



**Excipient Foods: Designing Food Matrices that Improve the Oral Bioavailability of Pharmaceuticals and Nutraceuticals**

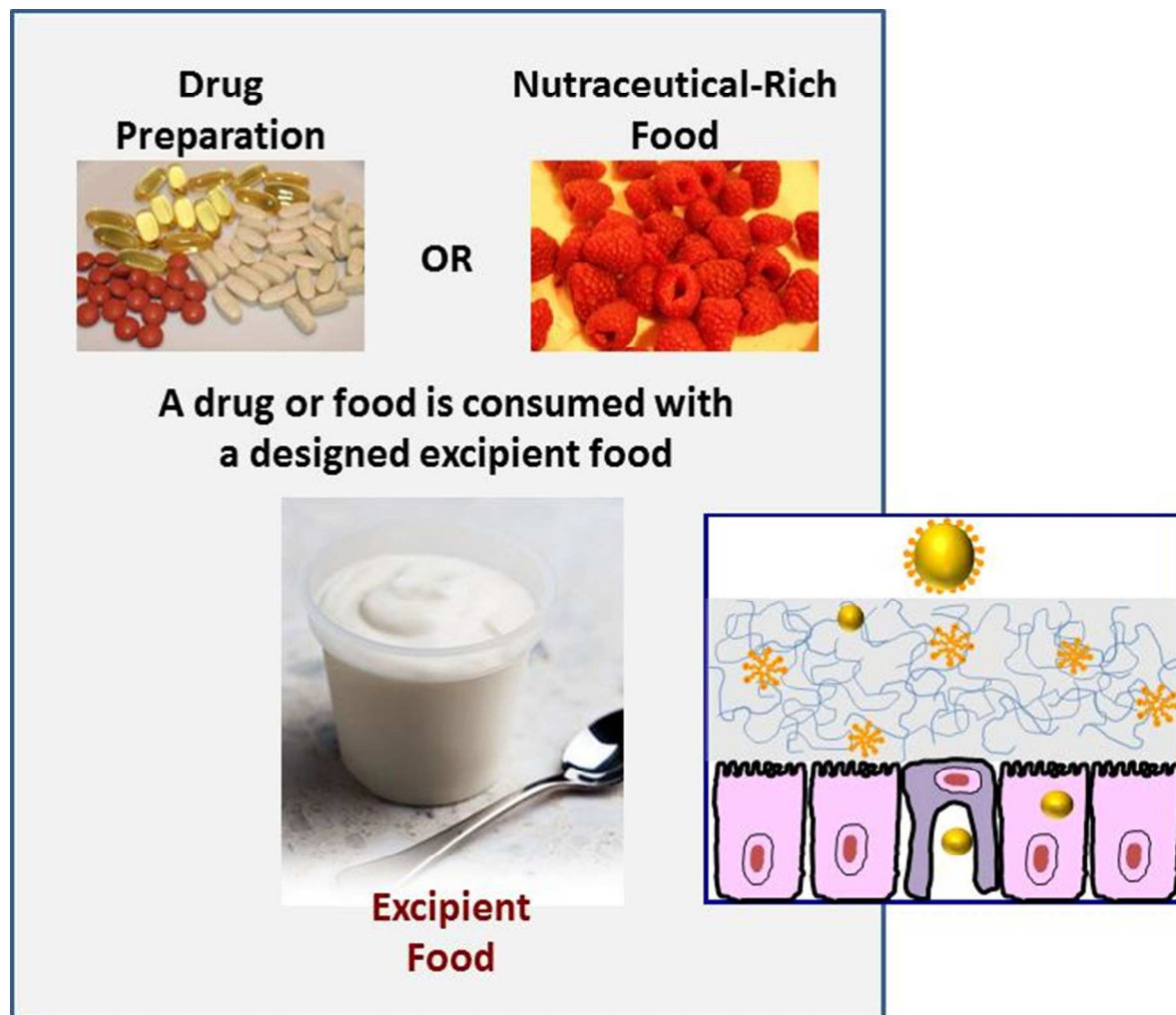
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# Excipient Foods: Designing Food Matrices that Improve the Oral Bioavailability of Pharmaceuticals and Nutraceuticals

David Julian McClements and Hang Xiao

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## Graphical Abstract:



1                   **Excipient Foods: Designing Food Matrices that**  
2                   **Improve the Oral Bioavailability of Pharmaceuticals**  
3                   **and Nutraceuticals**

4  
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21 **Abstract**

22 The oral bioavailability of many lipophilic bioactive agents (pharmaceuticals and  
23 nutraceuticals) is limited due to various physicochemical and physiological processes:  
24 poor release from food or drug matrices; low solubility in gastrointestinal fluids;  
25 metabolism or chemical transformation within the gastrointestinal tract; low epithelium  
26 cell permeability. The bioavailability of these agents can be improved by specifically  
27 designing food matrices that control their release, solubilization, transport, metabolism,  
28 and absorption within the gastrointestinal tract. This article discusses the impact of food  
29 composition and structure on oral bioavailability, and how this knowledge can be used to  
30 design *excipient foods* for improving the oral bioavailability of lipophilic bioactives.  
31 Excipient foods contain ingredients or structures that may have no bioactivity themselves,  
32 but that are able to promote the bioactivity of co-ingested bioactives. These bioactives  
33 may be lipophilic drugs in pharmaceutical preparations (such as capsules, pills, or syrups)  
34 or nutraceuticals present within food matrices (such as natural or processed foods and  
35 beverages).

36

37 *Keywords:* bioactives; lipophilic; nutraceuticals; pharmaceuticals; drugs; excipient  
38 foods; medical foods; functional foods; food effects; delivery systems.

39

## 40 1. Introduction

41 Many bioactive agents present in foods (nutraceuticals) or drugs (pharmaceuticals)  
42 intended for oral ingestion are highly lipophilic molecules with low and/or variable  
43 bioavailability<sup>1-6</sup>. The poor bioavailability characteristics of these bioactive agents may  
44 be the result of a number of physicochemical or physiological processes: restricted  
45 release from the product matrix<sup>7</sup>; low solubility in gastrointestinal fluids<sup>2,8</sup>; low  
46 permeability across intestinal epithelial cells<sup>6,9</sup>; and/or, enzymatic or chemical  
47 transformations within the gastrointestinal tract (GIT)<sup>3,10,11</sup>. Research in the food,  
48 nutrition, and pharmaceutical disciplines has established that the bioavailability of many  
49 bioactive agents depends strongly on the nature of the foods ingested with them<sup>2,3,5,8,</sup>  
50 <sup>12-16</sup>. Both the composition and the structural organization of the food matrix may  
51 influence the bioavailability of co-ingested bioactive agents<sup>17</sup>. The dependence of the  
52 oral bioavailability of lipophilic bioactive agents on food matrix characteristics means  
53 there is considerable opportunity for designing food-based delivery systems to improve  
54 the efficacy of lipophilic pharmaceuticals and nutraceuticals.

## 55 2. Medical, Functional and Excipient Foods

56 There is increasing convergence in the interests of the pharmaceutical and food  
57 industries in the development of products to prevent or treat human diseases (**Figure 1**).  
58 The pharmaceutical industry is developing drug preparations to combat chronic or acute  
59 diseases, whereas the food industry is developing food and beverage products whose  
60 purpose is to promote human health and wellbeing through diet. In particular, there is a  
61 considerable overlap in the development of food-based approaches to improve the  
62 bioavailability of lipophilic bioactive agents, such as nutraceuticals and pharmaceuticals.  
63 These approaches are based on the design of the composition or structure of food  
64 matrices to increase bioavailability and have led to new classes of foods: functional foods;

65 medical foods; and, excipient foods (**Figure 2**).

### 66 **2.1. Functional foods**

67 *A functional food* is fabricated from generally recognized as safe (GRAS) food  
68 ingredients, and typically contains one or more food-grade bioactive agent  
69 (“nutraceuticals”) dispersed within a food matrix. There are already many examples of  
70 functional food products that are commercially available, including milks fortified with  
71 vitamin D, yogurts fortified with probiotics, spreads fortified with phytosterols, and  
72 breakfast cereals fortified with  $\omega$ -3 fatty acids, vitamins, and minerals. A great deal of  
73 research is currently being carried out on identifying other kinds of nutraceuticals, and it  
74 will be important for the food industry to clearly demonstrate their health benefits before  
75 they can be successfully incorporated into functional food products and obtain regulatory  
76 and consumer acceptance.

### 77 **2.2. Medical foods**

78 *A medical food* contains one or more pharmaceutical-grade bioactive agents (drugs)  
79 dispersed within a food matrix. This food matrix may be a traditional food type (such as a  
80 beverage, yogurt, or confectionary) or it may be a nutritional fluid that is fed to a patient  
81 through a tube. A medical food is usually administered to treat a particular disease under  
82 medical supervision. A number of medical foods are commercially available that are  
83 specifically designed to manage or treat various diseases, such as Alzheimer’s, diarrhea,  
84 depression, diabetes, and osteoporosis.

### 85 **2.3. Excipient foods**

86 In this article, a new class of foods designed to improve the bioavailability of orally  
87 administered bioactive agents is introduced: *excipient foods*. An excipient is  
88 conventionally defined as a component that is not bioactive itself but is included in a  
89 pharmaceutical preparation to increase the efficacy of a drug<sup>2, 18-20</sup>. Some commonly

90 used excipients in the pharmaceutical industry include lipids, surfactants, synthetic  
91 polymers, carbohydrates, proteins, cosolvents, and salts. By analogy, an excipient food  
92 may not have any bioactivity itself, but it may increase the efficacy of any nutraceuticals  
93 or pharmaceuticals that are co-ingested with it. Excipient foods are therefore meant to  
94 be consumed with a conventional pharmaceutical dosage form (*e.g.*, capsule, pill, or  
95 syrup), a dietary supplement (*e.g.*, capsule, pill, or syrup), or nutraceutical-rich food (*e.g.*,  
96 fruits, vegetables, nuts, seeds, grains, meat, fish, and some processed foods). It is likely  
97 that different kinds of excipient foods will have to be designed for different types of  
98 bioactive agents or delivery matrices. Some examples of excipient foods that could be  
99 developed to increase the bioavailability of nutraceuticals in foods are shown in **Table 1**.  
100 For example, the bioaccessibility of carotenoids in a salad may be increased by  
101 consuming it with a specifically designed salad dressing. This dressing may contain  
102 various food components that increase the bioavailability of the nutraceuticals in the  
103 salad: lipids that increase intestinal solubility; antioxidants that inhibit chemical  
104 transformations; enzyme inhibitors that retard metabolism; permeation enhancers that  
105 increase absorption; efflux inhibitors. Indeed, previous studies have shown that the  
106 bioavailability of oil-soluble vitamins and carotenoids in salads can be increased by  
107 consuming them with dressings containing some fat <sup>14</sup>, which supports the concept of  
108 excipient foods.

109

110 **Table 1:** Examples of excipient foods that could be designed to improve the  
 111 bioactivity of nutraceuticals in foods.

Nutraceuticals	Food Source	Potential Excipient Foods
Carotenoids	Salad (lettuce, kale, carrot, tomato, peppers ...)	Salad Dressing
Carotenoids	Cooked vegetables (carrot, peppers, spinach, kale...)	Sauce
Carotenoids, Vitamins, Phytosterols/stanols	Nuts and Seeds (almonds, peanuts, sunflower seeds...)	Edible Coatings
Flavonoids, Vitamins	Fruits and Berries (blueberry, strawberry, raspberry, apple, pear..)	Cream, Ice Cream, Yogurt
Phytosterols/stanols	Nuts	Sauce, Edible Coatings
CLA	Meat and dairy products (beef, cheese, ...)	Sauce
$\omega$ -3 Oils	Fish	Sauce

112

113 In principle, a wide variety of different food products could be used as excipients to  
 114 increase the bioactivity of lipophilic bioactives, such as beverages, yogurts, dressings,  
 115 desserts, sauces, soups, dips, spreads, candies, and baked goods. These excipient foods  
 116 need to be selected so that they are economic, convenient, desirable, and effective, and  
 117 that can be regularly incorporated into a daily diet.

118 In the remainder of this review, we initially consider the design of excipient foods,  
 119 then we highlight the main factors limiting the bioavailability of lipophilic bioactive  
 120 components, and then we discuss the impact of food matrix composition and structure on  
 121 bioavailability and how this leads to the concept of excipient foods and excipient food



122 ingredients.

### 123 **3. Design of Excipient Foods**

124 Excipient foods may be fluids, semi-solids, or solids that may be consumed by  
125 drinking (beverages) or eating (foods). A number of different factors must be  
126 considered when designing excipient foods. First, the composition and structure of the  
127 food matrix should be designed to increase the bioavailability of co-ingested bioactive  
128 agents. This depends on knowledge of the influence of specific food components and  
129 structures on the biological fate of the bioactives. Second, the food matrix should be  
130 designed so that the product is desirable to consumers or patients to ensure good  
131 compliance, *e.g.*, the food should have a desirable appearance, texture, mouthfeel, and  
132 flavor<sup>21</sup>. Third, foods or beverages should be chosen so that they can be consumed on a  
133 regular basis with drugs or foods containing nutraceutical agents (such as fruits and  
134 vegetables). This restricts the type of products suitable for use as excipient foods to  
135 those that can easily be incorporated into a daily diet. Fourth, the product should have a  
136 sufficiently long shelf life and not take up too much storage space, since it is impractical  
137 for consumers to purchase a product too frequently. Some potential candidates for  
138 excipient foods that meet most or all of these requirements are fruit drinks, teas, coffees,  
139 dairy beverages, creams, yogurts, margarine, butter, cheese spreads, desserts,  
140 confectionary, and crackers. The nature of the excipient food might depend on the type  
141 of drug or nutraceutical-rich food that is being consumed. For example, an excipient  
142 food suitable for increasing the bioavailability of the nutraceuticals in fruits (such as  
143 apples, pears, blueberries, strawberries, or raspberries) might consist of a specially  
144 designed cream, yogurt, or ice cream. On the other hand, an excipient food suitable for  
145 increasing the bioavailability of nutraceuticals in cooked or raw vegetables (such as  
146 carrots, broccoli, spinach, or kale) might consist of a specially designed pouring sauce or  
147 salad dressing.

### 148 **3.1. Potential benefits of excipient foods**

149 There are several potential benefits of developing excipient foods to increase the  
150 bioavailability of nutraceuticals and drugs. The long-term consumption of low levels of  
151 nutraceuticals may improve human performance, enhance wellbeing, or inhibit the onset  
152 of chronic diseases, such as heart disease, diabetes, hypertension, and cancer<sup>5</sup>. This  
153 would increase the quality of life of the general population and reduce the costs of health  
154 care associated with treatment of these chronic diseases. At present, the bioavailability  
155 of the nutraceuticals in many natural sources, such as fruits and vegetables, is relatively  
156 low, and therefore their potential benefits on long-term human health are not being fully  
157 realized<sup>3</sup>. In addition, it is well established that the oral bioavailability of many  
158 lipophilic drugs is relatively low and variable, which reduces their efficacy and can lead  
159 to undesirable side effects<sup>2,22</sup>. The development of specially designed excipient foods  
160 that enhance bioavailability and bioactivity may be able to overcome these problems.

### 161 **3.2. Limitations of excipient foods**

162 The development of successful excipient foods faces a number of technical, legal,  
163 and commercial challenges. In particular, there are important differences in the ability  
164 to prove the impact of excipient foods on the bioactivity of drugs and nutraceuticals.  
165 Drugs can be administered in well-defined doses at specified times thereby enabling  
166 pharmaceutical researchers to carry out studies to establish their efficacy against specific  
167 disease symptoms or biomarkers. Thus the impact of excipient foods on drug  
168 bioactivity can be established using well-controlled experiments that involve taking the  
169 drug in the absence or presence of the excipient food. In contrast, nutraceuticals are  
170 typically consumed at relatively low levels as part of a complex diet over extended  
171 periods. Hence, it is often difficult to establish a strong correlation between the type and  
172 amount of nutraceutical consumed and a particular disease. This would make it  
173 challenging to prove the efficacy of excipient foods at improving human health and

174 wellness since long-term studies would be needed with well-controlled diets.  
175 Consequently, it would be difficult for food manufacturers to provide the scientific  
176 evidence required by regulators to make specific health claims about an excipient food  
177 product in their advertising or labeling. In the absence of this kind of competitive  
178 advantage food companies may be reluctant to spend research funds on developing and  
179 testing the efficacy of excipient foods. Nevertheless, one might be able to make the  
180 simpler claim that excipient foods increase the bioavailability of specific food  
181 components, such as carotenoids or oil-soluble vitamins. Another unique challenge that  
182 the food industry faces is in controlling the dose and timing that a nutraceutical  
183 containing food and an excipient food are consumed. Pharmaceuticals are usually  
184 taken in well-defined doses at specific times, whereas nutraceutical agents may be  
185 present in various types of foods that are consumed in different amounts by different  
186 individuals as part of a complex diet that contains other components that could affect  
187 bioavailability. The time that an excipient food is consumed relative to a  
188 nutraceutical-rich food may also be important for the efficacy in enhancing bioavailability,  
189 *e.g.*, before, during, or after consumption.

190 Another potential challenge is that an individual may consume a number of different  
191 kinds of foods containing nutraceuticals, or a patient may need to take more than one  
192 kind of drug per day. It may be necessary to design different kinds of food matrices in  
193 excipient foods for different kinds of nutraceutical-rich foods or drugs. In addition,  
194 different individuals or patients have different food preferences and so a range of  
195 different kinds of excipient product types may be required, *e.g.*, fruit drinks, yogurts,  
196 candies, deserts, spreads with different flavors.

197 Another potential issue with the development of excipient foods is their potential  
198 adverse side effects on human health. For example, the metabolizing enzymes and  
199 efflux transports in epithelial cells usually protect the human body from the effects of any

200 harmful substances that have been ingested<sup>23</sup>. If bioactive agents are incorporated into  
201 foods that appreciably alter these mechanisms, then they might increase the uptake of  
202 harmful substances that could have adverse effects on health and wellness. For example,  
203 some excipient food ingredients could increase the bioavailability of toxic substances  
204 found in foods. In addition, certain bioactive components may be beneficial to human  
205 health in relatively low doses, but have adverse effects at relatively high levels. In this  
206 case, the ability of an excipient food to greatly enhance the bioavailability of a bioactive  
207 component could be detrimental.

208 Finally, if excipient foods are going to be marketed to consumers, it will be  
209 important to educate them about their potential risk and benefits, and to provide advice  
210 about which excipient food should be consumed with which nutraceutical-rich food.  
211 For example, a dessert cream may be marketed as an excipient food to be consumed with  
212 berries, whereas a salad dressing may be marketed as an excipient food to be consumed  
213 with salads and vegetables (**Table 1**).

214

#### 215 **4. Bioavailability of lipophilic bioactive agents**

216 It is useful to highlight the major factors limiting the bioavailability of lipophilic  
217 bioactive agents since this information will aid in the successful development of  
218 efficacious excipient foods. The oral bioavailability of an ingested bioactive component  
219 depends on the fraction that reaches the target site-of-action in a biologically active form  
220<sup>5</sup>. The overall bioavailability ( $F$ ) of a lipophilic bioactive component depends on  
221 numerous factors (**Figure 3**)<sup>24, 25</sup>:

$$222 \quad F = F_L \times F_A \times F_D \times F_M \times F_E \quad (1)$$

223  $F_L$  is the fraction of bioactive agent *liberated* from its original environment, which  
224 may be a drug preparation or a food matrix, into the GIT so that it becomes bioaccessible  
225 *i.e.*, in a form suitable for absorption ( $F_L$ ).  $F_A$  is that fraction of the liberated bioactive

226 agent that is *absorbed* by the epithelial cells within the GIT.  $F_D$  is the fraction of absorbed  
227 bioactive agent that reaches the site of action after *distribution* amongst the various  
228 tissues of the body *e.g.*, blood, liver, kidney, heart, brain, muscles, adipose tissue *etc.*  
229  $F_M$  is the fraction of bioactive component that reaches the site of action in a *metabolically*  
230 active form, which depends on any chemical or enzymatic transformations that take place  
231 after ingestion *e.g.*, hydrolysis, oxidation, and conjugation.  $F_E$  is the fraction of  
232 metabolically active bioactive component that remains at the site of action, *i.e.*, has not  
233 been *excreted*. In reality, each of these parameters varies over time after a bioactive  
234 agent has been ingested to give a profile of bioavailability ( $F$ ) *versus* time ( $t$ ) at a  
235 specified site of action. Typically, the overall bioavailability increases sometime after  
236 ingestion, and then decreases as the bioactive agent is metabolized, stored, utilized,  
237 distributed, or excreted. Ultimately, the bioactivity of an ingested bioactive component  
238 depends on how its bioavailability changes over time in the target tissue. A number of  
239 physiological and physicochemical factors that influence the bioavailability of lipophilic  
240 bioactive components have been established<sup>26-28</sup>, and are summarized in the following  
241 sections.

#### 242 **4.1 Liberation**

243 A lipophilic bioactive agent must be liberated from a food matrix (*e.g.*, fruit,  
244 vegetable, fish, meat, processed food) or drug preparation (*e.g.*, pill or capsule) and then  
245 solubilized within mixed micelles in the small intestinal fluids before it becomes  
246 accessible for absorption (**Figure 3**). Mixed micelles are assembled from bile salts and  
247 phospholipids secreted by the body, as well as any lipid digestion products such as  
248 monoacylglycerols and free fatty acids. It should be stressed that the expression “mixed  
249 micelles” actually refers to a compositionally, structurally, and dynamically complex  
250 mixture within the GIT that may contain various colloidal structures, such as micelles,  
251 vesicles, and liquid crystals that changes over time during the digestion and absorption

252 processes<sup>29</sup>. The fraction of an ingested lipophilic bioactive agent that is solubilized  
253 within the mixed micelle phase of the small intestine is usually taken to be a measure of  
254 the fraction that is liberated ( $F_L$ ) in a form suitable for absorption.

#### 255 **4.2. Absorption**

256 Mixed micelles are able to transport solubilized lipophilic bioactive agents through  
257 the mucus layer and to the apical side of the intestinal epithelial cells (**Figure 3**). The  
258 bioactives may then be incorporated into the epithelial cells through various passive or  
259 active transfer mechanisms<sup>30</sup>. At present, it is not clear whether the bioactive agents  
260 are first released from the mixed micelles into the surrounding aqueous phase and then  
261 absorbed, or whether they are absorbed as part of the mixed micelles *e.g.*, by fusion with  
262 the cell membranes. In addition, it is also possible for bioactive molecules trapped  
263 within other types of colloidal particles (such as engineered nanoparticles) to be directly  
264 absorbed by intestinal epithelial cells. Overall, the fraction of the bioactive agent that is  
265 transported into the epithelial cells is usually taken as a measure of the fraction absorbed  
266 ( $F_A$ ) by the body.

#### 267 **4.3. Metabolism**

268 After ingestion, lipophilic bioactive agents may be transformed as they pass through  
269 the GIT or after they have been absorbed due to various chemical processes (such as acid  
270 hydrolysis or lipid oxidation)<sup>31</sup> or biochemical processes (such as digestive or metabolic  
271 enzyme activity)<sup>2, 5, 23, 32</sup>. The presence of digestive enzymes (such as lipases and  
272 phospholipases) may catalyze the breakdown of some lipophilic bioactive agents (such as  
273 triacylglycerols, phospholipids or Vitamin E acetate)<sup>33</sup>. The presence of metabolic  
274 enzymes changes the chemical structures of some ingested lipophilic bioactive agents,  
275 thereby altering their physicochemical and physiological characteristics. The extent of  
276 metabolism often depends on the route that the bioactive agents are transported into the

277 systemic circulation<sup>22,34</sup>. Strongly hydrophobic agents tend to be transported *via* the  
278 lymphatic route, whereas less hydrophobic agents tend to be transported *via* the portal  
279 vein and liver<sup>33</sup>. Lipophilic bioactives may be highly metabolized when they pass  
280 through the liver before reaching the systemic circulation, thereby altering their  
281 biological activity. In some cases, molecular transformations increase bioactivity,  
282 whereas in other cases they decrease it. The transformation of a lipophilic bioactive as it  
283 travels through the GIT and human body determine the fraction that arrives at the site of  
284 action in a metabolically active state ( $F_M$ ). The ability to alter the absorption pathway  
285 of bioactive agents by manipulating dietary composition or structure provides an  
286 important way of increasing the bioavailability of certain bioactives.

#### 287 **4.4. Distribution**

288 After a lipophilic bioactive agent has been absorbed it is usually distributed amongst  
289 various tissues within the human body (**Figure 3**), such as the systemic circulation, liver,  
290 kidney, muscles, adipose tissue, heart, lungs, brain, *etc.*<sup>35</sup>. The distribution of the  
291 bioactive agent depends on the molecular characteristics of the bioactive, as well as those  
292 of any co-ingested food components. The target tissue(s) for a bioactive agent depends  
293 on the nature of the biological response required, such as enhanced performance,  
294 maintenance of general wellbeing, prevention of chronic disease, or treatment of specific  
295 acute diseases.

#### 296 **4.5. Excretion**

297 Lipophilic bioactives and their metabolites are eventually removed from the human  
298 body through a variety of mechanisms, and often end up within the feces, urine, sweat, or  
299 breath<sup>36</sup>. It may therefore be possible to increase the bioavailability of an ingested  
300 bioactive by increasing its persistence within the human body. The rate of excretion  
301 determines the fraction of bioactive agent that remains at the site of action ( $F_E$ ) at a

302 particular time.

#### 303 **4.6. Improving oral bioavailability**

304 The oral bioavailability of ingested lipophilic agents can be improved by designing  
305 excipient foods that increase the fraction liberated ( $F_L$ ), absorbed ( $F_A$ ), and reaching the  
306 site of action ( $F_D$ ) in a metabolically active form ( $F_M$ ). This goal can be achieved by  
307 manipulating the composition and structure of food matrices based on knowledge of the  
308 impact of specific food matrix properties on the biological fate of ingested lipophilic  
309 bioactives (see Section 5).

### 310 **5. Impact of food matrix on bioavailability**

311 The oral bioavailability of lipophilic bioactives in drugs or foods may be increased  
312 by ingesting them with excipient foods with specifically designed compositions and  
313 structures. In this section, some of the major ways in which food components may alter  
314 the oral bioavailability of lipophilic bioactive agents is highlighted. It is assumed that  
315 an excipient food should be fabricated entirely from food-grade ingredients that are  
316 generally recognized as safe (GRAS). An excipient food could then be marketed and  
317 distributed as a conventional food product with additional health benefits.

#### 318 **5.1. Potential mechanisms of action**

319 The components within an excipient food may alter the oral bioavailability of  
320 co-ingested lipophilic bioactives through various physicochemical or biochemical  
321 mechanisms, which are highlighted in this section.

##### 322 **5.1.1. Bioactive liberation**

323 Prior to ingestion, lipophilic bioactive agents are typically trapped within some kind  
324 of fluid, semi-solid, or solid matrix in pharmaceutical or drug products. For example, a  
325 lipophilic drug may be present within a pill or capsule, whereas a lipophilic nutraceutical  
326 may be trapped inside the cells of a fruit or vegetable or within the fat droplets in a



327 processed food. The bioactive agents must therefore be liberated from their original  
328 location before they can be solubilized within intestinal fluids and absorbed by the body  
329 (**Figure 3**). An excipient food may therefore be designed so that it contains specific  
330 ingredients that facilitate the release and solubilization of bioactive agents. The design  
331 of this type of food requires knowledge of the physicochemical and physiological  
332 processes that occur within the human gastrointestinal tract after ingestion (**Figure 4**).

#### 333 *5.1.1.1. Release from Food or Drug Matrix*

334 The breakdown of the matrix surrounding a bioactive agent within the human GIT is  
335 usually carried out by mechanical, chemical, and enzymatic means<sup>37-39</sup>. Foods are  
336 usually masticated within the mouth to break them down into smaller fragments prior to  
337 swallowing, whereas pharmaceutical preparations (such as capsules and pills) are usually  
338 swallowed directly. After swallowing, pharmaceutical or drug matrices may be broken  
339 down in the stomach and small intestine due to the mechanical motions of the GIT, *e.g.*,  
340 peristalsis or grinding<sup>38,40,41</sup>. The high acidity and ionic strength of the stomach also  
341 facilitates the dissociation of certain structures, particularly those held together by  
342 electrostatic interactions<sup>39,42</sup>. Some matrix dissociation may also occur due to the  
343 simple fact that the material is dissolved within an aqueous environment, *e.g.*, pills,  
344 capsules, or powders formed from water-soluble substances such as carbohydrates or  
345 proteins. The activity of digestive enzymes (such as amylases, proteases, and lipases)  
346 stimulates the breakdown of major food components (such as starches, proteins, and  
347 lipids), which often play an important role in maintaining the matrix structure in foods  
348 and drug preparations. Secreted biological surfactants in the GIT, such as bile salts and  
349 phospholipids, may also facilitate the breakdown of matrix structures held together by  
350 hydrophobic interactions in foods and drug preparations, particularly those containing  
351 lipids or surface active agents.

352 Excipient foods may enhance one or more of these processes by numerous

353 mechanisms. Ingestion of an excipient food may stimulate the release of hormones that  
354 promote the release of acids, enzymes, or bile salts within the GIT<sup>43,44</sup>, thereby  
355 promoting the liberation of bioactive agents by facilitating the breakdown of matrix  
356 structures in foods or drug preparations. The co-ingestion of bioactive lipophilic agents  
357 with an excipient food may change their bioavailability by altering their transit time  
358 within the GIT. Food components that delay transit may lead to higher absorption of  
359 bioactive agents since then there is more time for them to be liberated and absorbed.  
360 The presence of fats within an excipient food may facilitate the release of lipophilic  
361 bioactive agents from co-ingested foods or pharmaceuticals by acting as an organic  
362 solvent. Salts, acids, bases, or chelating agents in an excipient food may contribute to  
363 the breakdown of matrix structures in foods or drug preparations by altering the  
364 molecular interactions between structural components. A number of food components  
365 may alter the intestinal pH due to their acidity, alkalinity or buffering capacity<sup>45</sup>. For  
366 example, ingestion of high amounts of protein may lead to a higher gastric pH due to the  
367 strong buffering capacity of some protein molecules. Changes in pH may alter the rate  
368 and extent of breakdown of food or pharmaceutical matrix structures and therefore the  
369 liberation of bioactive components.

#### 370 *5.1.1.2. Solubilization in Mixed Micelles*

371 After a lipophilic bioactive agent is liberated from the original food or  
372 pharmaceutical matrix it needs to be solubilized within the mixed micelle phase so that it  
373 can be transported to the intestinal epithelial cells. It is well established that  
374 co-ingestion of lipophilic drugs or nutraceuticals with lipids can greatly increase their  
375 oral bioavailability, which can be attributed to a number of factors<sup>2,13,22</sup>. First,  
376 ingestion of lipids stimulates the release of digestive enzymes and bile salts, as well as  
377 increasing GIT transit time. An increase in the bile salt levels increases the  
378 solubilization capacity of the intestinal fluids, whereas as an increase in GIT transit time

379 increases the time available for any ingested bioactive agents to be liberated, solubilized,  
380 and absorbed. Second, the digestion of co-ingested lipids (triglycerides) within the GIT  
381 leads to the formation of free fatty acids (FFA) and monoacylglycerols (MAG) that are  
382 incorporated into the mixed micelles in the small intestine thereby increasing their  
383 solubilization capacity for lipophilic bioactives (see later section). Third, ingestion of  
384 any surface active substances (such as phospholipids or surfactants) may also increase the  
385 solubilization capacity of the intestinal fluids due to their ability to be incorporated into  
386 mixed micelles<sup>46-48</sup>.

### 387 *5.1.1.3. Alteration of Mass Transport Processes*

388 The liberation of lipophilic bioactive agents within the GIT often depends on the  
389 mass transport of reactants, catalysts, and products from one location to another.  
390 Digestive enzymes must come into close proximity to their substrates before they can  
391 carry out their catalytic actions. Bioactive agents solubilized within mixed micelles  
392 must be transported through the lumen and across the mucous layer before they can be  
393 absorbed by epithelial cells (Figure 5). The rate and extent of liberation of bioactive  
394 agents from food or drug matrices may therefore be controlled by incorporating food  
395 ingredients within excipient foods that alter mass transport processes within the lumen of  
396 the GIT. In general, mass transport may occur by convective or diffusive processes,  
397 depending on the structural and physicochemical properties of the intestinal fluids and the  
398 flow profile within the region of the GIT involved<sup>49</sup>. The mechanical forces generated  
399 by the GIT mix components together and help move them from one location to another<sup>40,</sup>  
400<sup>50</sup>. Nevertheless, there are regions within the GIT where mass transport is primarily  
401 diffusion-limited, *e.g.*, the movement of small molecules through gelled phases.  
402 Excipient food components may be able to alter diffusion-limited or convection-limited  
403 processes by various mechanisms: binding to bioactive agents; altering the microscopic  
404 or macroscopic rheology of the intestinal fluids; altering GIT motility. For example,

405 some biopolymers are able to form viscous solutions or gels under simulated  
406 gastrointestinal conditions, and may therefore be able to alter mass transport and transit  
407 times, which in turn alter important events affecting the release and processing of  
408 bioactive agents<sup>51, 52</sup>. Cationic biopolymers, such as chitosan, are able to bind anionic  
409 bile salts and free fatty acids, and therefore alter their mass transport<sup>53, 54</sup>.

#### 410 *5.1.1.4. Alterations in Gut Motility*

411 Certain kinds of food components have been shown to alter the motility of the GIT,  
412 *e.g.*, gastric emptying time or the mechanical actions of the stomach and small intestine<sup>16,</sup>  
413 <sup>32, 38</sup>. The co-ingestion of a bioactive agent with a meal often increases the length of  
414 time it spends within the stomach<sup>16</sup>. Specific phytochemicals, such as piperine, have  
415 also been shown to inhibit gastric emptying<sup>55</sup>. The longer a food spends within the  
416 stomach the greater time there is for the breakdown of any matrices that normally inhibit  
417 the liberation of the bioactive agents into the intestinal fluids (*e.g.*, cell walls in plant  
418 tissues or solid drug forms). In addition, an increase in gastric emptying time may  
419 increase the amount of digestion, metabolism, or chemical transformation of a substance  
420 that occurs within the stomach. In some cases, this may increase the bioavailability of  
421 an ingested nutraceutical or pharmaceutical, *e.g.*, if the transformed form has a higher  
422 bioavailability than the original form, or if some of the components released from the  
423 food matrix increase the subsequent solubilization or absorption of the bioactive agents.  
424 In other cases, an increase in gastric retention may decrease bioavailability, *e.g.*, if the  
425 transformed form has a lower bioavailability than the original form, or if some of  
426 components released from the food matrix inhibit the subsequent solubilization or  
427 absorption of the bioactive agents. Furthermore, an increase in the gastric emptying time  
428 also slows down the rate at which bioactive agents are transported to small intestine,  
429 which may have a significant impact on their absorption and metabolism in the small  
430 intestine.

### 431 **5.1.2. Bioactive absorption**

432 There are numerous physicochemical and physiological mechanisms by which food  
433 matrix components could alter the absorption of co-ingested lipophilic bioactive agents.  
434 A number of the most important mechanisms that might be used in the development of  
435 excipient foods are highlighted in this section.

#### 436 *5.1.2.1. Increase in membrane permeability*

437 The bioavailability of some lipophilic bioactive agents is limited by their transport  
438 across the layer of epithelial cells surrounding the GIT<sup>56,57</sup>. When bioactive agents reach  
439 the apical side of the intestinal epithelial cells they may be transported into the systemic  
440 circulation by a number of passive or active transport processes (**Figure 5**). The precise  
441 mechanism(s) involved depend on the molecular characteristics of the bioactive, the  
442 nature of any particles that the bioactive might be trapped within or bound with, the  
443 composition and structure of the surrounding intestinal fluids, and the region of the GIT  
444 where absorption occurs.

445 The two major types of epithelial cells that line the gastrointestinal tract in regions  
446 where the majority of absorption occurs are enterocytes and M-cells<sup>58-61</sup>. Enterocytes  
447 are the most numerous type of cell lining the GIT, and they are where most of the  
448 absorption of molecular forms of drugs and nutraceuticals occur. Enterocytes also have  
449 ability to absorb certain types of particulate matter. Conversely, M-cells are much less  
450 numerous than enterocytes, typically occupying less than 1% of the epithelium surface,  
451 but they are much more efficient than enterocytes at absorbing particulate matter.  
452 M-cells are mainly found in specialized regions on the epithelium surface referred to as  
453 “Peyers patches”, which are primarily responsible for absorbing ingested antigens, such  
454 as macromolecules, microorganisms, and certain types of particles. The absorbed  
455 particles are then transported to the underlying lymphoid system where they promote  
456 immune responses<sup>58,62</sup>.

457 Molecules and particles reaching the epithelial cells may be absorbed through a  
458 number of mechanisms depending on their characteristics <sup>61-63</sup>:

459 *Paracellular*: Small molecules and particles are able to pass through the narrow gaps  
460 (“tight junctions”) that separate neighboring epithelial cells (**Figure 5**). Typically, only  
461 substances that are smaller than a few nanometers are able to pass through the tight  
462 junctions. However, some substances found in foods have been shown to be capable of  
463 increasing the dimensions of the tight junctions and may therefore be able to enhance  
464 transport by this mechanism <sup>64</sup>, e.g., some surfactants <sup>65,66</sup>, polymers <sup>67,68</sup>, minerals <sup>69</sup>,  
465 and chelating agents <sup>70</sup>. Specific examples of food-grade substances that might be used  
466 to increase the permeability of epithelial cells by increasing the dimensions of the tight  
467 junctions include the surfactant Tween 80 <sup>65</sup>, the polymer chitosan <sup>67,71</sup>, the mineral zinc  
468 <sup>69</sup>, and the chelating agent EDTA <sup>70</sup>.

469 *Transcellular* –Molecules and particles may also be transported through epithelial  
470 cell membranes by passive or active transport mechanisms (**Figure 5**). Many fairly  
471 lipophilic molecules are transferred across cell membranes by a passive mechanism.  
472 After encountering the epithelial cells, they are solubilized within the non-polar  
473 phospholipid tails that make up the phospholipid bilayer of the cell membrane. After  
474 moving across the cell membrane, they are incorporated into various vesicle-like  
475 structures on the other side, which then move them into the cell interior. Other types of  
476 molecules (particularly more hydrophilic ones) are transferred across the cell membrane  
477 by membrane protein-transporter systems. The absorption of particles that are small  
478 enough to travel through the mucus layer and reach the surface of the epithelial cells  
479 typically occurs by an “endocytosis” mechanism <sup>61</sup>. In this case, particles come into  
480 contact with the outer wall of the cell membrane, the membrane then wraps itself around  
481 the particle, and then part of the membrane buds-off to form a vesicle-like structure with  
482 a particle trapped inside that moves into the interior of the cell. This process may occur

483 in enterocyte cells, but is typically much more active in M-cells. The critical cut-off  
484 particle size for endocytosis has been estimated to be from less than 50 to around 100 nm  
485 for enterocyte cells, and to be from 20 to 500 nm for M-cells.

486 Certain types of molecules present in foods may be able to increase the transcellular  
487 uptake of lipophilic bioactive agents by epithelial cells by altering cell membrane  
488 permeability. Piperine (a compound found in black pepper) has been shown to be  
489 capable of increasing cell membrane permeability<sup>32</sup>. Food grade surfactants (sucrose  
490 monoesters) have also been shown to increase membrane permeability to model drugs<sup>72</sup>.  
491 Rhamnolipids have been shown to increase both transcellular and paracellular transport of  
492 model drugs<sup>73</sup>.

493 *Persorption*: Molecules or particles may also be absorbed through temporary pores  
494 formed in the layer of epithelial cells lining the GIT due to gaps formed when some of the  
495 cells are shed and replaced<sup>63</sup>.

#### 496 5.1.2.2. *Inhibition of efflux mechanisms*

497 The bioavailability of certain types of lipophilic bioactive agents is limited due to the  
498 presence of efflux mechanisms in the membranes of the intestinal epithelial cells<sup>57, 74, 75</sup>.  
499 After absorption by epithelial cells, some bioactives are transported back into the  
500 intestinal lumen by specific transports at the apical side of the cell membrane. For  
501 example, both P-glycoprotein (P-gp) and multidrug resistant protein (MRP) have been  
502 shown to pump out a wide range of lipophilic bioactives from epithelial cells lining the  
503 GIT<sup>57, 74</sup>. This efflux process can reduce the bioavailability of bioactive agents by two  
504 mechanisms: (i) decreasing the total amount absorbed; and, (ii) increasing the extent of  
505 metabolism within the GIT if the bioactive is pumped out and then reabsorbed, which  
506 increase exposure of the bioactive to metabolizing enzymes inside of the epithelial cells.  
507 Certain types of food-grade components have been shown to be able to block efflux  
508 mechanisms, and thereby increase the net absorption of lipophilic bioactive agents by

509 epithelial cells, *e.g.*, some surfactants, chelating agents, biopolymers, and phytochemicals  
510 <sup>5, 57, 76, 77</sup>. For example, resveratrol, quercetin and piperine have been shown to act as  
511 efflux inhibitors for certain kinds of drugs <sup>76, 78-81</sup>. In general, three different  
512 mechanisms have been proposed for the ability of these components to inhibit efflux  
513 processes: (i) blocking binding sites on the efflux protein surfaces; (ii) interference with  
514 ATP hydrolysis (which provides the energy needed for efflux protein action); (iii)  
515 alteration of cell membrane structure (which leads to alterations in efflux protein  
516 conformation and activity) <sup>57</sup>.

### 517 ***5.1.3. Bioactive metabolism or chemical transformation***

518 Numerous molecules isolated from plant and animal sources have been shown to  
519 enhance the bioavailability of nutraceuticals or pharmaceuticals due to their ability to  
520 interfere with chemical transformations that normally occur within the GIT or after  
521 absorption <sup>32</sup>. Some of these bioactivity enhancers act as antioxidants that retard the  
522 oxidation of nutraceuticals or pharmaceuticals, such as  $\omega$ -3 fatty acids, carotenoids, or  
523 conjugated linoleic acid <sup>82</sup>. For example, there are many natural and synthetic  
524 food-grade antioxidants that are effective at inhibiting oxidation reactions by mechanisms  
525 such as free radical scavenging, singlet oxygen quenchers, and chelating agents, *e.g.*,  
526 BHT, BHA, carotenoids, tocopherols, flavonoids, and grape seed extract <sup>83</sup>. Other  
527 bioactivity enhancers may inhibit the normal functioning of metabolic or digestive  
528 enzymes within the GIT or body <sup>23, 32</sup>. For example, piperine has been shown to retard  
529 the metabolism of certain drugs and nutraceuticals, such as ibuprofen, curcumin,  
530 resveratrol, EGCG, carotenoids, vitamins, and amino acids <sup>32</sup>. These affects have been  
531 partly attributed to its ability to inhibit metabolizing enzymes such as glucose  
532 dehydrogenase, cytochrome P450, and others <sup>32</sup>.

533



534 **Table 2.** Examples of phytochemicals from natural sources that may increase the  
 535 bioavailability of co-ingested lipophilic nutraceuticals and pharmaceuticals. Examples  
 536 taken from various sources: Dudharta<sup>32</sup>, Shimizu<sup>23</sup>, Choi<sup>84</sup>, Jia<sup>77</sup>.

<b>Bioavailability Enhancer</b>	<b>Nutraceuticals Enhanced</b>	<b>Mechanism</b>
Piperine	Vitamins A, D, E, K Carotenoids, Curcuminoids Coenzyme Q10 Hydrophobic drugs	Metabolizing Enzyme Inhibition Modulation of Gut Motility
Gingerols	Vitamins A and E Carotenoids, Curcumin	Modulation of Gut Motility
Curcumin	Hydrophobic drugs	Metabolizing Enzyme Inhibition Efflux Transporter Inhibition
Quercetin	Hydrophobic drugs	Efflux Transporter Inhibition

537

## 538 **5.2. Excipient food ingredients**

539 In this section, the potential influence of common food components that may be  
 540 incorporated into excipient foods on the oral bioavailability of lipophilic bioactive agents  
 541 is discussed. Those ingredients that appreciably increase the bioavailability of  
 542 nutraceuticals can be referred to as “excipient food ingredients”. An excipient food may  
 543 contain one or more of these ingredients so as to increase the bioavailability of one or  
 544 more nutraceuticals.

### 545 **5.2.1. Lipids**

546 Studies by pharmaceutical researchers have shown that co-ingestion of lipophilic

547 drugs with lipids improves their oral bioavailability by an amount that depends on the  
548 amount, type, and structure of the ingested lipids<sup>2</sup>. Food and nutrition research has also  
549 shown that the bioavailability of lipophilic nutraceuticals can be increased by  
550 co-ingestion with lipids<sup>85-87</sup>. *In vitro* studies have reported that the bioaccessibility  
551 (micelle solubilization) and absorption (cell culture uptake) of lipophilic bioactive agents  
552 from fruits and vegetables is greatly increased in the presence of lipids<sup>85,86</sup>. The extent  
553 of the increase in bioaccessibility and absorption depends on the amount and composition  
554 of the lipids used<sup>86,88,89</sup>. Bioaccessibility was higher for lipids containing long chain  
555 triglycerides (LCT) than those containing short or medium chain triglycerides (SCT or  
556 MCT), presumably because of differences in the solubilization capacity of the mixed  
557 micelles formed<sup>2,90</sup>. Lipophilic bioactives encapsulated within indigestible oils (flavor  
558 oils) have been shown to have low bioaccessibility using *in vitro* studies, which was  
559 attributed to the fact that some of them remained in the undigested oil droplets and there  
560 were fewer mixed micelles available to solubilize them<sup>90,91</sup>. In addition to their  
561 composition, the liberation of bioactives from emulsified lipids also depends on their  
562 particle size, physical state, and interfacial characteristics<sup>92,93</sup>. Typically, the release  
563 rate is faster for smaller particles, for liquid oils rather than solid fats, and for interfaces  
564 where bile salts and lipases can easily absorb.

565 Co-ingested lipids may also alter the bioavailability of lipophilic drugs or  
566 nutraceuticals through other mechanisms. When lipophilic bioactives are ingested with  
567 LCT they are packed into lipoprotein particles (chylomicrons) in the intestinal epithelial  
568 cells and then transported by the lymphatic route (thereby avoiding first pass metabolism  
569 in the liver), but when they are ingested with SCT or MCT they tend to be transported *via*  
570 the portal vein (where they must pass through the liver before entering the systemic blood  
571 circulation)<sup>34,94</sup>. Bioactives packaged in different vehicles (*e.g.* chylomicron *vs.*  
572 non-chylomicron) in the epithelial cells may have different metabolic fates due to

573 differences in their exposure to metabolizing enzymes present in different body tissues.

### 574 **5.2.2. Carbohydrates**

575 In general, food carbohydrates are classified as monosaccharides ( $n = 1$ ),  
576 oligosaccharides ( $n = 2$  to 20), or polysaccharides ( $n > 20$ ) depending on the number of  
577 monomers present <sup>95</sup>. Carbohydrates may also be classified as digestible or indigestible  
578 depending on their susceptibility to enzymatic hydrolysis in the upper GIT <sup>96,97</sup>. Starch  
579 is the most abundant digestible polysaccharide in foods, whereas there are many types of  
580 indigestible polysaccharides, such as cellulose, hemicellulose, pectin, alginate,  
581 carrageenan, xanthan gum, locust bean gum, and agar. Indigestible polysaccharides are  
582 part of a class of polymers known as dietary fibers, which vary according to their  
583 monomer type, distribution, and bonding, as well as their electrical charge,  
584 hydrophobicity, molecular weight, degree of branching, and conformation <sup>95,97</sup>.  
585 Co-ingested carbohydrates may influence the bioavailability of lipophilic bioactive drugs  
586 and nutraceuticals through various mechanisms. As mentioned earlier, many  
587 polysaccharides are able to increase the viscosity or form a gel within the GIT, thereby  
588 altering mass transport processes, *e.g.*, diffusion of enzymes to substrates in food  
589 matrices, or digestion products/bioactives to epithelial cells. Some dietary fibers may be  
590 able to form impermeable coatings around food matrix components that inhibit their  
591 digestion and therefore the release of bioactive agents <sup>92</sup>. Electrically charged  
592 polysaccharides are capable of binding oppositely charged molecular species in the GIT  
593 that may influence food matrix digestion and bioactive release. For example, cationic  
594 dietary fibers (such as chitosan) can bind anionic bile salts, fatty acids, or phospholipids,  
595 whereas anionic dietary fibers (such as alginate) can bind cationic calcium ions <sup>27, 98-100</sup>.  
596 Cationic dietary fibers have also been shown to inhibit lipase activity, and therefore  
597 reduce the rate of lipid digestion <sup>101</sup>. Some dietary fibers have been shown to alter cell  
598 membrane permeability through their effect on tight junction dimensions, *e.g.*, chitosan <sup>67</sup>,

599 <sup>68</sup>. Dietary fibers may also change the nature of the microbial population within the  
600 colon, which can alter the metabolism, activity, and absorption of lipophilic bioactives in  
601 the large intestine <sup>102</sup>.

### 602 **5.2.3. Proteins**

603 Food proteins exhibit a wide range of different molecular structures,  
604 physicochemical properties, and physiological effects <sup>103, 104</sup>. Co-ingested proteins can  
605 potentially alter the bioavailability of lipophilic bioactive agents through a number of  
606 mechanisms. Many food proteins and peptides have strong antioxidant activity and may  
607 therefore be able to inhibit the chemical degradation of nutraceuticals or drugs that are  
608 susceptible to oxidation within the GIT, such as  $\omega$ -3 fatty acids or carotenoids <sup>105</sup>. Some  
609 nutraceuticals may bind to proteins within the GIT <sup>106</sup>, which alters the location of their  
610 absorption within the GIT, *e.g.*, anthocyanins bound to proteins have been shown to  
611 travel further down the gastrointestinal tract <sup>107</sup>. Protein digestion within the  
612 gastrointestinal tract may generate hormonal responses that regulate food intake and  
613 processing <sup>108</sup>, thereby altering the way that a food or pharmaceutical matrix is broken  
614 down in the GIT and therefore the release of any trapped bioactive agents. Proteins and  
615 their digestion products may interact with various molecular species involved in the  
616 digestion of food matrices and the release and transport of bioactive agents, such as  
617 bioactives, mixed micelles, phospholipids, and enzymes <sup>109-112</sup>. For example, a recent  
618 study suggests that lactoferrin may reduce the bioavailability of  $\beta$ -carotene, which was  
619 attributed to the fact that it was positively charged and bound to negatively charged  
620 digestive components, such as bile salts or free fatty acids <sup>113</sup>. Some protein digestion  
621 products, for example those from casein and whey proteins, have been shown to alter  
622 (close) tight junction permeability, and may therefore alter the uptake of any  
623 nutraceuticals absorbed by this mechanism <sup>23</sup>.

### 624 **5.2.4. Surfactants**

625 Surfactants are commonly used in the food and pharmaceutical industries to form  
626 and stabilize colloidal delivery systems, such as microemulsions, nanoemulsions,  
627 emulsions, and solid lipid nanoparticles<sup>114, 115</sup>. Surfactants vary in the nature of their  
628 polar head groups and non-polar tail groups, which alters their behavior within foods and  
629 the GIT. The head group may be non-ionic, cationic, anionic, or zwitterionic, while the  
630 tail group may vary in the number, length and unsaturation of the non-polar chains.  
631 Synthetic or natural surfactants may be present within an ingested food *e.g.*, non-ionic  
632 surfactants (*e.g.*, Tweens, Spans, and sucrose esters), ionic surfactants (*e.g.*, DATEM and  
633 CITREM), phospholipids (*e.g.*, egg, soy, or sunflower lecithin), or monoacylglycerols<sup>116</sup>.  
634 Alternatively, they may be generated from ingested food components as a result of the  
635 digestion process, *e.g.*, monoacylglycerols from triacylglycerols or lysolecithin from  
636 phospholipids<sup>117</sup>. Surfactants can alter the bioavailability of lipophilic bioactives  
637 through a number of mechanisms: some surfactants bind to digestive enzymes (such as  
638 lipase or protease) and alter their activity<sup>118</sup>; surfactants may be incorporated into mixed  
639 micelles thereby increasing their solubilization capacity<sup>119</sup>; surfactants may inhibit lipase  
640 absorption to lipid surfaces through competitive absorption<sup>93, 120</sup>; surfactants may alter  
641 the permeability of enterocytes by interacting with transporters on cell membranes<sup>121</sup>;  
642 surfactants may increase cell permeability by increasing the dimensions of the tight  
643 junctions<sup>23, 65, 122</sup>.

#### 644 **5.2.5. Minerals**

645 Certain types of mineral ions also impact the liberation and absorption of lipophilic  
646 bioactives. For example, calcium ions may impact the rate and extent of lipid hydrolysis,  
647 which influences the release of bioactives from the lipid phase and their subsequent  
648 solubilization in the mixed micelle phase<sup>123, 124</sup>. In the absence of calcium, the  
649 digestion of triacylglycerols in the small intestine is inhibited by accumulation of  
650 long-chain fatty acids (LCFA) at the oil-water interface, since this restricts the access of

651 lipase to the lipid substrate<sup>27</sup>. Calcium ions precipitate accumulated LCFAs through  
652 complexation, thereby removing them from the interface and allowing the lipase to  
653 access the lipid substrate<sup>123, 125, 126</sup>. Calcium ions are therefore able to increase the rate  
654 and extent of lipid digestion through this mechanism<sup>127-130</sup>. Conversely, the formation  
655 of calcium-LCFA precipitates may reduce the solubilization capacity of the mixed micelle  
656 phase, thereby reducing the bioavailability of LCFAs and lipophilic bioactives<sup>123, 131-133</sup>.  
657 Calcium has also been shown to play an important role in the activity of pancreatic lipase,  
658 acting as a co-factor required for activity<sup>134-137</sup>. Multivalent mineral ions may promote  
659 the aggregation of oppositely charged lipid droplets<sup>114</sup>, thereby altering the surface area  
660 of lipid exposed to digestive enzymes. Mineral ions may also promote gelation of  
661 oppositely charged biopolymers (*e.g.*, calcium ions promote alginate gelation), which will  
662 also influence the accessibility of lipid phases to enzyme digestion<sup>138</sup>. Some minerals  
663 have been shown to influence the absorption of bioactive agents by altering cell  
664 membrane permeability, *e.g.*, zinc<sup>69</sup>.

#### 665 **5.2.6. Chelating agents**

666 Metal ion chelators (such as EDTA) have been shown to inhibit efflux transporters in  
667 the GIT, and may therefore increase the bioavailability of bioactive molecules that are  
668 susceptible to removal from enterocytes by this mechanism<sup>57</sup>. Metal ion chelators  
669 (such as EDTA and phosphates) may interfere with the various roles that calcium ions  
670 play in the digestion and release of lipids by complexing them – see section 5.2.5<sup>124</sup>.

#### 671 **5.2.7. Phytochemicals**

672 A number of phytochemicals derived from edible plant materials have been shown to  
673 be able to promote the bioavailability of certain bioactive food agents. For example,  
674 some polyphenols affect absorption and efflux transporters in enterocyte membranes thus  
675 altering the accumulation of bioactive agents within the body *e.g.*, quercetin, curcumin,  
676 piperine, and some catechins<sup>139-142</sup>. Specific phytochemicals may also be able to inhibit

677 chemical reactions (such as lipid oxidation) or biochemical reactions (such as digestion or  
678 metabolism) in the gastrointestinal tract<sup>23,32</sup>. For example, it has been reported that  
679 piperine reduced the metabolism of curcumin in the GIT by inhibiting metabolizing  
680 enzymes, thereby increasing bioavailability<sup>32</sup>. (See sections 5.1.2.2 and 5.1.3)

#### 681 **5.2.8. Excipient food ingredients**

682 Many of the food ingredients discussed in the previous sections have the ability to  
683 increase the oral bioavailability of co-ingested bioactive agents. These ingredients can  
684 therefore be used to construct excipient foods that are specifically designed to increase  
685 the overall oral bioavailability of one or more type of co-ingested bioactive agents. For  
686 example, an excipient food may contain lipids to increase the solubilization capacity of  
687 the intestinal fluids, a phytochemical to inhibit efflux mechanisms, and a surfactant to  
688 increase epithelium cell membrane permeability.

## 689 **6. Conclusions**

690 This article has introduced the concept of *excipient foods* that are specifically  
691 designed to enhance the oral bioavailability of lipophilic bioactive agents such as  
692 nutraceuticals in foods or drugs in pharmaceuticals. Knowledge of the influence of  
693 specific food components and structures on the bioavailability of specific lipophilic  
694 bioactive agents is increasing, which will facilitate the rational design of food matrices  
695 that can enhance the biological activity of nutraceuticals and drugs. A number of  
696 different approaches can be used, including increasing the release, solubilization,  
697 transport, and uptake of bioactive agents, while decreasing their metabolism or efflux.

698 There is a growing convergence in the interests of pharmaceutical and food  
699 companies. The food industry is increasingly focusing on the development of functional  
700 food and beverage products designed to improve performance, maintain wellbeing, and  
701 inhibit the onset of chronic diseases, such as osteoporosis, heart disease, cancer,

702 hypertension, and obesity. The pharmaceutical industry continues to develop products  
703 to prevent, manage, and cure chronic and acute diseases. Many of the biologically  
704 active substances present in foods and drugs are highly lipophilic agents that normally  
705 have poor oral bioavailability. The availability of a range of excipient foods specifically  
706 designed to increase the oral bioavailability of lipophilic bioactive molecules would  
707 therefore be beneficial to both the pharmaceutical and food industries. Nevertheless,  
708 further research is required to better understand the role of specific excipient food  
709 ingredients on the bioavailability of specific lipophilic bioactive agents, and to establish  
710 the influence of ingredient interactions on bioavailability when excipient foods are  
711 consumed as part of a complex diet that contains many other components.

712

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## 718 **8. References**

- 719 1. J. S. Jingling Tang, *Current Drug Therapy*, 2007, **2**, 85-93.
- 720 2. C. J. H. Porter, N. L. Trevaskis and W. N. Charman, *Nature Reviews Drug*  
721 *Discovery*, 2007, **6**, 231-248.
- 722 3. E. Fernandez-Garcia, I. Carvajal-Lerida, M. Jaren-Galan, J. Garrido-Fernandez,  
723 A. Perez-Galvez and D. Hornero-Mendez, *Food Research International*, 2012,  
724 **46**, 438-450.
- 725 4. A. R. Patel and K. P. Velikov, *Lwt-Food Science and Technology*, 2011, **44**,  
726 1958-1964.
- 727 5. M. J. Rein, M. Renouf, C. Cruz-Hernandez, L. Actis-Goretta, S. K. Thakkar and M.  
728 D. Pinto, *British Journal of Clinical Pharmacology*, 2013, **75**, 588-602.
- 729 6. D. Fleisher, C. Li, Y. Zhou, L. H. Pao and A. Karim, *Clin. Pharmacokinet.*, 1999,  
730 **36**, 233-254.



- 731 7. K. R. N. Moelants, L. Lemmens, M. Vandebroeck, S. Van Buggenhout, A. M. Van  
732 Loey and M. E. Hendrickx, *J. Agric. Food Chem.*, 2012, **60**, 11995-12003.
- 733 8. C. W. Pouton and C. J. Porter, *Advanced drug delivery reviews*, 2008, **60**,  
734 625-637.
- 735 9. M. N. Martinez and G. L. Amidon, *J. Clin. Pharmacol.*, 2002, **42**, 620-643.
- 736 10. S. Hurst, C. M. Loi, J. Brodfuehrer and A. El-Kattan, *Expert Opin. Drug Metab.*  
737 *Toxicol.*, 2007, **3**, 469-489.
- 738 11. D. N. D'Ambrosio, R. D. Clugston and W. S. Blaner, *Nutrients*, 2011, **3**, 63-103.
- 739 12. Y. Y. Yeap, N. L. Trevaskis, T. Quach, P. Tso, W. N. Charman and C. J. H. Porter,  
740 *Molecular Pharmaceutics*, 2013, **10**, 1874-1889.
- 741 13. W. N. Charman, C. J. H. Porter, S. Mithani and J. B. Dressman, *Journal of*  
742 *Pharmaceutical Sciences*, 1997, **86**, 269-282.
- 743 14. M. J. Brown, M. G. Ferruzzi, M. L. Nguyen, D. A. Cooper, A. L. Eldridge, S. J.  
744 Schwartz and W. S. White, *American Journal of Clinical Nutrition*, 2004, **80**,  
745 396-403.
- 746 15. A. Nagao, E. Kotake-Nara and M. Hase, *Bioscience Biotechnology and*  
747 *Biochemistry*, 2013, **77**, 1055-1060.
- 748 16. F. J. O. Varum, G. B. Hatton and A. W. Basit, *Int. J. Pharm.*, 2013, **457**, 446-460.
- 749 17. L. Salvia-Trujillo, C. Qian, O. Martin-Belloso and D. J. McClements, *Food*  
750 *Chemistry*, 2013, **141**, 1472-1480.
- 751 18. H. Kalasz and I. Antal, *Curr. Med. Chem.*, 2006, **13**, 2535-2563.
- 752 19. J. Hamman and J. Steenekamp, *Expert Opin. Drug Deliv.*, 2012, **9**, 219-230.
- 753 20. A. T. Florence and D. Attwood, *Physicochemical Principles of Pharmacy*,  
754 Pharmaceutical Press, London, U.K., 2011.
- 755 21. D. J. McClements, E. A. Decker, Y. Park and J. Weiss, *Critical reviews in food*  
756 *science and nutrition*, 2009, **49**, 577-606.
- 757 22. H. D. Williams, N. L. Trevaskis, S. A. Charman, R. M. Shanker, W. N. Charman, C.  
758 W. Pouton and C. J. H. Porter, *Pharmacological Reviews*, 2013, **65**, 315-499.
- 759 23. M. Shimizu, *Bioscience Biotechnology and Biochemistry*, 2010, **74**, 232-241.
- 760 24. J. A. Arnott and S. L. Planey, *Expert Opinion on Drug Discovery*, 2012, **7**,  
761 863-875.
- 762 25. D. J. McClements, *Expert Opin. Drug Deliv.*, 2013, **10**, 1621-1632.
- 763 26. E. Bauer, S. Jakob and R. Mosenthin, *Asian-Australasian Journal of Animal*  
764 *Sciences*, 2005, **18**, 282-295.
- 765 27. G. Fave, T. C. Coste and M. Armand, *Cellular and Molecular Biology*, 2004, **50**,  
766 815-831.
- 767 28. B. Bermudez, Y. M. Pacheco, S. Lopez, R. Abia and F. J. G. Muriana, *Grasas Y*  
768 *Aceites*, 2004, **55**, 1-10.
- 769 29. A. Mullertz, D. G. Fatouros, J. R. Smith, M. Vertzoni and C. Reppas, *Molecular*

- 770 *Pharmaceutics*, 2012, **9**, 237-247.
- 771 30. H. Singh, A. Ye and D. Horne, *Progress in Lipid Research*, 2008.
- 772 31. K. Larsson, L. Cavonius, M. Alminger and I. Undeland, *J. Agric. Food Chem.*,  
773 2012, **60**, 7556-7564.
- 774 32. G. B. Dudhatra, S. K. Mody, M. M. Awale, H. B. Patel, C. M. Modi, A. Kumar, D. R.  
775 Kamani and B. N. Chauhan, *Scientific World Journal*, 2012.
- 776 33. T. Y. Wang, M. Liu, P. Portincasa and D. Q. H. Wang, *Eur. J. Clin. Invest.*, 2013, **43**,  
777 1203-1223.
- 778 34. J. A. Yanez, S. W. J. Wang, I. W. Knemeyer, M. A. Wirth and K. B. Alton, *Advanced*  
779 *drug delivery reviews*, 2011, **63**, 923-942.
- 780 35. J. C. Espin, M. T. Garcia-Conesa and F. A. Tomas-Barberan, *Phytochemistry*,  
781 2007, **68**, 2986-3008.
- 782 36. A. Ruiz-Garcia, M. Bermejo, A. Moss and V. G. Casabo, *Journal of*  
783 *Pharmaceutical Sciences*, 2008, **97**, 654-690.
- 784 37. G. A. van Aken, *Food Biophysics*, 2010, **5**, 258-283.
- 785 38. M. Koziolok, G. Garbacz, M. Neumann and W. Weitschies, *Molecular*  
786 *Pharmaceutics*, 2013, **10**, 1610-1622.
- 787 39. R. G. Lentle and W. M. J. Patrick, *The Physical Processes of Digestion*, Springer,  
788 New York, N.Y., 2011.
- 789 40. F. Kong and R. P. Singh, *Journal of Food Science*, 2008, **73**, R67-R80.
- 790 41. R. G. Lentle and P. W. M. Janssen, *Critical reviews in food science and nutrition*,  
791 2010, **50**, 130-145.
- 792 42. A. Matalanis and D. J. McClements, *Food Biophysics*, 2012, **7**, 145-154.
- 793 43. J. L. Boyer, *Comprehensive Physiology*, 2013, **3**, 1035-1078.
- 794 44. M. Covasa, *Am. J. Physiol.-Regul. Integr. Comp. Physiol.*, 2010, **299**,  
795 R1423-R1439.
- 796 45. L. Kalantzi, K. Goumas, V. Kalioras, B. Abrahamsson, J. B. Dressman and C.  
797 Reppas, *Pharmaceutical Research*, 2006, **23**, 165-176.
- 798 46. D. M. Cirin, M. M. Posa and V. S. Krstonosic, *Ind. Eng. Chem. Res.*, 2012, **51**,  
799 3670-3676.
- 800 47. S. Rozner, D. E. Shalev, A. I. Shames, M. F. Ottaviani, A. Aserin and N. Garti,  
801 *Colloids and Surfaces B-Biointerfaces*, 2010, **77**, 22-30.
- 802 48. C. Rupp, H. Steckel and B. W. Muller, *Int. J. Pharm.*, 2010, **395**, 272-280.
- 803 49. J. M. Rabanel, V. Aoun, I. Elkin, M. Mokhtar and P. Hildgen, *Curr. Med. Chem.*,  
804 2012, **19**, 3070-3102.
- 805 50. M. J. S. Wickham, R. M. Faulks, J. Mann and G. Mandalari, *Dissolut. Technol.*,  
806 2012, **19**, 15-22.
- 807 51. J. F. Bradbeer, R. Hancocks, F. Spyropoulos and I. T. Norton, *Food Hydrocolloids*,  
808 2014, **35**, 522-530.

- 809 52. T. J. Wooster, L. Day, M. Xu, M. Golding, S. Oiseth, J. Keogh and P. Clifton, *Food*  
810 *Hydrocolloids*, 2014, **36**, 102-114.
- 811 53. T. Helgason, J. Weiss, D. J. McClements, J. Gislason, J. M. Einarsson, F. R.  
812 Thormodsson and K. Kristbergsson, *Journal of Aquatic Food Product*  
813 *Technology*, 2008, **17**, 216-233.
- 814 54. M. Thongngam and D. J. McClements, *Food Hydrocolloids*, 2005, **19**, 813-819.
- 815 55. S. Bajad, K. L. Bedi, A. K. Singla and R. K. Johri, *Planta Medica*, 2001, **67**,  
816 176-179.
- 817 56. A. Dahan and J. M. Miller, *Aaps Journal*, 2012, **14**, 244-251.
- 818 57. P. Fasinu, V. Pillay, V. M. K. Ndesendo, L. C. du Toit and Y. E. Choonara,  
819 *Biopharmaceutics & Drug Disposition*, 2011, **32**, 185-209.
- 820 58. A. des Rieux, V. Fievez, M. Garinot, Y. J. Schneider and V. Preat, *Journal of*  
821 *Controlled Release*, 2006, **116**, 1-27.
- 822 59. L. M. Ensign, R. Cone and J. Hanes, *Advanced drug delivery reviews*, 2012, **64**,  
823 557-570.
- 824 60. G. J. Doherty and H. T. McMahon, in *Annual Review of Biochemistry*, Annual  
825 Reviews, Palo Alto, 2009, vol. 78, pp. 857-902.
- 826 61. M. Bohdanowicz and S. Grinstein, *Physiol. Rev.*, 2013, **93**, 69-106.
- 827 62. E. Frohlich and E. Roblegg, *Toxicology*, 2012, **291**, 10-17.
- 828 63. J. J. Powell, N. Faria, E. Thomas-McKay and L. C. Pele, *J. Autoimmun.*, 2010, **34**,  
829 J226-J233.
- 830 64. S. Maher, D. J. Brayden, L. Feighery and S. McClean, *Crit. Rev. Ther. Drug Carr.*  
831 *Syst.*, 2008, **25**, 117-168.
- 832 65. F. Buyukozturk, J. C. Benneyan and R. L. Carrier, *Journal of Controlled Release*,  
833 2010, **142**, 22-30.
- 834 66. V. Gupta, B. H. Hwang, N. Doshi and S. Mitragotri, *Journal of Controlled Release*,  
835 2013, **172**, 541-549.
- 836 67. M. C. Chen, F. L. Mi, Z. X. Liao, C. W. Hsiao, K. Sonaje, M. F. Chung, L. W. Hsu and  
837 H. W. Sung, *Advanced drug delivery reviews*, 2013, **65**, 865-879.
- 838 68. V. Pillay, A. R. Hibbins, Y. E. Choonara, L. C. du Toit, P. Kumar and V. M. K.  
839 Ndesendo, *International Journal of Peptide Research and Therapeutics*, 2012,  
840 **18**, 259-280.
- 841 69. X. X. Wang, M. C. Valenzano, J. M. Mercado, E. P. Zurbach and J. M. Mullin,  
842 *Digestive Diseases and Sciences*, 2013, **58**, 77-87.
- 843 70. H. J. R. Lemmer and J. H. Hamman, *Expert Opin. Drug Deliv.*, 2013, **10**,  
844 103-114.
- 845 71. G. Di Colo, Y. Zambito and C. Zaino, *Journal of Pharmaceutical Sciences*, 2008,  
846 **97**, 1652-1680.
- 847 72. A. Yamamoto, H. Katsumi, K. Kusamori and T. Sakane, *Yakugaku Zasshi-J.*

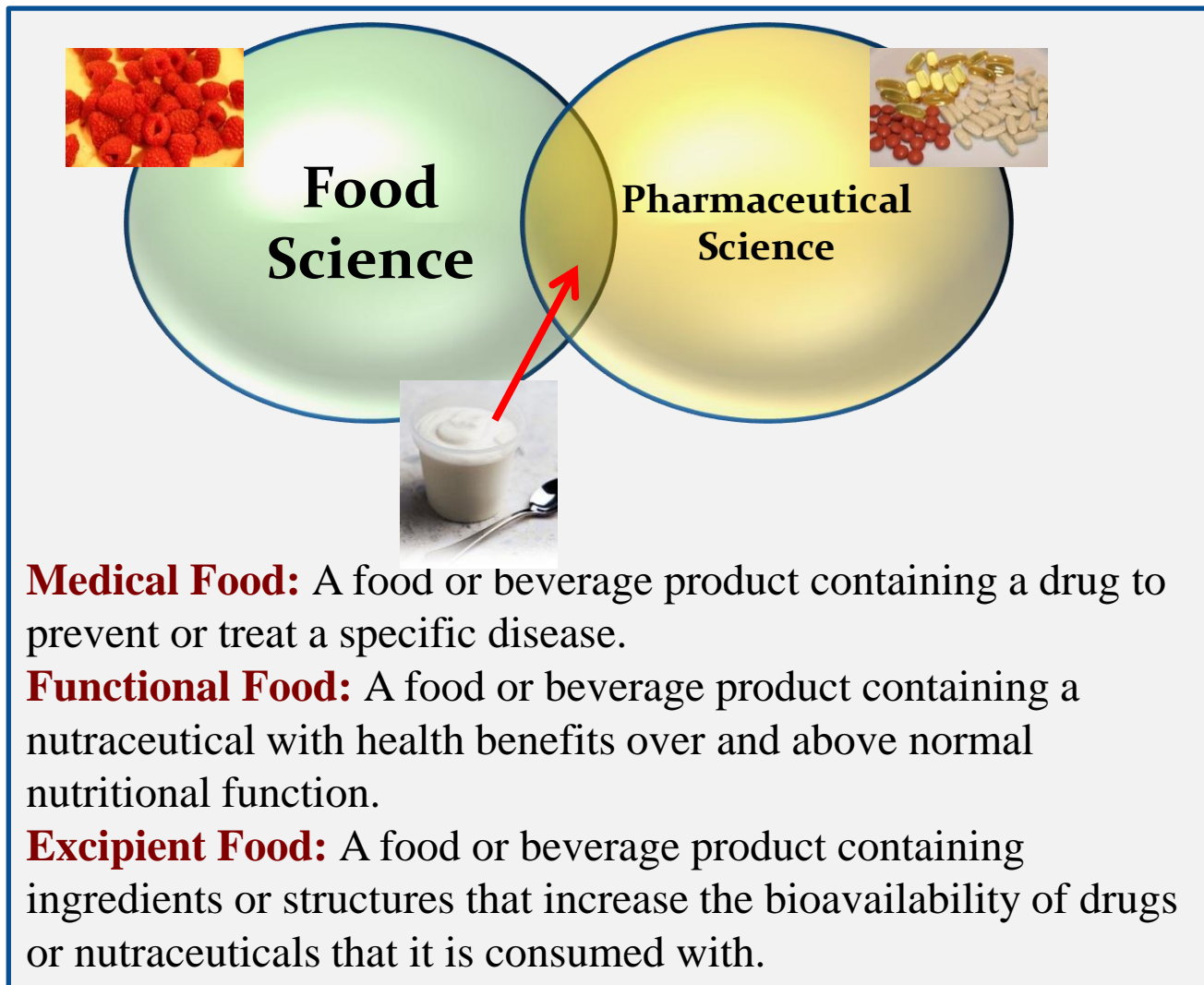
- 848 *Pharm. Soc. Jpn.*, 2014, **134**, 47-53.
- 849 73. L. F. Jiang, X. W. Long and Q. Meng, *Int. J. Pharm.*, 2013, **446**, 130-135.
- 850 74. P. P. Constantinides and K. M. Wasan, *Journal of Pharmaceutical Sciences*, 2007,  
851 **96**, 235-248.
- 852 75. J. M. Planas, I. Alfaras, H. Colom and M. E. Juan, *Archives of Biochemistry and*  
853 *Biophysics*, 2012, **527**, 67-73.
- 854 76. M. J. Jin and H. K. Han, *Journal of Food Science*, 2010, **75**, H93-H96.
- 855 77. J. X. Jia and K. M. Wasan, *J. Pharm. Pharm. Sci.*, 2008, **11**, 45-62.
- 856 78. Y. C. Chi, S. P. Lin and Y. C. Hou, *Toxicol. Appl. Pharmacol.*, 2012, **263**, 315-322.
- 857 79. J. S. Choi, B. C. Choi and K. W. Kang, *Pharmazie*, 2009, **64**, 49-52.
- 858 80. M. J. Kang, J. Y. Cho, B. H. Shim, D. K. Kim and J. Lee, *J. Med. Plants Res.*, 2009, **3**,  
859 1204-1211.
- 860 81. V. R. Challa, P. R. Babu, S. R. Challa, B. Johnson and C. Maheswari, *Drug Dev. Ind.*  
861 *Pharm.*, 2013, **39**, 865-872.
- 862 82. M. Tarvainen, A. Phuphusit, J. P. Suomela, A. Kuksis and H. Kallio, *J. Agric. Food*  
863 *Chem.*, 2012, **60**, 3564-3579.
- 864 83. E. A. Decker, B. Chen, A. Panya and R. J. Elias, in *Oxidation in Foods and*  
865 *Beverages and Antioxidant Applications, Vol 1: Understanding Mechanisms of*  
866 *Oxidation and Antioxidant Activity*, ed. E. A. Decker, R. J. Elias and D. J.  
867 McClements, 2011, pp. 225-248.
- 868 84. J. S. Choi, Y. J. Piao and K. W. Kang, *Arch. Pharm. Res.*, 2011, **34**, 607-613.
- 869 85. M. L. Failla, C. Chitchumroonchokchai and B. K. Ishida, *Journal of Nutrition*,  
870 2008, **138**, 482-486.
- 871 86. M. L. Failla, T. Huo and S. K. Thakkar, In vitro screening of relative  
872 bioaccessibility of carotenoids from foods, Taipei, TAIWAN, 2007.
- 873 87. V. Tyssandier, B. Lyan and P. Borel, *Biochimica Et Biophysica Acta-Molecular*  
874 *and Cell Biology of Lipids*, 2001, **1533**, 285-292.
- 875 88. T. Huo, M. G. Ferruzzi, S. J. Schwartz and M. L. Failla, *J. Agric. Food Chem.*, 2007,  
876 **55**, 8950-8957.
- 877 89. S. R. Goltz, W. W. Campbell, C. Chitchumroonchokchai, M. L. Failla and M. G.  
878 Ferruzzi, *Molecular Nutrition & Food Research*, 2012, **56**, 866-877.
- 879 90. C. Qian, E. A. Decker, H. Xiao and D. J. McClements, *Food Chemistry*, 2012, **135**,  
880 1440-1447.
- 881 91. J. Rao, E. A. Decker, H. Xiao and D. J. McClements, *Journal of the Science of Food*  
882 *and Agriculture*, 2013, **93**, 3175-3183.
- 883 92. D. J. McClements and Y. Li, *Advances in Colloid and Interface Science*, 2010,  
884 **159**, 213-228.
- 885 93. E. Troncoso, J. Miguel Aguilera and D. J. McClements, *Food Hydrocolloids*, 2012,  
886 **27**, 355-363.

- 887 94. P. Borel, V. Tyssandier, N. Mekki, P. Grolier, Y. Rochette, M. C.  
888 Alexandre-Gouabau, D. Lairon and V. Azais-Braesco, *Journal of Nutrition*, 1998,  
889 **128**, 1361-1367.
- 890 95. S. W. Cui, *Food Carbohydrates: Chemistry, Physical Properties and Applications*,  
891 Taylor and Francis, Boca Raton, FL, 2005.
- 892 96. J. N. BeMiller and K. C. Huber, in *Food Chemistry*, ed. S. Damodaran, K. L.  
893 Parkin and O. R. Fennema, CRC Press, Boca Raton, FL, Fourth Edition edn.,  
894 2008, ch. 4, pp. 83-154.
- 895 97. C. G. Biliaderis and M. S. Izydorczyk, *Functional Food Carbohydrates*, CRC  
896 Press, Boca Raton, FL., 2007.
- 897 98. M. Armand, *Sciences Des Aliments*, 2008, **28**, 84-98.
- 898 99. D. J. McClements, E. A. Decker and Y. Park, *Critical reviews in food science and*  
899 *nutrition*, 2009, **49**, 48-67.
- 900 100. M. V. Tzoumaki, T. Moschakis, E. Scholten and C. G. Biliaderis, *Food Funct.*,  
901 2013, **4**, 121-129.
- 902 101. T. Tsujita, H. Takaichi, T. Takaku, T. Sawai, N. Yoshida and J. Hiraki, *Journal of*  
903 *Lipid Research*, 2007, **48**, 358-365.
- 904 102. F. Fava, J. A. Lovegrove, R. Gitau, K. G. Jackson and K. M. Tuohy, *Curr. Med.*  
905 *Chem.*, 2006, **13**, 3005-3021.
- 906 103. S. Damodaran, in *Food Chemistry*, ed. S. Damodaran, K. L. Parkin and O. R.  
907 Fennema, CRC Press, Boca Raton, FL, Fourth Edition edn., 2008, ch. 5, pp.  
908 217-330.
- 909 104. G. O. Phillips and P. A. Williams, *Handbook of Food Proteins* Woodhead  
910 Publishing, Oxford, U.K., 2011.
- 911 105. M. C. O. Delgado, V. A. Tironi and M. C. Anon, *Lwt-Food Science and Technology*,  
912 2011, **44**, 1752-1760.
- 913 106. N. Bordenave, B. R. Hamaker and M. G. Ferruzzi, *Food Funct.*, 2014, **5**, 18-34.
- 914 107. D. M. Ribnicky, D. E. Roopchand, A. Oren, M. Grace, A. Poulev, M. A. Lila, R.  
915 Havenaar and I. Raskin, *Food Chemistry*, 2014, **142**, 349-357.
- 916 108. I. Depoortere, *Gut*, 2014, **63**, 179-190.
- 917 109. H. Singh and A. Sarkar, *Advances in Colloid and Interface Science*, 2011, **165**,  
918 47-57.
- 919 110. H. Singh, A. Q. Ye and D. Horne, *Progress in Lipid Research*, 2009, **48**, 92-100.
- 920 111. J. Maldonado-Valderrama, P. Wilde, A. Macierzanka and A. Mackie, *Advances in*  
921 *Colloid and Interface Science*, 2011, **165**, 36-46.
- 922 112. H. L. Yu and Q. R. Huang, *J. Agric. Food Chem.*, 2011, **59**, 9120-9126.
- 923 113. T. Tokle, Y. Mao and D. J. McClements, *Pharmaceutical Research*, 2013, **30**,  
924 3200-3213.
- 925 114. U. Lesmes, P. Baudot and D. J. McClements, *J. Agric. Food Chem.*, 2010, **58**,

- 926 7962-7969.
- 927 115. D. J. McClements, *Current Opinion in Colloid & Interface Science*, 2012, **17**,  
928 235-245.
- 929 116. I. Kralova and J. Sjoblom, *Journal of Dispersion Science and Technology*, 2009,  
930 **30**, 1363-1383.
- 931 117. D. J. McClements, E. A. Decker and Y. Park, *Critical reviews in food science and*  
932 *nutrition*, 2009, **49**, 48-67.
- 933 118. V. Delorme, R. Dhouib, S. Canaan, F. Fotiadu, F. Carriere and J. F. Cavalier,  
934 *Pharmaceutical Research*, 2011, **28**, 1831-1842.
- 935 119. Z. Vinarov, S. Tcholakova, B. Damyanova, Y. Atanasov, N. D. Denkov, S. D.  
936 Stoyanov, E. Pelan and A. Lips, *Langmuir*, 2012, **28**, 12140-12150.
- 937 120. Y. Li and D. J. McClements, *European Journal of Pharmaceutics and*  
938 *Biopharmaceutics*, 2011, **79**, 423-431.
- 939 121. E. Fernandez-Garcia, F. Rincon and A. Perez-Galvez, *J. Agric. Food Chem.*, 2008,  
940 **56**, 10384-10390.
- 941 122. B. Aspenstrom-Fagerlund, L. Ring, P. Aspenstrom, J. Tallkvist, N. G. Ilback and  
942 A. W. Glynn, *Toxicology*, 2007, **237**, 12-23.
- 943 123. R. Devraj, H. D. Williams, D. B. Warren, A. Mullertz, C. J. H. Porter and C. W.  
944 Pouton, *Int. J. Pharm.*, 2013, **441**, 323-333.
- 945 124. M. Hu, Y. Li, E. A. Decker and D. J. McClements, *Food Hydrocolloids*, 2010, **24**,  
946 719-725.
- 947 125. J. S. Patton and M. C. Carey, *Science*, 1979, **204**, 145-148.
- 948 126. J. S. Patton, R. D. Vetter, M. Hamosh, B. Borgstrom, M. Lindstrom and M. C.  
949 Carey, *Food Microstructure*, 1985, **4**, 29-41.
- 950 127. N. H. Zangenberg, A. Mullertz, H. G. Kristensen and L. Hovgaard, *European*  
951 *Journal of Pharmaceutical Sciences*, 2001, **14**, 237-244.
- 952 128. N. H. Zangenberg, A. Mullertz, H. G. Kristensen and L. Hovgaard, *European*  
953 *Journal of Pharmaceutical Sciences*, 2001, **14**, 115-122.
- 954 129. M. Armand, P. Borel, P. Ythier, G. Dutot, C. Melin, M. Senft, H. Lafont and D.  
955 Lairon, *Journal of Nutritional Biochemistry*, 1992, **3**, 333-341.
- 956 130. S. Hwang, S. Lee, I. S. Ahn and J. K. Yung, *Biocatal. Biotransform.*, 2009, **27**,  
957 290-295.
- 958 131. J. K. Lorenzen, S. Nielsen, J. J. Holst, I. Tetens, J. F. Rehfeld and A. Astrup,  
959 *American Journal of Clinical Nutrition*, 2007, **85**, 678-687.
- 960 132. T. Karupaiah and K. Sundram, *Nutrition and Metabolism*, 2007, **4**, 1-17.
- 961 133. K. E. Scholz-Ahrens and J. Schrezenmeir, *International Dairy Journal*, 2006, **16**,  
962 1399-1407.
- 963 134. M. Mukherjee, *Journal of Molecular Catalysis B-Enzymatic*, 2003, **22**, 369-376.
- 964 135. T. F. Whyne and J. M. Felts, *J. Am. Oil Chem. Soc.*, 1971, **48**, A101-&.

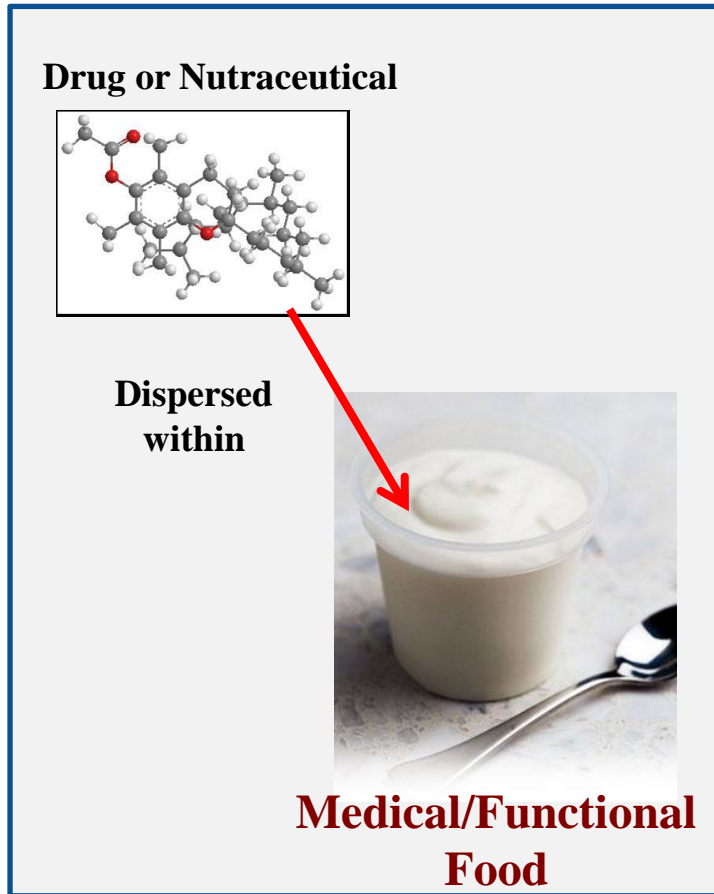
- 965 136. T. F. Whayne and J. M. Felts, *Circ.Res.*, 1971, **28**, 649-&.
- 966 137. H. Kimura, Y. Futami, S. Tarui and T. Shinomiya, *J. Biochem.*, 1982, **92**,  
967 243-251.
- 968 138. Y. Li and D. J. McClements, *Food Hydrocolloids*, 2011, **25**, 1025-1033.
- 969 139. T. Ranheim, A. Gedde-Dahl, A. C. Rustan and C. A. Drevon, *The Biochemical*  
970 *journal*, 1994, **303 ( Pt 1)**, 155-161.
- 971 140. F. Martel, R. Monteiro and C. Calhau, *Nutrition Research Reviews*, 2010, **23**,  
972 47-64.
- 973 141. K. Lohner, K. Schnabele, H. Daniel, D. Oesterle, G. Rechkemmer, M. Gottlicher  
974 and U. Wenzel, *Molecular Nutrition & Food Research*, 2007, **51**, 293-300.
- 975 142. S. F. Zhou, L. Y. Lim and B. Chowbay, *Drug Metab. Rev.*, 2004, **36**, 57-104.  
976

**Figure 1:** There is increasing convergence between the interests of the food and pharmaceutical industries in the development of products to prevent or treat diseases, particularly in the area of functional, medical, and excipient foods.

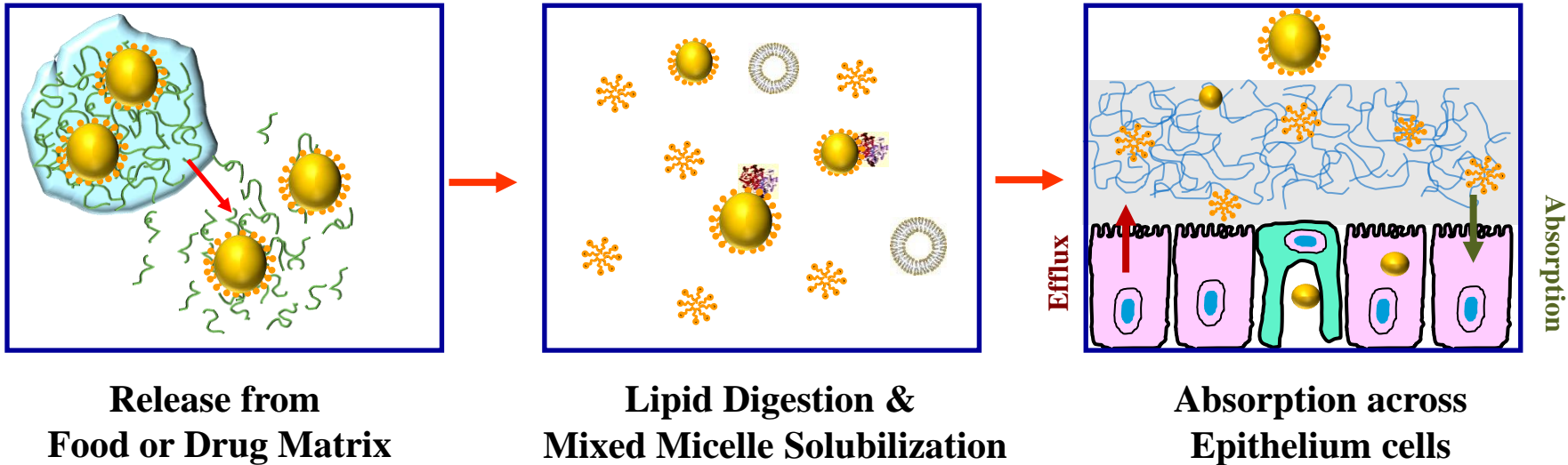




**Figure 2:** Schematic diagram of the difference between functional, medical, and excipient foods. The lipophilic bioactive component (pharmaceutical or nutraceutical) is usually encapsulated within the food matrix in medical and functional foods, but it is co-ingested with a different food matrix for excipient foods.

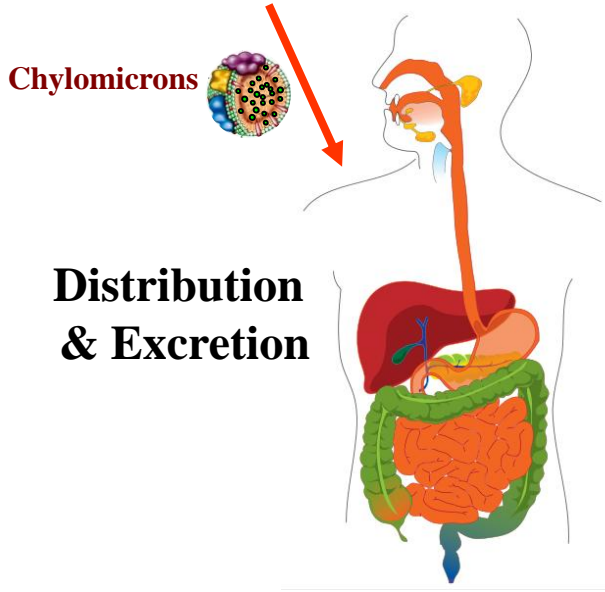


**Figure 3:** The oral bioavailability of a lipophilic bioactive agent depends on various liberation, absorption, distribution, metabolism, and excretion processes. Some of the key processes involved are shown schematically.

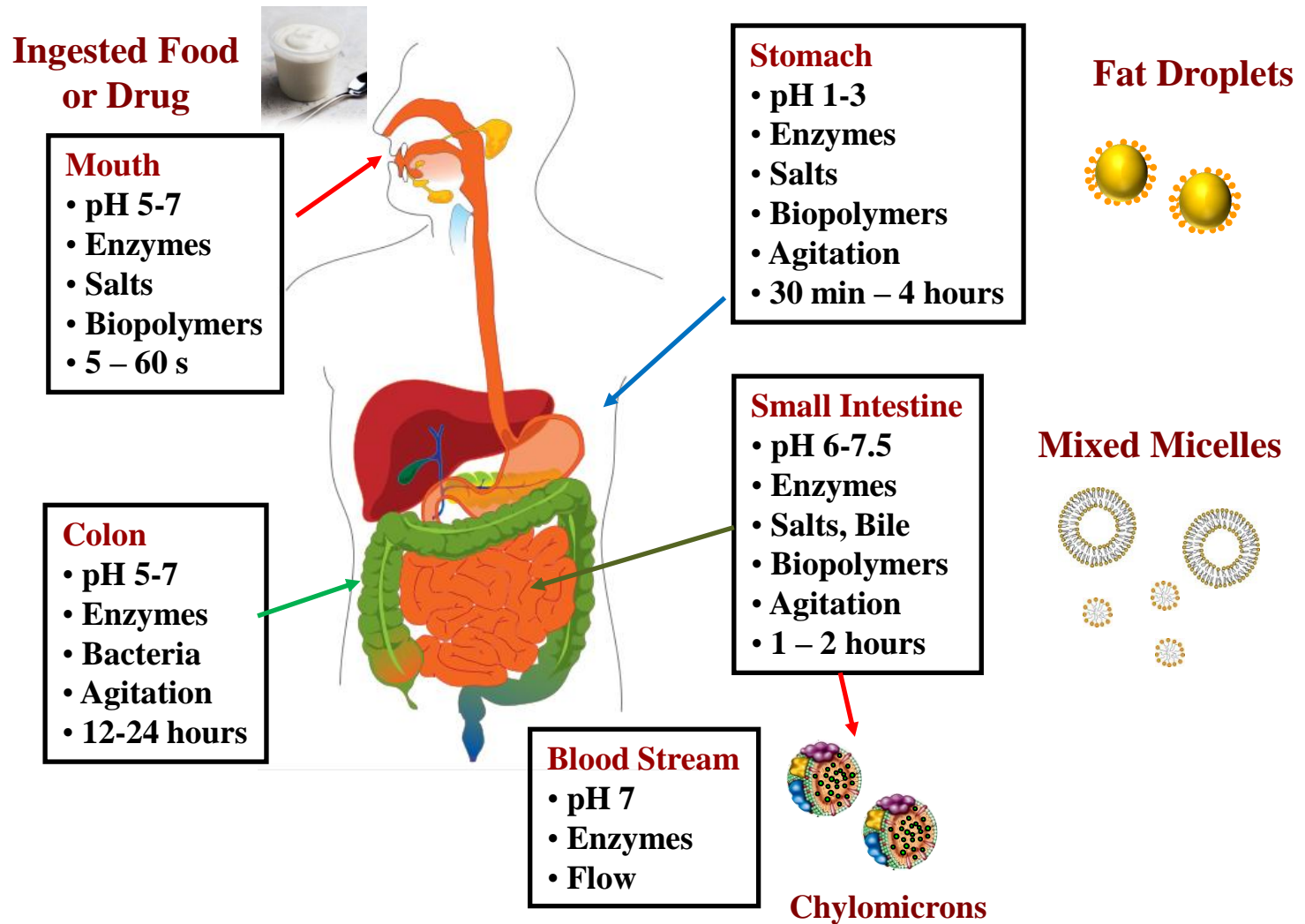


$$F = F_L \times F_A \times F_D \times F_M \times F_E$$

- $F_L$  = Fraction Liberated
- $F_A$  = Fraction Absorbed
- $F_D$  = Fraction Reaching Site of Action
- $F_M$  = Fraction Not Metabolized
- $F_E$  = Fraction Not Excreted



**Figure 4:** Schematic diagram of the physicochemical and physiological conditions in different regions of the human gastrointestinal tract that determine the liberation, absorption, metabolism and distribution of bioactives.



**Figure 5:** Bioactives in molecular form or trapped within small particles may penetrate through the mucus layer and be absorbed by epithelium cells by various mechanisms.

