

**Impact of mastication, salivation and food bolus formation
on salt release during bread consumption**

Journal:	<i>Food & Function</i>
Manuscript ID:	FO-ART-05-2014-000446.R1
Article Type:	Paper
Date Submitted by the Author:	23-Jul-2014
Complete List of Authors:	Tournier, Carole; INRA UMR CSGA, Grass, Manon; INRA UMR CSGA, Septier, Chantal; INRA UMR CSGA, bertrand, dominique; Data_frame, Salles, Christian; INRA UMR CSGA,

Impact of mastication, salivation and food bolus formation on salt release during bread consumption

Carole Tournier^{1,2,3}, Manon Grass^{1,2,3}, Chantal Septier^{1,2,3}, Dominique Bertrand⁴, Christian
Salles^{1,2,3}

¹ CNRS, UMR6265 Centre des Sciences du Goût et de l'Alimentation, F-21000 Dijon, France

² INRA, UMR1324 Centre des Sciences du Goût et de l'Alimentation, F-21000 Dijon, France

³ Université de Bourgogne, UMR Centre des Sciences du Goût et de l'Alimentation, F-21000
Dijon, France

⁴ Data_frame, 25 rue Stendhal, 44300 Nantes, France

1 **Abstract**

2 Health authorities recommend increasing fibre and decreasing salt content in bread
3 products. However, these basic ingredients of bread composition are multifunctional, and
4 important changes in their content influence the texture, flavour and acceptability of the
5 product.

6 This study was designed to investigate the link between oral processing, bolus formation and
7 sodium release during the consumption of four different breads that varied in composition
8 and structure. Chewing behaviour was determined using surface electromyography, and
9 salivation was quantified from the water content of the boluses collected. The kinetics of
10 bread degradation during food bolus formation was characterised by measuring bolus
11 heterogeneity using texture image analysis, and sodium release into the saliva was
12 quantified. Mastication and salivation varied between products and between subjects, thus
13 highlighting different bolus formation strategies. In vivo salt release was mainly explained by
14 mastication parameters. The initial slope of sodium release increased when the chewing
15 muscles' activity increased, and the maximum sodium concentration was reached later when
16 more masticatory cycles were required to reach the swallowing point.

17

18

19 **Keywords:** Bread, Salt Release, Food Bolus, Mastication, Salivation

20 Introduction

21 Cereals are an important source of dietary fibres. Beside insoluble fibres, soluble fibres have
22 beneficial effects on health through conferring a low glycaemic index to foods high in soluble
23 fibres, in particular. The glycaemic index of bread is a specific dietary feature that may
24 influence metabolic and cardiovascular risk factors long-term.¹⁻³ This index is known to
25 depend on the bread's composition and processing. For example, the traditional baguette was
26 found to have a lower index than that of the classic baguette,⁴ and pumpernickel bread had a
27 lower index than white bread.⁵ These last years, researchers aimed to develop new cereal
28 products with higher dietary fibre content and reduced glycaemic index.^{6,7}

29 After processing steps, the final bread texture, governed by the mechanical properties of
30 the crumb and the crust, and flavour are formed. These properties mainly depend on the
31 matrix material, such as cross-linked gluten, starch, of which the dispersion and structure are
32 a function of both hydration and thermal history during the process, and other minor flour
33 components. The mechanical properties of the bread depend on the density and cellular
34 structure⁸⁻¹⁰, and changes in the amount and nature of dietary fibres affect dough rheological
35 behaviour and water binding, and thus significantly modify bread texture.^{11, 12} The texture of
36 food influences the oral behaviour of the consumer during eating. In particular, the
37 mastication and saliva parameters are affected, with important consequences on the release of
38 tastants, odourants and temporal perception during the in-mouth process.¹³

39 During mastication, bread is progressively hydrated by saliva and broken into particles of
40 different sizes, increasing the interface area between the food matrix and the saliva phase and
41 favouring the transfer of stimuli from the food bolus to the saliva phase. The saliva is a
42 complex viscous aqueous medium containing various salts and proteins, and enzymes that are
43 able to partially modify the initial structure and composition of the food and the availability of
44 some flavour components during the in-mouth process. As an example, a direct relation was

45 found between the alpha-amylase activity level in saliva and saltiness perception of starchy
46 matrices.¹⁴ The breakdown of the matrix due to the action of alpha-amylase favours the
47 release of sodium in saliva and consequently increases the perception of saltiness.

48 Sodium salt is another bread component linked to health issues¹⁵. International public
49 health policies recommend that average population daily salt intake should be lower than 5 - 6
50 g per day. In most of the industrialised countries, bread, and more generally bakery products,
51 contribute in a large part to the daily sodium intake and is the first major sodium source. As
52 an example, in European countries, bread accounts for a range of 19-26% of the total salt
53 intake according to the countries and the optimal salt content in white bread is between 1.29
54 and 1.43%.¹⁶ However, in bakery products, salt is a common multifunctional ingredient
55 contributing to both sensory and technological properties, such as impacting the development
56 of gluten structures in the mixing of bread¹⁷ and more generally fermented bakery products,
57 inhibiting bakers' yeasts in the fermentation of bread dough and controlling the water activity
58 in the baked products.^{15, 18} In particular, the control of water activity in bakery products is
59 critical to both product quality and safety.

60 In bread, many strategies have already been explored to reduce sodium content, such as
61 partial substitution of sodium salt by potassium salt,^{19, 20} potentiation of saltiness by the
62 substitution of salt with fermented ingredients,²¹ progressive reduction of salt in bread over
63 time²² and changes in the sodium concentration distribution.²³ These strategies only allow for
64 partial sodium reductions. However, in-mouth salt release and saltiness perception depends on
65 both food characteristics and oral parameters.¹³ It was reported that the combined effects of
66 food composition and chewing behaviour affected salt release in model cheeses.²⁴⁻²⁶ In
67 particular, fat influenced in-mouth salt release and saltiness perception differently depending
68 on the fat level.²⁶ Moreover, most people develop an individual oral strategy consisting of an
69 adaptation of oral behaviour to the food characteristics. In cheeses products, it was reported

70 that among the 70% of subjects who adapted their chewing behaviour, 57% adapted their
71 behaviour via chewing time, and 40% adapted their behaviour via chewing time and muscular
72 contraction amplitude.²⁷ Few studies have been reported about breads. As soon as the salt
73 concentration is lowered in crumbs, differences in salt content can be distinguished by
74 consumers. Moreover, salt influences not only saltiness but also aroma perception, and it
75 masks unwanted flavour attributes, such as mustiness and flouriness.²⁸ Saltiness in bread is
76 influenced by both the velocity of sodium release and the texture of the crumb.²⁹ Bread
77 texture also influences saltiness perception. Among three breads of different structures and
78 textures, the denser bread was perceived as the least salty.³⁰ Bread texture is also important for
79 bolus formation and swallowing. For two breads of different textures, small particle size and
80 an appropriate amount of saliva are both important to give to the bread bolus the adequate
81 rheological properties to be swallowed.³¹ The physical structure of bread influenced the
82 mastication time, which lasted until the water uptake was appropriate for swallowing. In
83 particular, the researchers reported that the plasticising effect of water on starch, which
84 influenced the gradual decrease of viscosity of the bread bolus during chewing, is more
85 important than particle fragmentation for the rheological behaviour of the bread bolus.
86 However, bolus viscoelasticity did not seem to be a key parameter to trigger swallowing, and
87 the panellists exhibited different masticatory and bolus hydration behaviour.³⁰

88 The aim of this study is to evaluate the influence of bread characteristics and oral
89 characteristics of individuals on in-mouth sodium release. This study, taking into account the
90 interactions between individuals and products, should provide information on higher in-mouth
91 sodium release for low-salt reformulated breads, thus compensating for a lower saltiness
92 intensity.

93

94

95 **Experimental**

96 **Breads**

97 Four commercial breads with different textures and compositions were studied (Table 1). Two
98 breads were French baguettes. Baguettes are made from wheat flour, water, salt and yeast.
99 The texture is characterised by a crisp eggshell crust 3 - 4 mm thick and an open and random
100 crumb cell structure.³² They were provided by a local supermarket (industrial manufacturing
101 sector) and by a local bakery (artisan manufacturing sector). An industrial toast bread (white-
102 wheat pan bread) was also studied. This type of industrial bread also contains fats and sugars.
103 Toast bread crumb is characterised by a flexible and soft texture and small cells with thin
104 borders. The crust is very thin and soft.³³ Finally, a German rye bread (pumpernickel) made
105 from whole rye grains, water, salt and sourdough was selected. The texture is usually dense
106 and composed of rye grains.

107 These products were characterised for their density, water content, hardness (maximum
108 force), crust/crumb ratio and sodium content. Bread densities were measured in five replicates
109 according to the rapeseed displacement method.³⁴ Water content was determined in triplicate
110 after drying in an oven as described below. Maximum force was determined from a
111 compression test developed to characterise the mechanical behaviour of products with
112 different crust/crumb textures (whole breads).³⁵ Baguette samples were obtained by cutting
113 the baguettes into 11-cm length slices. Samples of similar size (around 11 × 5 × 5 cm) were
114 prepared from the toast bread and rye bread by superimposing several slices. All samples
115 were compressed using a TAXT2 Texture Analyser (Swantech International, Gennevilliers,
116 France) fitted with a multi-puncture probe (two series of 8 spikes). Compression was set at
117 60% strain of the initial height and at a speed of 0.8 mm sec⁻¹. The maximum compression
118 force for each type of bread was determined. Eight replicates were run per type of product.
119 The approximate crust/crumb ratio was determined by carefully separating the crust and the

120 crumb and weighting each portion. Sodium content was determined by atomic absorption
121 spectrometry on the ash content (Tests carried out by Laboratoires Agrobio, Vezin le Coquet,
122 France).

123 In the following experiments, breads were presented to subjects in 5 ± 0.1 g slices in a
124 format representative of how that bread is usually consumed.³⁶ The breads were consumed
125 fresh; the baguettes had been produced a maximum of 10 hours earlier, and the samples (5 g)
126 were prepared a maximum of 15 min before the tests.

127

128 **Subjects**

129 Five subjects (two female and three male, aged 32 to 50 years, coded A to E) with good dental
130 status participated in the study. They were selected to represent different chewing
131 efficiencies.³⁶ Subjects gave their written informed consent after receiving a full explanation
132 of the goals and schedule of the study. This study was approved by the local Ethic Committee
133 and by ANSM (ID RCB: 2013-A01084-41).

134 Subjects were characterised for their salivary flow rates and amylase concentration in
135 saliva. Salivary flow rates were determined as described elsewhere,²⁴ over collection periods
136 of 10 min for saliva at rest and 1 min for mechanically stimulated saliva. Alpha-amylase
137 levels were determined in the saliva collected at rest using a commercial kit from Biolabo
138 (ref: 99523/99123, Maizy, France). Group averages for unstimulated and stimulated salivary
139 flow rates were found to be 0.43 ± 0.15 and 2.14 ± 0.29 mL min⁻¹, respectively, and average
140 amylase concentration was $10.1 (\pm 1.4) \times 10^3$ IU L⁻¹. No significant differences were observed
141 between subjects for any of these three parameters.

142

143 **Experimental procedure**

144 Mastication, salivation, bolus heterogeneity and salt release in the oral cavity were followed
145 during bread consumption. All of these measurements were made in independent sessions.
146 The subjects took part in eight sessions of one-hour duration each. They had eaten 2 hours
147 before the sessions. During the experiment, they were seated comfortably in an air-
148 conditioned room (21 ± 1 °C). Sessions 1 and 2 were devoted to the measurement of the
149 activity of the chewing muscles. Subjects were asked to consume bread samples (5 g) in their
150 usual manner, and chewing activity was measured throughout the chewing sequence
151 (including swallowing) by surface electromyography (EMG). Sessions 3 through 5 were
152 devoted to the collection of bread boluses for the study of saliva uptake and bolus
153 heterogeneity. Subjects were asked to chew bread samples, without swallowing, in their usual
154 manner and on a signal of the experimenter to spit out the bread boluses into a container (lid
155 of a glass Petri dish). Boluses were collected at three different stages: 1) at 10 masticatory
156 cycles, 2) at 20 cycles and 3) after complete mastication, when the subject felt the need to
157 swallow (SW). For all of the experiments, the number of cycles and chewing time that
158 induced swallowing (SW) was recorded by the researcher. Initially, boluses were also
159 collected at 30 cycles, but as this number exceeded the number of chews for swallowing in
160 some cases, there were some missing data for this chewing period.³⁶ Therefore, data collected
161 at 30 cycles are not presented in this paper.

162 Subjects were trained in the procedure at the beginning of session 3 with a few samples.
163 Boluses collection started once the subjects felt comfortable with the protocol. Boluses were
164 collected at different stages for given bread before the next bread was offered. With a
165 particular bread type, the subjects always started with the SW bolus, followed by other
166 chewing durations in a random order. At the end of session 5, 180 bread boluses (5 subjects \times
167 4 breads \times 3 replicates \times 3 chewing stages) were obtained.

168 Sessions 6 through 8 were devoted to saliva collection for further sodium analysis. The
169 general procedure was similar to the procedure presented for bolus collection. The
170 instructions given to subjects were to move the bolus to one side of the mouth and spit out a
171 sample of saliva (approximately 0.5 mL) into a 1.5 mL Eppendorf® tube. At the end of
172 session 8, 180 saliva samples were collected (5 subjects \times 4 breads \times 3 chewing stages \times 3
173 replicates). A saliva sample was also collected before eating each sample (stage 0 masticatory
174 cycle = blank). Over the eight individual sessions and the five subjects, the four breads were
175 tested in random order. The subjects cleansed their palates with Evian® mineral water and
176 ensured that their mouths were completely clear of any bread particles before starting the next
177 sample. An interval of 1 minute was taken between the chewing of different samples.

178 **Chewing activity recorded by Electromyography.** The activity of masticatory muscles
179 (superficial masseter and right and left anterior temporalis) was monitored during natural
180 bread consumption.³⁷ From the electromyographic signal collected, eight variables were
181 analysed.³⁸ The chewing sequence was characterised by (a) chewing time (CT in s), (b)
182 number of bursts (i.e., masticatory cycles, BN) and (c) chewing rate (CR). Moreover, muscle
183 activity was characterised by (d) mean and (e) maximum voltage of bursts (V_m and V_{max} ,
184 respectively, in mV), (f) burst duration (BD in s), (g) total EMG activity (sum of the
185 integrated areas of all individual masticatory cycles of the sequence; W_{tot} , expressed in mV
186 s) and (h) mean EMG activity per cycle (W_c , mV s). To study chewing process dynamics, the
187 chewing sequence was also divided into chewing periods of 10 cycles; for each one, the
188 average EMG activity (W_c) was also calculated. Four replicates (two per session) were
189 performed for each bread.

190 **Heterogeneity of bread boluses.** Heterogeneity was studied using image texture analysis as
191 described in a previous study.³⁶ The Petri dish was closed immediately after bolus collection
192 and four images of each bolus were acquired. These images, acquired in colour (RGB

193 system), were first converted into YCbCr images. Only the channel (plan) associated with Y
194 (luminance) was processed for texture analysis. Images were analysed using the Grey level
195 co-occurrence matrix (GLCM) method.³⁹ The co-occurrence matrix describes the second-
196 order statistics in the images and allows for the calculation of textural features that are
197 expected to represent the textural characteristics of the image studied. For a given image, we
198 extracted 5 textural features. Among these textural features, contrast feature represents the
199 heterogeneity of the image and was found to be a suitable feature to characterise bread
200 degradation during the chewing sequence.³⁶

201 **Water uptake of bread bolus.** The saliva taken up by the breads during oral processing was
202 determined from the water content of the breads and of the boluses. The water content of the
203 boluses was determined directly after image acquisition. The water content corresponded to
204 the mass loss obtained after drying in an oven 24 h at 100 °C. For each subject, each bread
205 and each chewing period, the percentage of saliva incorporated in the bolus was calculated as
206 follows: $\text{Saliva (\%)} = ((W1 / D1) \times D2) - W2$ (with W1: water content of the bolus (%), D1 :
207 dry matter of the bolus (%), D2 : dry matter of the bread (%) and W2 : water content of the
208 bread (%)).

209

210 **Sodium content in saliva**

211 Salt release in the mouth during bread consumption was characterised from the sodium
212 content in saliva. The saliva samples were centrifuged directly after collection at 10,000 g for
213 10 min at 4 °C (2-16K, Sigma –Aldrich, St Quentin Fallavier, France). The supernatants were
214 collected, diluted to 1/25 in MilliQ® water and filtered (pore size: 0.45 µm, C.I.L, Sainte Foy
215 La Grande). The amount of sodium in each sample was determined by high performance
216 liquid chromatography using an ISC3000 Dionex system (Voisins le Bretonneux, France)
217 whose individual components included a GP quaternary pump, an AS50 autosampler and a

218 detector used in conductance mode with a CSRS 300 suppressor. The loop injection was set at
219 20 μL (sample volume). The sodium content was analysed using a Dionex IonPac CS12A and
220 an IonPac CG12A guard column at 25 $^{\circ}\text{C}$. Elution was achieved with 11 mM sulphuric acid at
221 a flow rate of 0.5 mL min^{-1} . System controls and data acquisitions were accomplished using
222 Dionex Chromeleon 6.8 software. Quantifications were performed versus standard sodium
223 solutions ranging from 0 to 3 mM.

224

225 **Data analysis**

226 One-way analysis of variance (ANOVA) was performed on the physical properties of the
227 breads to assess the differences between breads. The variability in EMG parameters between
228 subjects and products were studied using Principal Component Analysis (PCA). Variables
229 collected for the entire chewing process, such as EMG data, number of cycles, saliva uptake
230 of bolus at swallowing and image textural contrast of boluses collected at swallowing, were
231 studied using two-way ANOVA with bread, random subject and the bread \times subject
232 interaction as factors. The dynamic evolution of variables collected at different stages during
233 the chewing process (EMG data, saliva uptake and image contrast) was studied using three-
234 way ANOVA. The factors in the model included subject (random), bread, chewing stage and
235 interactions between these factors. Sodium release curve parameters were defined: slope
236 (initial slope of the curve between 0 and 10 chewing cycles), C_{max} (maximum concentration
237 in sodium) corrected for initial differences in the sodium content between different breads and
238 T_{max} (number of cycles corresponding to C_{max}). These parameters were analysed using 2-
239 way ANOVA (random subjects, breads and interactions). ANOVAs were performed using the
240 GLM procedure in SAS Software (SAS Institute Inc., Cary, NC, USA), and the LSMEANS
241 statement was used for a post-hoc multiple comparison test. Correlation coefficients were
242 calculated between different variables. Partial Least Squares (PLS) regression was used to

243 explain salt release parameters (slope, Cmax and Tmax: Block Y) in terms of oral parameters
244 (burst duration, number of masticatory cycles observed in EMG, saliva and salt experiments,
245 chewing rate, chewing time, mean and maximum amplitudes of muscle contraction, EMG
246 activity per chewing cycle, total EMG activity, saliva uptake in bolus, bolus homogeneity
247 (contrast textural feature, Block X)). As a data pre-treatment, normalisation using the 1/SDEV
248 transform to treat all parameters as having equal potential influence was used. A full cross-
249 validation procedure to determine the maximum number of significant dimensions was
250 applied. The Nonlinear Iterative Partial Least Squares (NIPALS) algorithm was used.
251 Discriminant variable selection was performed using variable importance in the projection
252 (VIP)⁴⁰ with a threshold of 0.8.

253

254 **Results**

255 **Bread characteristics**

256 The breads significantly differed in terms of both composition and physicochemical
257 properties (Table 1).

258

259 <Table 1>

260

261 Rye bread was significantly denser, more humid and harder than other breads. Baguettes
262 were the least dense and the least humid products. Artisan and industrial baguettes were
263 significantly different only in water content and hardness. Salt content was higher in
264 baguettes. Our data were globally consistent with density values for industrial and artisan
265 French baguettes and sodium contents in breads reported elsewhere.^{41, 42}

266

267 **Chewing activity and bolus properties**

268 **Chewing activity.** EMG data, chewing efficiency and salivary flow rates obtained for each
269 subject and each type of bread were analysed using PCA (Fig. 1).

270

271 <Fig. 1>

272

273 The first two components accounted for 70% of the total variance. Inter-individual
274 differences were more important than differences between products. PC1 was negatively
275 correlated with the mean and maximum amplitudes of the EMG signal, chewing rate and
276 salivary flow rate and positively correlated with the chewing time, burst duration and chewing
277 efficiency on the left and discriminated between the subjects. Subjects E and C chewed breads
278 faster, and their muscle signals were stronger than those of subjects A and B. The latter
279 chewed for longer and with more chews than the former. Subjects were involved in this study
280 on the basis of their differences in chewing efficiency. It is interesting to note that no
281 correlations were observed between chewing efficiency measured by dental polymer
282 breakdown and any of the EMG parameters measured during bread consumption ($p > 0.05$).
283 PC2 was mainly correlated with EMG activities and discriminated between the products for
284 each subject. From the ANOVA results, the breads differed significantly for the mean (V_m :
285 $F_{(3, 12)} = 26.7$, $p < 0.001$) and maximum (V_{max} : $F_{(3, 12)} = 18.3$, $p < 0.0001$) amplitudes, EMG
286 activity per cycle (W_c : $F_{(3, 12)} = 8.6$, $p < 0.001$) and total activity (W_{tot} : $F_{(3, 12)} = 18.81$, $p <$
287 0.001). Baguettes required stronger muscle contractions (work and amplitude) and longer
288 chewing times than the rye bread and toast bread. These parameters were significantly higher
289 for the baguettes than for the two other breads (data not shown). Breads varied also in
290 parameters characterising the chewing sequences. Baguettes were chewed for a significantly
291 greater number of masticatory cycles (BN : $F_{(3, 12)} = 23.9$, $p < 0.0001$) and consequently for a
292 longer period (CT : $F_{(3, 12)} = 18.9$, $p < 0.0001$) than the rye bread and the toast bread. The

293 average numbers of cycles per bread were as follows: 43.7 (average subject values ranged
294 from 36.2 to 51) for the bakery baguette (BB), 42.5 (31.2 - 50.2) for the supermarket baguette
295 (BS), 33.5 (28.2 - 44.7) for the rye bread (RB) and 32.3 (25.5 - 38.0) for the toast bread (TB).
296 No significant differences ($p > 0.05$) were observed between breads for the chewing rate (CR)
297 or for burst duration (BD).

298 Division of the EMG signal into chewing periods enabled the study of the evolution of
299 chewing activity throughout the chewing sequence. EMG activity significantly decreased
300 between chewing periods ($F_{(3, 18.5)} = 7.5$, $p < 0.001$) from 10 cycles to swallowing (Fig. 2).
301 Moreover, the chewing period \times subject and chewing period \times bread interactions were both
302 significant ($p < 0.001$). This suggests that the profile of chewing activity adapted depending
303 both on the subject and the type of bread, as highlighted on Fig. 2. Only subject E and Bread
304 TB are represented as an example on Fig. 2 but the same trends were observed for the others.

305

306 <Fig. 2>

307

308 **Saliva uptake in boluses.** The numbers of masticatory cycles leading to natural swallowing
309 varied between subjects ($F_{(4, 12)} = 8.93$, $p < 0.001$). On average, subjects A, B and C were
310 found to produce significantly more cycles (35.2 ± 4.8 , 32.2 ± 6.9 and 38.6 ± 8.4 ,
311 respectively) than subjects D and E (28.4 ± 7.5 and 26.3 ± 7.5 , respectively). The number of
312 cycles also varied between products. More cycles were necessary to reach swallowing for the
313 baguettes ($F_{(3, 12)} = 10.35$, $p < 0.001$) than the rye bread and the toast bread. No significant
314 subject \times bread interaction was found ($p > 0.05$).

315 The amounts of saliva obtained for the different breads at three stages of the chewing
316 process are presented Fig. 3. The amount of saliva increased with the number of cycles ($F_{(2, 8)}$
317 $= 11.25$, $p < 0.01$) and depended on the type of bread ($F_{(3, 12)} = 19.90$, $p < 0.0001$). Boluses

318 made from rye bread had a significantly higher saliva uptake than the other breads (Fig. 3).
319 Differences between breads were observed at 10 and 20 cycles but were more marked at
320 swallowing, as already observed for other foods.^{37, 43}

321 At swallowing, boluses made from the bakery baguette had higher saliva content than boluses
322 prepared from the supermarket baguette and toast bread.

323

324 <Fig. 3>

325

326 Saliva uptake increased throughout the chewing process as the number of cycles increased.
327 Nevertheless, we found no correlation between saliva uptake at swallowing and the number of
328 cycles required to reach swallowing ($r < 0.2$).

329 Depending on the type of bread studied, the differences between subjects varied (bread \times
330 subject interaction: $F_{(12,24)} = 2.35$, $p < 0.05$). Saliva content in the swallowable bolus prepared
331 from different subjects and different breads are presented Fig. 4. Globally, the amount of
332 saliva taken up by swallowable boluses ranged from 13.6 to 66.6%. Our results are consistent
333 with other studies from the literature studying salivary impregnation/uptake in bread and
334 cereals.^{44, 45}

335

336 <Fig. 4>

337

338 For the rye bread, subjects A, C and E produced significantly more saliva (up to 50%
339 more) than subjects B and D. Moreover, subject A produced significantly more saliva than 1)
340 subject C for the supermarket baguette and 2) all of the other subjects for the toast bread.
341 Looking at bread differences at the individual level seems to suggest two types of salivary
342 behaviours. Subjects A, C and E adapted their salivary production to the type of bread,

343 whereas the amount of saliva produced did not vary significantly between breads for subjects
344 B and D.

345

346 **Bolus homogeneity.** The image texture heterogeneity of each bolus collected after a fixed
347 number of cycles and at the swallowing threshold were determined using image texture
348 analysis.³⁶ During the chewing process, the breads were transformed into a bolus that lost its
349 heterogeneity (decrease in the contrast values) as the number of chewing cycles increased
350 ($F_{\text{cycle}(2,8)} = 101.9, p < 0.0001$). In our previous study, the analysis of contrast values had
351 revealed specific patterns of bread degradation between breads and between subjects.³⁶ TB
352 and RB reached a homogeneity suitable for swallowing more rapidly than the baguettes.
353 Boluses prepared from different breads had different heterogeneity depending on the subject
354 considered (subject \times bread interaction ($F_{(12,24)} = 6.91, p < 0.0001$)). Differences between
355 subjects were further studied for each individual bread and are presented Table 2.

356

357 <Table 2>

358

359 Contrast values varied greatly between subjects for the baguettes. Boluses collected from
360 subject E were significantly more heterogeneous (higher contrast values) than boluses of other
361 subjects. Inversely, Subject A (and C and D for the BS bread) produced the most
362 homogeneous boluses. These data can be partly explained by individual chewing parameters.
363 Indeed subject E had the lowest chewing efficiency and used fewer cycles to form a
364 swallowable bolus. These chewing parameters may partly explain differences observed
365 between subjects for the RB bread, too. Indeed, the most heterogeneous boluses were
366 produced by the subject (subject D) who applied the smallest number of cycles and who also
367 presented a relatively low chewing efficiency. In the case of this bread, we might also

368 suppose that the higher saliva uptake previously observed for subject E may have helped in
369 preparing a more homogeneous bolus for this subject (as compared to subject D).
370 Nevertheless, differences between subjects in chewing parameters and saliva uptake
371 parameters did not explain all of the contrast results. Indeed, in the case of TB, no differences
372 in contrast values were observed between boluses collected from different subjects despite the
373 fact that subjects varied in mastication and salivation.

374

375 **Sodium content in saliva**

376 The numbers of masticatory cycles leading to natural swallowing varied between breads
377 (Table 3) and between subjects (Table 4). As observed previously, the baguettes required
378 more cycles to form a swallowable bolus than the rye bread and the toast bread (Table 4). The
379 number of cycles applied to reach the swallowing stage was significantly smaller for subject
380 E. Sodium content in saliva varied between breads (Table 3). A higher sodium concentration
381 was observed for toast bread and rye bread than for the baguettes, and the higher
382 concentration was also reached faster (T_{max}) in those products. These effects were not
383 subject-dependent (bread \times subject interaction: $p > 0.05$).

384

385 <Table 3>

386

387 Large inter-individual differences were observed for the three parameters studied,
388 suggesting different dynamics of the sodium release profile between subjects. Subject A
389 released significantly more sodium than subjects B and C. Subject E had a faster release at the
390 beginning of chewing (slope) and at the maximum concentration (T_{max}).

391

392 <Table 4>

393

394 To gain deeper insight into the mechanisms influencing sodium release, a PLS regression
395 was performed to explain the release parameters in terms of the subjects' chewing behaviour,
396 saliva uptake in the bolus and bolus heterogeneity (Figure 5).

397

398 <Fig. 5>

399

400 On the PLS biplot associated with the two first dimensions, 40% of the variability in oral
401 parameters explained 65% of the variability in sodium release parameters. The first axis
402 shows Tmax on the left-hand side and Cmax on the right-hand side. These variables were
403 well-explained by VIP. The slope variable was not explained as well by VIP and was
404 separated on the projection along the second axis. From the beta-weight coefficients, Tmax
405 was mainly explained by the number of masticatory cycles observed in all experiments
406 (EMG, saliva and salt release) and the chewing time. The higher the number of cycles applied
407 to the bread, the later the maximum salt concentration was released. Cmax was related to low
408 cycles and high chewing rate. For the slope parameters, the EMG parameters Wtot, Wc and
409 Vmax were the most important variables, as determined by beta-weight coefficients. The
410 saliva uptake in bolus and bolus heterogeneity did not explain salt release parameters.

411

412 **Discussion**

413 **Methodological considerations**

414 Mastication, bolus properties (saliva uptake and image textural heterogeneity) and salt release
415 were investigated in 3 independent studies dealing with the same subjects and the same
416 products. We chose this set-up to avoid any potential interference between bolus collection
417 and the natural chewing behaviour of subjects and between saliva collection and the structural

418 properties of collected food boluses. For all of the experiments, subjects were introduced to
419 the protocol via a short training on a few products at the beginning of the first session. They
420 were not intensively trained because we did not want to induce a stereotyped chewing
421 behaviour. Unexpectedly, the number of masticatory cycles required for swallowing was
422 significantly higher in EMG experiments than in other experiments (data not shown). This
423 result may be related to the use of surface electrodes in EMG experiments, which unavoidably
424 attracts the subjects' attention to mastication and may result in emphasis of their chewing
425 behaviour. Despite this difference in chewing parameters, products were discriminated in the
426 same way (baguettes required more cycles than the other products) in all experiments.
427 Differences between subjects were also globally similar; subject A is always classified with
428 the subjects producing the most cycles, and subject E systematically produced fewer cycles
429 than subjects A, B and C. Therefore, because the numbers of cycles were different but the
430 conclusions in term of product and subject differences were in agreement, we concluded that
431 the data from the different experiments could be compared together.

432 The difficulty in exactly quantifying the salt released in saliva when chewing bread also
433 bears mentioning. In this study, we determined salt release from the concentration in sodium
434 measured in saliva swabs collected at different stages during the chewing process. However,
435 saliva naturally contains sodium in low concentrations at rest but with a high inter-individual
436 variability (i.e. 11.5 to 217.3 mmol L⁻¹ according to Kallapur et al.⁴⁶). We quantified intrinsic
437 sodium concentrations from saliva samples collected at rest (data not shown). These
438 concentrations were rather small compared to those obtained while chewing breads. In the
439 literature, other authors have suggested that intrinsic sodium concentration in saliva increases
440 when the salivary flow rate increases.⁴⁷ Sodium content in stimulated saliva by chewing
441 parafilm on forty eight subjects has been shown to be on average four times higher than in
442 resting saliva.⁴⁸ Because chewing real food is known to stimulate salivary flow rate, it is

443 actually difficult to quantify the exact contribution of intrinsic sodium and sodium originated
444 from the bread. Whatever the saliva sampling technique, it seems difficult at the moment to
445 be able to access to the exact sodium released from food product in a natural chewing context.

446

447 **In-mouth processing: variability between breads and between subjects**

448 **Evolution of mastication, salivation and bolus homogeneity during the chewing period.**

449 The effect of chewing cycles was observed for EMG activity, saliva uptake in boluses and
450 bolus heterogeneity. A decrease in chewing muscle activity during the chewing sequence has
451 been reported several times for different products^{43, 49, 50} and is explained by an adaptation of
452 chewing behaviour to the changes in food structure during the bolus formation process. As
453 expected, saliva uptake in the bolus increased during the chewing process as a result of
454 continuous saliva production⁵¹. We observed different saliva uptake in boluses collected at
455 swallowing from the different breads. This result is rather in contradiction with another study
456 reporting no significant differences in bolus water content between the three bread types
457 despite their difference in structure and composition.³⁰ This result can be explained by the
458 greater difference between the structure of baguettes, toast breads and rye breads used in our
459 study. No correlation was found between the amount of saliva in the bolus at swallowing and
460 the number of masticatory cycles leading to swallowing. This observation is consistent with
461 other studies, according to which the number of chewing cycles until swallowing and salivary
462 flow rate were independent.^{45, 52} This suggests that salivation also depends on other
463 parameters, such as product properties. In particular, the perception of texture can influence
464 salivary flow rate,^{53, 54} but in the case of bread, different crumb textures were reported to have
465 no influence on salivary flow rate during chewing.²⁹ In parallel to the dynamic evolution of
466 mastication and salivation, breads were transformed into boluses that increase in homogeneity
467 as the number of chewing cycles increased. This result is in agreement with recent data

468 showing a continuous reduction of bread into many small particles throughout the chewing
469 process.³¹

470 **Food oral processing of different breads.** Chewing behaviour was adapted to the textural
471 properties of the food. In the literature, it has been shown that dry and hard products usually
472 require more chewing cycles to be broken down and to capture saliva before swallowing^{52, 55}
473 and that hardness is a key parameter influencing chewing muscle activities measured by
474 EMG.⁵⁶ In our study, toast bread was significantly less hard; it was thus quickly broken down
475 and required less than 30 cycles to be swallowed. Toast bread also contains fat, which may
476 have helped in-mouth breakdown. Instrumental hardness cannot account for differences
477 observed in in-mouth processes for the other breads. The baguettes are slightly harder, less
478 dense and less humid but need significantly more cycles, muscle activity and chewing time to
479 be swallowed. We believe it can be explained by the presence of a thick crust, which provides
480 resistance to biting the baguettes. The rye bread was the hardest and densest product but
481 required similar chewing behaviour (EMG) as toast bread. We argue that it is linked to its
482 specific structure (rye grain agglomerates), which most likely easily separate under chewing,
483 higher humidity and higher saliva uptake. It is important to note that the higher saliva
484 production of rye bread could also be related to the taste of the bread. Indeed, during the
485 experiment some subjects reported that this bread had a sour taste. We can therefore
486 hypothesise that the greater sourness might also have stimulated saliva production during
487 eating.⁵⁷

488 **Food oral processing in different subjects.** Variation between subjects was large in term of
489 mastication and salivation. The subject's characterisation parameters did not predict
490 individual mastication and salivation behaviours in a real (food product) eating context. The
491 huge differences in chewing efficiencies (i.e., ability to break down a silicone rubber into
492 particles in a given number of masticatory cycles³⁶) between subjects was not related to

493 individual EMG activity or number of chewing cycles required to form a swallowable bolus
494 from breads. Similarly, subjects did not differ in their initial salivary flow rate collected at rest
495 and under mechanical (Parafilm®) stimulation but produced bread boluses with different
496 saliva content. Finally, individual parameters, such as chewing efficiency or numbers of
497 cycles required to reach swallowing, could explain the inter-individual differences in bolus
498 homogeneity (contrast textural feature of images), but only for a few breads.

499 Interestingly, at the end of mastication, we did not find similar saliva content and similar
500 homogeneity for boluses collected from different subjects. In hard and brittle products
501 (carrots, nuts,...) a narrow inter-individual variability in particle size distribution has been
502 observed.⁵⁸⁻⁶⁰ Recent studies suggest that the swallowing threshold of cereal products may be
503 more multi-components.^{45, 61} A further characterisation of the physical properties of boluses,
504 using for example the methods recently proposed,³¹ seems necessary to improve our
505 understanding of the mechanisms controlling bolus formation and swallowing in breads.

506

507 **In-mouth salt release from different breads**

508 **Role of bread characteristics on salt release.** More salt is released from toast bread and rye
509 bread than from the baguettes, and two main reasons should be considered.

510 First, breads varied in their composition (wheat vs. rye, presence of fat and sugars, etc),
511 which induced different chemical properties in the breads. We could suppose different
512 interactions between salt sodium ions and bread components, such as proteins. Nevertheless,
513 sodium-protein interactions have been reported to be sufficiently weak in breads to allow
514 complete extraction during in-mouth processing.²⁸

515 Second, breads also varied in their physical properties. Among the compositional factors
516 affecting in vivo sodium release, water content has been the most frequently cited in the case
517 of cheese products with a 50% humidity content^{25, 26} and in the case of sausages.⁶² Similarly,

518 we observed a higher level of release with a higher water content (Table 1) in the case of drier
519 products, such as breads. The kinetics of sodium release may be related to the mobility of
520 sodium within a food product.⁶³ In the case of cheeses, increasing the water content was
521 found to increase sodium mobility within the product (relaxation time measured by NMR)⁶⁴
522 and the release of NaCl from the product to an aqueous phase, observed in vitro.⁶⁵ The lower
523 sodium release observed in baguettes could be linked to physical properties of the crust. More
524 than half of a baguette's weight is composed of a dry and crisp eggshell crust. Because of
525 higher dryness, salt may be more concentrated in the crust than in the crumb. The crust is
526 more difficult to break down and to impregnate with saliva, leading to a smaller salt
527 extraction.

528 In this study, it is actually difficult to further explain how bread factors control temporal
529 sodium release. The development of model breads with a controlled formulation and process
530 seems necessary to further understand their relative contributions.

531 **Role of food oral processing on salt release.** Salt release parameters were mainly explained
532 by chewing parameters. The maximum sodium concentration was reached later when subjects
533 applied a large number of chews and a long chewing duration, as observed in others studies.⁶⁶
534 Rapid initial sodium release was linked to high EMG activity and signal amplitude. This is in
535 agreement with a study reporting that rapid sodium release is linked to high bite force in
536 model cheeses.²⁵ The effect of chewing activity on the rate of sodium release could be
537 explained by a greater breakdown of the product due to the application of stronger mechanical
538 forces between the teeth and more chews. However, this only explains the beginning of
539 release as the maximum concentration of release was related to short chewing time only. We
540 did not observe any influence of the type of bread on the sodium release velocity despite the
541 differences in bread structure and composition, even though significantly faster sodium
542 release has been reported in another study for the more coarse-pored breads.²⁹ These authors

543 reported that this faster release mainly occurred during the beginning of chewing that is
544 crucial for saltiness perception. According to literature, sodium release from model cheeses
545 was partly related to saliva parameters, but conflicting results were observed. High salivary
546 flow rates were linked to a high level of sodium release in one study²⁵ but low levels in other
547 studies.^{24, 66} In our study, saliva uptake in the bolus was found to vary between 15 and 65%
548 between subjects and breads overall, but was not an important parameter for explaining salt
549 release from breads.

550 Saliva enzymatic composition needs to be mentioned as a parameter affecting food
551 breakdown and salt release. Indeed, several studies have shown that salivary alpha-amylase
552 has an impact on bread digestion.^{44, 67, 68} In an in vivo study, alpha-amylase was able to
553 hydrolyse 50% of bread starch during bread mastication for chewing periods shorter or
554 similar to those observed during our study.⁴⁴ In our study, alpha-amylase concentration did
555 not explain inter-individual differences, as similar concentrations were observed between
556 subjects for saliva collected at rest. Nevertheless, it is possible that salivary alpha-amylase
557 accounted for differences observed between breads. Indeed, enzymatic degradation of starch
558 may have been different depending on the chemical and physical properties of the bread,
559 leading to variation in salt release in the saliva. This result would be in line with other studies
560 that showed that in-mouth alpha-amylase activity affected flavour release and saltiness
561 perception.^{14, 69} Further experiments using more subjects and well-designed model bread
562 composition are required to better understand the link between food oral processing including
563 salivary composition, bread properties and salt release in cereal products.

564

565 **Conclusion**

566 Mastication and salivation are two complementary oral mechanisms that lead to food bolus
567 formation. During this study, these mechanisms were adapted to the composition and

568 structural properties of the breads. Marked inter-individual differences were observed in oral
569 parameters, leading to differences in bolus formation. Salt release in saliva was mainly
570 explained by mastication parameters. Higher chewing muscle activity induced a rapid initial
571 rate of sodium release, and longer chewing time induced later sodium release.

572

573 **Acknowledgement**

574 We would like to thank Christine Achilleos (INRA URTAL, Poligny, France), H el ene
575 Brignot and Gilles Feron (INRA CSGA, Dijon, France) for their help with the rheological
576 analyses, alpha-amylase determination and assistance with PLS analysis, respectively.

577

578 **References**

- 579 1 H. G. Coleman, C. M. Kitahara, L. J. Murray, K. W. Dodd, A. Black, R. Z. Stolzenberg-
580 Solomon and M. M. Cantwell, *Am. J. Epidemiol.*, 2014, **179**, 75-84.
- 581 2 S. M. Liu, W. C. Willett, M. J. Stampfer, F. B. Hu, M. Franz, L. Sampson, C. H.
582 Hennekens and J. E. Manson, *Am. J. Clin. Nut.*, 2000, **71**, 1455-1461.
- 583 3 D. Yu, X.-O. Shu, H. Li, Y.-B. Xiang, G. Yang, Y.-T. Gao, W. Zheng and X. Zhang, *Am.*
584 *J. Epidemiol.*, 2013, **178**, 1542-1549.
- 585 4 S. W. Rizkalla, M. Laromiguiere, M. Champ, F. Bruzzo, J. Boillot and G. Slama, *Eur. J.*
586 *Clin. Nut.*, 2007, **61**, 175-183.
- 587 5 C. Breen, M. Ryan, M. J. Gibney, M. Corrigan and D. O'Shea, *Diabetes Educator*, 2013,
588 **39**, 376-386.
- 589 6 P. M. Burton, J. A. Monro, L. Alvarez and E. Gallagher, *Crit. Rev. Food Sci.*, 2011, **51**,
590 965-982.
- 591 7 F. Scazzina, S. Siebenhandl-Ehn and N. Pellegrini, *Brit. J. Nut.*, 2013, **109**, 1163-1174.
- 592 8 M. G. Scanlon and M. C. Zghal, *Food Res. Int.*, 2001, **34**, 841-864.
- 593 9 M. Shibata, M. Tsuta, J. Sugiyama, K. Fujita, M. Kokawa, T. Araki and H. Nabetani, *Int.*
594 *J. Food Eng.*, 2013, **9**, 115-119.
- 595 10 M. C. Zghal, M. G. Scanlon and H. D. Sapirstein, *J. Cereal Sci.*, 2002, **36**, 167-176.
- 596 11 C. M. Courtin and J. A. Delcour, *J. Cereal Sci.*, 2002, **35**, 225-243.

- 597 12 H. Goesaert, K. Brijs, W. S. Veraverbeke, C. M. Courtin, K. Gebruers and J. A. Delcour,
598 *Trends Food Sci. Tech.*, 2005, **16**, 12-30.
- 599 13 C. Salles, M. C. Chagnon, G. Feron, E. Guichard, H. Laboure, M. Morzel, E. Semon, A.
600 Tarrega and C. Yven, *Crit. Rev. Food Sci.*, 2011, **51**, 67-90.
- 601 14 A. L. S. Ferry, J. R. Mitchell, J. Hort, S. E. Hill, A. J. Taylor, S. Lagarrigue and B.
602 Valles-Pamies, *J. Agric. Food Chem.*, 2006, **54**, 8869-8873.
- 603 15 S. P. Cauvain, in *Reducing salt in foods: practical strategies*, eds. D. Kilcast and F.
604 Angus, Woodhead Publishing Limited, Cambridge, UK, 2007, pp. 283-295.
- 605 16 J. Quilez and J. Salas-Salvado, *Nutr. Rev.*, 2012, **70**, 666-678.
- 606 17 T. Ukai, Y. Matsumura and R. Urade, *J. Agric. Food Chem.*, 2008, **56**, 1122-1130.
- 607 18 T. H. McCann and L. Day, *J. Cereal Sci.*, 2013, **57**, 444-452.
- 608 19 A. Braschi, L. Gill and D. J. Naismith, *Int. J. Food Sci.*, 2009, **60**, 507-521.
- 609 20 A. K. F. Ignacio, J. T. d. D. Rodrigues, P. Y. Niizu, Y. Chang and C. J. Steel, *Braz. J.*
610 *Food Technol.*, 2013, **16**, 1-11.
- 611 21 L. A. Jimenez-Maroto, T. Sato and S. A. Rankin, *J. Cereal Sci.*, 2013, **58**, 313-317.
- 612 22 S. Girgis, B. Neal, J. Prescott, J. Prendergast, S. Dumbrell, C. Turner and M. Woodward,
613 *Eur. J. Clin. Nut.*, 2003, **57**, 616-620.
- 614 23 K. Konitzer, T. Pflaum, P. Oliveira, E. Arendt, P. Koehler and T. Hofmann, *J. Agric.*
615 *Food Chem.*, 2013, **61**, 10659-10669.
- 616 24 G. Lawrence, C. Septier, C. Achilleos, P. Courcoux and C. Salles, *J. Agric. Food Chem.*,
617 2012, **60**, 5299-5306.
- 618 25 V. A. Phan, C. Yven, G. Lawrence, C. Chabanet, J. M. Reparet and C. Salles, *Int. Dairy*
619 *J.*, 2008, **18**, 956-963.
- 620 26 G. Lawrence, S. Buchin, C. Achilleos, F. Bérodiér, C. Septier, P. Courcoux and C. Salles,
621 *J. Agric. Food Chem.*, 2012, **60**, 5287-5298.
- 622 27 C. Yven, J. Patarin, A. Magnin, H. Laboure, M. Repoux, E. Guichard and G. Feron, *J.*
623 *Texture Stud.*, 2012, **43**, 309-318.
- 624 28 T. Pflaum, K. Konitzer, T. Hofmann and P. Koehler, *J. Agric. Food Chem.*, 2013, **61**,
625 6485-6494.
- 626 29 T. Pflaum, K. Konitzer, T. Hofmann and P. Koehler, *J. Agric. Food Chem.*, 2013, **61**,
627 10649-10658.
- 628 30 M. Panouillé, A. Saint-Eve, I. Deleris, F. Le Bleis and I. Souchon, *Food Res. Int.*, 2014,
629 DOI: 10.1016/j.foodres.2014.02.031, accepted.

- 630 31 F. Le Bleis, L. Chaunier, G. Della Valle, M. Panouille and A. L. Reguerre, *Food Res.*
631 *Int.*, 2013, **50**, 308-317.
- 632 32 P. Baardseth, K. Kvaal, P. Lea, M. R. Ellekjaer and E. M. Faergestad, *J. Cereal Sci.*,
633 2000, **32**, 73-87.
- 634 33 P. Roussel and H. Chiron, *Les pains français - Evolution, qualité, production*, MAE-
635 ERTI, Conflandey, France, 2nd edn., 2005.
- 636 34 AACC, in *Approved methods of the American Association of Cereal Chemists*, St. Paul,
637 MN, 2000, 10th edn., p. 349.
- 638 35 L. Chaunier, H. Chiron, G. Della Valle and L. Saulnier, *Industries des céréales*, 2008,
639 **160**, 2-8.
- 640 36 C. Tournier, M. Grass, D. Zope, C. Salles and D. Bertrand, *J. Food Eng.*, 2012, **113**, 615-
641 622.
- 642 37 L. Mioche, P. Bourdiol, S. Monier and J. F. Martin, *Food Qual. Prefer.*, 2002, **13**, 583-
643 588.
- 644 38 K. Kohyama, L. Mioche and J. F. Martin, *J. Texture Stud.*, 2002, **33**, 269-283.
- 645 39 R. M. Haralick, K. Shanmugam and I. Dinstein, *IEEE Trans. Syst. Man Cybern.*, 1973, **3**,
646 610-621.
- 647 40 I. G. Chong and C. H. Jun, *Chemometr. Intell. Lab. Syst.*, 2005, **78**.
- 648 41 C. Martin, H. Chiron and S. Issanchou, *J. Food Qual.*, 2013, **36**, 324-333.
- 649 42 Observatoire du pain, 2006. <http://www.observatoiredupain.com>.
- 650 43 A. Tarrega, C. Yven, E. Semon and C. Salles, *Int. Dairy J.*, 2011, **21**, 358-364.
- 651 44 C. Hoebler, A. Karinithi, D. M.F., F. Guillon, D. J. Gallant, B. Bouchet, C. Melegari and
652 J. L. Barry, *Brit. J. Nut.*, 1998, **80**, 429-436.
- 653 45 C. Loret, M. Walter, N. Pineau, M. A. Peyron, C. Hartmann and N. Martin, *Physiol.*
654 *Behav.*, 2011, **104**, 855-864.
- 655 46 B. Kallapur, K. Ramalingam, Bastian, A. Mujib, A. Sarkar and S. Sethuraman, *J. nat.*
656 *sci., biol. Med.*, 2013, **4**, 341-345.
- 657 47 L. H. Schneyer and C.A. Schneyer, in *Handbook of physiology. Sect 6. Alimentary canal*,
658 ed. C.F. Code, Americal Physiological Society, Washington, 1967, vol. 2, pp. 497-530.
- 659 48 G. Feron, C. Ayed, E.M. Qannari, P. Courcoux, H. Labouré and E. Guichard, *Plos One*,
660 2014, 9(4), e93113.
- 661 49 K. Kohyama and L. Mioche, *J. Texture Stud.*, 2004, **35**, 395-414.
- 662 50 C. Yven, L. Bonnet, D. Cormier, S. Monier and L. Mioche, *Eur. J. Oral Sci.*, 2006, **114**,
663 184-190.

- 664 51 J. F. Prinz and P. W. Lucas, *Proc. R. Soc. London, Ser. B*, 1997, **264**, 1715-1721.
- 665 52 M. B. D. Gavião, L. Engelen and A. van der Bilt, *Eur. J. Oral Sci.*, 2004, **112**, 19-24.
- 666 53 D. J. Anderson and M. P. Hector, *J. Dent. Res.*, 1987, **66**, 518-523.
- 667 54 M. P. Hector and R. W. A. Linden, *Q. J. Exp. Physiol.*, 1987, **72**, 285-301.
- 668 55 L. Engelen, A. Fontijn-Tekamp and A. van der Bilt, *Arch. Oral Biol.*, 2005, **50**, 739-746.
- 669 56 A. Woda, K. Foster, A. Mishellany and M. A. Peyron, *Physiol. Behav.*, 2006, **89**, 28-35.
- 670 57 E. Neyraud, C. I. Heinzerling, J. H. F. Bult, C. Mesmin and E. Dransfield, *Chemosens.*
671 *Percep.*, 2009, **2**, 108-116.
- 672 58 M.-L. Jalabert-Malbos, A. Mishellany-Dutour, A. Woda and M.-A. Peyron, *Food Qual.*
673 *Prefer.*, 2007, **18**, 803-812.
- 674 59 A. Mishellany, A. Woda, R. Labas and M. A. Peyron, *Dysphagia*, 2006, **21**, 87-94.
- 675 60 M. A. Peyron, A. Mishellany and A. Woda, *J. Dent. Res.*, 2004, **83**, 578-582.
- 676 61 M. A. Peyron, I. Gierczynski, C. Hartmann, C. Loret, D. Dardevet, N. Martin and A.
677 Woda, *Plos One*, 2011, **6**, e21167.
- 678 62 C. Chabanet, A. Tarrega, C. Septier, F. Siret and C. Salles, *Meat Sci.*, 2013, **94**, 253-261.
- 679 63 L. Boisard, I. Andriot, C. Arnould, C. Achilleos, C. Salles and E. Guichard, *Food Chem.*,
680 2013, **136**, 1070-1077.
- 681 64 M. Gobet, M. Mouaddab, N. Cayot, J.-M. Bonny, E. Guichard, J.-L. Le Quéré, C.
682 Moreau and L. Foucat, *Magn. Reson. Chem.*, 2009, **47**, 307-312.
- 683 65 C. Lauverjat, I. Deleris, C. I. Trelea, C. Salles and I. Souchon, *J. Agric. Food Chem.*,
684 2009, **57**, 9878-9887.
- 685 66 E. Pionnier, C. Chabanet, L. Mioche, A. J. Taylor, J. L. Le Queré and C. Salles, *J. Agric.*
686 *Food Chem.*, 2004, **52**, 565-571.
- 687 67 G. M. Bornhorst and R. P. Singh, *Food Biophys.*, 2013, **8**, 50-59.
- 688 68 C. Hoebler, M. F. Devaux, A. Karinthe, C. Belleville and J. L. Barry, *Int. J. Food Sci.*,
689 2000, **51**, 353-366.
- 690 69 A.-L. Ferry, J. Hort, J. R. Mitchell, S. Lagarrigue and B. V. Pamies, *J. Texture Stud.*,
691 2004, **35**, 511-524.
- 692
- 693
- 694

Figure captions

Fig. 1 Principal Component Analysis biplot of oral parameters obtained for each subject (A-E) and each bread (BB: bakery baguette, BS: supermarket baguette, TB: toast bread, RB: rye bread). Parameters: EMG parameters (burst duration (BD), number of bursts (i.e., masticatory cycle; BN), chewing rate (CR), chewing time (CT), mean and maximum amplitudes (V_m and V_{max} , respectively), EMG activity per chewing cycle (W_c), total EMG activity (W_{tot}), salivary flow rate (SF), and chewing efficiency (CE).

Fig. 2 (a) Evolution of mean EMG activity per cycle during eating of the TB bread by five different subjects (A to E). **(b)** Evolution of mean EMG activity per cycle during eating of four breads (BB, BS, TB, RB) observed for subject E. SW: average number of chews inducing swallowing. Average value \pm standard deviation ($n = 4$).

Fig. 3 Saliva uptake of boluses made from 4 breads (BB, BS, TB and RB) expectorated at three periods during the chewing process: at 10 (\square) cycles, 20 (\boxtimes) cycles and at swallowing point (\blacksquare). a, b, c, d: Mean values with different letters are significantly different ($p < 0.05$).

Fig. 4 Saliva content of boluses collected at swallowing from different subjects (A, B, C, D, E) eating different breads (BB; BS, RB, TB). a, b, c, d, e, f: Mean values with different letters are significantly different ($p < 0.05$).

Figure 5 The Partial Least Square (PLS) regression explaining salt release parameters (initial slope of release between 0 and 10 chews (Slope), maximum concentration in sodium (C_{max}) and number of cycles corresponding to C_{max} (T_{max}); block Y) in terms of oral parameters

(EMG parameters (burst duration (BD), number of masticatory cycles (Chews-EMG), chewing rate (CR), chewing time (CT), mean and maximum amplitudes (V_m and V_{max} , respectively), EMG activity per chewing cycle (W_c), total EMG activity (W_{tot})), number of chews observed in saliva and salt experiments (Chews-saliva and Chews-salt, respectively), contrast textural feature from bolus images at swallowing, and saliva uptake in the bolus at swallowing (Saliva)). The method was applied to all subject \times bread combinations ($n = 20$). **(a)** Correlation plot, **(b)** biplot of observations (subject (A-E) and bread (BB, BS, TB, RB)), plan #1-#2.

Fig. 1

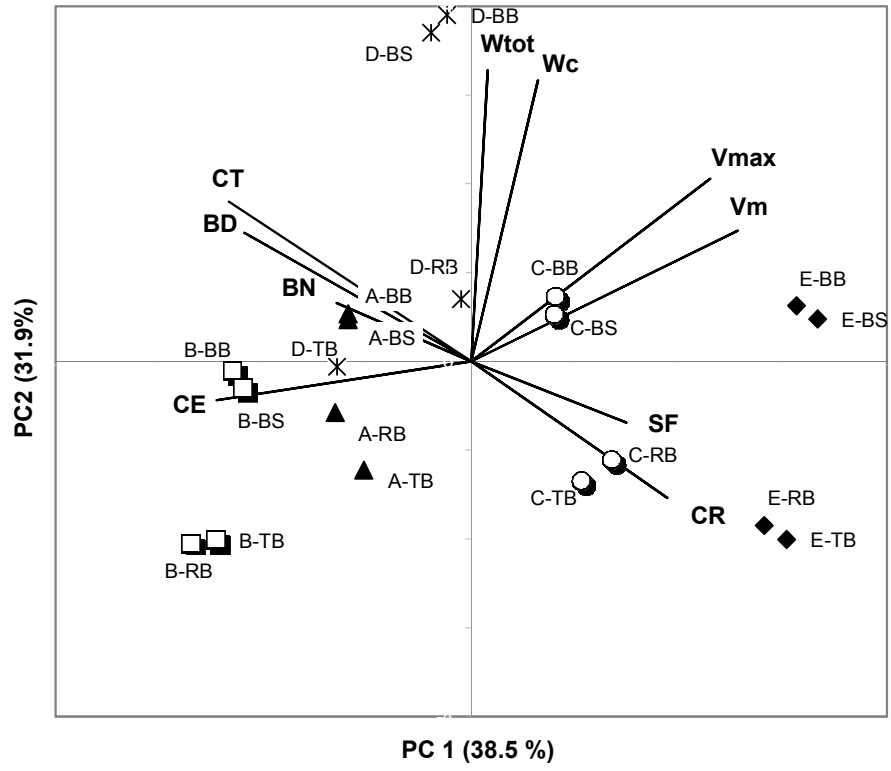


Fig. 2

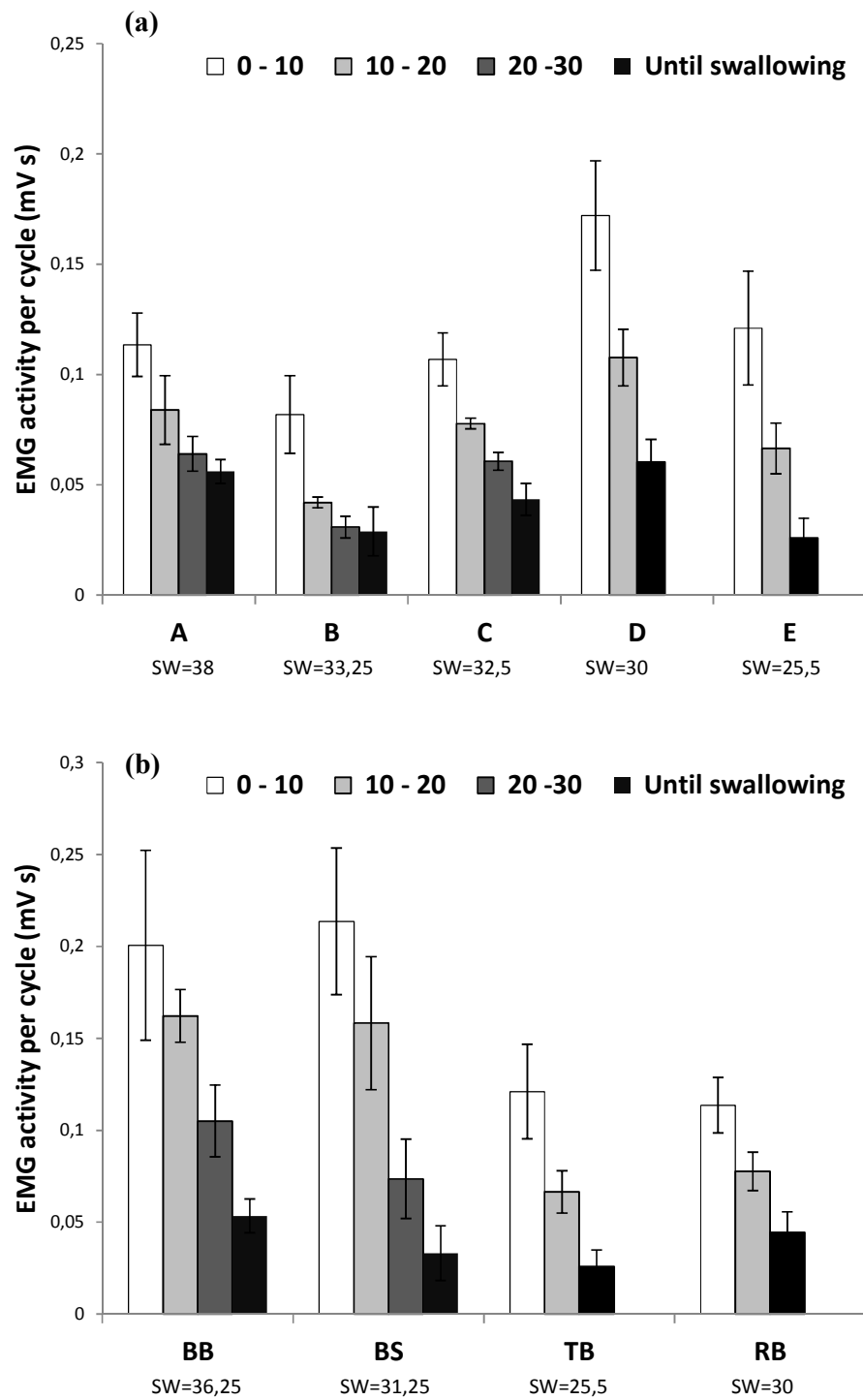


Fig. 3

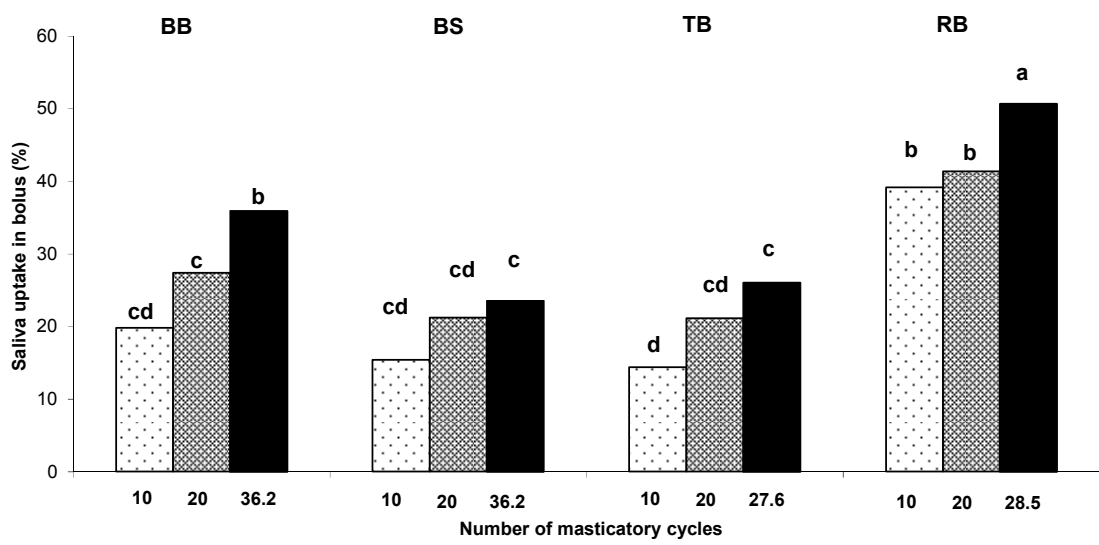


Fig. 4

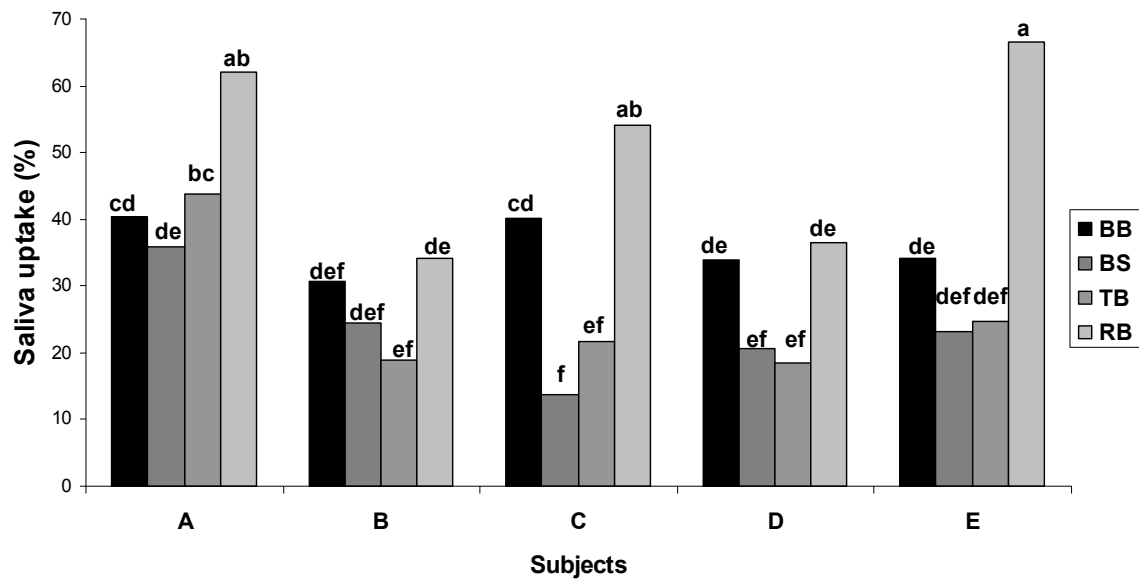


Fig. 5

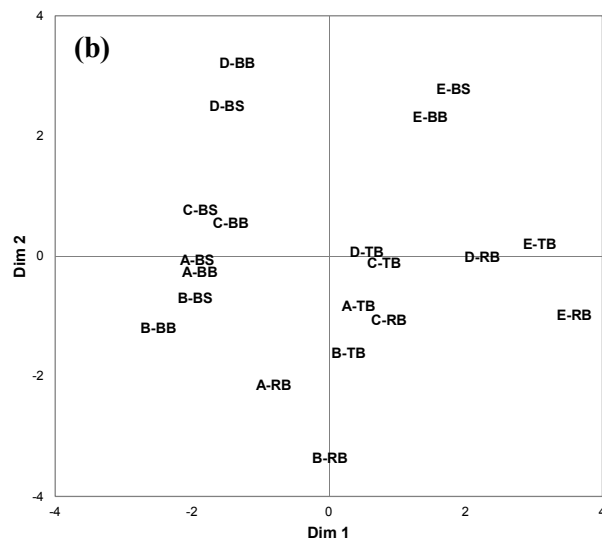
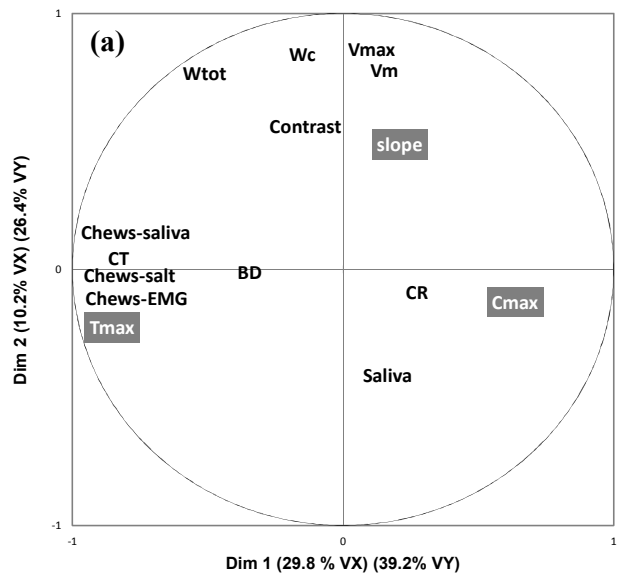


Table 1 Physical properties of the four breads studied. Different letters indicate significant differences between products ($p < 0.05$)

Code	Breads	Supplier	Brand	Density (g / mL)	Water content (g H ₂ O / 100 g total weight)	Approximate crust/crumb weight ratio (%)	Maximum force (N)*	Sodium content (mg / 100 g)
BB	Baguette	Artisan baker	'banette'	0.194 ± 0.01 ^c	23.2 ± 1.1 ^d	69/31	30.8 ± 6.5 ^b	575.3 ± 13.6 ^a
BS	Baguette	Supermarket	standard	0.180 ± 0.02 ^c	29.6 ± 0.9 ^c	68/32	23.7 ± 7.2 ^c	596.6 ± 10.2 ^a
TB	Toast bread	Supermarket	'Jaquet'	0.270 ± 0.03 ^b	31.3 ± 1.4 ^b	24/76	16.4 ± 1.4 ^d	438.6 ± 51.0 ^b
RB	Rye bread	Organic shop	'Pural'	0.602 ± 0.03 ^a	42.1 ± 1.9 ^a	0/100	93.4 ± 3.5 ^a	408.3 ± 41.6 ^b

*maximum force was obtained by uniaxial compression using a TAXT2 texture analyser.

Table 2 Chewing efficiency of subjects and contrast (features from textural image analysis) values of boluses collected at swallowing and the number of masticatory cycles required to form a bolus obtained from different subjects (A, B, C, D, E) eating 4 different breads (BB, BS, TB and RB). Average values \pm standard deviation

		$F(p)^1$	A	B	C	D	E
Contrast ²	BB	13.99***	2.60 \pm 0.07 ^c	2.75 \pm 0.11 ^b	2.71 \pm 0.11 ^b	2.79 \pm 0.07 ^b	2.88 \pm 0.11 ^a
	BS	28.23***	2.69 \pm 0.11 ^c	2.86 \pm 0.15 ^b	2.59 \pm 0.08 ^c	2.63 \pm 0.13 ^c	3.03 \pm 0.11 ^a
	TB	2.27 NS	2.73 \pm 0.05	2.84 \pm 0.19	2.70 \pm 0.09	2.84 \pm 0.22	2.88 \pm 0.25
	RB	6.51***	2.17 \pm 0.06 ^b	2.19 \pm 0.12 ^b	2.16 \pm 0.07 ^b	2.35 \pm 0.012 ^a	2.22 \pm 0.14 ^b
Nb cycle required to form a bolus	BB	52.91***	35.7 \pm 1.8 ^b	40.3 \pm 1.8 ^a	40.4 \pm 0.5 ^a	34.7 \pm 3.4 ^b	30.0 \pm 1.7 ^c
	BS	31.24***	38.3 \pm 3.0 ^b	34.3 \pm 1.3 ^b	46 \pm 8.13 ^a	34.6 \pm 2.1 ^b	27.7 \pm 2.1 ^c
	TB	15.22***	31.3 \pm 2.5 ^a	25.7 \pm 3.2 ^b	32 \pm 4.5 ^a	24.3 \pm 3.4 ^b	24.6 \pm 2.6 ^b
	RB	19.35***	35.6 \pm 6.9 ^a	27.7 \pm 4.0 ^b	36 \pm 8.9 ^a	20 \pm 4.26 ^c	23 \pm 1.5 ^{bc}
Chewing efficiency (%)		9.4***	25.4 \pm 6.9 ^a	20.0 \pm 5.5 ^{ab}	15.6 \pm 4.9 ^{ab}	10.8 \pm 0.7 ^{bc}	2.4 \pm 0.9 ^c

¹ *F* value and associated significance effect (NS non-significant: $p > 0.05$, *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$) obtained from ANOVAs testing the subject effect,

² contrast: textural feature obtained from texture image analysis (a high value represents a high level of heterogeneity). a, b, c: Mean values with different letters are significantly different ($p < 0.05$).

Table 3 Effect of the type of bread (BB, BS, TB, RB) on average (n = 15) temporal sodium release parameters

	$F_{product}, P^a$	BB	BS	TB	RB
Number of chews	9.9**	34.2 ± 5.2 ^a	34.9 ± 6.1 ^a	28.3 ± 5.19 ^b	29.4 ± 5.9 ^b
Cmax (mg 100 g ⁻¹ saliva)	8.7 **	166.1 ± 51.4 ^b	175.3 ± 69.1 ^b	217.8 ± 59.3 ^a	232.5 ± 57.7 ^a
Tmax (no. of chews)	5.7 *	29.7 ± 8.9 ^{ab}	32.8 ± 6.2 ^a	26.2 ± 4.5 ^b	27.7 ± 5.0 ^b
Slope (mg 100 g ⁻¹ saliva s ⁻¹)	1.8 ^{NS}	7.0 ± 3.9	7.2 ± 4.1	6.0 ± 3.0	7.8 ± 4.4

^a F and p values (*: $p < 0.05$, **: $p < 0.01$, NS: $p > 0.05$) obtained from the analysis of two-way ANOVA (random subjects, breads). ^{a, b}: average values (± standard deviation) associated with the same letters are not significantly different ($p = 0.05$)

Table 4 Effect of the subjects (A - E) on average (n = 12) temporal sodium release parameters

	$F_{subjects} P^a$	A	B	C	D	E
Number of chews	15.5**	35.0 ± 4.9 ^a	33.0 ± 4.9 ^a	33.6 ± 5.7 ^a	32.0 ± 4.5 ^a	23.5 ± 2.8 ^b
Cmax (mg 100 g ⁻¹ saliva)	7.2 **	237.3 ± 62.0 ^a	159.4 ± 48.1 ^b	169.7 ± 68.6 ^b	204.1 ± 56.1 ^{ab}	219.2 ± 60.7 ^{ab}
Tmax (no. of chews)	13.8 ***	30.5 ± 4.9 ^a	32.3 ± 4.6 ^a	29.5 ± 5.9 ^a	32.0 ± 5.8 ^a	20.6 ± 4.2 ^b
Slope (mg 100 g ⁻¹ saliva s ⁻¹)	20.5***	7.7 ± 2.2 ^b	6.2 ± 2.8 ^b	3.2 ± 1.2 ^c	6.7 ± 2.9 ^b	11.4 ± 4.3 ^a

^a *F* and *p* values (*: *p* < 0.05, **: *p* < 0.01, NS: *p* > 0.05) obtained from the analysis of two-way ANOVA (random subjects, breads). ^{a, b}: average values (± standard deviation) associated with the same letters are not significantly different (*p* > 0.05)

