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ARTICLE TYPE

Depth-selective microscopic observation of photomobile liquid crystal polymer under UV illumination

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By using the depth selective imaging method, we studied the UV induced change in a photomobile liquid crystalline polymer film. With 1 µm depth resolution, each slice inside the film was selectively observed. A network-like structure mixed with the ordered and disordered regions of molecules in the middle of the 10 film, and a rubbed polymer layer at the bottom of the film were observed. In each slice of the film, the phase change induced by UV light was observed strongly dependent on the director direction, which indicates the ordering change of the liquid crystalline molecules in the director direction. It took several tens of seconds for the ordering change caused by the collaborative interaction between the molecules. Furthermore, it was suggested that the UV induced change travelled from the bottom layer to the middle 15 layer on the micron order.

Introduction

Liquid crystalline polymers have promising properties for various applications due to both the properties of liquid crystals (LCs) and polymers. Molecular alignment can be controlled by stimuli 20 such as electric field, while elasticity is given by polymerization. In cross-linked LC polymers, the orientation of mesogens is strongly coupled with the conformation of the polymer chains, and macroscopic deformation can be induced by heat, electricity and light. 1, 2 When the photochromic molecules are incorporated 25 into the cross-linked LC polymers, the deformation was controlled by light. By introducing azobenzene as a photochromic molecule, the macroscopic motion is controllable by the combination of the UV and visible lights, because the trans isomer is photo-isomerized by UV light and it is returned back by 30 a visible light. By coupling the photo-isomerization and the surrounding polymer, not only contraction or expansion,³ but also three-dimensional motion such as a film bending4, 5 have been demonstrated. Furthermore, a plastic motor, ⁶ an inch-size worm, ⁷ and a robotic arm⁸ only by using light have been developed.

The overall mechanism for the three-dimensional motions was understood that the surface region of the film was contracted more than the bottom side of it by the excess UV absorption in the surface region.4 However, the understanding from the viewpoint of the molecular interactions is still under investigation, 40 although much effort has been made for it. 9 In the photomobile films in motion, the initial nanometer-scale change due to the photo-isomerization should be expanded into the micron- or milli- scale region via the collaborative interactions between molecules, polymer chains, domains. This kind of spatial 45 expansion of the molecular interactions is difficult to understand because both temporal and spatial changes range over many

orders. In the previous paper, 10 we studied the UV-induced dynamics from the viewpoint of the temporal change, using the heterodyne transient grating method, which gives the dynamics 50 from a few nanoseconds to a hundred seconds after a single pulse-light irradiation. 11, 12 Since the refractive index change was in this method, it is suitable for LC or polymer samples because they typically have no optical absorption bands in the visible wavelength region.^{13, 14} We could observe the initial film 55 contraction on the order of several hundreds of nanoseconds, the subsequent reorientation and molecular rotation dynamics from a few microseconds to a hundred milliseconds, and the return of the cis isomer to the trans isomer on tens of seconds. Furthermore, we figured out that the molecular orientation by the UV 60 irradiation was highly anisotropic under the conditions preferred for the mobility.10

In this study, we studied the dynamics from the viewpoint of the spatial change. For depth-selective measurements, we used speckle correlation reflection phase microscopy (SpeCRPM), ¹⁵ 65 with the depth resolution of about 1 µm and a field of view up to about hundred micron. The phase microscopy is suitable for the observation of the LC polymer, because the structural or orientational change induce the variation in the refractive index. Furthermore, the depth selective observation and the wide field of 70 view of SpeCRPM is beneficial for the LC polymer sample, because it is consisted of different layers with an inhomogeneous structure inside on the order of nano- to micron- scale and the UV induced change could be observed at a video rate. Here we studied on the polarization dependent change induced by UV 75 irradiation using SpeCRPM, and also the structure was studied with a Raman microscopy.

Experimental

The general principle and setup of SpeCRPM was described in detail recently, 15 and a brief outline is given. In this technique, a time-varying speckle-field is used as an illumination source. By constructing Linnik-type configuration, identical speckle-fields 5 are introduced on two arms of the interferometry. The detection port of the microscope is based on the off-axis geometry offering single-shot imaging capability. By using the digital holographic imaging method complex electric field images, amplitude and phase, for objects can be retrieved. When the speckle-field is 10 changing fast enough within a camera acquisition time, a number of independent speckle patterns are averaged out leaving clear interference fringes overlaying object images. The strength of the interference is highly sensitive to the optical path length difference introduced between the two interferometric arms due 15 to the 3-D nature of the speckle-field. The interference quickly decorrelated along the optical axis of the microscope allowing us the confocal equivalent axial resolution, 1.03 µm. More details about SpeCRPM can be found in Ref. [15].

A mode-locked Ti: Sapphire laser was used as a light source, 20 which has a center wavelength of 800 nm and a spectral bandwidth of 17 nm. The speckle-field was circular polarized on the sample plane, thus each polarization component was selectively measured using a polarizer. The UV light source was a UV curing device (UV75, Thorlabs, <40 mW), which was fixed 25 at the side of the sample, and the irradiation intensity was about 10 mW/cm^2 .

A custom-built Raman NIR confocal Raman microscopv^{16, 17} was also used to study the molecular orientation in the film. Briefly, 785nm wavelength from Ti: Sapphire laser (3900S, 30 Spectra-Physics) was used as an excitation source. High-speed XY scanning was performed by the galvanometer mirrors (CT-6210, Cambridge Technology). A 1.2 NA water immersion objective lens (UPLSAPO60XWIR 60X/1.20, Olympus) was used to both focus the laser light onto the sample and to collect 35 the back-scattered light. The axial resolution was around 2 µm. Linear polarizer was inserted in the beam path to control the polarization. From the film, 100×100 spectra were acquired from $150 \mu m \times 150 \mu m$ area with 15mW power and the integration time was 50ms/pixel. The total measurement time was 40 approximately 8.5 minutes...

The sample preparation procedure was described in the previous paper, 10 and it is briefly explained here. The monomer molecule was 6-[4-(4-hexyloxyphenylazo)phenoxy]hexylacrylate (A6AB6), and 4,4'-bis[6-(acryloyloxy)hexyloxy]azobenzene 45 (DA6AB) was used as a crosslinker. (Supporting Information, Fig.S1) The monomer and crosslinker were solved in chloroform. After that, Irgacure®784(2 mol%) was added as an initiator for polymerization, and chloroform was removed by evaporation. The mixture sample was solved at 110 °C and put in a LC cell (15 50 mm × 20 mm), which has an alignment polyimide layer with a thickness of a few microns on the internal side of the cell. The temperature was decreased to 88 °C, and it was polymerized by irradiating a high-pressure mercury lamp (USHIO, OPM2-502HQ) with the wavelength > 540 nm with the intensity of 2 55 mW/cm² for 2 hrs. The final thickness of the film was 16 μm. For the microscopic measurements, only the top glass was peeled off from the LC cell to get a better image. After taking it out from the cell, it was confirmed that it could be bent under a UV

illumination typically at 10 mW/cm², and returns back to the 60 original position by illumination of a visible light (> 540 nm) typically at 40 mW/cm².

Results and Discussions

First, the sample was scanned by SpeCRPM in the depth direction, and the typical result is shown in Fig. 1. The sample 65 was imaged from the top surface to the internal slices downward with a step of 1 μm. There were debris and scratches on the top surface. Further going into the film, we could find domain-like structure in the region of 5-15 microns typically. In the 15-20 micron region, striped structure in the same direction as the 70 director was observed.

The striped shape around the 15-20 micron region is supposed to be an alignment polymer layer because of the agreement with its rubbed direction and the depth region observed. The domainlike structure in the 5-15 micron region was observed everywhere 75 in the film and the size ranged from a few to tens of microns. Since the light scattering was higher at such domain positions and also they were not dependent on the polarization direction of the illumination light, it is supposed that the domains were composed of the disordered region of molecules.

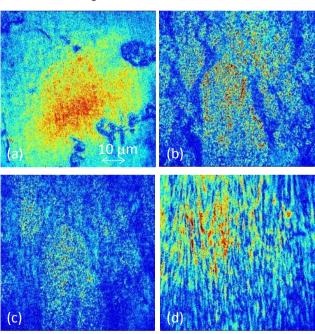
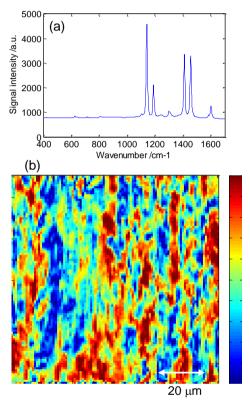


Fig.1 Depth scan of a photomobile liquid crystalline polymer film by using SpeCRPM. The images shown here correspond to those at 0, 8.0, 12.0 and 18.4 µm from the top surface for (a), (b), (c) and (d), respectively.

To investigate the domain-like structure and its ordering condition, the polarized Raman measurements were performed for the same sample. A typical Raman spectrum is shown in Fig.2(a). Based on the assignments reported, 18, 19 most of the 90 strong peaks, 1140, 1188, 1410, 1455, 1602 cm⁻¹ were assigned to aromatic CH ring mode. By measuring Ix and Iy (Ix, Iy: the Raman intensities when excited and detected in x-polarized or ypolarized directions, respectively), denoting x and y as the parallel and perpendicular directions to the director axes, we

evaluated the molecular anisotropy in the film by calculating, $||\mathbf{l}_{x} - \mathbf{l}_{y}|/(|\mathbf{l}_{x} + \mathbf{l}_{y})$, which is shown in Fig.2(b). The images obtained by using the different peaks showed almost the same images. (Supporting Information, Fig.S2) We could find 5 inhomogeneous distribution of anisotropic and isotropic regions with their sizes similar as the domain-like structures observed by SpeCRPM. The scan in the wider area with 120 microns square revealed that the structure looked like a network structure aligned in the director axis. The isotropic regions correspond to the 10 domain-like structure observed by SpecRPM. It is considered that the film has a network-like structure consisted of two different regions; isotropically and anisotropically aligned molecules, and that each region was likely to be aligned in the director axis.



15 Fig.2 (a) A Raman spectrum for a photomobile liquid crystalline polymer film. (b) Anisotropy imaging of the film using the peak at 1140 cm⁻¹, with a scan step of 1 µm with a field of view of 120x120 µm².

Next, we observe the dynamic change in the images by UV 20 irradiation at the depth position at 8.0 and 18.4 µm from the surface. Hereinafter, the layers were called a domain layer and alignment layer, respectively. The UV light was irradiated for the initial 1 min, and the subsequent change was observed for another 4 min. The results are shown in Fig.3. In both the domain and 25 alignment layers, apparent change was observed when the image was observed under the polarization parallel with the director direction, while only a slight change was observed for the polarization in the perpendicular direction. (Compare with the images in Supporting Information, Fig.S3)

At the alignment layer, the image change was observed throughout the image 30 seconds after starting the UV irradiation, while the change at the domain layer was observed 60 seconds

after it. Looking at the domain layer carefully, the image change was mainly observed in the region except the domain-like islands. 35 Furthermore we could confirm the shape of stripes after a few minutes,(Fig.3 top (e)-(h)) which was very similar structure as observed at the alignment layer. (Easily noticed by the videos in Supporting Information.)

The strong polarization dependence of the image change 40 suggests that the signal enhancement was caused by the ordering of molecular alignment, not by the isotropic change such as the thermal expansion. Since it is known that the photoisomerization of azobenzne itself occurs very fast on the picosecond order, ^{20, 21} the image change was not caused by the structure change in 45 azobenzene, and the alignment of LC polymer network would be the origin of the image change, which would take tens of seconds, judging from the time lag for the image change after the UV irradiation. The delay of the image enhancement for the domain layer compared to that for the alignment layer indicates that the 50 molecular ordering initially occurred at the alignment layer and then the ordering was travelled to the upper layer. This is supported by the fact that the structure observed after a few minutes showed a similar stripe pattern in the domain layer as observed in the alignment layer.

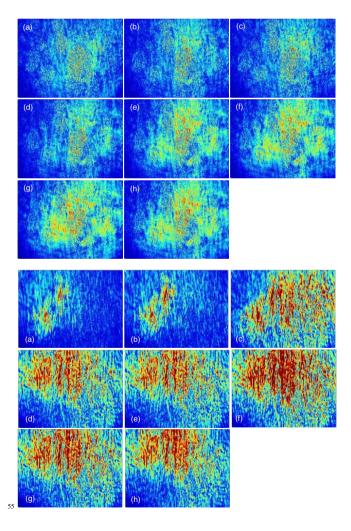


Fig.3 Sequences of the images of a photomobile liquid crystalline polymer film by irradiation of UV light, and the above and bottom sequences correspond to the images at 8.0 and 18.4 μm from the top surface. (a)-(h) corresponds to the images, 0, 15, 30, 45, 60, 90, 120, 180, 5 240 seconds after the UV irradiation was started, which was topped after 60 seconds.

Conclusions

We studied the UV induced change in the photomobile LC polymer films using the microscopic techniques with a depth resolution about 1 µm. The observed change at each slice of the depth suggested a network-like structure inside, which responded anisotropically to the UV light tens of seconds after the irradiation, and the change stayed a while on minute order. Since the rubbed surface at the alignment layer, which was the bottom of the layer, was gradually observed even in the middle of the film, it is suggested that the collaborative interaction travelled inside the film on the order of microns. This kind of collaborative molecular interactions is very important in the devices using LCs and polymers, and the microscopic observation with a depth resolution and with a high speed detection would give much insight of the collaborative interactions, which would beneficial for the development of new devices.

Notes and references

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- † Electronic Supplementary Information (ESI) available: [The molecular structure of the monomer and crosslinker, the Raman imaging using the other peaks, the SpeCRPM images for the polarization perpendicular with the director are shown. The movies corresponding to Fig.3 can be found online.]. See DOI: 10.1039/b000000x/
 - 1. R. Zentel, 1989, -1, -329.
 - 2. S. M. Kelly, Journal of Materials Chemistry, 1995, 5, 2047-2061.
- 3. H. Finkelmann, E. Nishikawa, G. G. Pereira and M. Warner, *Physical Review Letters*, 2001, **87**, 015501.
- 4. Y. Yu and T. Ikeda, *Journal of Photochemistry and Photobiology C: Photochemistry Reviews*, 2004, **5**, 247-265.
- 5. T. Seki, *Polym J*, 2004, **36**, 435-454.
- M. Yamada, M. Kondo, J. I. Mamiya, Y. L. Yu, M. Kinoshita, C. J.
 Barrett and T. Ikeda, Angewandte Chemie-International Edition, 2008, 47, 4986-4988.
 - M. Yamada, M. Kondo, R. Miyasato, Y. Naka, J.-i. Mamiya, M. Kinoshita, A. Shishido, Y. Yu, C. J. Barrett and T. Ikeda, *Journal of Materials Chemistry*, 2009, 19, 60-62.
- M. Yamada, M. Kondo, J.-i. Mamiya, Y. Yu, M. Kinoshita, C. J. Barrett and T. Ikeda, Angewandte Chemie, 2008, 120, 5064-5066.
 - 9. J.-i. Mamiya, *Polym J*, 2013, **45**, 239-246.
- 10. T. Fujii, S. Kuwahara, K. Katayama, K. Takado, T. Ube and T. Ikeda, *Physical Chemistry Chemical Physics*, 2014, **16**, 10485-10490.
- 11. M. Okuda and K. Katayama, *Chemical Physics Letters*, 2007, **443**, 158-162.

- M. Okuda and K. Katayama, Journal of Physical Chemistry A, 2008, 112, 4545-4549.
- 60 13. T. Chiba, H. Inoue, S. Kuwahara and K. Katayama, Journal of Photochemistry and Photobiology a-Chemistry, 2013, 266, 1-5.
 - M. Arai, T. Fujii, H. Inoue, S. Kuwahara and K. Katayama, Analytical Sciences, 2013, 29, 401-404.
- C. Youngwoong, P. Hosseini, W. Choi, R. R. Dasari, P. T. C. So, and
 Z. Yaqoob, *Optics Letters*, 39, 6062-6065.
- J. W. Kang, N. Lue, C.-R. Kong, I. Barman, N. C. Dingari, S. J. Goldfless, J. C. Niles, R. R. Dasari and M. S. Feld, *Biomedical Optics Express*, 2011, 2, 2484-2492.
- 17. J. W. Kang, F. T. Nguyen, N. Lue, R. R. Dasari and D. A. Heller,
 Nano Letters, 2012, **12**, 6170-6174.
- 18. F. Lagugné Labarthet and C. Sourisseau, *Journal of Raman Spectroscopy*, 1996, **27**, 491-498.
- F. L. Labarthet, J. L. Bruneel, T. Buffeteau and C. Sourisseau, *The Journal of Physical Chemistry B*, 2004, 108, 6949-6960.
- 75 20. T. Fujino, S. Y. Arzhantsev and T. Tahara, The Journal of Physical Chemistry A, 2001, 105, 8123-8129.
 - 21. N. Tamai and H. Miyasaka, *Chemical Reviews*, 2000, **100**, 1875-