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ARTICLE TYPE

Regioselective acetylation of carbohydrates and diols catalyzed by tetramethyl-ammonium hydroxide in water

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A novel method for an efficient regioselective acetylation of carbohydrates and diols in aqueous solution is described. Treatment of substrates with 1-acetylimidazole, tetramethylammonium hydroxide (TMAH) in water under mild 10 conditions gave highly regioselective acetylation for primary

hydroxyl groups. This discovery provides an eco-friendly way

for selective acetylation of non-protected glycosides and diols in water, avoiding the use of toxic organic solvents and the necessity of pre-protection of secondary hydroxyl groups.



Regioselective protection of polyols and diols under mild and eco-friendly conditions represents a central challenge in chemical ²⁰ synthesis.^[1] Carbohydrates, which are important and classic polyols, contain hydroxyl groups with similar reactivities. Regioselective protection of carbohydrates with several hydroxyl groups is a long-standing problem.^[2] Recently, several protection methods have been developed to this end by using certain 25 reagents as promoters to enlarge small differences between reactivities of hydroxyl groups in carbohydrates and diols. include reactions employing organotin,^[3] Examples organoboron,^[4] organosilicon,^[5] and metal salts,^[6] organocatalytic method,^[7] and enzymatic method.^[8] However, many of 30 these promoters have short-comings in selective protection of carbohydrates and diols, including environmental toxicity, expensive reagents, moisture intolerance, and the necessity of pre-protection of secondary hydroxyl groups. Hence, there is an urgent need to find a mild, non-toxic and eco-friendly method for

³⁵ selective protection of carbohydrates and diols, where the reaction is best carried out in water since reactions in aqueous solution have recently attracted considerable attention from the standpoint of green chemistry.^[9]

In this study, we sought to develop a green approach for ⁴⁰ selective acetylation of primary hydroxyl groups in carbohydrates and diols. It was known that carbonylimidazole derivatives constitute a class of highly active acetylation reagents, particularly in preparation of esters and amides.^[10] In several cases, the carbonylimidazole derivatives have shown superior ⁴⁵ selectivity for N-acetylation of oxindoles and indoles (Scheme 1).^[11] However, the reactions mentioned above were run in organic solvents because of poor water solubility of carbonylimidazole derivatives. Inspired by this work, we decided to explore the possibility of 1-acetylimidazole (AI) as an ⁵⁰ acetylation reagent for replacing toxic and unstable acid halides in water.



Scheme 1 N-acetylation of heterocycles with carbonylazoles

In another aspect, we sought to find an eco-friendly basic catalyst for the acetylation of carbohydrates and diols. The quaternary ammonium hydroxide as a classical strong organic alkali attracted our attention due to its unique features: $R_4N^+OH^-$ s zwitterionic form, non-toxicity, easy degradation, good water solubility, cheap and commercial availability. Earlier, Wang used tetraethylammonium hydroxide (TEAH) as catalyst for

- selectively protecting the primary hydroxyl group of ribonucleotides and ribonucleosides in DMF.^[12] Recently, tetramethylammonium hydroxide (TMAH) has been used in several important organic transformations.^[13] All these previous
- several important organic transformations.^[13] All these previous studies encouraged us to investigate TMAH as a basic catalyst in the selective acetylation of carbohydrates and diols with AI in water.



Scheme 2 Regioselective acetylation of methyl α-D-glucopyranoside

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Taking methyl α -D-glucopyranoside (1) as substrate, the regioselective acetylation of 1 in water was catalyzed by base in the presence of AI (Scheme 2). To improve water solubility of AI, ²⁰ the reaction temperature was increased to 50 °C. The catalytic effects of a range of bases, including TMAH, have been investigated (Table 1). As can be seen in Table 1, DBN, NaOH, and Na₂CO₃ led to low yields (Entry 2, 4 and 5). However, as we hoped, the best yield was obtained with TMAH as catalyst.

25 Table 1 Optimization of methyl α-D-glucopyranoside acetylation ^a

Entry	Base	Yield ^c (%)
1	DBU	30
2	DBN	18
3	LiOH	35
4	NaOH	23
5	Na ₂ CO ₃	26
6	K_2CO_3	32
7	TMAH	45 (52 ^b)
8	TEAH	33

^{*a*}AI: 2.0 equiv., base: 0.5 equiv., Temp: 50 °C, time: 24 h; ^{*b*}AI: 3.0 equiv., TMAH: 1.2 equiv., Temp: 60 °C, time:16 h; ^cIsolated yield. DBU: 1,8diazabicyclo[5.4.0]-undec-7-ene, DBN: 1,5-diazabicyclo-[4.3.0]non-5ene, TMAH: tetramethylammonium hydroxide, TEAH: tetraethylam-³⁰ monium hydroxide.

In a series of optimization studies, methyl α -D-glucopyranoside was allowed to react with various quantities of TMAH (0.2 to 2.0 equiv) in the presence of AI (1.2 to 4.0 equiv) in water at temperature range from rt to 80 °C. It was found that ³⁵ methyl 6-*O*-acetyl- α -D-glucopyranoside (2) was synthesized in

- the presence of TMAH (1.2 equiv) and 1-acetylimidazole (3.0 equiv) at 60 $^{\circ}$ C in water for 16 h with an acceptable yield of 52%. It was also observed that an amount of TMAH more than 2.0 equiv disfavored the formation of product **2**.
- With these findings in hand, the optimized catalyst system found above was further used for selective monoacetylation with a wide range of substrates (Table 2), including non-protected methyl D-pyranoside (entry 1-5), partially protected methyl Dpyranoside (entry 6-8) and diols (entry 9-16). The common
- 45 feature of the substrates is that they all contain a primary and one

or more secondary hydroxyl groups. The reaction condition has been successfully applied for the selectively monoacetylating carbohydrates and diols, where only minute quantities of byproducts were detected by ¹H NMR (Fig. 1). Especially, the non-50 protected methyl D-pyranosides showed high regioselectivity for primary hydroxy group and had almost no secondary acetylated product (entry 1-3 and 5, only trace amount of byproduct detected). The partially protected methyl D-pyranosides had good selectivity for primary hydroxy group as well. The byproduct 55 yield was no higher than 7% (see Fig. 1a, methyl 3-O-benzyl-α-D-galactopyranoside acetylated byproduct 12b only about 5%; Fig. 1b, methyl 3-O-benzyl-β-D-galactopyranoside byproduct 14b only about 7%). When the method was expanded to protect the primary hydroxyl group of various diols, good regioselectivity 60 and yields were also obtained. The byproduct yield is less than 8% (Fig. 1c, 1-phenyl-1, 2-ethanediol acetylated byproduct 20b only about 8%; Fig. 1d guaifenesin acetylated byproduct 30b only about 6%).



⁶⁵ Fig. 1 ¹H NMR spectra of product mixtures resulting from different substrates: (a) methyl 3-*O*-benzyl-6-*O*-acetyl-α-D-galactopyranoside (11), (b) methyl 3-*O*-benzyl-6-*O*-acetyl-β-D-galactopyranoside (13), (c) 1-phenyl-1,2-ethanediol (19), (d) guaifenesin (29).

Because the ¹H NMR spectra of product mixture resulting 70 from substrate 21 (entry 11) and 25 (entry 13) lack suitable characteristic peaks for comparison, we isolated their byproducts (22b yield 5% and 26b yield 6%) by chromatography. Moreover, no byproducts were detected by ¹H NMR when the substrates were 7 (methyl B-D-galactopyranoside, entry 4), 17, 23, and 31 75 (diols, entry 9, 12 and 16). To the best of our knowledge, this is the first report for regioselective acetylation of carbohydrates and diols in water with AI as acetylation reagent in the presence of TMAH, where acetylation was performed by transacylation reactions of imidazolides under strong basic condition.^[14] Based so on the previous studies.^[12a,15] we proposed a mechanism for the regioselective acetylation of primary hydroxyl group using TMAH as a catalyst in water. In this hypothesis, both primary and secondary hydroxyl groups can be deprotonated in the presence of strong base TMAH. The acylation of primary 85 hydroxyl group are more preferably produced due to its less steric hindrance and stronger nucleophilicity than the other hydroxyl groups.^[12a,15] Moreover, Edgar and co-worker reported that

Table 2 1-acetylimidazole used for selective acetylation of primary hydroxyl group in carbohydrates and diols

но НО	° ∕_N∕∼N ∖/	AcO HO	
$R_1 R_2$	TMAH, H ₂ O, 16 h, 60 °C	$R_1 R_2$	

Entry	Substrates	Major products	Byproducts	Conversion (%) ^c Isola	ated yield (%)	Ref
1		HO HO HO HO HO OCH ₃	Trace amount	60	52	18
2		HO HO HO HO HO HO	Trace amount	62	56	9d
3		HO OAC HO HO OCH ₃	Trace amount	64	59	19
4			-	74	70	20
5		HO HO HO HO 10 OCH	Trace amount	50	44	21
6	HO OH BnO HO OCH	HO OR_2 BnO R_1O OCH . R_1O OCH . $R_2=OAc$ $12a:R_1=H, R_2=OAc$ $12b:R_1=OAc, R_2=H$	5% of 12b based on NMR	58	50	_
7		HO OR_2 14a:R ₁ =H, R ₂ =OAc 14b:R ₁ =OAc, R ₂ =H BnO OCH_3	7% of 14b based on NMR	61	52	21
8	HO BnO 15 OCH ₃	HO BnO 16 OCH ₃	Trace amount	48	43	21
9	он 17 — Он		_	39	36	22
10	он 19 Рh ОН	OR1 20a:R1=H, R2=Ac 20b:R1=Ac, R2=H Ph OR2	8% of 20b based on NMR	70	60	19
11 2	OH OLOH	$\bigcirc \begin{array}{c} OR_1 & 22a:R_1=H, R_2=Ac\\ O & OR_2 & 22b:R_1=Ac, R_2=H \end{array}$	5% of 22b product was isolated.	64	56	23
12	23 HO OH	24 HO OAc	-	73 ^b	70	24
13		$\begin{array}{c} OR_1 & 26a:R_1=H, R_2=Ac \\ O & OR_2 \end{array}$	6% of 26b product was isolated	65	55	25
14	ОН 27 — ООН	0H 28OOAc	Trace amount	54	50	25
15		OH 30a:R ₁ =H, R ₂ =Ac 30b:R ₁ =Ac, R ₂ =H	6% of 30b based on NMR	71	62	26
16		OCOH 32 OH OAc	_	60 ^b	55	27

⁵ Notes: ^a reaction conditions: AI (3.0 equiv.); TMAH (1.2 equiv.); ^bTMAH (1.5 equiv.); ^cNMR ratio.

deacylation of cellulose triesters in DMSO by adding TBAOH aqueous solution, mainly affording cellulose-6-O-esters, which demonstrated primary ester is more stable than secondary esters in the basic condition.^[16] Leino and co-worker studied the acyl

s groups migration for β-D-galactopyranosides in D₂O, which demonstrated the secondary esters is easier to migration to produce primary ester at basic condition $(C2 \leftrightarrows C3 \boxdot C4 \rightarrow C6 \text{ migration})$.^[17]

Unlike the known methods, where the selective acetylation ¹⁰ required pre-preparation of catalysts, toxic organic solvents, or pre-protection of secondary hydroxide groups, the method we developed in this work has no such disadvantages, and is therefore simple, eco-friendly, and highly regioselective. While yields are moderate, they compare quite favorably in general to

- ¹⁵ the overall yields of the complex protection, acylation, and deprotection sequences that they can replace. For example, regioselective acetylation of 7 and 3-methyl-1,3-butanediol **23** under the optimized conditions yielded the corresponding acetylated products in 70% yield (see Entry 4 and 12). These
- ²⁰ results indicated that TMAH was successfully used to regioselectively acylate carbohydrates as well as diols.

Conclusions

For the first time, an eco-friendly approach for regioselective acetylation of carbohydrates and diols in aqueous ²⁵ solution has been developed. Highly regioselective acetylation for

- 25 solution has been developed. Highly regisselective acetylation for primary hydroxyl groups can be obtained by using 1acetylimidazole as an acetylation reagent in the presence of TMAH, avoiding the complex, costly, time-consuming protection and deprotection schemes. The acetylation reaction can be
- ³⁰ conveniently carried out in water, avoiding the use of toxic and flammable organic solvents. With these particular merits, this method will provide a novel approach to an environmentally benign process for regioselective acetylation of primary hydroxyl groups in carbohydrates and diols.

35 Experimental

General Acetylation procedure of substrates (methyl Dpyranosides or diols)

To a solution of substrate (1 equiv.) in water (0.2 mL) was added TMAH (25% aq. 1.2 equiv.), and the mixture was allowed ⁴⁰ to stir at 60 °C for 5 min. 1-acetylimidazole (3.0 equiv.) was added to the reaction mixture in two portions and it was allowed to stir at 60 °C for 16 h. Water was removed under reduced pressure and the resulting mixture was purified by flash column chromatography (ethyl acetate/petroleum ether: 30% to 100% ⁴⁵ gradient) to afford acetylated products.

Methyl 3-O-benzyl-6-O-acetyl-a-D-galactopyranoside (14):

¹H NMR (CDCl₃, 500 MHz) δ 7.37-7.32 (m, 5H, Ph), 4.83 (d, J = 3.9 Hz, 1H, H₁), 4.73 (m, 2H, PhCH₂), 4.38-4.24 (m, 2H, H_{6a}, H_{6b}), 4.08-3.89 (m, 3H, H₂, H₄, H₅), 3.63 (d, J = 9.6 Hz, 1H, H₃), ⁵⁰ 3.41 (s, 3H, OMe), 2.44 (s, 1H, OH), 2.15 (d, J = 8 Hz, 1H, OH), 2.08 (s, 3H, OAc).; ¹³C NMR (CDCl₃, 125 MHz) δ 170.8, 137.8, 128.7, 128.2, 127.9, 99.5, 78.2, 72.3, 68.5, 67.8, 67.0, 63.6, 55.4, 20.9; $[\alpha]^{20}{}_{D} = +53.5(c = 1.6, CH_3OH)$. HRMS calcd for C₁₆H₂₂O₇ [M + Na] 349.1263. Found 349.1261.

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60 Notes and references

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‡ Footnotes should appear here. These might include comments relevant 70 to but not central to the matter under discussion, limited experimental and spectral data, and crystallographic data.

- (a) A. Ferry, X. Guinchard, P. Retailleau and C. David, *J. Am. Chem. Soc.*, 2012, **134**, 12289-12301; (b) Y. X. Zhou, M. Rahm, B. Wu and H. Dong, *J. Org. Chem.*, 2013, **78**, 11618-11622.
- 75 2 (a) R.U. Lemieux, H. Driguez, J. Am. Chem. Soc., 1975, 97, 4063-4069; (b) K. C. Nicolaou and H. J. Mitchell, Angew. Chem., Int. Ed., 2001, 40, 1576-1624; (c) W. Muramatsu, J. M. William and O. Onomura, J. Org. Chem., 2012, 77, 754-759.
- 3 (a) S. David. S. Hanessian, *Tetrahedron*, 1985, 41, 643-663; (b) T. B.
 Grindley, Adv. Carbohyd. Chem. Bi., 1998, 53, 17-142; (c) W.
 Muramatsu, S. Tanigawa, Y. Takemoto and O. Onomura, *Chem.-Eur.* J., 2012, 18, 485-4853; (d) W. Muramatsu, Y. Takemoto, J. Org. Chem., 2013, 78, 2336-2345; (e) H. Dong, Y. X. Zhou, X. L. Pan, F.
 C. Cui and O. Ramström, J. Org. Chem., 2012, 77, 1457-1467; (f) H.
 F. Xu, Y. C. Lu, Y. X. Zhou, B. Ren, Y. X. Pei, H. Dong and Z. C.
 Pei, Adv. Synth. Catal., 2014, 8,1735-1740.
- 4 (a) D. Lee, M. S. Taylor, J. Am. Chem. Soc., 2011, 133, 3724-3727; (b)
 C. Gouliaras, D. Lee, L. Chan and M. S. Taylor, J. Am. Chem. Soc., 2011, 133, 13926-13929; (c) D. Lee, C. L. Williamson, L. Chan and
 M. S. Taylor, J. Am. Chem. Soc., 2012, 134, 8260-8267.
- 5 (a) C. C. Wang, J. C. Lee, S. Y. Luo, S. S. Kulkarni, Y. W. Huang, and S. C. Hung, *Nature.*, 2007, 446, 896-899; (b) Y. Bourdreux, A. Lemétais, D. Urban and J. M. Beau, *Chem. Commun.*, 2011, 47, 2146-2148; (c) Y. X. Zhou, O. Ramström and H. Dong, *Chem Commun.*, 2012, 48, 5370-5700.
- 6 (a) H. Wang, J. She, L. H. Zhang and X. S. Ye, J. Org. Chem., 2004,
 69, 5774-5777; (b) I. J. Gray and R. Kluger, Carbohydr. Res., 2007,
 342, 1998-2002; (c) R. S. Dhiman and R. Kluger, Org. Biomol. Chem., 2010, 8, 2006-2008.
- (a) K. S. Griswold and S. J. Miller, *Tetrahedron.*, 2003, **59**, 8869-8875; (b) T. Kawabata, W. Muramatsu, T. Nishio, T. Shibata and H. Schedel, *J. Am. Chem. Soc.*, 2007, **129**, 12890-12895; (c) X. Sun, H. Lee, S. Lee and K. L. Tan, *Nat. Chem.*, 2013, **5**, 790-795.
- 8 (a) M. Therisod and A. M. Klibanov, J. Am. Chem. Soc., 1987, 109, 3977-3981; (b) F. Theil and H. Schick, Synthesis., 1991,7,533-535; (c) M. C. Roux, J. S. Pene and L. Wartski, J. Org. Chem., 1993, 58, 3969-3973; (d) B. Danieli, M. Luisetti, G. Sampognaro, G. Carrea and S. Riva, J. Mol. Catal. B: Enzym., 1997, 3, 193-201.
- 9 (a) B. Feng, Z. S. Hou, X. R. Wang and Y. X. Qiao, *Green Chem.*, 2009, 11, 1446-1452; (b) J. L. Song, H. L. Fan, J. Ma and B. X. Han, *Green Chem.*, 2011, 15, 2619-2635; (c) A. Santra, G. Guchhait and A. Kumar Misra, *Green Chem.*, 2011, 13, 1345-1351; (d) M. Osada, K. Kikuta, K. Yoshida, M. Oqata and T. Usui, *Green Chem.*, 2013, 15, 2960-2966; (e) J. E. Andreasen, B. Shimpukade and T. Ulven, *Green Chem.*, 2013, 15, 336-340.
 - (a) Z. Simone, S. Roger and Y. Esther, *Eur. J. Org. Chem.*, 2002, 15, 2633-2639;
 (b) P. A. Duspara, M. S. Islam and R. A. Batey, *J. Org. Chem.*, 2012, 22, 10362–10368.

- (a) S. T. Heller, E.E. Schultz and R. Sarpong, *Angew. Chem., Int. Ed.*, 2012, **51**, 8304-8308; (b) S. T. Heller, T. Fu and R. Sarpong, *Org. Lett.*, 2012, **14**, 1970-1973
- 12 (a) Y. Wang, Y. Z. Xu and Z. W. Yang, *Acta Chim. Sinica.*, 1988, 3, 244-250; (b) Y. Wang, X. Y. Liu and Z. W. Yang, *Nucleic Acids Res.*, 1987, **15**, 4291-4035.
- 13 F. Shadkami and R. Helleur, J. Anal. Appl. Pyrol., 2010, 89, 2-16.
- 14 H. A. Staab, Angew. Chem., Int. Ed., 1962, 7, 351-367.
- 15 Y. Sawada, N. Nanboku, E. Yanase and S. Nakatsuka, *Heterocycl.* 10 *Commun.*, 2010, **1**, 21-24;
- 16 (a) X. Zheng, R. D. Gandour and K. J. Edgar, *Carbohyd. Polym.*, 2014, **111**, 25-32; (b) X. Zheng, R. D. Gandour and K. J. Edgar, *Biomacromolecules.*, 2013, **14**, 1388–1394.
- 17 M. U. Roslund, O. Aitio, J. Warna, H. Maaheimo, D. Y. Murzin and 15 R. Leino, *J. Am. Chem. Soc.*, 2008, **130**, 8769-8772.
- 18 H. Tina, C. H. Tran and D. Crout, J. Chem. Soc. Perkin Trans. 1, 1998, 6, 1069-1080.
- 19 P.E. Jansson, L. Kenne and E. Schweda, *Tetrahedron Lett.*, 1987,4, 651-652.
- 20 20 B. Danieli, M. Luisetti, G. Sampognaro, G. Carrea and S. Riva, J. Mol. Catal. B: Enzym., 1997, 3, 193-201.
- 21 H. Dong, Y. X. Zhou and H. Y. Liu, *Faming Zhuanli Shenqing Gongkai Shuomingshu*. CN 103193570 (2013).
- 22 M. Šato, K. Matsushima, H. Kawanami and R. Sarpong, *Angew. Chem., Int. Ed.*, 2007, **46**, 6284-6288.
 - 23 M. N. S. Rad and S. Behrous, Mol. Divers., 2013, 17, 9-18
 - 24 W. C. Lin and T. H. Morton, J. Org. Chem., 1991, 56, 6850-6856
 - 25 J. Otera, Y. Niibo, N. Tatsumi and H. Nozaki, J. Org. Chem., 1988, 53, 275-278.
- ³⁰ 26 B. K. Liu, Q. Wu, D. S. Lv and X. F. Lin, *J. Biotechnol.*, 2011, **153**, 111-115
 - 27 P. Cesti, A. Zaks and A. M. Klibanov, *Appl. Biochem. Biotech.*, 1985, 11, 401-406