most
55 successful application of a chelating ligand as an activator of MPs for therapeutic treatment of a CD
56
57
58
59
60

1 is probably the use of PBT2. Indeed, this drug has completed phase IIa clinical trials and is
2 currently in phase IIb for the treatment of AD^{56,57} and phase IIa against Huntington Disease.
3 Although PBT2 and its predecessor clioquinol (see Figure 5) have been initially designed as metal
4 chelators to target the ascertained metal ion dyshomeostasis involved in AD,⁵⁸ it has been later
5 recognized that their success as modulators of the A β protein level is due to their ability to work as
6 ionophores. Indeed, they are able to shuttle metal ions inside the cell, i.e., to re-establish their
7 correct spatial distribution, which seems to be one of the major problems in AD rather than a simple
8 alteration of the metal ion levels.⁵⁹ Once the metal ions are brought back inside the cell, it has been
9 demonstrated that they activate PI3K (phosphoinositide 3-kinase), which leads to the downstream
10 phosphorylation of GSK3 (glycogen synthase kinase 3) and potentiation of the MAPK
11 (mitogenactivated protein kinase) JNK (c-Jun N-terminal kinase). The stimulation of this pathway
12 culminates in up-regulation of cellular MP activity and degradation of extracellular A β .^{60,61} This
13 delicate mechanism of action is very much dependent on the coordinating ligand (PBT2) that
14 functions as a metal “chaperone”. Therefore, extensive screening studies have been carried out in
15 order to decide which ligand has the best impact on metal uptake, MAPK signalling pathways, and
16 A β levels.⁶² Indeed, an important feature of these compounds is their not-too-high affinity towards
17 metal ions. Such moderate metal affinity allows them to remove metals from extracellular amyloid
18 aggregates without chelating metals from the catalytic sites of MPs.⁶³ Therefore, in our opinion this
19 laborious but successful study represents an important milestone in the field of applied medicinal
20 bioinorganic chemistry,⁶⁴ which demonstrates the potentiality of MCs in tackling so far incurable
21 CDs.
22

23 An indirect up-regulation of MMPs has even been reported to occur with the well-known anti-
24 tumor drug cisplatin.⁶⁵ In the investigated case of thyroid PC Cl3 cells, cisplatin seems to provoke
25 an oxidant-induced MMP-2-dependent EGFR transactivation responsible for the induction of cell
26 apoptosis, a process ascribable to the intracellular signaling of PKC-e and MAPK/p38. Overall,
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 these biological studies highlight the intricate character of MC/MP interaction: even in cases when
2 a direct chemical interaction does not occur, indirect biological interaction is possible.
3

4
5
6 Finally, it is important to highlight that even endogenous MCs have been reported to stimulate MPs
7 activity.⁶⁶ Glycyl-L-histidyl-L-lysine (GHK) is a tripeptide which was isolated from human plasma
8 for the first time in 1973.⁶⁷ Either alone or in combination with copper(II) ions it was initially used
9 to make hair- and skin-care products. More recently it has also been proposed as a possible
10 therapeutic agent against age-associated neurodegeneration and cognitive decline.⁶⁸ Interestingly,
11 its copper(II) complex seems to be involved in wound remodeling, and the mechanism of action has
12 been demonstrated to be the modulation of MMP-2 expression.⁶⁶ Another MC based on zinc(II)
13 ions bound to a naturally occurring ligand, kinetin (N6-furfuryladenine), has also been
14 demonstrated to change noticeably the pro-MMP-2/MMP-2 ratio towards a higher amount of
15 mature MMP-2, therefore modulating the inflammatory response in THP-1 cells.⁶⁹ Overall,
16 although the above-mentioned examples of MCs used as MPs activators show the possibility to
17 develop new therapeutic agents, the prediction of their efficiency *in vivo* is very difficult because of
18 the indirect mechanisms of modulation of MMPs by MCs, which is often unveiled *a posteriori*. For
19 this reason, a more versatile approach that, on paper, seems to have a more predictable output, is
20 presented hereafter, which consists in the use of MCs as artificial MPs.
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42

43 **4. Metal complexes as artificial metalloproteases**

44
45 The lack of specificity of MCs as MPis spurred another area of investigation in the field of
46 bioinorganic chemistry, that is, the development of artificial proteases capable of degrading specific
47 proteins involved in CDs. Providing that reasonable target specificity could be attained, such
48 molecules could represent a new and promising class of molecular therapeutics.⁷⁰ The major
49 advantage of this approach is the catalytic nature of these drugs, which would allow for using lower
50 drug dosages and, consequently, for minimizing side effects. In the literature, very recent and
51 comprehensive reviews on this subject have been published,⁷⁰⁻⁷² so that we will only indicate the
52
53
54
55
56
57
58
59
60

1 main features allowing a MC for appearing as an artificial MP, and elucidate the biomolecular
2 mechanisms of action of the most commonly used artificial MPs.
3

4
5
6 First of all, it is important to highlight that the selective hydrolysis of peptide bonds by artificial
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

MPs is a very challenging task. Indeed, despite their name “peptides” comes from the Greek *πεπτος* – “digestable”, this kind of bonds are rather unreactive if compared for examples with esters, as they have a very long half-life in solution (at pH 4–8 their spontaneous hydrolysis would take several hundred years at room temperature!). For this reason, nature has equipped living organisms with MPs, which are highly specialized enzymes that specifically interact with the protein that has to be degraded. It has been recently recognized that the microenvironment of the active site of the various MPs is the crucial feature that determines the height of the energetic barrier for the hydrolytic cleavage of a peptide bond.⁷³ This energy barrier eventually determines the hydrolysis rate, and so far artificial MPs are well below natural MPs performances. However, artificial MPs have some advantages on natural ones, which spur the scientific community to develop efficient artificial MPs as therapeutic drugs in many of the above-listed CDs. Such advantages are: i) a better stability to heat; ii) a better compatibility with organic solvents; iii) their applicability to abiotic reactions; and iv) sometimes a better selectivity.⁷⁴ As this area progresses, research interests now focus on artificial MPs that can solve biological problems that cannot be solved by natural enzymes. Indeed, the medicinal application of natural enzymes is limited because of their special requirements, notably their sensitivity to temperature and pH. Also, many natural MPs are poorly selective and cleave peptide bond indiscriminately, to give many unwanted short fragments. On the contrary, artificial MPs can in principle be designed to be highly specific toward any particular target. Moreover, artificial MPs may also provide a new method for the detoxification of many soluble protein toxins.

An artificial MP should have key features to be an efficient drug against a CD. A first requirement is to have at least two coordination sites occupied by weak ligands that can be readily replaced, one for anchoring to the side chain of the amino acid in the peptide, and a second one for interaction

1 with the nearby peptide bond to hydrolyse.⁷⁵ The remaining coordination sites should be occupied
2
3 by one or several polydentate ligand(s) that clearly define a stable coordination sphere for the metal
4
5 ion. The perfect candidate for the metal ion should be a strong Lewis acid with limited redox
6
7 activity in biological conditions in order to avoid side reactions such as the production of reactive
8
9 oxygen species (ROS). In any case, the metal ion should be as oxophilic as possible in order to
10
11 interact with an amide carbonyl oxygen in the peptide backbone, which activates the amide
12
13 quaternary carbon toward nucleophilic attack (Figure 6a). Simultaneously, oxophilic metal ions
14
15 may deliver a hydroxide nucleophile to cleave the amide bond (Figure 6b). In addition to the metal
16
17 complex, a targeting domain should also be present in the artificial MP to control the localization of
18
19 each reactive catalyst to the protein target.⁷⁶ In some cases, an aldehyde group has been employed
20
21 as the binding site of artificial proteases because of its ability to form imine bonds with the ϵ -amino
22
23 groups of Lys residues exposed on the surface of proteins. Since the imine bonds are readily
24
25 hydrolysed, the artificial protease equipped with an active site containing the aldehyde group may
26
27 be able to form complexes with a variety of proteins reversibly.⁷⁷ More recently, a series of amyloid
28
29 peptide-binding groups were covalently attached to Co(III) or Cu(II) complexes to obtain catalytic
30
31 cleavage of the peptide responsible for AD (Figure 6c). In one case, apoptosis of INS-1 cell in the
32
33 presence of preformed polymeric aggregates of h-IAPP was even inhibited.^{72,78} Other strategies can
34
35 be mentioned here. In order to increase the intrinsic proteolytic activity of a metal centre it is also
36
37 possible to alter the polarity of its microenvironment.⁷⁹ For example, it was found that the
38
39 proteolytic activity of the Cu(II) complex of 1,4,7,10-tetraazacyclododecane (cyclen) is enhanced
40
41 significantly when attached to crosslinked polystyrene.⁷⁷ Another mechanism of action reported in
42
43 the literature consists in the abstraction of an α -carbon hydrogen atoms of the peptide backbone by
44
45 hydroxyl radicals and other ROS generated by a redox-active metal center. Subsequent degradation
46
47 of the NH-C α and C α -C(O) bonds yields multiple, fragmented peptide products.⁸⁰ However, as it
48
49 was mentioned above, in the latter case damage of the protein could be a problem. An interesting
50
51 but unique example also reports the attachment of an iron(III) peptide-cleaving complex to
52
53
54
55
56
57
58
59
60

1 phospholipids, which allowed for cleaving specifically integral membrane proteins (Figure 6d).⁸¹
2
3 Finally, sequence specific peptide hydrolysis can be achieved if the metal ion is capable of
4 coordinating to anchoring side chains in amino acid residues such as cysteine, aspartate, histidine,
5
6 or methionine. One for all, it is instructive to mention the results reported by A. Erxleben⁸² showing
7
8 the reaction of Cp_2MoCl_2 with cysteine-containing di- and tri-peptides, where the coordination of
9
10 the $\text{Cp}_2\text{Mo}^{2+}$ unit to the thiol group of X-Cys-Y peptides assists the release of the amino acid at the
11
12 carboxyl end of the cysteine residue.
13
14
15
16
17
18
19
20
21

22 4. Conclusions and perspective

23
24
25 In the current bioinorganic literature concerning CDs, most attention is given to the interaction
26
27 between metal complexes and the proteins whose misfolding or dyshomeostasis is considered as
28
29 responsible for the development of the disease. On the other hand, although it has been recognized
30
31 that MPs play an important role in the development of many CDs (see Table 1), the inhibiting
32
33 properties that MCs might have towards MPs and the therapeutic consequences of those inhibitions
34
35 properties, have not received, in our opinion, enough attention. Meanwhile, although MCs have
36
37 been considered for a long time as possible drugs for CD, their mechanism of action and the
38
39 question of whether they interact with MPs, have only been recognized at a second, much more
40
41 recent glance (see the case of clioquinol or cisplatin discussed above). The possibility to modulate
42
43 MPs activity by MCs opens fascinating therapeutic strategies, in particular because several MPs
44
45 play a role in different CDs. Thus, compounds that are active in one CD might be active as well in
46
47 other CDs. Of course, this interpenetration of MPs inhibition also represents a challenge for
48
49 medicinal chemistry, as ultimately only drugs that show a selective therapeutic action in a single
50
51 type of disease can be retained. The few published cases in which MCs have been shown to act
52
53 either as direct inhibitors of endogenous MPs, or as indirect activators of MPs, can be taken as a
54
55 proof-of-concept that inorganic complexes can be complementary to organic inhibitors in
56
57
58
59
60

1 modulating MPs activity. Meanwhile, the more numerous cases of artificial MPs that could
2 substitute malfunctioning MPs and lower the concentration of misfolded, aggregate-prone proteins
3 or peptides, have already opened new routes for bioinorganic chemistry. Overall, only a better
4 understanding of the action of MPs in CDs, and of course *in vivo* data on the selectivity of existing
5 and future metal-containing MP inhibitors, will allow for developing new and efficient drugs for so
6 far incurable diseases.
7
8
9
10
11
12
13
14
15
16
17
18

19 **6. Acknowledgements**

20 We thank FIRB 'RINAME' RBAP114AMK and PRIN 2010 2010M2JARJ_001 for partial
21 financial support.
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Tables

Conformational Disease	Protein	MPs involved	Ref.
Hypercholesterolaemia,	low-density lipoprotein	MMP-12	83
Atherosclerosis	receptor		
Cystic fibrosis	cystic fibrosis trans-membrane regulator	MMP-9, MMP-13	84-86
Phenylketonuria	phenylalanine hydroxylase	Not identified	87
Huntington's disease	huntingtin	MMP-9	88
Marfan syndrome	fibrillin	MMP-2, ADAMTSL6 β	89,90
Osteogenesis imperfecta	procollagen	BMP1	91
Sickle cell anaemia	hemoglobin	MMP-2, MMP-9	92
Scurvy	collagen	MMP-1	93
Alzheimer's disease	β -amyloid, presenilin	NEP, IDE, ECE-1, ECE-2, ACE, BACE2, BACE1 MMPs, PreP. plasmin, APH, MBP, CatB, CatD, the proteasome	94
Parkinson's disease	α -synuclein, neuromelanin, lactoferrin, ferritin, ceruloplasmin, bivalent cation transporters	MMPs, ADAMs,	94-96
Scrapie/Creutzfeldt- Jakob disease	prion protein, ferritin	ADAM 8, ADAM9, ADAM 10	94,97-99

Familial amyloidoses	transthyretin/lysozyme	MMP-9	100
Retinitis pigmentosa	rhodopsin	ADAM-9	101
Cataracts	Crystallins	MMPs, ADAM,	102,103
		ADAMTS	
Cancer	p53	MMPs, IDE, ADAMs	104,105
Friedreich's ataxia	Frataxin, aconitase	MPP	106
Progressive supranuclear palsy	α-Synuclein	MMP-1, MMP-9	107
Wilson's disease	Ceruloplasmin, Wilson's protein	MMP-2, MMP-12	108
Type II diabetes	Insulin, amylin	IDE, NEP	109,110
Carotid atherosclerosis	Proteins in vessel walls	MMP-9, MMP-12	111,112
Lewy-body dementia	α-Synuclein	MMPs	113
Familial amyotrophic lateral sclerosis	Superoxide dismutase 1	MMP-3, MMP-9	114,115

Table 1: aggregation prone proteins and MPs involved with various CDs

References

- 1 R. R. Crichton, *Biological Inorganic Chemistry An Introduction*, Elsevier, Amsterdam, The Netherlands, 2008.
- 2 K. M. Taylor and R. I. Nicholson, The LZT proteins; the LIV-1 subfamily of zinc transporters, *BBA-Biomembranes* 2003, **1611**, 16–30.
- 3 N. M. Hooper, *Zinc Metalloproteases in Health and Disease*, Taylor & Francis, London, UK, 1996.
- 4 B. L. Vallee and D.S. Auld, Active-site zinc ligands and activated H₂O of zinc enzymes, *Proc. Natl. Acad. Sci. U.S.A.* 1990, **87**, 220.
- 5 Y. Chen, Y. Huang, Y. Huang, X. Xia, J. Zhang, Y. Zhou, Y. Tan, S. He, F. Qiang, A. Li, O. D. Re, G. Li and J. Zhou, JWA suppresses tumor angiogenesis via Sp1-activated matrix metalloproteinase-2 and its prognostic significance in human gastric cancer, *Carcinogenesis* 2014, **35**, 442–451.
- 6 B. Detry, C. Erpicum, J. Paupert, S. Blacher, C. Maillard, F. Bruyere, H. Pendeville, T. Remacle, V. Lambert, C. Balsat, S. Ormenese, F. Lamaye, E. Janssens, L. Moons, D. Cataldo, F. Kridelka, P. Carmeliet, M. Thiry, J.-M. Foidart, I. Struman and A. Noel, Matrix metalloproteinase-2 governs lymphatic vessel formation as an interstitial collagenase, *Blood* 2012, **119**, 5048-5056.
- 7 P. K. Gajendrareddy, C. G. Engeland, R. Junges, M. P. Horan, I. G. Rojas and P. T. Marucha, MMP-8 overexpression and persistence of neutrophils relate to stress-impaired healing and poor collagen architecture in mice, *Brain Behav. Immun.* 2013, **28**, 44–48.
- 8 J. Liu, X Cheng, Z. Guo, Z. Wang, D. Li, F. Kang, H. Li, B. Li, Z. Cao, M. Nassal and D. Sun, Truncated active human matrix metalloproteinase-8 delivered by a chimeric adenovirus-hepatitis B virus vector ameliorates rat liver cirrhosis, *PLoS One* 2013, **8**, e53392.

- 1
2
3 9 W.-j. Peng, J.-w. Yan, Y.-n. Wan, B.-x. Wang, J.-h. Tao, G.-j. Yang, H.-f. Pan and J. Wang,
4
5 Matrix metalloproteinases: a review of their structure and role in systemic sclerosis, *J. Clin.*
6
7 *Immunol.* 2012, **32**, 1409-1414.
8
9
10 10 L. A. S. Moss, S. Jensen-Taubman and W. G. Stetler-Stevenson, Matrix metalloproteinases:
11
12 changing roles in tumor progression and metastasis, *Am. J. Pathol.* 2012, **181**, 1895–1899.
13
14 11 J. D. Raffetto and R. A. Khalil, Matrix metalloproteinases and their inhibitors in vascular
15
16 remodeling and vascular disease, *Biochem. Pharmacol.* 2008, **75**, 346–359.
17
18 12 A. Tiiman, P. Palumaa and V. Tougu, The missing link in the amyloid cascade of Alzheimer's
19
20 disease - Metal ions, *Neurochem. Int.* 2013, **62**, 367–378.
21
22
23 13 A. Loaiza, J. A. Ronau, A. Ribbe, L. Stanciu, J. W. 2nd Burgner, L. N. Pauland and M. M. Abu-
24
25 Omar, Folding dynamics of phenylalanine hydroxylase depends on the enzyme's metallation state:
26
27 the native metal, iron, protects against aggregate intermediates, *Eur. Biophys. J.* 2011, **40**, 959–968.
28
29 14 S. Waermlaender, A. Tiiman, A. Abelein, J. Luo, J. Jarvet, K. L. Soederberg, J. Danielsson and
30
31 A. Graeslund, Biophysical studies of the amyloid β -peptide: interactions with metal ions and small
32
33 molecules, *ChemBioChem* 2013, **14**, 1692–1704.
34
35
36 15 Y. Li, W. Xu, Y. Mu and J. Z. H. Zhang, Acidic pH retards the fibrillization of human islet
37
38 amyloid polypeptide due to electrostatic repulsion of histidines, *J. Chem. Phys.* 2013, **139**,
39
40 055102/1–055102/9.
41
42
43 16 V. Bellotti, M. Nuvolone, S. Giorgetti, L. Obici, G. Palladini, P. Russo, F. Lavatelli, V. Perfetti
44
45 and G. Merlini, The workings of the amyloid diseases, *Ann. Med.* 2007, **39**, 200–207.
46
47
48 17 G. Grasso and G. Spoto, Plasmonics for the study of metal ion–protein interactions, *Anal.*
49
50 *Bioanal. Chem.* 2013, **405**, 1833–1843.
51
52
53 18 C. M. Dobson, The structural basis of protein folding and its links with human disease. *Philos.*
54
55 *Trans. R. Soc. Lond. B* 2001, **356**, 133–145.
56
57
58 19 G. Grasso, E. Rizzarelli and G. Spoto, How the binding and degrading capabilities of insulin
59
60 degrading enzyme are affected by ubiquitin, *BBA-Proteins Proteom.* 2008, **1784**, 1122–1126.

1
2
3 20 G. Grasso, F. Salomone, G. R. Tundo, G. Pappalardo, C. Ciaccio, G. Spoto, A. Pietropaolo, M.
4 Coletta and E. Rizzarelli, Metal ions affect insulin-degrading enzyme activity, *J. Inorg. Biochem.*
5 2012, **117**, 351–358.

6
7
8
9 21 G. Grasso, M. L. Giuffrida and E. Rizzarelli, Metallostatics and amyloid β -degrading enzymes,
10 *Metalloomics* 2012, **4**, 937–949.

11
12
13 22 G. Grasso, A. Pietropaolo, G. Spoto, G. Pappalardo, G. R. Tundo, C. Ciaccio, M. Coletta and E.
14 Rizzarelli, Copper(I) and copper(II) inhibit A β peptides proteolysis by insulin-degrading enzyme
15 differently: implications for metallostatics alteration in Alzheimer's disease, *Chem.- Eur. J.* 2011,
16 **17**, 2752–2762.

17
18
19
20 23 G. Grasso, M. Fragai, E. Rizzarelli, G. Spoto and K. J. Yeo A new methodology for monitoring
21 the activity of cdMMP-12 anchored and freeze-dried on Au (111), *J. Am. Soc. Mass Spectrom.*
22 2007, **18**, 961–969.

23
24
25
26 24 G. Grasso, A. I. Bush, R. D'Agata, E. Rizzarelli and G. Spoto, Enzyme solid-state support
27 assays: a surface plasmon resonance and mass spectrometry coupled study of immobilized insulin
28 degrading enzyme, *Eur. Biophys. J.* 2009, **38**, 407–414.

29
30
31
32 25 G. Grasso, E. Rizzarelli and G. Spoto, The proteolytic activity of insulin-degrading enzyme: a
33 mass spectrometry study, *J. Mass Spectrom.* 2009, **44**, 735–741.

34
35
36
37 26 F. Bellia, A. Pietropaolo and G. Grasso, Formation of insulin fragments by insulin-degrading
38 enzyme: the role of zinc(II) and cystine bridges, *J. Mass Spectrom.* 2013, **48**, 135–140.

39
40
41
42 27 K. J. Kilpin and P. J. Dyson, Enzyme inhibition by metal complexes: concepts, strategies and
43 applications, *Chem. Sci.* 2013, **4**, 1410–1419.

44
45
46
47 28 N. P. E. Barry and P. J. Sadler, Challenges for metals in medicine: how nanotechnology may
48 help to shape the future, *ACS Nano* 2013, **7**, 5654–5659.

49
50
51
52 29 G. Lupidi, M. Angeletti, A. M. Eleuteri, E. Fioretti, S. Marini, M. Gioia, M. Coletta, Aluminum
53 modulation of proteolytic activities, *Coordin. Chem. Rev.* 2002, **228**, 263–269.

- 1
2
3 30 A. Yiotakis and V. Dive, Synthetic active site-directed inhibitors of metzincins: achievement and
4 perspectives, *Mol. Aspects Med.*, 2008, **29**, 329–338.
5
6
7 31 M. Flipo, J. Charton, A. Hocine, S. Dassonneville, B. Deprez and R. J. Deprez-Poulain,
8 Hydroxamates: relationships between structure and plasma stability, *J. Med. Chem.* 2009, **52**, 6790–
9 6802.
10
11
12
13 32 J. Jacobsen, J. L. Fullagar, M. T. Miller and S. M. Cohen, Identifying chelators for
14 metalloprotein inhibitors using a fragment-based approach, *J. Med. Chem.* 2011, **54**, 591–602.
15
16
17 33 A. Tanakit, M. Rouffet, D. P. Martin and S. M. Cohen, Investigating chelating sulfonamides and
18 their use in metalloproteinase inhibitors, *Dalton Trans.* 2012, **41**, 6507–6515.
19
20
21
22 34 M. A. Esteves, O. Ortet, A. Capelo, C. T. Supuran, S. M. Marques and M. A. Santos, New
23 hydroxypyrimidinone-containing sulfonamides as carbonic anhydrase inhibitors also acting as
24 MMP inhibitors, *Bioorg. Med. Chem. Lett.* 2010, **20**, 3623–3627.
25
26
27
28 35 S. M. Marques, T. Tuccinardi, A. Martinelli, S. Santamaria, E. Nuti, A. Rossello, V. André and
29 M. A. Santos, Novel 1-hydroxypiperazine-2,6-diones as new leads in the inhibition of
30 metalloproteinases, *J. Med. Chem.* 2011, **54**, 8289–8298.
31
32
33
34 36 L. J. McCawley and L. M. Matrisian, Matrix metalloproteinases: multifunctional contributors to
35 tumor progression, *Mol. Med. Today*, 2000, **6**, 149–156.
36
37
38
39 37 M. Pavlaki and S. Zucker, Matrix metalloproteinase inhibitors (MMPIs): the beginning of Phase
40 I or the termination of Phase III clinical trials, *Cancer Metastas. Rev.* 2003, **22**, 177–203.
41
42
43
44 38 M. Mori, A. Massaro, V. Calderone, M. Fragai, C. Luchinat and A. Mordini, Discovery of a new
45 class of potent MMP inhibitors by structure-based optimization of the arylsulfonamide scaffold,
46 *ACS Med. Chem. Lett.* 2013, **4**, 565–569.
47
48
49
50 39 B. G. Rao, Recent developments in the design of specific matrix metalloproteinase inhibitors
51 aided by structural and computational studies, *Curr. Pharm. Des.* 2005, **11**, 295–322.
52
53
54
55 40 J. Wang, C. Medina, M. W. Radomski and J. F. Gilmer, N-Substituted homopiperazine
56 barbiturates as gelatinase inhibitors, *Bioorg. Med. Chem.* 2011, **19**, 4985–4999.
57
58
59
60

- 1
2
3 41 A. Bergamo, A. Masi, A. F. A. Peacock, A. Habtemariam, P. J. Sadler, G. Sava, In vivo tumour
4 and metastasis reduction and in vitro effects on invasion assays of the ruthenium RM175 and
5 osmium AFAP51 organometallics in the mammary cancer model, *J. Inorg. Biochem.* 2010, **104**,
6 79–86.
7
8
9
10
11 42 R. Sasanelli, A. Boccarelli, D. Giordano, M. Laforgia, F. Arnesano, G. Natile, C. Cardellicchio,
12 M. A. M. Capozzi and M. Coluccia, Platinum complexes can inhibit matrix metalloproteinase
13 activity: platinum-diethyl[(methylsulfinyl)methyl]phosphonate complexes as inhibitors of matrix
14 metalloproteinases 2, 3, 9, and 12, *J. Med. Chem.* 2007, **50**, 3434–344.
15
16
17
18
19 43 B. D. Belviso, R. Caliandro, D. Siliqi, V. Calderone, F. Arnesano and G. Natile, Structure of
20 matrix metalloproteinase-3 with a platinum-based inhibitor, *Chem. Comm.* 2013, **49**, 5492–5494.
21
22
23
24 44 J. -Y. Lee, Cobalt (III) complexes as novel matrix metalloproteinase-9 inhibitors, *B. Kor. Chem.*
25 *Soc.* 2012, **33**, 2762–2764.
26
27
28
29 45 A. S. Harney, L. B. Sole and T. J. Meade, Kinetics and thermodynamics of irreversible
30 inhibition of matrix metalloproteinase 2 by a Co(III) Schiff base complex, *J. Biol. Inorg. Chem.*
31 2012, **17**, 853–860.
32
33
34
35 46 A. R. Farina and A. R. Mackay, Gelatinase B/MMP-9 in tumour pathogenesis and progression,
36 *Cancers* 2014, **6**, 240–296.
37
38
39
40 47 B. Fingleton, Matrix metalloproteinases as valid clinical targets, *Curr. Pharm. Design* 2007, **13**,
41 333–346
42
43
44 48 C. S. Anthony, G. Masuyer, E. D. Sturrock and K. R. Acharya, Structure based drug design of
45 angiotensin-I converting enzyme inhibitors, *Curr. Med. Chem.*, 2012, **19**, 845–855.
46
47
48
49 49 V. Dive, C.-F. Chang, A. Yiotakis and E. D. Sturrock, Inhibition of zinc metalloproteinases in
50 cardiovascular disease - from unity to trinity, or duality?, *Curr. Pharm. Design* 2009, **15**, 3606–
51 3621.
52
53
54
55 50 S. Laurent, M. Schlaich and M. Esler, New drugs, procedures, and devices for hypertension,
56 *Lancet* 2012, **380**, 591–600.
57
58
59
60

1
2
3 51 A. Agrawal, D. Romero-Perez, J. A. Jacobsen, F. J. Villarreal and S. M. Cohen, Zinc-binding
4 groups modulate selective inhibition of MMPs, *ChemMedChem* 2008, **3**, 812–820.

5
6
7 52 C. Gros, N. Noël, A. Souque, J. C. Schwartz, D. Danvy, J. C. Plaquevent, L. Duhamel, P.
8 Duhamel, J. M. Lecomte and J. Bralet, Mixed inhibitors of angiotensin-converting enzyme (EC
9 3.4.15.1) and enkephalinase (EC 3.4.24.11): rational design, properties, and potential cardiovascular
10 applications of glycopril and alatriopril. *Proc. Natl. Acad. Sci. U.S.A.* 1991, **10**, 4210–4214.

11
12
13 53 L.-B. Zou, A. Mouri, N. Iwata, T.C. Saido, D. Wang, M.-W. Wang, H. Mizoguchi, Y. Noda and
14 T. Nabeshima, Inhibition of neprilysin by infusion of thiorphan into the hippocampus causes an
15 accumulation of amyloid Beta and impairment of learning and memory, *J. Pharmacol. Exp. Ther.*
16 2006, **317**, 334–340.

17
18
19 54 C. Ciaccio, G. F. Tundo, G. Grasso, G. Spoto, D. Marasco, M. Ruvo, S. Marini, E. Rizzarelli and
20 Massimo Coletta, Somatostatin: a novel substrate and a modulator of insulin degrading enzyme
21 activity, *J. Mol. Biol.* 2009, **385**, 1556–1567.

22
23
24 55 K. Zou and M. Michikawa, Angiotensin-converting enzyme as a potential target for treatment of
25 Alzheimer's disease: inhibition or activation?, *Rev. Neurosci.* 2008, **19**, 203–212.

26
27
28 56 L. Lannfelt, K. Blennow, H. Zetterberg, S. Batsman, D. Ames, J. Harrison, C. L. Masters, S.
29 Targum, A. I. Bush, R. Murdoch, J. Wilson and C. W. Ritchie, Safety, efficacy, and biomarker
30 findings of PBT2 in targeting Abeta as a modifying therapy for Alzheimer's disease: a phase IIa,
31 double-blind, randomised, placebo-controlled trial, *Lancet Neurol.* 2008, **7**, 779–786.

32
33
34 57 N. G. Faux, C. W. Ritchie, A. Gunn, A. Rembach, A. Tsatsanis, J. Bedo, J. Harrison, L.
35 Lannfelt, K. Blennow, H. Zetterberg, M. Ingelsson, C. L. Masters, R. E. Tanzi, J. L. Cummings, C.
36 M. Herd and A. I. Bush, PBT2 rapidly improves cognition in Alzheimer's Disease: additional phase
37 II analyses, *J. Alzheimers Dis.* 2010, **20**, 509–516.

38
39
40 58 M. Kawahara, D. Mizuno, H. Koyama, K. Konoha, S. Ohkawara and Y. Sadakane, Disruption of
41 zinc homeostasis and the pathogenesis of senile dementia, *Metallomics* 2014, **6**, 209–219.

- 1
2
3 59 P. J. Crouch and K. J. Barnham, Therapeutic redistribution of metal ions to treat Alzheimer's
4 disease, *Acc. Chem. Res.* 2012, **45**, 1604–1611.
- 5
6
7 60 A. R. White, T. Du, K. M. Laughton, I. Volitakis, R. A. Sharples, M. E. Xilinas, D. E. Hoke, R.
8 M. D. Holsinger, G. Evin, R. A. Cherny, A. F. Hill, K. J. Barnham, Q.-X. Li, A. I. Bush and C. L.
9 Masters, Degradation of the Alzheimer disease amyloid beta-peptide by metal-dependent up-
10 regulation of metalloprotease activity, *J. Biol. Chem.* 2006, **281**, 17670–17680.
- 11
12
13 61 P. J. Crouch, M. S. Savva, L. W. Hung, P. S. Donnelly, A. I. Mot, S. J. Parker, M. A.
14 Greenough, I. Volitakis, P. A. Adlard, R. A. Cherny, C. L. Masters, A. I. Bush, K. J. Barnham and
15 A. R. White, The Alzheimer's therapeutic PBT2 promotes amyloid- β degradation and GSK3
16 phosphorylation via a metal chaperone activity, *J. Neurochem.* 2011, **119**, 220–230.
- 17
18
19 62 A. Caragounis, T. Du, G. Filiz, K. M. Laughton, I. Volitakis, R. A. Sharples, R. A. Cherny, C. L.
20 Masters, S. C. Drew, A. F. Hill, Q. X. Li, P. J. Crouch, K. J. Barnham and A. R. White, Differential
21 modulation of Alzheimer's disease amyloid beta-peptide accumulation by diverse classes of metal
22 ligands, *Biochem. J.* 2007, **407**, 435–450.
- 23
24
25 63 C. Duncan and A. R. White, Copper complexes as therapeutic agents, *Metalloomics* 2012, **4**, 127–
26 138.
- 27
28
29 64 K. D. Mjos and C. Orvig, Metallodrugs in medicinal inorganic chemistry, *Chem. Rev.* 2014,
30 **114**, 4540–4563.
- 31
32
33 65 A. Muscella, L. Urso, N. Calabriso, C. Vetrugno, F. P. Fanizzi, C. Storelli and S. Marsigliante,
34 Functions of epidermal growth factor receptor in cisplatin response of thyroid cells, *Biochem.*
35 *Pharmacol.* 2009, **77**, 979–992.
- 36
37
38 66 A. Siméon, F. Monier, H. Emonard, P. Gillery, P. Birembaut, W. Hornebeck, F. X. Maquart,
39 Expression and activation of matrix metalloproteinases in wounds: modulation by the tripeptide-
40 copper complex glycyl-L-histidyl-L-lysine-Cu²⁺, *J. Invest. Dermatol.* 1999, **112**, 957–964.
- 41
42
43 67 L. Pickart and M. M. Thaler, Tripeptide in human serum which prolongs survival of normal liver
44 cells and stimulates growth in neoplastic liver, *Nature New Biol.* 1973, **243**, 85–87.
- 45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 68 L. Pickart, J. M. Vasquez-Soltero, A. Margolina, The human tripeptide GHK-Cu in prevention
4 of oxidative stress and degenerative conditions of aging: implications for cognitive health, *Oxid.*
5
6
7 *Med. Cell. Longev.* 2012, **324832**, 8pp.

8
9
10 69 J. Hošek, R. Novotná, P. Babula, J. Vančo, Z. Trávníček, Zn(II)-Chlorido complexes of
11 phytohormone kinetin and its derivatives modulate expression of inflammatory mediators in THP-1
12 cells, *PLoS One* 2013, **8**, e65214.

13
14
15 70 J. Suh, W. S. Chei, Metal complexes as artificial proteases: toward catalytic drugs, *Curr. Opin.*
16
17
18 *Chem. Biol.* 2008, **12**, 207–213.

19
20
21 71 T. Y. Lee and J. Suh, Target-selective peptide-cleaving catalysts as a new paradigm in drug
22 design, *Chem. Soc. Rev.* 2009, **38**, 1949–1957.

23
24
25 72 J. Suh, Progress in designing artificial proteases: a new therapeutic option for amyloid diseases,
26
27
28 *Asian J. Org. Chem.* 2014, **3**, 18–32.

29
30
31 73 R. P. Bora, A. Barman, X. Zhu, M. Ozbil and R. Prabhakar, Which one among aspartyl protease,
32 metallopeptidase, and artificial metallopeptidase is the most efficient catalyst in peptide
33 hydrolysis?, *J. Phys. Chem. B* 2010, **114**, 10860–10875.

34
35
36 74 J. Suh, Synthetic artificial peptidases and nucleases using macromolecular catalytic systems,
37
38
39 *Acc. Chem. Res.* 2003, **36**, 562–570.

40
41
42 75 V. P. Petrović, D. Simijonović and Z. D. Petrović, Use of diethanolammonium–
43 tetrachloridopalladate(II) complex in bioorganic modelling as artificial metallopeptidase in the
44 reaction with N-acetylated L-methionylglycine dipeptide. NMR and DFT study of the hydrolytic
45 reaction, *J. Mol. Struct.* 2014, **1060**, 38–41.

46
47
48 76 J. C. Joyner and J. A. Cowan, Target-directed catalytic metallodrugs, *Braz. J. Med. Biol. Res.*
49
50
51 2013, **46**, 465–485.

52
53
54 77 S. H. Yoo, B. J. Lee, H. Kim and J. Suh, Artificial metalloprotease with active site comprising
55 aldehyde group and Cu(II)cyclen complex, *J. Am. Chem. Soc.* 2005, **127**, 9593–9602.

1
2
3 78 J. Suh, S. H. Yoo, M. G. Kim, K. Jeong, J. Y. Ahn, M.-S. Kim, P. S. Chae, T. Y. Lee, J. Lee, J.
4
5 Lee, Y. A. Jang and E. H. Ko, Cleavage agents for soluble oligomers of amyloid β peptides, *Angew.*
6
7 *Chem. Int. Ed.* 2007, **46**, 7064–7067.

8
9
10 79 M. G. Kim, S. H. Yoo, W. S. Chei, T. Y. Lee, H. M. Kim and J. Suh, Soluble artificial
11
12 metalloproteases with broad substrate selectivity, high reactivity, and high thermal and chemical
13
14 stabilities, *J. Biol. Inorg. Chem.* 2010, **15**, 1023–1031.

15
16 80 K. B. Grant and M. Kassai, Major advances in the hydrolysis of peptides and proteins by metal
17
18 ions and complexes, *Curr. Org. Chem.* 2006, **10**, 1035–1049.

19
20
21 81 T. Furuta, M. Sakai, H. Hayashi, T. Asakawa, F. Kataoka, S. Fujii, T. Suzuki, Y. Suzuki, K.
22
23 Tanaka, N. Fishkin, K. Nakanishi, Design and synthesis of artificial phospholipid for selective
24
25 cleavage of integral membrane protein, *Chem. Commun.* 2005, 4575–4577.

26
27 82 A. Erxleben, Interaction of molybdocene dichloride with cysteine-containing peptides:
28
29 coordination, regioselective hydrolysis, and intramolecular aminolysis, *Inorg. Chem.* 2005, **44**,
30
31 1082–1094.

32
33
34 83 V. Subramanian, H. A. Uchida, T. Ijaz, J. J. Moorleggen, D. A. Howatt and A. Balakrishnan,
35
36 Calpain inhibition attenuates angiotensin II-induced abdominal aortic aneurysms and
37
38 atherosclerosis in low-density lipoprotein receptor-deficient mice, *J. Cardiovasc. Pharm.* 2012, **59**,
39
40 66–76.

41
42
43 84 T. J. Bensman, A. N. Nguyen, A. P. Rao and M. P. Beringer, Doxycycline exhibits anti-
44
45 inflammatory activity in CF bronchial epithelial cells, *Pulm. Pharmacol. Ther.* 2012, **25**, 377–382.

46
47 85 B.-H. Qu, E. Strickland and P. J. Thomas, Localization and suppression of a kinetic defect in
48
49 cystic fibrosis transmembrane conductance regulator folding, *J. Biol. Chem.* 1997, **272**, 15739–
50
51 15744.

52
53
54 86 T. Nkyimbeng, C. Ruppert, T. Shiomi, B. Dahal, G. Lang, W. Seeger, Y. Okada, J. D'Armiento
55
56 and A. Guenther, Pivotal role of matrix metalloproteinase 13 in extracellular matrix turnover in
57
58 idiopathic pulmonary fibrosis, *PLoS One* 2013, **8**, e73279.

- 1
2
3 87 S. Koizumi, H. Iinuma, T. Takeuchi, H. Umezawa and T. Nagatsu, Activation of phenylalanine
4 hydroxylase by a protease from *Bacillus subtilis*, *Biogenic Amines* 1988, **5**, 495–503.
- 5
6
7 88 J. Duran-Vilaregut, J. del Valle, G. Manich, A. Camins, M. Pallas, J. Vilaplana and C. Pelegri,
8 Role of matrix metalloproteinase-9 (MMP-9) in striatal blood-brain barrier disruption in a 3-
9 nitropropionic acid model of Huntington' disease, *Neuropath. Appl. Neuro.* 2011, **37**, 525–537.
- 10
11
12
13 89 W. Xiong, T. Meisinger, R. Knispel, J. M. Worth and B. T. Baxter, MMP-2 regulates erk1/2
14 phosphorylation and aortic dilatation in Marfan syndrome, *Circ. Res.* 2012, **110**, e92–e101.
- 15
16
17
18 90 M. Saito, M. Kurokawa, M. Oda, M. Oshima, K. Tsutsui, K. Kosaka, K. Nakao, M. Ogawa, R.-i.
19 Manabe, N. Suda, G. Ganjargal, Y. Hada, T. Noguchi, T. Teranaka, K. Sekiguchi, T. Yoneda and
20 T. Tsuji, ADAMTSL6 β protein rescues fibrillin-1 microfibril disorder in a marfan syndrome mouse
21 model through the promotion of fibrillin-1 assembly, *J. Biol. Chem.* 2011, **286**, 38602–38613,
22 S38602/1–S38602/14.
- 23
24
25
26
27
28 91 V. Martínez-Glez, M. Valencia, J. A. Caparrós-Martín, M. Aglan, S. Temtamy, J. Tenorio, V.
29 Pulido, U. Lindert, M. Rohrbach, D. Eyre, C. Giunta, P. Lapunzina and V. L. Ruiz-Perez,
30 Identification of a mutation causing deficient BMP1/mTLD proteolytic activity in autosomal
31 recessive osteogenesis imperfecta, *Hum. Mutat.* 2012, **33**, 343–350.
- 32
33
34
35
36
37 92 S. P. Lee, K. I. Ataga, M. Zayed, J. M. Manganello, E. P. Orringer, D. R. Phillips and L. V.
38 Parise, Phase I study of eptifibatide in patients with sickle cell anaemia, *Brit. J. Haematol.* 2007,
39 **139**, 612–620.
- 40
41
42
43
44
45 93 B. V. Nusgens, P. Humbert, A. Rougier, A. C. Colige, M. Haftek, C. A. Lambert, A. Richard, P.
46 Creidi and C. M. Lapiere, Topically applied vitamin C enhances the mRNA level of collagens I and
47 III, their processing enzymes and tissue inhibitor of matrix metalloproteinase 1 in the human
48 dermis, *J. Invest. Dermatol.* 2001, **116**, 853–859.
- 49
50
51
52
53
54 94 G. Malgieri and G. Grasso, The clearance of misfolded proteins in neurodegenerative diseases
55 by zinc metalloproteases: an inorganic perspective, *Coordin. Chem. Rev.* 2014, **260**, 139–155.
- 56
57
58
59
60

- 1
2
3 95 G. A. Rosenberg, Matrix metalloproteinases and their multiple roles in neurodegenerative
4 diseases, *Lancet Neurol.* 2009, **8**, 205–216.
5
6
7 96 V. Gupta, M. K. Singh, R. K. Garg, K. K. Pant and S. Khattri, Evaluation of peripheral matrix
8 metalloproteinase-1 in Parkinson's disease: a case-control study, *Int. J. Neurosci.* 2014, **124**, 88–92.
9
10
11 97 H. C. Altmeyden, B. Puig, F. Dohler, D. K. Thurm, C. Falker, S. Krasemann and M. Glatzel,
12 Proteolytic processing of the prion protein in health and disease, *Am. J. Neurodegener. Dis.* 2012, **1**,
13 15–31.
14
15
16 98 D. R. Taylor, E. T. Parkin, S. L. Cocklin, J. R. Ault, A. E. Ashcroft, A. J. Turner and N. M.
17 Hooper, Role of ADAMs in the ectodomain shedding and conformational conversion of the prion
18 protein, *J. Biol. Chem.* 2009, **284**, 22590–22600.
19
20
21 99 J. Liang, W. Wang, D. Sorensen, S. Medina, S. Ilchenko, J. Kiselar, W. K. Surewicz, S. A.
22 Booth and Q. Kong, Cellular prion protein regulates its own α -cleavage through ADAM8 in
23 skeletal muscle, *J. Biol. Chem.* 2012, **287**, 16510–16520.
24
25
26 100 M. R. Almeida and M. J. Saraiva, Clearance of extracellular misfolded proteins in systemic
27 amyloidosis: Experience with transthyretin, *FEBS Lett.* 2012, **586**, 2891–2896.
28
29
30 101 D. A. Parry, C. Toomes, L. Bida, M. Danciger, K. V. Towns, M. McKibbin, S. G. Jacobson, C.
31 V. Logan, M. Ali, J. Bond, R. Chance, S. Swendeman, L. L. Daniele, K. Springell, M. Adams, C.
32 A. Johnson, A. P. Booth, H. Jafri, Y. Rashid, E. Banin, T. M. Strom, D. B. Farber, D. Sharon, C. P.
33 Blobel, E. N. Jr Pugh, E. A. Pierce and C. F. Inglehearn, Loss of the metalloprotease ADAM9 leads
34 to cone-rod dystrophy in humans and retinal degeneration in mice, *Am. J. Hum. Genet.* 2009, **84**,
35 683–691.
36
37
38 102 J. V. Robertson, A. Siwakoti and J. A. West-Mays, Altered expression of transforming growth
39 factor beta 1 and matrix metalloproteinase-9 results in elevated intraocular pressure in mice, *Mol.*
40 *Vis.* 2013, **19**, 684–695.
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 103 L. M. Hodgkinson, L. Wang, G. Duncan, D. R. Edwards and I. M. Wormstone, ADAM and
4
5 ADAMTS gene expression in native and wound healing human lens epithelial cells, *Mol. Vis.* 2010,
6
7 **16**, 2765–2776.
8
9
10 104 G. R. Tundo, D. Sbardella, C. Ciaccio, A. Bianculli, A. Orlandi, M. G. Desimio, G. Arcuri, M.
11
12 Coletta and S. Marini, Insulin-degrading enzyme (IDE), *J. Biol. Chem.* 2013, **288**, 2281–2289.
13
14 105 N. Moro, C. Mauch and P. Zigrino, Metalloproteinases in melanoma, *Eur. J. Cell Biol.* 2014,
15
16 **93**, 23–29.
17
18 106 D. M. Gordon, Q. Shi, A. Dancis and D. Pain, Maturation of frataxin within mammalian and
19
20 yeast mitochondria: one-step processing by matrix processing peptidase, *Hum. Mol. Genet.* 1999, **8**,
21
22 2255–2262.
23
24 107 S. Lorenzl, D. S. Albers, J. W. Chirichigno, S. J. Augood and M. F. Beal, Elevated levels of
25
26 matrix metalloproteinases-9 and -1 and of tissue inhibitors of MMPs, TIMP-1 and TIMP-2 in
27
28 postmortem brain tissue of progressive supranuclear palsy, *J. Neurol. Sci.* 2004, **218**, 39–45.
29
30 108 A. V. Sokolov, M. O. Pulina, K. V. Ageeva, O. S. Tcherkalina, E. T. Zakharova and V. B.
31
32 Vasilyev, Identification of complexes formed by ceruloplasmin with matrix metalloproteinases 2
33
34 and 12, *Biochemistry* 2009, **74**, 1388–1392.
35
36 109 F. Bellia and G. Grasso, The role of copper(II) and zinc(II) in the degradation of human and
37
38 murine IAPP by insulin-degrading enzyme, *J. Mass Spectrom.* 2014, **49**, 274–279.
39
40 110 C. Oefner, S. Pierau, H. Schulz and G.E. Dale, Structural studies of a bifunctional inhibitor of
41
42 neprilysin and DPP-IV, *Acta Crystallogr. D* 2007, **63**, 975–981.
43
44 111 D. Silvello, L. B. Narvaes, L. C. Albuquerque, L. F. Forgiarini, L. Meurer, N. C. Martinelli, M.
45
46 E. Andrades, N. Clausell, K. Goncalves dos Santos and L. E. Rohde, Serum levels and
47
48 polymorphisms of matrix metalloproteinases (MMPs) in carotid artery atherosclerosis: higher
49
50 MMP-9 levels are associated with plaque vulnerability, *Biomarkers* 2014, **19**, 49–55.
51
52 112 V. P. W. Scholtes, J. L. Johnson, N. Jenkins, G. B. Sala-Newby, J.-P. P. M. de Vries, G. J. de
53
54 Borst, D. P. V. de Kleijn, F. L. Moll, G. Pasterkamp and A. C. Newby, Carotid atherosclerotic
55
56
57
58
59
60

1
2
3 plaque matrix metalloproteinase-12-positive macrophage subpopulation predicts adverse outcome
4
5 after endarterectomy, *J. Am. Heart Assoc.* 2012, **1**, 001040/1–001040/13.

6
7 113 S. Lorenzl, K. Buerger, H. Hampel and M. F. Beal, Profiles of matrix metalloproteinases and
8
9 their inhibitors in plasma of patients with dementia, *Int. Psychogeriatr. / IPA* 2008, **20**, 67–76.

10
11 114 M. Dewil, C. Schurmans, S. Starckx, G. Opdenakker, L. Van Den Bosch and W. Robberecht,
12
13 Role of matrix metalloproteinase-9 in a mouse model for amyotrophic lateral sclerosis,
14
15 *NeuroReport* 2005, **16**, 321–324.

16
17 115 J. K. Lee, J. H. Shin, J.H. Suh, I. S. Choi, K. S. Ryu and B. J. Gwag, Tissue inhibitor of
18
19 metalloproteinases-3 (TIMP-3) expression is increased during serum deprivation-induced neuronal
20
21 apoptosis in vitro and in the G93A mouse model of amyotrophic lateral sclerosis and tential
22
23 modulator of Fas-mediated apoptosis, *Neurobiol. Dis.* 2008, **30**, 174–185.
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Figures and Tables Legends

Table 1: Representative conformational diseases. The aggregation-prone protein and the MPs capable of degrading the latter are also reported. Readapted from ref. 17.

Figure 1: The ligand smp (bottom) and the structures of catalytic domains of MMPs and model of their interaction with $[\text{PtCl}_2(\text{smp})]$. Ribbon representation of the structures of catalytic domains of MMP-2 (PDB ID 1HOV), MMP-9 (1GKC), MMP-3 (1SLN), and MMP-12 (theoretical model from reference 42 with $[\text{PtCl}_2(\text{smp})]$). Helices and strands are coloured in grey. The zinc atoms are shown as cyan spheres and the calcium atoms as magenta spheres. The catalytic and structural zinc sites as well as the loops S and S1' are indicated on the structure of MMP-2. Fully conserved His and Met residues are shown as yellow sticks, while non-conserved His and Met residues are shown as blue and green sticks, respectively. The smp ligand of $[\text{PtCl}_2(\text{smp})]$ bound to MMP-12 is shown in red and the platinum atom as an orange sphere.

Figure 2: MMP inhibitors based on a) cobalt(III) and b) platinum(II) MCs. $[\text{Co}(\text{acacen})(\text{NH}_3)_2]\text{Cl}$: $\text{R}=\text{CH}_3$, $\text{Y}=\text{H}$, $\text{X}=\text{NH}_3$.

Figure 3: Metal-based MMP inhibitors: reversible inhibition with an organic, zinc-binding moiety can occur first (a), followed by irreversible coordination of an MMP aminoacid residue to the inhibitor metal center (b). In the final, irreversible complex additional secondary interactions such as H-bonding, π - π interaction, etc. can occur between the secondary ligands coordinated to the inhibitor metal ion, and either nearby residue of the MMP, or the catalytic zinc ion itself.

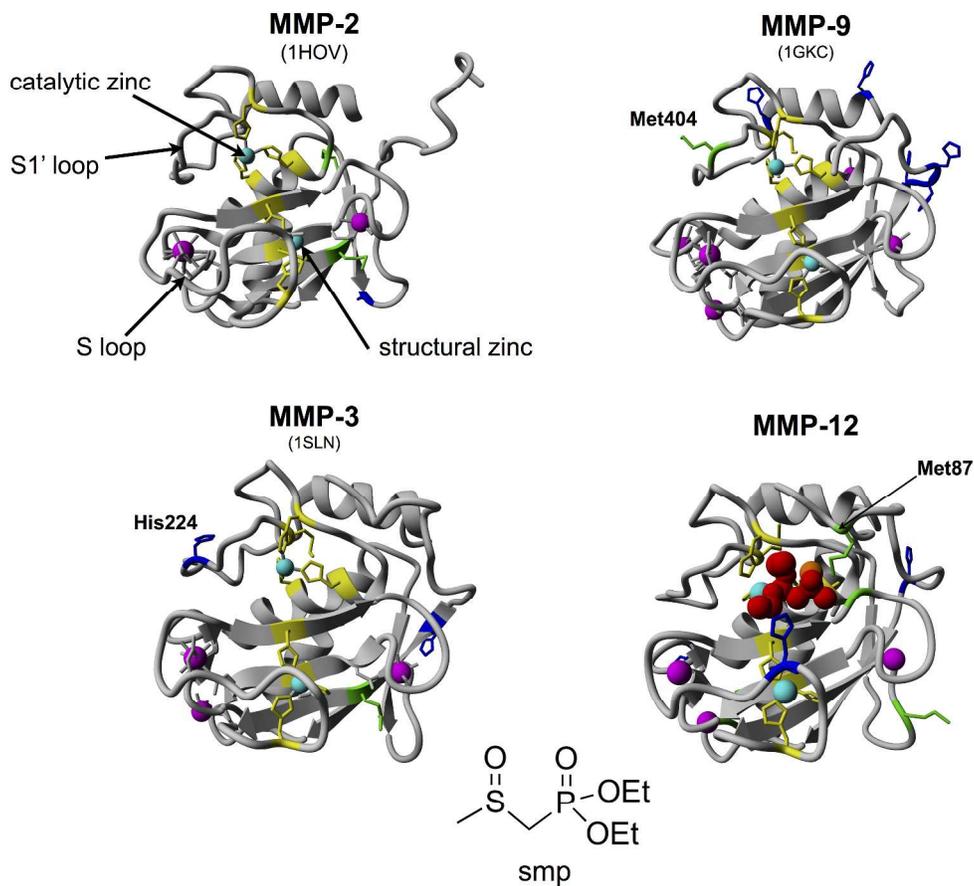
Figure 4: Zoom on the crystal structure of phosphoramidon-NEP (PDB code 1DMT). The inhibitor phosphoramidon in the centre of the molecule is in stick-and-ball form (color code: orange=C,

1
2 magenta=N, red=O, and yellow=P) and the zinc ion is represented by a cyan sphere. The
3
4 coordinated His and Glu residues are colored in blue and green, respectively.
5
6
7

8 **Figure 5:** Two metal chelators in clinical trials for the treatment of neurodegenerative diseases.
9

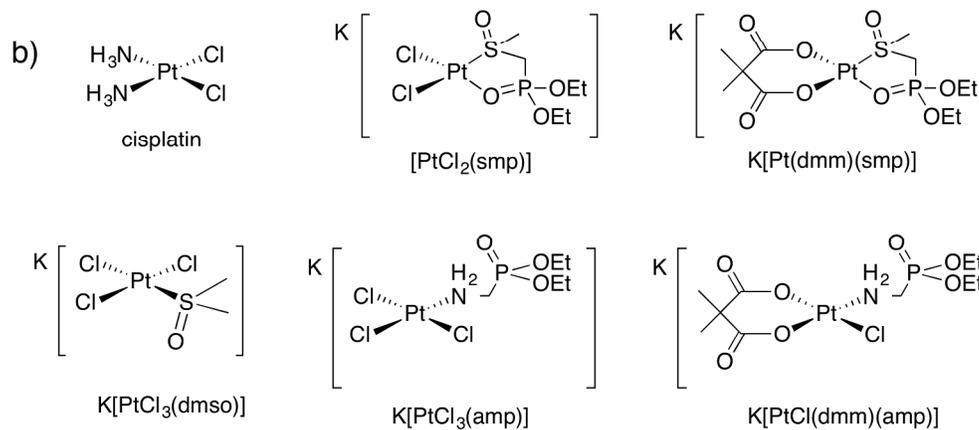
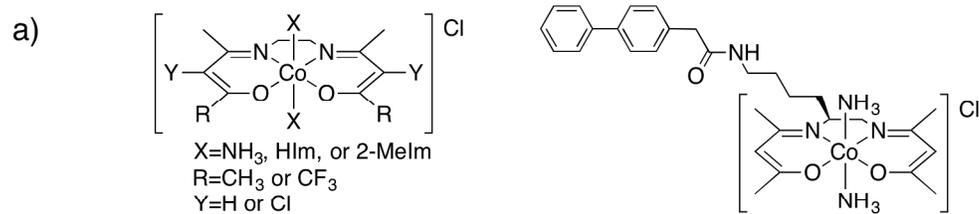
10 Reprinted with permission from Ref. 64.
11
12
13
14

15 **Figure 6:** Artificial metalloproteases. Two mechanism for peptide cleavage: a) chelation of peptides
16 followed by nucleophilic attack of OH⁻; or b) coordination of peptides followed by nucleophilic
17 attack of a metal-bound OH⁻ ligand. c) Amyloid beta peptide-targeted artificial metalloprotease
18 (readapted from Ref. 78, peptide taken from PDB 2LFM). d) Membrane-bound artificial
19 metalloprotease for cleavage of integral membrane proteins.
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

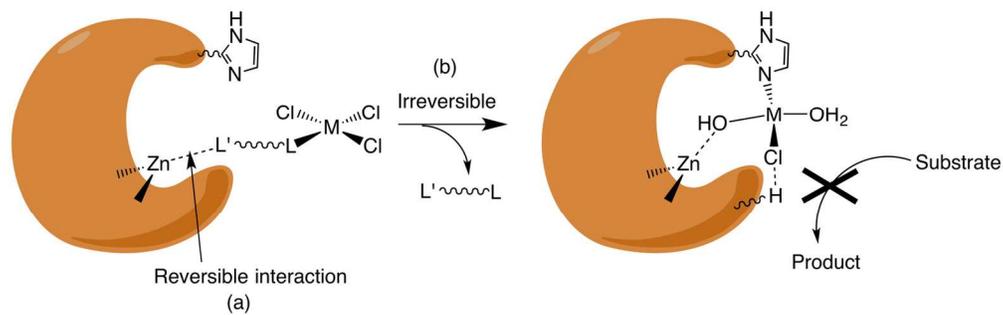


1550x1435mm (72 x 72 DPI)

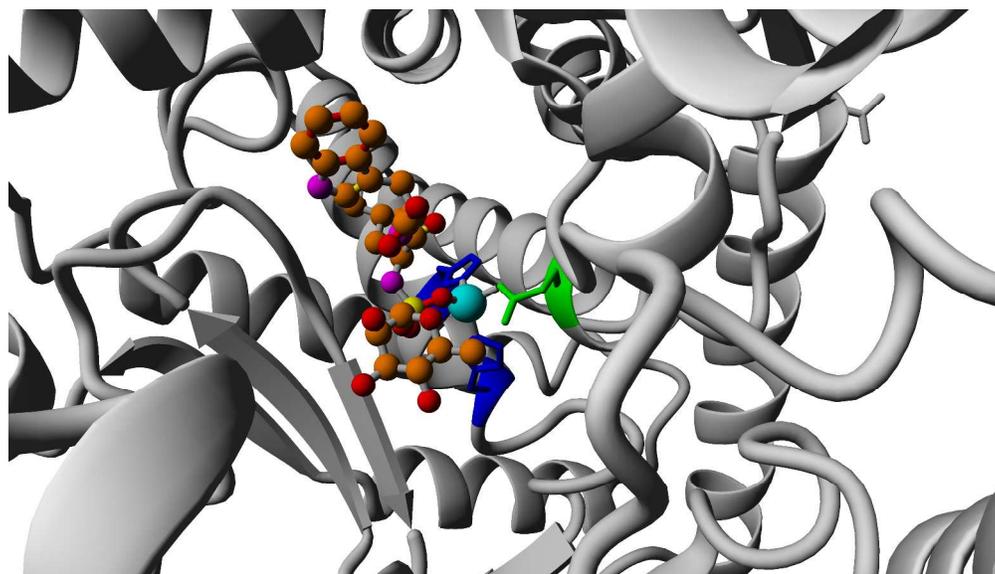
1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



242x167mm (300 x 300 DPI)

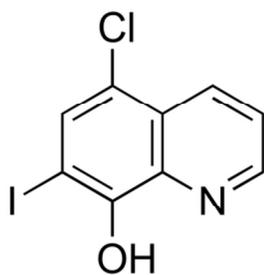
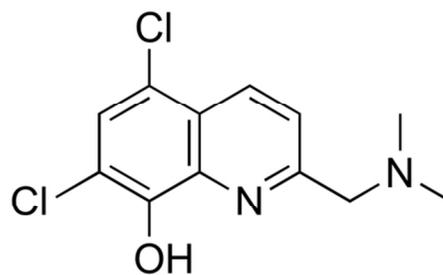


76x23mm (600 x 600 DPI)



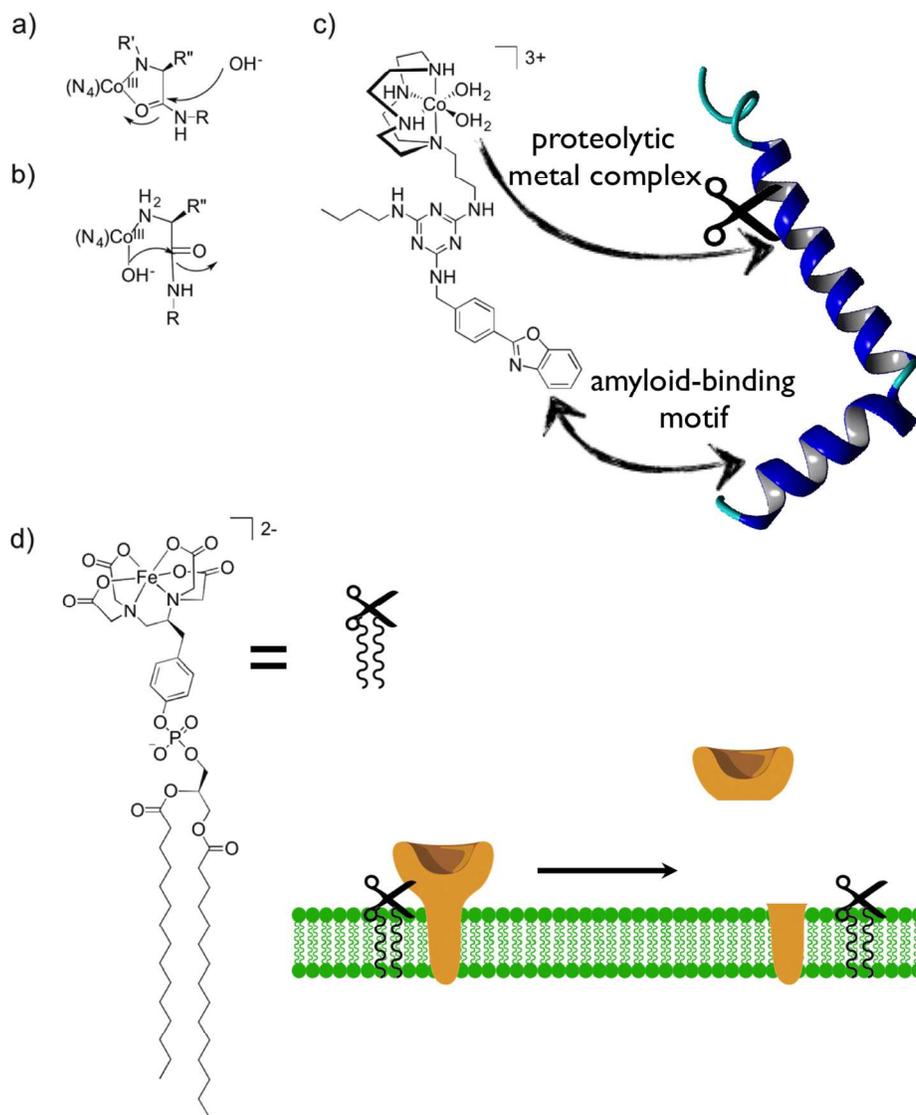
1411x805mm (72 x 72 DPI)

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

**clioquinol****PBT2**

42x17mm (600 x 600 DPI)

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



570x673mm (72 x 72 DPI)