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3 **1 Magnesium nitrate as a chemical modifier to improve**
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6 **2 sensitivity in manganese determination in plant material by**
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9 **3 tungsten coil atomic emission spectrometry**
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13 Sidnei G. Silva^{a*}, Joaquim A. Nóbrega^a, Bradley T. Jones^b and George L. Donati^b
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21 ^aGroup of Applied Instrumental Analysis, Department of Chemistry, Federal University
22
23 of São Carlos, P.O. Box 676, São Carlos SP13560-970, Brazil.
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27 ^bDepartment of Chemistry, Wake Forest University, Winston-Salem NC 27109, USA.
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* Corresponding author: sidgons@gmail.com
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56 FAX: +55-16-33518350, Phone: +55-16-3351-8058
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3 **Abstract**
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6 In this work, magnesium nitrate is used for the first time as a chemical modifier
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8 in atomic emission spectrometry to improve sensitivity and minimize matrix effects in
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10 Mn determinations. Magnesium/Mn interactions in the condensed and the gas phases
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12 contribute to increasing the population of excited-state Mn atoms, which result in
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14 improved sensitivity, precision and accuracy in plant analysis by tungsten coil atomic
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16 emission spectrometry (WCAES). The limit of detection calculated for Mn
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18 determinations in the presence of 750 mg L⁻¹ Mg was 0.05 mg L⁻¹, which is a 17-fold
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20 improvement when compared to determinations without using the chemical modifier.
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22 Repeatabilities also improved from 6.4 to 3.6 % (RSD, n = 10, Mn 2.5 mg L⁻¹) when
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24 Mg was used, and linear responses were observed in the 0.1–7.5 mg L⁻¹ Mn range.
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26 Manganese concentrations in standard reference materials of plant were 2-fold higher
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28 than the certified values for determinations without the modifier. For determinations
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30 using Mg(NO₃)₂, no statistically significant difference was observed between
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32 determined and certified Mn values at the 95% confidence level.
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55 **Keywords:** Charge-transfer reactions; Chemical modifier; Manganese; Tungsten coil
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57 atomizer; Atomic emission spectrometry; Magnesium.
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43 Introduction

44 Devices that use tungsten coils as atomizers are characterized as open systems
45 with pronounced temperature gradients between the coil surface and the gas phase.^{1,2}
46 Due to this inherent characteristic of open systems, some interference process and
47 analyte atomization mechanisms may be affected during instrumental measurement.³
48 One of the reasons for such interfering processes is associated to the formation of stable
49 compounds between analyte and concomitant elements, such as mixed oxides, in the
50 condensed phase.⁴

51 An alternative to minimize matrix effects in electrothermal atomization-based
52 methods is the application of chemical modifiers.^{2,5} Commonly used in high
53 concentrations, chemical modifiers provide thermal and temporal separation between
54 analytes and concomitants during the pyrolysis step.⁵⁻¹³ Often, chemical modifiers
55 thermally stabilize the analyte and allow the removal of matrix components by
56 volatilization, which contributes to minimizing analyte-concomitant interactions and
57 their consequent interfering effects.¹⁰ Different chemical modifiers have been employed
58 in GFAAS measurements, however the mixture Pd + Mg, also known as universal
59 modifier, is used for most analytes.¹⁴ In this case, the effect of Mg is related to thermal
60 stabilization by imbedding analyte atoms in a matrix of MgO, which delays analyte
61 vaporization.¹⁵⁻¹⁷ There still are some controversy regarding the mechanisms involved
62 in chemical modification with palladium. One hypothesis is that intermetallic
63 compounds and stable phases are formed from the interaction between Pd and the
64 analyte.⁷ Some authors attribute the analyte thermal stability in presence of Pd to the
65 formation of intercalation compounds, followed by their activation. Finally, the active
66 metal forms strong covalent bonds with the volatile analyte.¹⁸

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3 67 The first work employing a chemical modifier in a tungsten coil-based procedure
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5 68 was reported in 1994, for the determination of Ba in water samples.¹⁹ In this work, the
6
7 69 authors used EDTA as chemical modifier to eliminate interferences caused by Ca. Due
8
9 70 to the high concentrations of EDTA used, the atomizer lifetime was significantly
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11 71 reduced, probably because of the formation of tungsten carbides. Other chemical
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13 72 modifiers were also evaluated for W-coil atomic absorption spectrometry (WCAAS),
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15 73 such as $\text{NH}_4\text{H}_2\text{PO}_4$,^{20,21} ascorbic acid,²² and even the mixture of Pd and Mg.²³ However,
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17 74 the use of Pd as well as the mixture Pd + Mg are usually avoided in WCAAS due to
18
19 75 pitting effects on the coil surface observed after only a few runs.²³
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23 76 The use of chemical modifiers in tungsten coil-based devices has been mostly
24
25 77 restricted to WCAAS. On the other hand, this strategy remains mainly unexplored for
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27 78 tungsten coil atomic emission spectrometry (WCAES), with only two studies published
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29 79 to date. Salido and Jones proposed a procedure using Al or K as chemical modifiers to
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31 80 improve sensitivity in Sr determinations.²⁴ In a recent work, Silva *et al.* described the
32
33 81 use of Co as a chemical modifier to improve sensitivity and minimize matrix effects in
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35 82 Cr determinations by WCAES.²⁵ In this work, the presence of Co contributes to
36
37 83 increasing the population of Cr^+ by charge-transfer reactions in the gas phase. In a
38
39 84 second step, Cr^+/e^- recombination takes place, which results in a larger population of
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41 85 excited-state Cr atoms. The charge-transfer reactions are more effective for elements
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43 86 with ionization potential differences (ΔIE) of 2.0 eV or lower.^{26–28} This same
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45 87 mechanism could be used to explain the emission signal enhancements observed by
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47 88 Salido and Jones, and Silva *et al.*^{24,25} In addition to this two-step mechanism, the
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49 89 analytical signal enhancements observed in WCAES are also depend on similar boiling
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51 90 points (BP) for analyte and chemical modifier. Elements with large BP differences
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53 91 ($\Delta\text{BP} \geq \pm 600$ K) from the analyte cause no significant improvement in the analytical
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3 92 signal, probably because they have little interaction in the gas phase due to temporal
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5 93 separation of atomization processes.²⁵
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8 94 In the present work, magnesium nitrate is evaluated to improve sensitivity and
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10 95 minimize matrix effects in Mn determinations by WCAES. Considering Mg effects as
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12 96 chemical modifier,¹⁵ and the gas-phase mechanisms proposed in a previously published
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14 97 work,²³ this strategy may significantly improve limits of detection and accuracy in
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16 98 WCAES determinations.
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22 100 **Experimental**

23 24 25 101 **Apparatus**

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28 102 The WCAES device employed in this work has been previously described in the
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30 103 literature.²⁹ The atomizer consists of a tungsten coil 15 V, 150 W (Osram, Pullach,
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32 104 Germany) housed in a customized glass cell equipped with 2 quartz windows. To
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34 105 control the output current in the filament, a solid state power supply (BatMod, Part
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36 106 Number VI-LU1-EU-BM, Vicor, Andover, MA, USA) was used. A digital-to-analog
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38 107 converter (Measurement Advantage MiniLab 1008, Measurement Computing, Norton,
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40 108 MA, USA), connected to a USB 2.0 port of a Pentium 4 microcomputer (Intel[®]), in
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42 109 combination with a lab-written Visual Basic 6.0 (Microsoft, Seattle, WA, USA)
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44 110 program, were used to control current and time. Atomic emission signals were collected
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46 111 by a fused silica lens and focused as a 1:1 image onto the entrance slit of a CCD-based
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48 112 spectrometer (Ocean Optics, USB 4000, Dunedin, FL, USA), powered by the same
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50 113 computer used to control the filament power supply.
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3 114 For Mn ($\lambda = 403.1$ nm) determination, 25 μL of sample or reference solutions
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5 115 were directly deposited on the tungsten coil atomizer using an automatic micropipette
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7 116 (Eppendorf 10-100 μl , Brinkman, Westbury, NY, USA). The heating program presented
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9 117 in Table 1 was used in all experiments. The inverted ramp drying cycle was adopted to
10
11 118 keep a constant temperature during and right after the complete vaporization of the
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13 119 solvent.^{30,31} Vaporization takes place during steps 1-3. By the end of step 3, all solvent
14
15 120 has been vaporized and the applied current is reduced further to prevent the atomizer
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17 121 from overheating, which would result in analyte losses. Sample pyrolysis takes place
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19 122 during steps 4 and 5, and a short step with no applied current allows the sample to cool
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21 123 down near room temperature, which can minimize atomization differences due to
22
23 124 physical properties of the analytes. Finally, a high current, high temperature atomization
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25 125 step (step 7) is employed to atomize and excite the analytes. Ten consecutive spectra,
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27 126 each one with a 0.4 s integration time, were collected for each measurement during the
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29 127 atomization step. For background correction, the average between background signals
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31 128 recorded on each side of the Mn analytical peak was used.
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37 **Reagents and solutions**

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39 130 All solutions were prepared with analytical grade chemicals and distilled-
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41 131 deionized water (18.2 $\text{M}\Omega$ cm, Milli-Q[®], Millipore, Bedford, MA, USA). Manganese
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43 132 reference solutions were prepared by diluting a single-element 1000 mg L^{-1} stock
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45 133 solution (SPEX CerPrep, Metuchen, NJ, USA). A 10 g L^{-1} Mg stock solution (SPEX
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47 134 CerPrep) was used as chemical modifier. For measurements employing chemical
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49 135 modification, the analytical blank consisted of a 750 mg L^{-1} Mg solution.
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54 136 Trace metal grade concentrated nitric acid (Fischer, Pittsburgh, PA, USA) and
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56 137 hydrogen peroxide 30 % m m^{-1} (Acros, Morris Plains, NJ, USA) were employed for
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58 138 sample digestion. Peach Leaves (SRM 1547, National Institute of Standards and
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3 139 Technology, NIST, Gaithersburg, MD, USA) and Tomato Leaves (SRM 1573, NIST)
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5 140 were used to check the procedure's accuracy.
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8 141 **Sample preparation**

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10 142 Aliquots of approximately 250 mg of reference plant material were accurately
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12 143 weighted directly in plastic extraction containers. After this, a volume of 2 mL of
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14 144 concentrated HNO₃ was added to the sample and the mixture was left to react for 1 h.
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16 145 Then, 3 mL of H₂O₂ plus 2 mL of deionized water were added, and the mixtures were
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18 146 heated at 100 °C in an aluminum block for 2 h. The flasks were then allowed to cool
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20 147 down and reach room temperature before transferring blanks and sample digests to 15
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22 148 mL polypropylene tubes. The volumes were then made up to 10 mL with distilled-
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24 149 deionized water. Finally, aliquots of chemical modifier were added to each solution to a
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26 150 final concentration of 750 mg L⁻¹ Mg, and the volumes were made up to 15 mL with
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28 151 distilled-deionized water.
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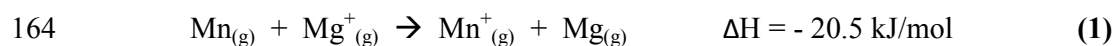
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34 153 **Results and Discussion**

35 154 **General Aspects**

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38 155 In preliminary studies, it was observed that the presence of Mg at high
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40 156 concentrations cause significant increase in Mn emission signals ($\lambda = 403.1$ nm). This
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42 157 effect may be related to charge-transfer and cation/electron recombination reactions in
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44 158 the gas-phase, which increase the population of Mn excited atoms, ultimately increasing
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46 159 atomic emission intensity.²⁸ A similar effect has been previously observed in both
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48 160 inductively coupled plasma mass and optical emission spectrometries (ICP-MS and ICP
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50 161 OES).^{27,28;32-35} In this case, Mg has an ionization potential only 0.21 eV higher than Mn.
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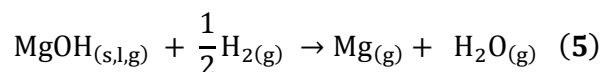
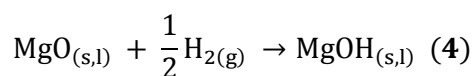
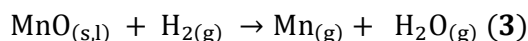
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3 162 Such small difference may facilitate charge-transfer reactions involved in a two-step
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5 163 excitation mechanism (Eqns. 1-2),³⁶ as described previously for Cr.²⁵
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14 166 Magnesium nitrate has been widely used as chemical modifier to increase
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16 167 analytical signals in GF AAS.^{5,37} Some authors attribute the thermal stabilization of
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18 168 analytes to their physical occlusion in the presence of high concentrations of Mg.^{11,15}
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20 169 Slavin *et al.* evaluated the presence of Mg as a chemical modifier for the determination
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22 170 of Mn.¹⁵ It was observed that Mn atomization is delayed to a higher temperature in the
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24 171 presence of Mg, which, in combination to the use of a L'vov platform, allows
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26 172 atomization to take place in a more thermally homogeneous environment, minimizing
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28 173 matrix effects.¹⁵ In this case, Mg is present as an oxide even in the solid phase.
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30 174 Manganese atoms are then trapped in a matrix of MgO, which delays their atomization
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32 175 until the MgO matrix is vaporized.¹⁵ This process, combined to the two-step mechanism
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34 176 described previously may explain the significant signal improvements observed for Mn
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36 177 when using Mg(NO₃)₂ as chemical modifier in WCAES determinations (Fig. 1). For the
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38 178 two-step mechanism to be effective, both analyte and chemical modifier atoms must be
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40 179 in contact in the gas-phase during the atomization step, which requires similar boiling
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42 180 points.²⁵ Although the BP difference between Mn and Mg is relatively large ($\Delta\text{BP} = 971$
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44 181 K),³⁶ the fact that Mn atoms are trapped in a MgO matrix, and that they most likely
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46 182 atomize at the same time enables a better interaction between gas-phase species
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48 183 probably formed after the MgO vaporization.¹⁵
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54 184 Muzgin *et al.*³⁸ evaluated some atomization mechanisms taking place on
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56 185 tungsten coil atomizers under an atmosphere of Ar/H₂ (such as the one used in the
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3 186 present work). According to the authors, free Mn atoms are mainly generated by
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5 187 thermal dissociation of MnO (Eqn. 3). On the other hand, Queiroz *et al.*³⁹ proposed a
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7 188 mechanism for Mg atomization in which mono-magnesium hydroxide is generated in
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9 189 the condensed phase, followed by reduction by hydrogen and generation of free Mg
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11 190 atoms (Eqns. 4-5). Considering these mechanisms, it is reasonable to assume that MnO
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13 191 and MgO/MgOH atomizing at the same time would both be reduced by H₂, and the
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15 192 resulting gas-phase species would participate in the two-step excitation mechanism
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17 193 represented in Eqns. 1-2.



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32 194 Considering the signal improvement provided by the use of magnesium nitrate
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34 195 (Fig. 1), a procedure employing this chemical modifier to increase sensitivity and
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36 196 minimize matrix effects was developed and applied to Mn determination by WCAES in
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38 197 certified reference samples of plant materials.

39 40 41 42 198 **Effect of chemical modifier concentration**

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45 199 The effect of modifier concentration on Mn emission signal was evaluated
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47 200 aiming to establish the minimum reagent concentration to yield maximum sensitivity.
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49 201 According to the results presented in Figure 2, the highest emission signals at 403.1 nm
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51 202 were obtained for Mg concentrations above 750 mg L⁻¹, which resulted in analytical
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53 203 signals 3.7-fold higher than the ones obtained without chemical modification. In these
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55 204 conditions, 19 µg of Mg were consumed per determination, which represents a minimal
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3 205 Mg/Mn ratio of 170. Therefore, a 750 mg L⁻¹ Mg solution was used as chemical
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5 206 modifier in all following studies.
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8 207 **Analytical figures of merit and application**

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10 208 The analytical features for Mn solutions containing 750 mg L⁻¹ of Mg were
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12 209 estimated and compared with those without the chemical modifier (Table 2). A linear
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14 210 relationship between emission intensity (*I*) and Mn concentration (*C*, mg L⁻¹) was
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16 211 observed in the range 0.1–7.5 mg L⁻¹ ($I = 2262C + 399$) for solutions containing the
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18 212 chemical modifier. The limit of detection was estimated as 0.05 mg L⁻¹, which is 17-
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20 213 fold lower than the one achieved in the absence of Mg. Comparing the calibration curve
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22 214 slopes for solutions with and without Mg, a 15-fold sensitivity improvement was
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24 215 observed when using the modifier. The precisions calculated as % RSD for a solution
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26 216 containing 2.5 mg L⁻¹ of Mn (*n* = 10) with or without the chemical modifier were 3.6
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28 217 and 6.4%, respectively. As it can be seen, in addition to increasing sensitivity, the
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30 218 application of Mg(NO₃)₂ can also improve repeatability in Mn determinations by
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32 219 WCAES.
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38 220 The procedure's accuracy was evaluated by analyzing certified reference
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40 221 materials of plant with and without Mg addition (Table 3). In the absence of modifier,
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42 222 Mn values obtained were 2-fold higher than the certified ones. On the other hand, no
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44 223 statistically significant differences between determined and certified Mn values were
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46 224 observed, at a 95 % confidence level, for determinations using Mg. These results
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48 225 indicate that high concentrations of Mg also contribute to minimizing matrix effects in
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50 226 Mn determinations.
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54 227 It is interesting to note that the high recoveries observed when no chemical
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56 228 modifier was employed may be related to the natural presence of high concentrations of
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3 229 Mg in plant materials, which would cause the same effect observed when using the
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5 230 chemical modifier. Only in this case, signal increases would be observed for the
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7 231 digested sample, but not for the standard reference solutions. Considering the sample
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9 232 mass initially used and dilutions carried out after digestion, Mg concentrations in the
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11 233 sample solutions were approximately 100 mg L⁻¹. On the other hand, this and other
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13 234 matrix effects may be minimized by the overwhelming effect of adding even higher
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15 235 concentrations of Mg to both sample and standard reference solutions. In this case, in
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17 236 addition to the matrix matching effect, high concentrations of Mg may cause the two-
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19 237 step process represented in Eqns. 1-2 to be dominant and override other matrix effects.
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21 238 Therefore, one may say that the effect caused by Mg on Mn emission is in fact a matrix
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23 239 effect, and that in this case, it can be used to the benefit of the analytical procedure (*e.g.*
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25 240 a Mn signal is only observed in the presence of Mg in Fig. 1).
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242 **Conclusions**

243 The use of magnesium nitrate as a chemical modifier in Mn determinations by
244 WCAES results in improved sensitivity, precision, accuracy and power of detection.
245 Manganese atoms are trapped in a MgO matrix, which allows the interaction between
246 Mn and Mg⁺ species in the gas phase. The slightly larger Mg ionization potential
247 contributes to increasing the population of excited-state Mn atoms following charge-
248 transfer and electron recombination reactions. In the presence of 750 mg L⁻¹ of Mg, a
249 17-fold limit of detection improvement was achieved when compared to determinations
250 without the chemical modifier. The high concentrations of Mg added to both samples
251 and standard reference materials also contributed to minimizing matrix effects and
252 improving sensitivity in plant material analysis.

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3 253 This is the first time that $\text{Mg}(\text{NO}_3)_2$ is employed as chemical modifier in atomic
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5 254 emission spectrometry. This strategy is easily implemented and may result in significant
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7 255 improvements for methods prone to severe matrix effects such as WCAES.
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11 12 13 257 **Acknowledgments**

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16 258 The authors would like to thank the financial support from the Conselho
17
18 259 Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and Wake Forest
19
20 260 University. Sidnei G. Silva is grateful to the grants 2011/13288-7 and 2012/13647-0,
21
22 261 São Paulo Research Foundation (FAPESP).
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26 27 28 29 263 **References**

- 30
31
32 264 1. Z. F. Queiroz, P. V. Oliveira, J. A. Nóbrega, C. S. Silva, I. A. Rufini, S. S. Souza and
33
34 265 F. J. Krug, *Spectrochim. Acta Part B*, 2002, **57**, 1789–1799.
- 35
36
37 266 2. G. L. Donati and B. T. Jones, *J. Anal. At. Spectrom.*, 2011, **26**, 838-844.
- 38
39
40 267 3. J. A. Nóbrega, J. Rust, C. P. Calloway Jr. and B. T. Jones, *Spectrochim. Acta Part B*,
41
42 268 2004, **59**, 1337–1345.
- 43
44
45 269 4. C. G. Bruhn, J. Y. Neira, G. D. Valenzuela and J. A. Nóbrega, *J. Anal. At. Spectrom.*,
46
47 270 1998, **13**, 29–35.
- 48
49
50 271 5. Y. Terui, K. Yasuda and K. Hirokawa, *Anal. Sci.*, 1991, **7**, 397–402.
- 51
52
53 272 6. I. C. F. Damin, M. G. R. Vale, M. M. Silva, B. Welz, F. G. Lepri, W. N. L. Santos
54
55 273 and S. L. C. Ferreira, *J. Anal. At. Spectrom.*, 2005, **20**, 1332-1336.
56
57
58
59
60

- 1
2
3 274 7. B. Welz, G. Schlemmer and J. R. Mudakavi, *J. Anal. At. Spectrom.*, 1992, **7**, 1257–
4
5 275 1271.
6
7
8 276 8. A. B. Volynsky, *Spectrochim. Acta Part B*, 2000, **55**, 103–150.
9
10
11 277 9. I. Havezov, A. Detcheva and J. Rendl, *Microchim. Acta*, 1995, **155**, 147–155.
12
13
14 278 10. Y. Morishige, K. Hirokawa and K. Yasuda, *Anal. Bioanal. Chem.*, 1994, **350**, 410–
15
16 279 412.
17
18
19 280 11. M. Y. Shiue, J. Mierzwa and M. H. Yang, *J. Anal. At. Spectrom.*, 2001, **16**, 1172–
20
21 281 1179.
22
23
24 282 12. A. B. Volynsky, *Spectrochim. Acta Part B*, 1998, **53**, 1607–1645.
25
26
27 283 13. B. V. L'vov, *Spectrochim. Acta Part B*, 2000, **55**, 1659–1668.
28
29
30 284 14. W. Penninckx, D. L. Massart and J. Smeyers-Verbeke, *Anal. Bioanal. Chem.*, 1992,
31
32 285 **343**, 526–531.
33
34
35 286 15. W. Slavin, G. R. Carnrick and D. C. Manning, *Anal. Chem.*, 1982, **54**, 621–624.
36
37
38 287 16. D. L. Styris and D. A. Redfield, *Anal. Chem.*, 1987, **59**, 2891–2897.
39
40
41 288 17. D. L. Styris and D. A. Redfield, *Anal. Chem.*, 1987, **59**, 2897–2903.
42
43
44 289 18. H. M. Ortner, E. Bulska, U. Rohr, G. Schlemmer, S. Weinbruch and B. Welz,
45
46 290 *Spectrochim. Acta Part B*, 2002, **57**, 1835–1853.
47
48
49 291 19. M. M. Silva, R. B. Silva, F. J. Krug, J. A. Nóbrega and H. Berndt, *J. Anal. At.*
50
51 292 *Spectrom.*, 1994, **9**, 861–865.
52
53
54 293 20. P. J. Parsons, H. Qiao, K. M. Aldous, E. Millis and W. Slavin, *Spectrochim. Acta*
55
56 294 *Part B*, 1995, **50**, 1475–1480.
57
58
59
60

- 1
2
3 295 21. F. J. Krug, M. M. Silva, P. V. Oliveira and J. A. Nóbrega, *Spectrochim. Acta Part*
4
5 296 *B*, 1995, **50**, 1469–1474.
6
7
8 297 22. J. C. J. Silva, E. E. Garcia, A. R. A. Nogueira and J. A. Nóbrega, *Talanta*, 2001, **55**,
9
10 298 847–854.
11
12
13 299 23. Y. Zhou, P. J. Parsons, K. M. Aldous, P. Brockman and W. Slavin, *J. Anal. At.*
14
15 300 *Spectrom.*, 2001, **16**, 82–89.
16
17
18 301 24. A. Salido and B. T. Jones, *Anal. Lett.*, 2011, **44**, 2760–2767.
19
20
21 302 25. S. G. Silva, G. L. Donati, L. N. Santos, B. T. Jones and J. A. Nóbrega, *Anal. Chim.*
22
23 303 *Acta*, 2013, **780**, 7–12.
24
25
26 304 26. G. C.Y. Chan and W.T. Chan, *Spectrochim. Acta Part B*, 2003, **58**, 1301–1317.
27
28
29 305 27. J. Machat, V. Kanicky and V. Otruba, *Anal. Bioanal. Chem.*, 2002, **372**, 576–81.
30
31
32 306 28. J. Machat, V. Otruba and V. Kanicky, *J. Anal. At. Spectrom.*, 2002, **17**, 1096–1102.
33
34
35 307 29. J. A. Rust, J. A. Nóbrega, C. P. Calloway Jr. and B. T. Jones, *Spectrochim. Acta*
36
37 308 *Part B*, 2006, **61**, 225–229.
38
39
40 309 30. P. V. Oliveira, M. Catanho, J. A. Nóbrega and P. O. Luccas, *Quim. Nova*, 2000, **23**,
41
42 310 706–708.
43
44
45 311 31. G. L. Donati, J. Gu, J. A. Nóbrega, C. P. Calloway Jr. and B. T. Jones, *J. Anal. At.*
46
47 312 *Spectrom.*, 2008, **23**, 361.
48
49
50 313 32. P. Allain, L. Jaunault, Y. Mauras, J.M. Mermet and T. Delaporte, *Anal. Chem.*,
51
52 314 1991, **63**, 1497–1498.
53
54
55 315 33. C. D. Pereira, E. E. Garcia, F. V. Silva, A. R. A. Nogueira and J. A. Nóbrega, *J.*
56
57 316 *Anal. At. Spectrom.*, 2010, **25**, 1763–1768.
58
59
60

- 1
2
3 317 34. M. Pettine, B. Casentini, D. Mastroianni and S. Capri, *Anal. Chim. Acta*, 2007, **599**,
4
5 318 191–198.
6
7
8 319 35. G. Grindlay, L. Gras, J. Mora and M. T. C. de Loos-Vollebregt, *Spectrochim. Acta*
9
10 320 *Part B*, 2008, **63**, 234–243.
11
12
13 321 36. *Natl. Inst. Stand. Technol. Chem. WebBook, NIST Stand. Ref. Database*,
14
15 322 <http://webbook.nist.gov/chemistry/form-ser.html> (accessed February, 2013).
16
17
18 323 37. A. B. Volynsky, *Spectrochim. Acta Part B At. Spectrosc.*, 1998, **53**, 139–149.
19
20
21 324 38. V. Muzgin, Y. Atnashev, V. Korepanov and A. Pupyshev, *Talanta*, 1987, **34**, 197–
22
23 325 200.
24
25
26 326 39. Z. F. Queiroz, F. J. Krug, P. V Oliveira, M. M. Silva and J. A. Nóbrega,
27
28 327 *Spectrochim. Acta Part B*, 2002, **57**, 49-61.
29
30
31 328
32
33
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3 329 **Figures captions**
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5 330 **Figure 1** – Transients emission signals for manganese solutions (5 mg L^{-1}) in presence
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8 331 (—) and absence (---) of magnesium nitrate (750 mg L^{-1}) as chemical modifier.
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10 332 **Figure 2** - Effect of Mg concentration on Mn emission signals ($\text{Mn} = 5 \text{ mg L}^{-1}$).
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Tables

Table 1 - W-coil heating program for Mn determination.

Step	Applied Current (A)	Time (s)
1	3.0	40
2	2.7	30
3	2.5	20
4	2.0	15
5	1.5	10
6	0	10
7	10	4

Table 2 – Analytical features for Mn solutions in presence and absence of magnesium as chemical modifier.

Analytical Features	Mn/Mg	Mn
Linear range (mg L ⁻¹)	0.1 – 7.5	2 - 20
Linear equation	S = 2262C + 399	S = 147C + 4
R	0.998	0.997
LOD (mg L ⁻¹)	0.05	0.86
RSD (%)	3.6	6.4

Table 3 - Manganese determination (mean \pm confidence interval, n = 3) in certified reference materials in presence and absence of magnesium nitrate as chemical modifier.

CRM	Certified value	With chemical modifier	Without chemical modifier
Peach Leaves (SRM 1547)	98 \pm 3	94 \pm 4	219 \pm 25
Tomato Leaves (SRM 1573)	238 \pm 7	224 \pm 13	448 \pm 13

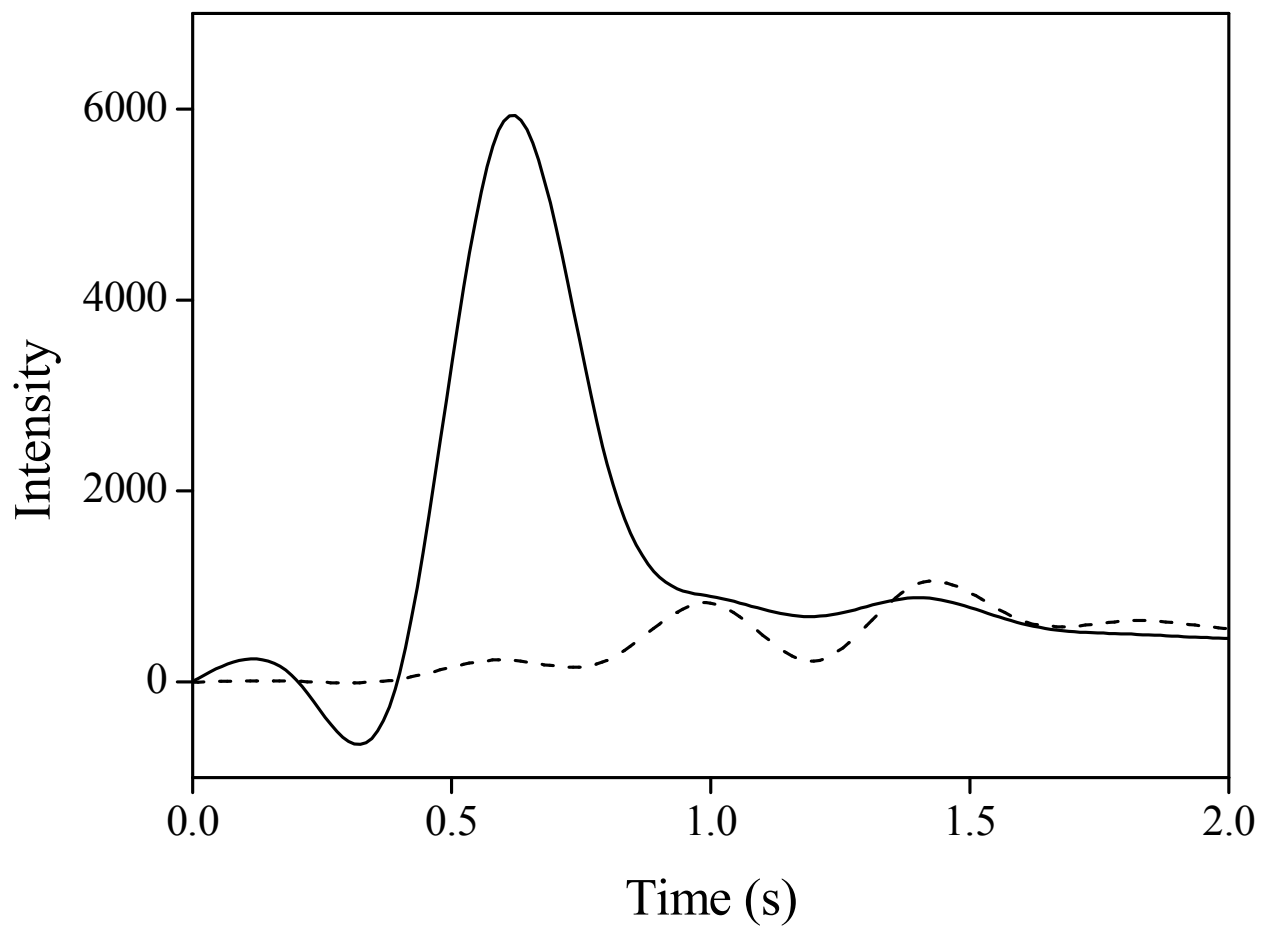


Figure 1

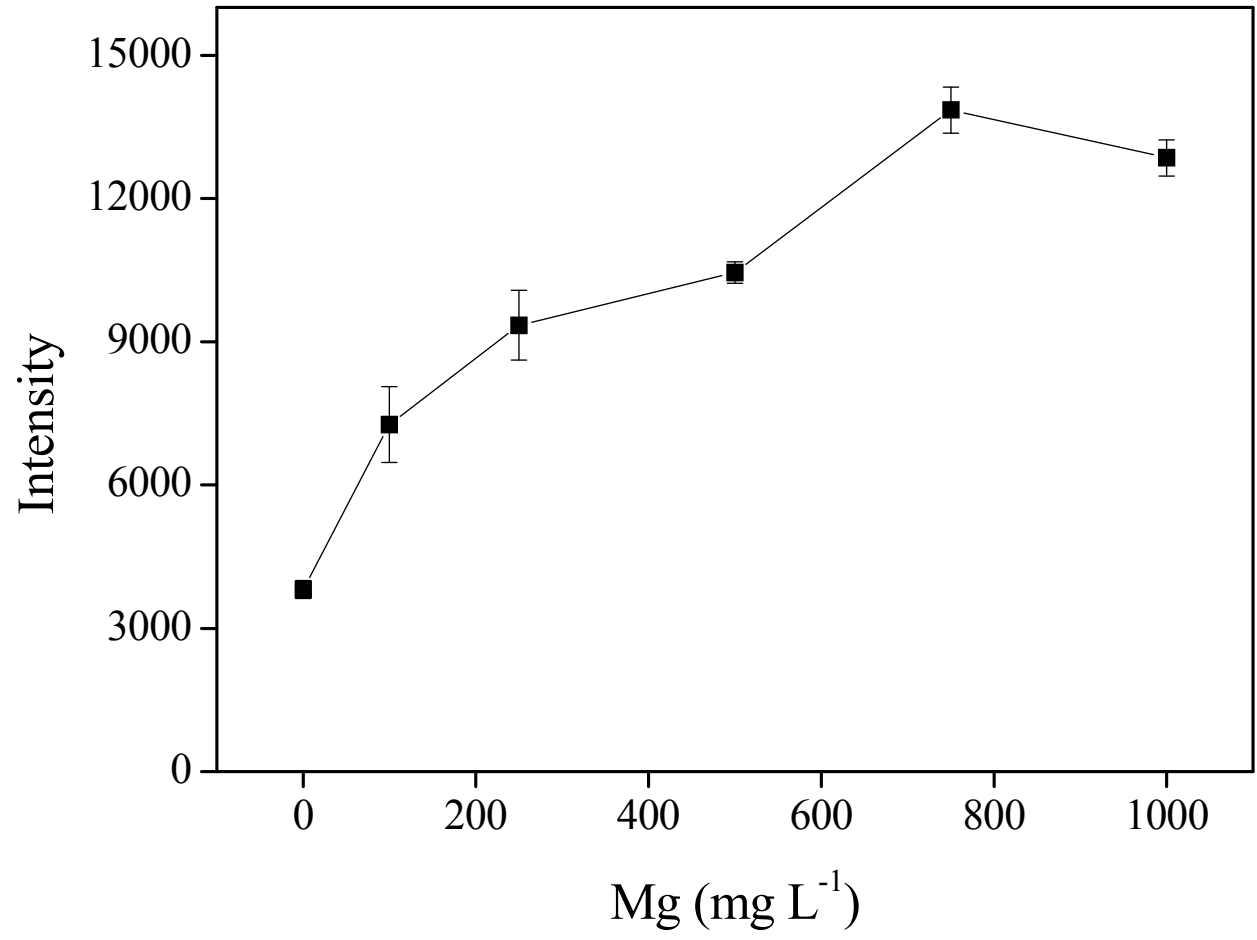


Figure 2

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Table of contents entry

The use of magnesium nitrate as a chemical modifier in atomic emission spectrometry to improve sensitivity and minimize matrix effects in Mn determinations is here demonstrated.

