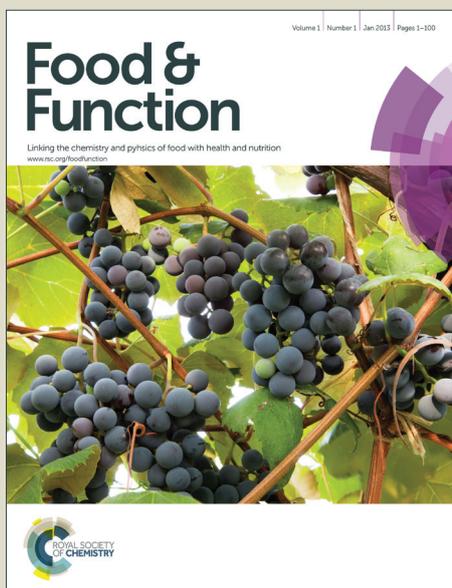


# Food & Function

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

# Absorption of Caffeine in Fermented Pu-er tea is Inhibited in Mice

Ye-wei Huang<sup>1,2,3</sup>, Huan-huan Xu<sup>2,3</sup>, Su-min Wang<sup>2,3</sup>, Yi Zhao<sup>1,2,3</sup>, Yu-min Huang<sup>2,3</sup>,  
Run-bo Li<sup>2,3</sup>, Xuan-jun Wang<sup>2,3\*</sup>, Shu-mei Hao<sup>4\*</sup> and Jun Sheng<sup>2,3\*</sup>

<sup>1</sup>College of Life Science, Jilin University, Changchun, 130012, People's Republic of China.

<sup>2</sup>Key Laboratory of Pu-erh Tea Science, Ministry of Education, Yunnan Agricultural University, Kunming, 650201, People's Republic of China.

<sup>3</sup>Tea research Center of Yunnan, Kunming, 650201, People's Republic of China.

<sup>4</sup>Yunnan University, Kunming, 650091, People's Republic of China.

## 1 **Abstract**

2 Caffeine present in number of dietary sources consumed worldwide.  
3 Although its pharmacokinetics has been intensively explored, little is known  
4 about complexed caffeine (C-CAF) in aqueous extraction of fermented Pu-er  
5 tea. The major components of C-CAF are oxidative tea polyphenols (OTP) and  
6 caffeine. Furthermore, the C-CAF can be precipitated in low pH solution. After  
7 administrating the same amount of total caffeine and comparing the peak level  
8 of plasma caffeine with the coffee (contains  $0.11 \pm 0.01$  % C-CAF) group, the  
9 results showed that the caffeine/OTP (contains  $66.67 \pm 0.02$  % C-CAF) group  
10 and the instant Pu-er tea (contains  $23.18 \pm 0.02$  % C-CAF) group were 33.39  
11 % and 25.86 % lower, respectively. The concentration of the metabolites of  
12 caffeine supports the idea that the absorption of the C-CAF was inhibited in  
13 mice. Congruent with this result, the amount of caffeine detected in mice  
14 excrement showed that more caffeine was eliminated in the caffeine/OTP  
15 group and the Pu-er tea group. The locomotor activity tests of mice  
16 demonstrated that the caffeine/OTP group and Pu-er tea group were more  
17 peaceful than coffee group. Our findings demonstrated that caffeine can be  
18 combined with OTP and the absorption of C-CAF is inhibited in mice, thus  
19 decreasing the irritation effect of caffeine. This may also be developing as a  
20 slow release formulation of caffeine.

21 **Key words:** absorption, caffeine, complexed caffeine, fermented Pu-er  
22 tea, oxidative tea polyphenols

## 23 Introduction

24 Caffeine (1, 3, 7-trimethylxanthine), a naturally occurring plant xanthine  
25 alkaloid<sup>1</sup>, was first isolated in the eighteenth century<sup>2</sup>. Caffeine is present in a  
26 number of dietary sources consumed worldwide (i.e., coffee, tea, soft drinks –  
27 cola, cocoa beverages, and chocolate bars)<sup>3-5</sup>. About 87 % of the world's  
28 population consumes caffeine with an average of 193 mg per day. Among  
29 adults 18 years and older, the daily intake ranges between 166 and 336 mg per  
30 day<sup>1,6,7</sup>. Caffeine is considered as a stimulant of the central nervous system for  
31 its ability to enhance alertness and can inhibit lipid absorption in  
32 ovariectomized rats<sup>2,4,8</sup>. The efficacy and side effects of caffeine have been  
33 extensively studied<sup>3,6,9-12</sup>, caffeine is generally recognized as safe by the Food  
34 and Drug Administration. However, excessive caffeine use can result in  
35 serious health problems and, in rare cases, death<sup>13,14</sup>.

36 One of the most common side effects of caffeine consumption is sleep  
37 loss. Some reported experiments about the negative effects of caffeine on  
38 sleep have consistently found that caffeine shortens total sleep time, prolongs  
39 sleep latency, and changes the duration of light and deep sleep<sup>15</sup>. Sleep loss is  
40 a major reason that people are reluctant to consume caffeinated beverages,  
41 namely tea and coffee. Fermented Pu-er tea is a unique fermented tea  
42 produced in Yunnan province of China<sup>16</sup>, it has been consumed by Chinese  
43 people for centuries and has recently become more popular functional  
44 beverage in Asian, Europe and America<sup>17,18</sup>. A mass of oxidative tea

45 polyphenols (OTP) in fermented Pu-er tea are produced during the  
46 fermentation process<sup>19,20</sup>. During the stage of fermentation, caffeine content  
47 has been found to increase<sup>21</sup>. Contrary to assumption, it is believed that the  
48 influence of fermented Pu-er tea on sleep is significantly less in comparison with  
49 other kinds of tea or coffee. Unlike coffee or other kinds of tea, many people  
50 can enjoy Pu-er tea before sleep.

51 The above discrepancy laid the groundwork for us to investigate the  
52 effects of caffeine in fermented Pu-er tea. We previously found that caffeine in  
53 fermented Pu-er tea can be divided into two kinds as complexed caffeine  
54 (C-CAF) and free caffeine (F-CAF), and that the C-CAF content can be  
55 evaluated by Low-pH Precipitate Method<sup>22</sup>. In this study, our primary objective  
56 was to compare the content of C-CAF in different kinds of tea and coffee  
57 samples, and then probe into the material basis of C-CAF formation. On  
58 account that mouse stomach acidity has a pH of about 2.0, we postulated that  
59 caffeine from Pu-er tea can be precipitated with other components in stomach  
60 when administered orally. Resultantly, the absorption of caffeine in Pu-er tea  
61 may be unavoidably influenced. To test this hypothesis, our second objective  
62 was to investigate the difference between the absorption of caffeine in coffee,  
63 fermented Pu-er tea and OTP. Our experiment results demonstrated that: one,  
64 fermented Pu-er tea has a larger amount of C-CAF than non-fermented Pu-er  
65 tea, green tea, black tea, oolong tea and coffee; two, C-CAF was principally  
66 combined with OTP; and three, the absorption of C-CAF in fermented Pu-er

67 tea was inhibited in mice and the effect of caffeine was influenced significantly.  
68 Our findings provide a theoretical basis for the phenomenon of the mitigation  
69 insomnia effects of fermented Pu-er tea in comparison with other kinds of tea  
70 or coffee.

## 71 **Materials and methods**

72 ***Chemicals and reagents.*** Caffeine was purchased from Aladdin.  
73 Theobromine was purchased from Shanghai Yuanye biological technology co.,  
74 LTD. Paraxanthine, theophylline and sodium dihydrogen phosphate were  
75 purchased from Sigma-Aldrich. 8-chlorotheophylline was purchased from  
76 Tokyo chemical industry co., LTD. Methanol used in the mobile phases were of  
77 HPLC-grade reagent purchased from Tedia Co. Inc.. Deionized water was  
78 prepared using the Purifier (FST-UV-20, Shanghai Fushite instrument  
79 equipment Co., Ltd. Shanghai, China) and was degassed by ultrasonic  
80 cleaning machine (SK-18TC, Shanghai Kudos ultrasonic instrument Co., Ltd.  
81 Shanghai, China).

82 ***Samples and preparation of HPLC infusion.*** Fourteen Chinese tea samples  
83 of various types, including three fermented Pu-er tea, two Non-fermented  
84 Pu-er tea, three black tea, two oolong tea, and four green tea, which were  
85 made into aqueous extract powder. Moreover, three coffee samples, one  
86 instant essence of Pu-er tea (Deepure, Yunnan tasly deepure biological tea  
87 group co., LTD.), one OTP, one TPS, one Pu-er tea theabrownin (PTTB), and  
88 one Pu-er tea polysaccharide (PTPS) sample were also made into aqueous

89 extract power. Supporting information of all samples is listed in **Table 1**. The  
90 preparation process of OTP, TPS, PTPS and PTTB was showed in the  
91 “Supplementary Experimental Procedures” of Electronic Supplementary  
92 Information.

93 Preparation of HPLC infusion: 800mg of the sample was dissolved in 40  
94 mL ultrapure water and was shocked by Vortex Mixer for 2 min and then was  
95 centrifuged at 800 ×g for 10 min. The resulting supernatant was diluted  
96 10-folds and filtered through a 0.45µm membrane and subjected to HPLC to  
97 determine the content of caffeine and its metabolites of the sample. In addition,  
98 the 20 g/L sample solution would be used for Low-pH Precipitate Method and  
99 Chloroform Extraction as described below.

100 **HPLC-VWD analysis.** A 50 µL sample was analyzed using an autosampler  
101 (G1329B, 1260ALS, Agilent, USA), a ultraviolet detector (G1314F, 1260VWD,  
102 Agilent, USA) at wavelength 274 nm, and HPLC pump (G1311B, 1260Quat  
103 Pump, Agilent, USA) at 0.6 mL/min flow at 30°C (G1316A, 1260TCC, Agilent,  
104 USA) through a C18 ODS column (ZORBAX SB-C18 4.6mm × 250mm, 5  
105 Micron, Agilent, USA) with a guard column (ZORBAX Eclipse Plus-C18,  
106 4-Pack, Analytical Guard Column, 4.6mm×12.5mm, 5-Micron, Agilent, USA).  
107 The mobile phase was 73 % 0.1 mol/L sodium dihydrogen phosphate and 27  
108 % methanol, filtered through a 0.45 micron filter. Concentrations of caffeine  
109 and its metabolites were determined in mg/L for all of the samples<sup>3</sup>.

110 **Determination of C-CAF content of the samples by Low-pH Precipitate**

111 **Method.** The C-CAF content of the samples was evaluated by Low-pH  
112 Precipitate Method as previously described<sup>23</sup>. In brief, the poly-phenolic  
113 hydroxyl contained components were precipitated under acidic conditions (pH  
114  $\leq 2$ ) and then dissolved with alkaline solution. Conveniently, if caffeine was  
115 combined with the Poly-phenolic hydroxyl contained components, the C-CAF  
116 content of the samples can be determined in the precipitate dissolved solution  
117 by HPLC.

118 Sample preparation: Hydrochloric acid was added to 12mL of 20 g/L  
119 sample solution for adjusting acidity to pH 1.5 and was shocked by Vortex  
120 Mixer for 2min and centrifuged at  $1600 \times g$  at  $25 \text{ }^\circ\text{C}$  for 30 min. The precipitate  
121 and supernatant were separated and the precipitate was weighed by sensitive  
122 balance. The precipitate was dissolved NaOH solution (pH = 10). Then, the  
123 supernatant and precipitate solution were diluted with deionized water to 13mL.  
124 The caffeine content of the supernatant and precipitate was determined by  
125 HPLC.

126 **Checkout of C-CAF content of the samples through chloroform**  
127 **extraction.** The linear regression equation of the balance line of caffeine  
128 distribution between water and chloroform was  $y = 15.02x$  at  $25 \text{ }^\circ\text{C}$ .  $X$  and  $y$   
129 mean that caffeine concentration (g/L) of aqueous phase and chloroform  
130 phase respectively when the distribution achieves equilibrium. The correlation  
131 coefficient  $R = 0.9942$ . It has no significant effect on the caffeine extraction  
132 between pH = 2.28 ~ 11.37 when  $x < 0.6 \text{ g/L}$ <sup>24</sup>. To checkout content of C-CAF

133 of the samples, we determined caffeine distribution ratio of the samples  
134 between chloroform phase and aqueous phase.

135 Sample preparation: 5mL chloroform was added to 5 mL of 20 g/L sample  
136 solution and was shocked by Vortex Mixer for 2 min and centrifuged at 800 ×g  
137 at 25 °C for 15 min. Then, the aqueous phase solution was diluted 10 folds with  
138 deionized water. The caffeine content was determined by HPLC.

139 **Mice.** Healthy virgin inbred BALB/c mice (7–8 weeks old) composed of male  
140 and female in equal numbers from Nanjing Peng-sheng biotechnology Co., Ltd,  
141 China, were used in the animal experiments. Mice were maintained in a  
142 controlled environment (12 h light/12 h dark cycle; humidity 50–60 %; ambient  
143 temperature 24°C ± 1°C) and were administered standard laboratory food and  
144 water *ad libitum*. All mice experiments were performed in the animal facility  
145 according to institutional guidelines and were approved by the  
146 Institutional Animal Care and Use Committee of Yunnan Agricultural University.  
147 Adverse events were not observed.

148 **Group designations and caffeine administration.** The BALB/c mice  
149 composed of male and female in equal numbers were divided at random into  
150 coffee group, fermented Pu-er tea group, and caffeine added OTP group.  
151 Coffee (85.91 g/L), fermented Pu-er tea (31.77 g/L) and OTP (31.77 g/L) with  
152 additional caffeine were prepared in deionized water (the caffeine content of  
153 the three samples are equivalent to 2.5 g/L) and administered via intragastric  
154 administration (0.2 mL/10g). The caffeine dose (50 mg/kg) was selected based

155 on its pharmacokinetic profile in mice compared with humans, which is well  
156 below the lethal dose of approximately 10 g in humans<sup>11</sup>. Caffeine doses were  
157 also based on the finding that 50 mg/kg of caffeine in rodents corresponds to  
158 approximately 10 cups of coffee for humans, but caffeine is metabolized up to  
159 6 times faster in mice than in humans<sup>5,25</sup>. The mice were feed-deprived for 12  
160 hours before the intragastric administration.

161 **Levels of caffeine and its metabolites in plasma.** Levels of caffeine and its  
162 metabolites in plasma were determined by HPLC as described by Youngberg<sup>3</sup>  
163 et al. with modifications. In brief, the blood samples were taken at various  
164 intervals (5, 10, 20, 40, 60, 120, 240, 360, 480 min) after the caffeine  
165 administration. As the internal standard, 3.33 mg/L of 8-chlorotheophylline was  
166 added to each sample. 150  $\mu$ l samples of subject plasma were extracted in 1  
167 mol/L HCl and 3 mL methylene chloride. The organic layer was evaporated to  
168 dryness and reconstituted in 0.1 mol/L HCl solution. Plasma levels of caffeine  
169 and its metabolites were determined by HPLC with the same chromatographic  
170 condition as “HPLC-VWD analysis” part. For caffeine, paraxanthine,  
171 theophylline and theobromine in plasma: the limit of quantitation (mg/L) was  
172 0.05, 0.10, 0.105 and 0.035; the limit of detection (mg/L) was 0.025, 0.045,  
173 0.060, 0.015; the response factor was 1.005, 0.456, 0.514 and 0.906; and the  
174 overall extraction recovery from human plasma was  $85.28 \pm 2.31$  %,  $57.13 \pm$   
175  $0.87$  %,  $60.10 \pm 0.39$  % and  $72.84 \pm 0.87$  %, respectively. The recovery for the  
176 internal standard (8-chlorotheophylline) was  $90.89 \pm 1.07$  %. The precision and

177 accuracy for the analytes were within the acceptable range (< 7 %). The assay  
178 validation was showed in the “supplemental data” of Electronic Supplementary  
179 Information.

180 **Excrement sample extraction.** The excrement samples were taken 8 hours  
181 after the caffeine administration, and then dried and ground to powder. 5 mL  
182 methanol (70 %) was added to a 200 mg powder sample, shocked by Vortex  
183 Mixer for 2min, and then extracted by ultrasound-assisted for 30 min at room  
184 temperature. Finally, the caffeine concentration was determined by HPLC.

185 **Evaluation of locomotor activity.** Locomotor activity was monitored with a  
186 mouse activity monitor system (ZZ-6, Taimeng, Chengdu Technology & Market  
187 Co. Ltd., Chengdu, China), which monitored the horizontal (locomotion) and  
188 vertical (rearing) movements of the mice. The individual compartments (L=15;  
189 W=12; H=10 cm) were put in a dimly lit and quiet room. The mice were  
190 feed-deprived for 12 hours before the experiment. Groups of female BALB/c  
191 mice were first thoroughly habituated to the test environment over a 30 min  
192 period. Then, they were removed from the open field, administrated with coffee,  
193 fermented Pu-er tea, and caffeine added OTP aqueous solution (which were  
194 equivalent to caffeine at 5 mg/kg) or water respectively via intragastric  
195 administration (0.2 mL/10g), and replaced in the compartments for an  
196 additional 90 min. The 5 mg/kg test dose of caffeine was chosen on the basis  
197 of previous dose-effect studies performed in our laboratory. This dose is a little  
198 higher than the threshold of stimulant effect of caffeine for the female mice.

199 Locomotor activity was recorded during the 90 min following the caffeine  
200 administration. The floor and walls of the chamber were cleaned with ethanol  
201 (70%) and dried with paper towels between each mouse exposure<sup>26-28</sup>.

202 **Statistical analyses.** All values were presented as mean  $\pm$  the standard error  
203 of the mean (SEM). Differences within groups were analyzed with repeated  
204 measures one-way ANOVA, Two-tailed  $p < 0.05$  was considered to be  
205 statistically significant. All analyses were performed using SPSS 17.0 (Chicago,  
206 IL, USA) and GraphPad Prism 5 (GraphPad Software, Inc., La Jolla, CA, USA).

## 207 Results

208 **C-CAF content of the samples.** Previous research has shown that tea  
209 contains theobromine and theophylline in addition to caffeine<sup>29</sup>. Therefore, in  
210 the current study, we set out to observe plasma levels of caffeine and its  
211 metabolites, by initially evaluating the content of caffeine, theophylline,  
212 theobromine and paraxanthine of the samples (**Supplemental Table 1**). There  
213 was a large amount of caffeine in the tea samples, whereas, there was slight  
214 amount of caffeine in the OTP. In addition, the tea and coffee samples contain  
215 small amounts of theophylline and theobromine. There was a relatively high  
216 level of theobromine in the fermented Pu-er tea samples, but only trace  
217 amounts of paraxanthine in the tea and coffee samples.

218 To evaluate and compare the C-CAF content of all samples, we next  
219 determined the C-CAF content by Low-pH Precipitate Method (**Table. 1**).  
220 Nearly a quarter of the caffeine in fermented Pu-er tea was C-CAF. What's

221 more, about three-quarters of the caffeine in the caffeine added OTP sample  
222 was C-CAF. Only about one-tenth of caffeine in the black tea samples,  
223 however, was C-CAF. In the non-fermented Pu-er tea, oolong tea, green tea  
224 and caffeine added TPS there was just a moderate level of C-CAF. Curiously,  
225 the C-CAF levels of PTPS and caffeine added PTTB were also high. In  
226 addition to fermented Pu-er tea, the C-CAF percent of the caffeine added OTP,  
227 PTPS and caffeine added PTTB samples were also obviously greater than the  
228 percent of precipitate volume (**Table. 1 and Supplemental Table 2**). These  
229 results indicate that, compared to the other samples, fermented Pu-er tea,  
230 caffeine added OTP, PTPS, and caffeine added PTTB samples contained  
231 more C-CAF, and caffeine in these samples abounded in the precipitate  
232 components; while there was hardly any C-CAF in the coffee samples.

233 To further characterize the combination of caffeine with other components  
234 of the samples, the fermented Pu-er tea, OTP and coffee samples were  
235 examined through Chloroform Extraction (**Fig. 1**). The OTP were mainly  
236 distributed in the aqueous phase. The caffeine concentration ratio of the  
237 Chloroform phase to aqueous phase was determined with significant  
238 differences ( $P < 0.001$ ) among the coffee, fermented Pu-er tea and OTP  
239 samples. For the fermented Pu-er tea and OTP samples, the balance of  
240 caffeine distribution between water and chloroform was shifted severely. The  
241 caffeine concentration ratio of the Chloroform phase to aqueous phase in  
242 descending order was as follows: coffee samples, fermented Pu-er tea

243 samples and OTP samples. These results indicate that caffeine in the OTP and  
244 fermented Pu-er tea samples was combined with OTP which belonged to the  
245 aqueous phase.

246 ***Effect of pH on the formation of caffeine contained precipitate.*** To further  
247 probe into whether or not fermented Pu-er tea aqueous extract or OTP can be  
248 precipitated in the condition of mouse stomach, we examined the association  
249 between the formation of precipitate and solution acidity (**Fig. 2 A, B**). Plenty of  
250 precipitate was formed at pH = 1.5 in the OTP and fermented Pu-er tea  
251 solution (**Fig. 2 A**) and the C-CAF percent of the OTP and fermented Pu-er tea  
252 samples was high (**Fig. 2 B**). Compared to the pH of 1.5, in spite of the  
253 precipitate quantity, the percentage of C-CAF was less at pH 3.5, but obvious  
254 precipitate was still formed. Consequently, the fermented Pu-er tea and OTP  
255 might be precipitated by internal conditions of the mouse stomach.

256 ***Plasma levels of caffeine and its metabolites.*** The results above suggest  
257 that the caffeine in fermented Pu-er tea was combined with OTP and the  
258 C-CAF may be precipitated in mouse gastric juice. Nearly the same as pure  
259 caffeine, there is hardly any C-CAF in coffee. Therefore, coffee was selected  
260 as a control group to investigate whether the absorption of caffeine was  
261 influenced by combining with OTP. Following the coffee, fermented Pu-er tea  
262 and caffeine added OTP administration, plasma levels of caffeine, and its  
263 metabolites were determined at different time points (**Fig. 3 A, B, C, D**).

264 Following the sample solutions administration, plasma levels were not

265 significantly different 5 or 10 min later. However, compared to the coffee group,  
266 plasma caffeine levels of the fermented Pu-er tea group and the caffeine  
267 added OTP group were significantly reduced at 20 min and 40 min after  
268 ingestion ( $P < 0.05$ ). Moreover, there were significant differences of plasma  
269 caffeine levels between the coffee group and the caffeine added OTP group at  
270 60 min and 120 min after ingestion ( $P < 0.01$ ). There were not significant  
271 differences between the coffee group and the fermented Pu-er tea group 40  
272 min after ingestion (**Fig. 3 A**).

273 For paraxanthine and theophylline, the plasma levels of the fermented  
274 Pu-er tea group and the caffeine added OTP group were higher than the coffee  
275 group in the early stage (5 min to 60 min). In contrast, the plasma levels of the  
276 two groups above were lower than the coffee group after 120 min (**Fig. 3 B, C**).  
277 Noteworthy, the plasma theobromine levels of the fermented Pu-er tea group  
278 were far higher than the other two groups before 360 min ( $P < 0.05$ ) (**Fig. 3 D**).  
279 Compared to the coffee group, the plasma theobromine levels of the caffeine  
280 added OTP group were higher in the early stage (5 min to 60 min), with the  
281 most significant difference occurring 10 min after ingestion ( $P < 0.01$ ).  
282 Interestingly, the plasma levels of theobromine of the caffeine added OTP  
283 group were also lower than the coffee group after 120 min.

284 ***Excrement levels of caffeine.*** Given that the absorption of C-CAF in  
285 fermented Pu-er tea and OTP was decreased within a prescribed time limit, we  
286 speculated that the absorption of C-CAF must be either delayed or inhibited in

287 mice. To ascertain the cause of this, the excrement levels of caffeine were  
288 examined (**Fig. 4**).

289 Excrement levels of caffeine from the fermented Pu-er tea group and  
290 caffeine added OTP group were significantly higher than that of the coffee  
291 group ( $P < 0.0001$ ). The result indicates that more caffeine was excreted in the  
292 fermented Pu-er tea group and the caffeine added OTP group compared to the  
293 coffee group. In all probability, the absorption of C-CAF was inhibited.

294 ***Locomotor activity levels.*** The result above suggested that caffeine can be  
295 combined with OTP, and the absorption of C-CAF can be inhibited in mice. To  
296 better understand the biological significance of that, we then performed  
297 another experiment for monitoring the locomotor activity of mice to elucidate  
298 the effects of C-CAF (**Fig. 5**).

299 The locomotor activity levels of the water and caffeine added OTP groups  
300 were lower than the coffee group, significantly ( $P < 0.05$ ). The fermented Pu-er  
301 tea group was also lower than the coffee group ( $P = 0.14$ ). These results  
302 indicate that the effect of caffeine in vivo was inhibited in the fermented Pu-er  
303 tea group and the caffeine added OTP group.

## 304 **Discussion**

305 To clarify the absorption of C-CAF in mice, it is necessary to discuss the  
306 total caffeine, C-CAF and caffeine metabolites content of the samples, and  
307 probe into which components can combine with caffeine. Previous studies<sup>34-36</sup>  
308 have reported that total sugar (14.47-27.62 %), caffeine (2.35-10.43 %) and

309 catechins (29.86-78.66 %) were found to be the main chemical components of  
310 tea cream in the green tea concentrate. Furthermore, the effect of caffeine on  
311 tea cream and haze of tea infusions is dependent on the presence of sufficient  
312 substances within pyrogallol groups<sup>37</sup>. Our previous study<sup>22</sup> also showed that  
313 the caffeine in fermented Pu-er tea can be categorized into F-CAF and C-CAF.  
314 However, fermented Pu-er tea contained trace free tea catechins. The major  
315 catechins were transformed to OTP due to the specific feature of the catechin  
316 – a strong anti-oxidant and easily oxidated in the fermentation process.  
317 Theaflavins, thearubigins and theabrownins are the main complex phenolic  
318 compounds deriving from the oxidation of catechins and their gallates during  
319 the fermentation process. Theaflavins undergo further oxidation during  
320 fermentation to form more polymerized thearubigins, and then condensed  
321 theabrownins<sup>18,38</sup>. The OTP still contained a large amount of benzene groups  
322 even though their chemical structure is not cleared, because the original  
323 catechins retain at least one benzene ring and the absorption curve of the OTP  
324 has characteristic peaks of benzene. The OTP was characterized as polymeric  
325 organic acid, which can be precipitated by adjusting the pH value to 2.  
326 Coincidentally, an increased level of C-CAF in tea is associated with the  
327 increased degree of oxidization or fermentation during the processing of tea.  
328 There is abundant OTP in fermented Pu-er tea, which is closely associated  
329 with a high level of C-CAF<sup>39</sup>. Accordant with previous studies<sup>22</sup>, the C-CAF  
330 content of fermented Pu-er tea is far higher than black tea, green tea, and

331 instant coffee.

332 In order to investigate the material basis of the C-CAF we attempted to  
333 separate PTPS and PTTB – two main components in Pu-er tea<sup>33</sup>. In doing so,  
334 we found that PTTB can be precipitated with PTPS during the alcohol  
335 sedimentation process, and PTPS and PTTB are difficult to separate. In  
336 essence, OTP is the principal component of the PTPS and PTTB samples. The  
337 significantly lower C-CAF content of PTTB to that of OTP may result from the  
338 fact that PTTB was mixed with other components, TPS for example. As  
339 another major component in the extraction of tea, TPS is a kind of  
340 poly-sacchride that may be, during the fermentation process, hydrolyzed to  
341 oligosaccharides and monosaccharides. In keeping with this interpretation,  
342 results of Chloroform Extraction for caffeine are consistent with the results of  
343 the Low-pH Precipitate Method. These results highlight the importance of OTP  
344 in the formation of C-CAF and demonstrated that OTP is the main material  
345 basis for the formation of C-CAF in fermented Pu-er tea.

346 As the major stimulant compound to human central nervous system<sup>30</sup>,  
347 caffeine is known to form complexes by self-association and with tea catechins.  
348 The Crystalline structure of the complexes of (-)-catechin (CA) and  
349 (-)-catechin-3-O-gallate (Cg) with caffeine have shown that CA formed a 1 : 1  
350 complex with caffeine by intermolecular hydrogen bonds, whereas Cg formed  
351 a 2 : 4 complex with caffeine by face-to-face and offset  $\pi$ - $\pi$  interactions and  
352 intermolecular hydrogen bonds<sup>31,32</sup>. The <sup>1</sup>H-NMR spectra data indicated that

353 the chemical shift for caffeine between reference sample (free caffeine) and  
354 the complex of OTP & caffeine sample changed little (Supplemental Figure 2,  
355 Supplemental Table 3 and Supplemental Figure 3). These results suggest that  
356 OTP and caffeine formed a complex by intermolecular forces but without  
357 covalent bonds combination.

358 Moreover, we should note that the effect of pH on tea solids extraction  
359 yield was significant. Total solids extracted between pH levels 2 to 11 varied  
360 from 270 to 290 g/kg tea, but figures were doubled at pH 1.1<sup>40</sup>. One can  
361 speculate that pH is also a key factor to the precipitate quantity of the  
362 fermented Pu-er tea and OTP aqueous solution. Considering that caffeine can  
363 be combined with OTP and can form a precipitate with OTP in a Low-pH  
364 solution, we sought to determine if the C-CAF can be precipitated in the  
365 condition of mouse stomach. For this purpose, we examined the precipitate  
366 quantity and percent of C-CAF of the samples at the pH similar to the stomach.  
367 The results showed that the caffeine in fermented Pu-er tea and OTP samples  
368 might be precipitated in the condition of mouse stomach. In addition, it's worth  
369 noting that the content of C-CAF in the coffee samples was very low.

370 After ascertaining the above results, we examined whether the absorption  
371 of the C-CAF was influenced in mice. To address this issue, we investigated  
372 the plasma levels of caffeine and its metabolites after the mice were  
373 administrated caffeine contained samples. In agreement with previous reports<sup>4</sup>,  
374 our study showed that the mice plasma caffeine concentration of the coffee

375 group peaked 20min after the ingestion of caffeine (50 mg/kg). In marked  
376 contrast, the plasma levels of caffeine of the fermented Pu-er tea and caffeine  
377 added OTP group varied marginally between 10 and 40 min, and were  
378 significantly lower than the coffee group. The simplest explanation for this  
379 observation is that the absorption of caffeine in fermented Pu-er tea and OTP  
380 was reduced, significantly.

381 Caffeine from coffee or other beverages is absorbed by the small intestine  
382 within 45 min of ingestion and moves through cellular membranes with the  
383 same efficiency that it is absorbed and circulated to tissues of the body<sup>41</sup>. It is  
384 eliminated by first-order kinetics<sup>42</sup>. Caffeine is metabolised by CYP enzymes  
385 (de-methylation), xanthine oxidase (formation of uric acid metabolites) and/or  
386 N-acetyltransferase (acetylation) in the liver, and through enzymatic action  
387 results in three metabolites: paraxanthine, theophylline, and theobromine<sup>43</sup>. In  
388 contrast to caffeine, the plasma levels of paraxanthine and theophylline in the  
389 fermented Pu-er tea group and caffeine added OTP group were higher than  
390 the coffee group in the early stage (5min to 60min), but were lower than the  
391 coffee group after 120 min. Nevertheless, the paraxanthine and theophylline  
392 intake from the fermented Pu-er tea group and caffeine added OTP group is  
393 not more than the coffee group (**Table 1**). One postulate for the above results  
394 is that some components (probably OTP) influenced the distribution or  
395 metabolism of caffeine in the mice. However, owing to more caffeine was  
396 absorbed and metabolized by the mice in the coffee group, the plasma levels

397 of paraxanthine and theophylline in the fermented Pu-er tea group and caffeine  
398 added OTP group were lower than the coffee group after 120 min. As a result  
399 of the dramatically higher theobromine content of fermented Pu-er tea in  
400 comparison to the coffee and OTP groups (**Table 1**), the fermented Pu-er tea  
401 group plasma levels of theobromine were far higher than the other two groups  
402 before 360min.

403 To assist in determining if the significantly lower plasma levels of caffeine  
404 found in the fermented Pu-er tea and caffeine added OTP group were the  
405 result of caffeine metabolism, we tested the mice's excrement levels of  
406 caffeine. Our findings confirmed that the absorption of C-CAF was inhibited in  
407 mice and was excreted, in part. The popularity of investigating caffeine has  
408 generated more interest from scientists of late due to the potentially harmful  
409 effects it has on our health<sup>2</sup>. Caffeine is a non-selective antagonist for  
410 adenosine receptors<sup>27</sup>. Several studies confirm that the activation of adenosine  
411 receptors A<sub>1</sub> and A<sub>2A</sub>, as well as the regulation of adenosine production and  
412 degradation, is essential for sleep induction and proper control of the  
413 sleep-wakefulness cycle<sup>44</sup>. The results of locomotor activity of mice indicate  
414 that the irritation effect of caffeine was inhibited when complexed with OTP.

415 To our knowledge, this is the first report that investigated the absorption of  
416 C-CAF in mice. Several main findings were obtained that might advance the  
417 theoretical framework by potentially illustrating why different sources of  
418 caffeine have different effects on humans. If the total caffeine intake is

419 consistent, the high level of C-CAF may be beneficial for people with sensitivity  
420 to caffeine to reduce the side effects of caffeine, such as insomnia. Moreover,  
421 this may be meaningful for developing some low F-CAF level products.

## 422 **Conclusions**

423 In conclusion, our results showed that OTP formed with caffeine to  
424 become the C-CAF contained in fermented Pu-er tea. The absorption of this  
425 C-CAF is inhibited in mice and may be the same in humans. What's more, the  
426 effect of caffeine was influenced significantly when it was combined with OTP.  
427 Taken together, this offers insight into the influences of fermented Pu-er tea on  
428 sleep loss are diminished in comparison with other forms of caffeine and/or  
429 caffeine containing products.

## 430 **Associated content**

### 431 **Abbreviations Used**

432 C-CAF, complexed caffeine; F-CAF, free caffeine; OTP, oxidative tea  
433 polyphenols; PTPS, Pu-er tea polysaccharide; PTTB, Pu-er tea theabrownin;  
434 TPS, tea polysaccharides.

### 435 **Acknowledgments**

436 J.S., S.H., X.W. and Y.H. designed the research; Y.H., H.X., S.W., Y.Z., Y.M.H.  
437 and R.L. conducted the research; Y.H., H.X. and S.W. analyzed the data; Y.H.  
438 performed statistical analysis and wrote the manuscript; and J.S., S.H. and  
439 X.W. had primary responsibility for final content. All authors read and approved  
440 the final manuscript. The financial supports are the Natural Science

441 Foundation of Yunnan Province, China (no. 2012FB151) and foundation of  
442 Scientific and Technical Innovation of Yunnan Province, China (no.  
443 2009AB021).

#### 444 **Supplementary Material**

445 Refer to Web version on PubMed Central for the preparation process of OTP,  
446 TPS, PTPS and PTTB; the content of caffeine, theophylline, theobromine and  
447 paraxanthine of the samples (Supplemental Table 1); the percent of precipitate  
448 volume of the samples (Supplemental Table 2); “Assay Validation” for Levels of  
449 caffeine and its metabolites in plasma (Supplemental Figure 1); and <sup>1</sup>H NMR  
450 spectra data for caffeine (Supplemental Figure 2, Supplemental Table 3 and  
451 Supplemental Figure 3).

#### 452 **Author information**

##### 453 **Corresponding Author**

454 \*(X.W.) Phone: +8615912579655. Fax: +8687165226711. E-mail:  
455 wangxuanjun@gmail.com; haosm@sina.com (S. H.); shengj@ynau.edu.cn (J.  
456 S.).

#### 457 **Notes**

458 The authors declare no competing financial interest.

## References

1. Addicott, M. A.; Yang, L. L.; Peiffer, A. M.; Burnett, L. R.; Burdette, J. H.; Chen, M. Y.; Hayasaka, S.; Kraft, R. A.; Maldjian, J. A.; Laurienti, P. J. The effect of daily caffeine use on cerebral blood flow: How much caffeine can we tolerate? *Hum. Brain Mapp.* 2009, 30, 3102-3114.
2. Ma, Z. L.; Qin, Y.; Wang, G.; Li, X. D.; He, R. R.; Chuai, M.; Kurihara, H.; Yang, X. Exploring the caffeine-induced teratogenicity on neurodevelopment using early chick embryo. *PLoS One* 2012, 7, e34278.
3. Youngberg, M. R.; Karpov, I. O.; Begley, A.; Pollock, B. G.; Buysse, D. J. Clinical and physiological correlates of caffeine and caffeine metabolites in primary insomnia. *J Clin. Sleep Med.* 2011, 7, 196-203.
4. Alvi, S. N.; Hammami, M. M. Validated HPLC method for determination of caffeine level in human plasma using synthetic plasma: application to bioavailability studies. *J. Chromatogr Sci.* 2011, 49, 292-296.
5. Fredholm, B. B.; Battig, K.; Holmen, J.; Nehlig, A.; Zvartau, E. E. Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev.* 1999, 51, 83-133.
6. Jura, Y. H.; Townsend, M. K.; Curhan, G. C.; Resnick, N. M.; Grodstein, F. Caffeine intake, and the risk of stress, urgency and mixed urinary incontinence. *J. Urol.* 2011, 185, 1775-1780.
7. Laughon, S. K.; Powers, R. W.; Roberts, J. M.; Parana, S.; Catov, J. Caffeine and insulin resistance in pregnancy. *Am. J. Perinatol.* 2011, 28, 571-578.
8. Wang, S.; Noh, S. K.; Koo, S. I. Epigallocatechin gallate and caffeine differentially inhibit the intestinal absorption of cholesterol and fat in ovariectomized rats. *J. Nutr.* 2006, 136, 2791-2796.
9. Lucas, M.; Mirzaei, F.; Pan, A.; Okereke, O. I.; Willett, W. C.; O'Reilly, E. J.; Koenen, K.; Ascherio, A. Coffee, caffeine, and risk of depression among women. *Arch. Intern. Med.* 2011, 171, 1571-1578.
10. Lu, Y. P.; Lou, Y. R.; Peng, Q. Y.; Nghiem, P.; Conney, A. H. Caffeine decreases phospho-Chk1 (Ser317) and increases mitotic cells with cyclin B1 and caspase 3 in tumors from UVB-treated mice. *Cancer Prev. Res. (Phila)* 2011, 4, 1118-1125.
11. Lusardi, T. A.; Lytle, N. K.; Szybala, C.; Boison, D. Caffeine prevents acute mortality after TBI in rats without increased morbidity. *Exp. Neurol.* 2012, 234, 161-168.
12. Sharmin, S.; Guan, H.; Williams, A. S.; Yang, K. Caffeine reduces 11beta-hydroxysteroid dehydrogenase type 2 expression in human trophoblast cells through the adenosine A(2B) receptor. *PLoS One* 2012, 7, e38082.
13. Temple, J. L. Caffeine Use in Children: What we know, what we have left to learn, and why we should worry. *Neurosci Biobehav Rev.* 2009, 6, 793-806.
14. Gandhi, K. K.; Williams, J. M.; Menza, M.; Galazyn, M.; Benowitz, N. L. Higher serum caffeine in smokers with schizophrenia compared to smoking controls. *Drug Alcohol Depend* 2010, 110, 151-155.
15. Byrne, E. M.; Johnson, J.; McRae, A. F.; Nyholt, D. R.; Medland, S. E.; Gehrman, P. R.; Heath, A. C.; Madden, P. A.; Montgomery, G. W.; Chenevix-Trench, G.; Martin, N. G. A

- genome-wide association study of caffeine-related sleep disturbance: confirmation of a role for a common variant in the adenosine receptor. *Sleep* 2012, 35, 967-975.
16. Xie, G.; Ye, M.; Wang, Y.; Ni, Y.; Su, M.; Huang, H.; Qiu, M.; Zhao, A.; Zheng, X.; Chen, T.; Jia, W. Characterization of pu-erh tea using chemical and metabolic profiling approaches. *J. Agric. Food Chem.* 2009, 57, 3046-3054.
  17. Abe, M.; Takaoka, N.; Idemoto, Y.; Takagi, C.; Imai, T.; Nakasaki, K. Characteristic fungi observed in the fermentation process for Puer tea. *Int. J. Food Microbiol.* 2008, 124, 199-203.
  18. Zhang, L.; Li, N.; Ma, Z. Z.; Tu, P. F. Comparison of the chemical constituents of aged pu-erh tea, ripened pu-erh tea, and other teas using HPLC-DAD-ESI-MSn. *J. Agric. Food Chem.* 2011, 59, 8754-8760.
  19. Zhao, L.; Jia, S.; Tang, W.; Sheng, J.; Luo, Y. Pu-erh Tea Inhibits Tumor Cell Growth by Down-Regulating Mutant p53. *Int. J. Mol. Sci.* 2011, 12, 7581-7593.
  20. Xu X.; Yan M.; Zhu Y. Influence of Fungal Fermentation on the Development of Volatile Compounds in the Puer Tea Manufacturing Process. *Eng.Life.Sci.* 2005, 5, 382-386.
  21. Wang, X.; Hu, S.; Wan, X.; Pan, C. Effect of microbial fermentation on caffeine content of tea leaves. *J. Agric. Food Chem.* 2005, 53, 7238-7242.
  22. Song, S.; Huang, Y.; Wang X.; Yu H.; Fang, C.; Sheng J.; Hao, S. Determination of Conjugated Caffeine in Tea by Low-pH Precipitation Method. *J.Tea Sci.* 2013, 33, 322-326.
  23. Wang T.; Wang, X.; Huang Y.; Fang, C.; Zhu, Q.; Yang, J.; Sheng, J.; Hao, S. Effect of fermentation on content of free caffeine in Puer tea. *Chin. J. Biologicals* 2013, 26, 509-511.
  24. Hu, X.; Dai, W.; Wang X.; Guo, Z.; Liao, B.; Bai, R. Study of extraction process of caffeine by chloroform. *J. Tsinghua University(Sci and Tech)* 1995, 35, 13-19.
  25. Gulick, D.; Gould, T. J. Effects of ethanol and caffeine on behavior in C57BL/6 mice in the plus-maze discriminative avoidance task. *Behav. Neurosci.* 2009, 123, 1271-1278.
  26. El Yacoubi, M.; Ledent, C.; Menard, J. F.; Parmentier, M.; Costentin, J.; Vaugeois, J. M. The stimulant effects of caffeine on locomotor behaviour in mice are mediated through its blockade of adenosine A(2A) receptors. *Br. J. Pharmacol.* 2000, 129, 1465-1473.
  27. Okuro, M.; Fujiki, N.; Kotorii, N.; Ishimaru, Y.; Sokoloff, P.; Nishino, S. Effects of paraxanthine and caffeine on sleep, locomotor activity, and body temperature in orexin/ataxin-3 transgenic narcoleptic mice. *Sleep* 2010, 33, 930-942.
  28. Zancheta, R.; Possi, A. P.; Planeta, C. S.; Marin, M. T. Repeated administration of caffeine induces either sensitization or tolerance of locomotor stimulation depending on the environmental context. *Pharmacol. Rep.* 2012, 64, 70-77.
  29. Hiroshi Ashihara, F. M. G. a. A. C. Metabolism of Caffeine and Related Purine Alkaloids in Leaves of Tea.(*Camellia sinensis* L.). *Plant Cell Physiol.* 1997, 38, 413-419.
  30. Jobstl, E.; Fairclough, J. P.; Davies, A. P.; Williamson, M. P. Creaming in black tea. *J. Agric. Food Chem.* 2005, 53, 7997-8002.
  31. Yin, J.; Xu, Y.; Yuan, H.; Luo, L.; Qian, X. Cream formation and main chemical components of green tea infusions processed from different parts of new shoots. *Food Chem.* 2009, 114, 665-670.
  32. Xu, Y. Q.; Chen, S. Q.; Yuan, H. B.; Tang, P.; Yin, J. F. Analysis of cream formation in green

- tea concentrates with different solid concentrations. *J. Food Sci. Technol.* 2012, 49, 362-367.
33. Liang, Y.; Xu, Y. Effect of extraction temperature on cream and extractability of black tea [*Camellia sinensis* (L.) O. Kuntze]. *Int. J. Food Sci. Technol.* 2003, 1, 37-45.
34. Zhao, H.; Zhang, M.; Zhao, L.; Ge, Y. K.; Sheng, J.; Shi, W. Changes of constituents and activity to apoptosis and cell cycle during fermentation of tea. *Int. J. Mol. Sci.* 2011, 12, 1862-1875.
35. Tan C.; Guo, G.; Li, B.; Zhou, H.; Gong, J. Physico-chemical and Spectral Properties of Theabrownin from Pu-erh Tea. *Chemistry and Industry of Forest Products* 2010, 30, 53-58.
36. Wang, Q.; Peng, C.; Gong, J. Effects of enzymatic action on the formation of theabrownin during solid state fermentation of Pu-erh tea. *J. Sci. Food Agric.* 2011, 91, 2412-2418.
37. Buscariollo, D. L.; Breuer, G. A.; Wendler, C. C.; Rivkees, S. A. Caffeine acts via A1 adenosine receptors to disrupt embryonic cardiac function. *PLoS One* 2011, 6, e28296.
38. Tsutsumi, H.; Kinoshita, Y.; Sato, T.; Ishizu, T. Configurational studies of complexes of various tea catechins and caffeine in crystal state. *Chem. Pharm. Bull (Tokyo)* 2011, 59, 1008-1015.
39. Sato, T.; Kinoshita, Y.; Tsutsumi, H.; Yamamoto, H.; Ishizu, T. Characterization of creaming precipitate of tea catechins and caffeine in aqueous solution. *Chem. Pharm. Bull (Tokyo)* 2012, 60, 1182-1187.
40. Liang, Y.; Xu, Y. Effect of pH on cream particle formation and solids extraction yield of black tea. *Food Chemistry* 2001, 74, 155-160.
41. Anthony Liguori, J. R. H., Jacob A. Grass Absorption and Subjective Effects of Caffeine from Coffee, Cola and Capsules. *Pharmacology Biochemistry and Behavior* 1997, 58, 721-726.
42. Newton R, B. L., Lind MJ, Morrison PJ, Rogers HJ, Bradbrook ID Plasma and salivary pharmacokinetics of caffeine in man. *Eur. J. Clin. Pharmacol.* 1981, 21, 45-52.
43. Anthe S. Zandvliet, A. D. R. H., Milly E. de Jonge, Rob den Hoed, Rolf W . Sparidans, Vincent M. Hendriks, Wim van den Brink, Jan M. van Ree and Jos H. Beijnen Population Pharmacokinetics of Caffeine and its Metabolites Theobromine, Paraxanthine and Theophylline after Inhalation in Combination with Diacetylmorphine. *Basic & Clinical Pharmacology & Toxicology* 2005, 96, 71-79.
44. Mazzotti, D. R.; Guindalini, C.; Pellegrino, R.; Barrueco, K. F.; Santos-Silva, R.; Bittencourt, L. R.; Tufik, S. Effects of the adenosine deaminase polymorphism and caffeine intake on sleep parameters in a large population sample. *Sleep* 2011, 34, 399-402.

TABLE 1. The content and percent of C-CAF of the samples<sup>1</sup>

sample type	sample name	F-CAF, mg/L	C-CAF, mg/L	C-CAF, %
fermented pu-erh tea	Gold bud tribute pu-erh tea	1716.13±0.93	486.54±3.81	22.09±0.14
	TAETEA pu-erh tea (7592)	1318.77±0.48	378.47±2.68	22.30±0.13
	Brick pu-erh tea	1370.85±0.22	267.63±1.13	16.33±0.06
non-fermented pu-erh tea	Moonlight White pu-erh tea	2530.20±0.50	60.23±0.47	2.32±0.02
	Bangwei pu-erh tea	1179.72±1.25	11.27±0.20	0.94±0.02
black tea	Changning black tea(first grade)	1100.19±1.41	115.78±0.64	9.52±0.04
	MAKEMY black tea	1830.36±1.22	261.60±0.52	12.50±0.02
	Dianhong black tea	1443.38±0.11	118.37±0.49	7.58±0.03
oolong tea	Tieguanyin oolong tea	1033.37±7.51	6.29±0.47	0.60±0.04
	Da Hung Pao oolong tea	1460.97±4.83	69.08±0.51	4.52±0.05
green tea	Longjing green tea	1474.15±0.46	13.63±0.24	0.92±0.02
	Biluochun green tea	1399.68±2.39	20.78±0.84	1.46±0.06
	Huilong green tea	1387.57±4.29	8.80±0.06	0.63±0.00
	Yunnan green tea	1411.13±1.57	10.91±0.27	0.77±0.02
coffee	Nescafe coffee	550.58±0.40	0.63±0.06	0.11±0.01
	Maxwell House coffee	664.36±0.19	0.98±0.02	0.15±0.00
	Yunnan Pasteral coffee	702.01±0.45	0.76±0.03	0.11±0.00
instant fermented pu-erh tea	Deepure instant essence of pu-erh tea	1162.04±0.65	350.63±0.62	23.18±0.02
OTP	OTP	45.92±0.44	146.95±0.61	76.19±0.16
PTPS	PTPS	262.13±0.73	165.80±2.06	38.74±0.34
mixture of TPS and caffeine	caffeine added TPS	1085.51±0.99	34.92±0.07	3.12±0.01
mixture of OTP and caffeine	caffeine added OTP	415.83±0.58	831.85±0.42	66.67±0.02
mixture of PTTB and caffeine	caffeine added PTTB	825.93±0.23	317.33±0.24	27.76±0.02
caffeine	caffeine	1234.68±0.63	0.70±0.05	0.06±0.00

<sup>1</sup>Data were presented as mean ± SEM of three independent experiments. C-CAF, complexed caffeine; F-CAF, free caffeine; OTP, oxidative tea polyphenols; PTPS, Pu-er tea polysaccharide; PTTB, Pu-er tea theabrownin; TPS, tea polysaccharides.

Figures

FIGURE 1

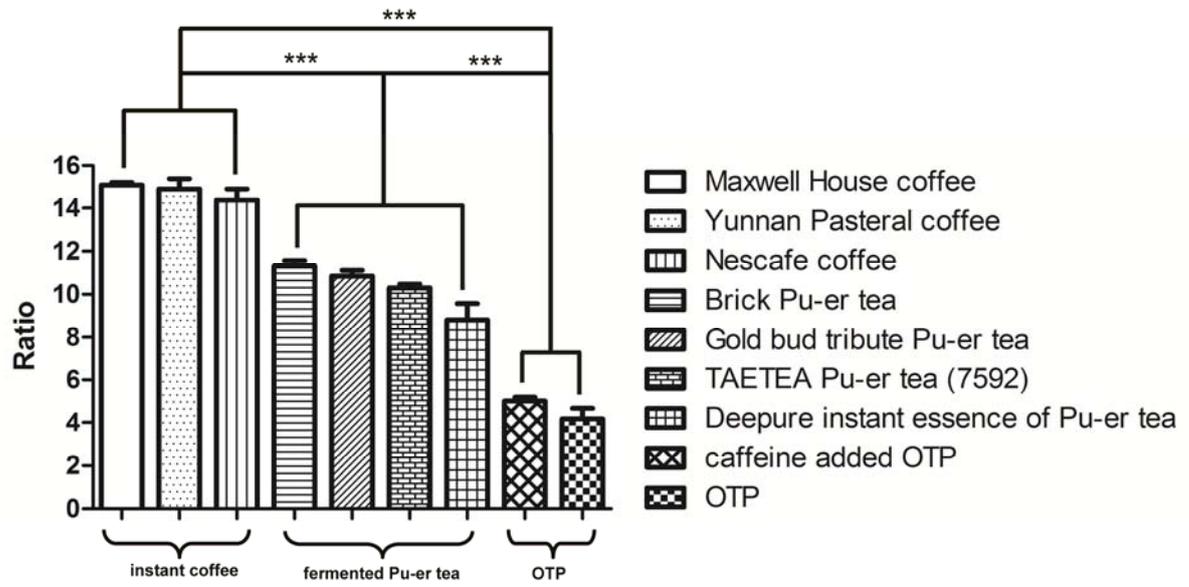
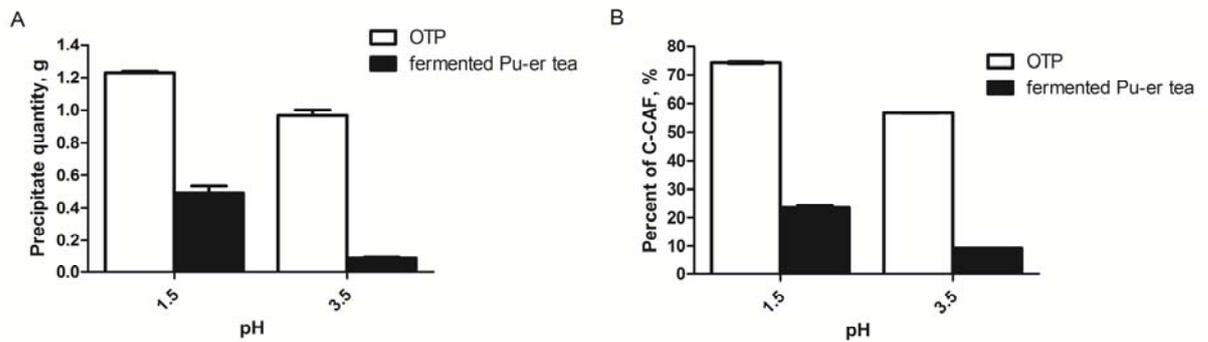


FIGURE 2



Food & Function Accepted Manuscript

FIGURE 3

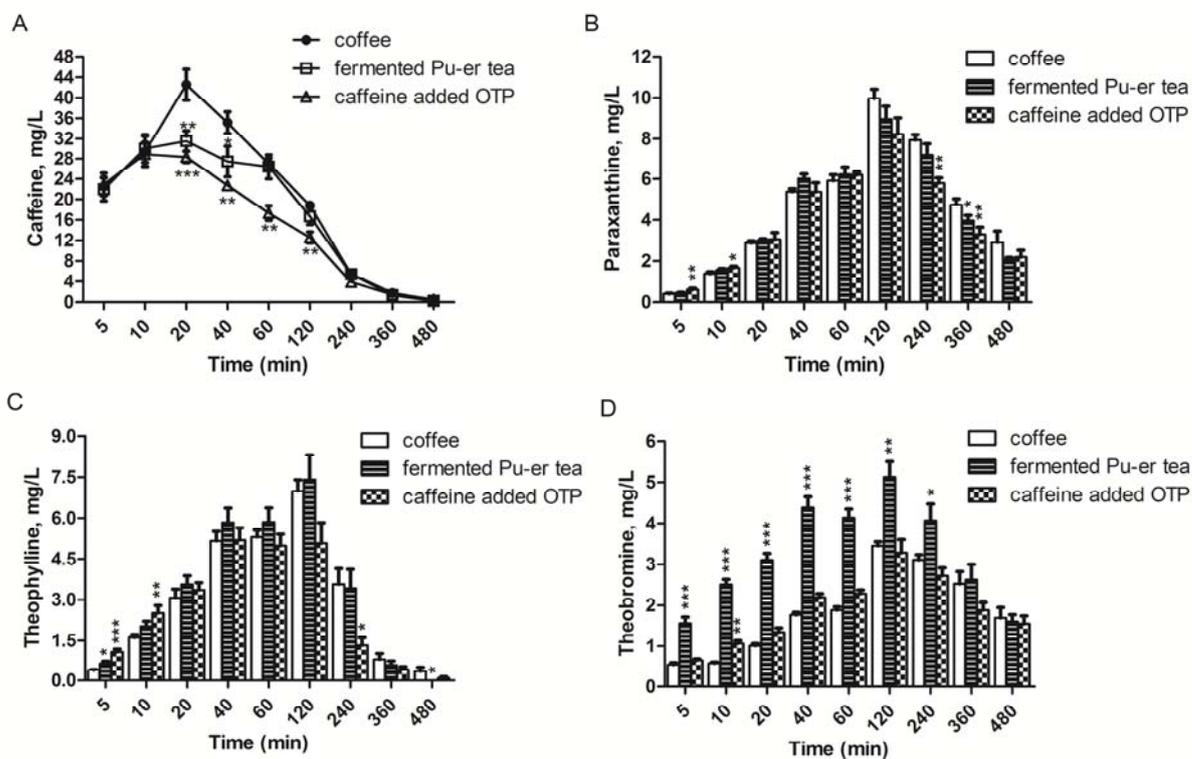


FIGURE 4

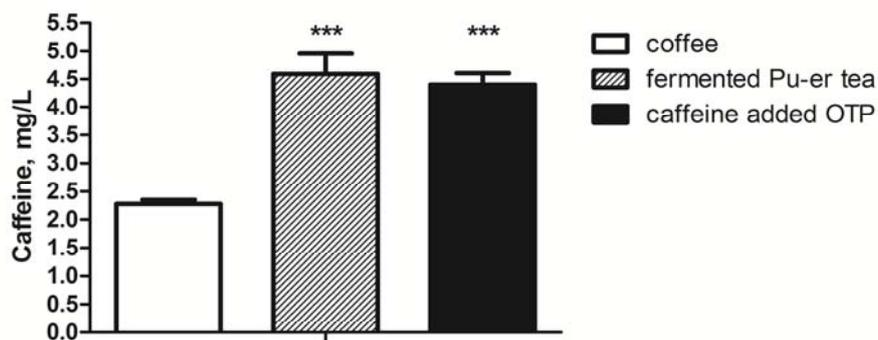
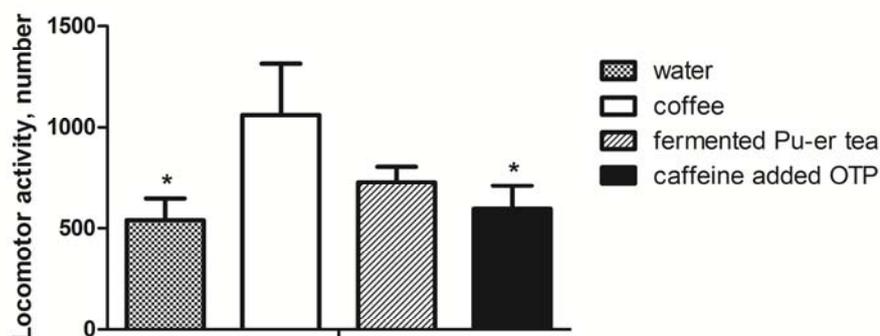


FIGURE 5



### Figure captions

**FIGURE 1. Caffeine concentration ratio of Chloroform phase to aqueous phase.**

Data were presented as mean  $\pm$  SEM and were analyzed with repeated measures (n=4) one-way ANOVA, Two-tailed  $p < 0.05$  was considered to be statistically significant (\*\* $P < 0.001$  vs. coffee samples). OTP, oxidative tea polyphenols.

**FIGURE 2. Effect of pH on the formation of precipitate.** Data were presented as mean

$\pm$  SEM and were analyzed with repeated measures (n=3) one-way ANOVA. OTP, oxidative tea polyphenols.

**FIGURE 3. Plasma levels of caffeine (A), paraxanthine (B), theophylline (C) and theobromine (D).** Data were presented as mean  $\pm$  SEM. Differences within groups at

different time points were analyzed with repeated measures (n=6) one-way ANOVA,

Two-tailed  $p < 0.05$  was considered to be statistically significant. (\* $P < 0.05$ , \*\* $P < 0.01$  and

\*\*\* $P < 0.001$  vs. coffee group). OTP, oxidative tea polyphenols.

**FIGURE 4. Excrement levels of caffeine.** Data were presented as mean  $\pm$  SEM. Differences within groups were analyzed with repeated measures (n=9) one-way ANOVA, Two-tailed  $p < 0.05$  was considered to be statistically significant. (\*\* $P < 0.001$  vs. coffee samples). OTP, oxidative tea polyphenols.

**FIGURE 5. Locomotor activity levels after caffeine administration.** Data were presented as mean  $\pm$  SEM. Differences within groups were analyzed with repeated measures (n=6) one-way ANOVA, Two-tailed  $p < 0.05$  was considered to be statistically significant. ( $P < 0.05$ , vs. coffee group). OTP, oxidative tea polyphenols.