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The synthesis of novel, chignolin-derived peptides comprising the azobenzene photoswitch [3-(3-aminomethyl)phenylazo]-phenylacetic acid (AMPP) is reported. Reversible photoswitching behavior led to folding into β-hairpin-like structures, as unequivocally demonstrated by CD, FT-IR and NMR spectroscopy.

Understanding the mechanisms, by which particular sequences of amino acids are folded into well-defined three-dimensional protein structures, is a challenging problem in the field of molecular biology. The complexity of possible conformational transitions and different timescales of folding prevents accurate predictions and simulations for larger amino acid ensembles. Moreover, it is believed that certain key secondary motifs, e.g., β-hairpins and β-sheets, serve as nucleation sites for protein folding, providing insight into the early events of secondary structure formation. In particular β-hairpin-forming peptides have received much attention as model systems for both experimental and theoretical studies of the initial folding steps. For instance, light-triggered folding of two amino acids strands into a β-hairpin peptide has been investigated by means of azobenzene photoswitches, such as [3-(3-aminomethyl)phenylazo]phenylacetic acid (AMPP). Upon incorporation into the peptide backbone, the latter offers the possibility to control the hairpin structure by initiating a reversible folding (cis-form) or unfolding (trans-form) transition.

Herein, we present a peptidomimetic model system for photo-controlled reversible β-hairpin formation based on chignolin and AMPP. The decapeptide chignolin (GYDPETGTWG) was designed by Honda et al. on the basis of the central part of the GB1 hairpin and is the smallest β-hairpin known to be stable in solution. As such and in combination with ultrafast initiation of structural changes upon photoswitching, chignolin derivatives are particularly attractive targets for folding studies of protein nuclei. Therefore, the peptide AzoChig1 (GYDP-AMPP-GTWG) was designed, in which the two central amino acids Glu5 and Thr6 of chignolin’s four-residue turn sequence were substituted by AMPP (Fig. 1). Besides, peptidomimetic AzoChig2 (GYDP-AMPP-GT(5FW)G) with a 5-fluoro-L-tryptophan (5FTrp, 5FW) residue instead of Trp9 was synthesized to enhance key hydrophobic interactions between aromatic Tyr2 and Trp9. Fluorinated amino acid residues are known to be tolerated by a variety of proteins without introducing much steric perturbation and usually favor protein folding and stability due to their increased hydrophobicity. For instance, the hydrophobicity of 5FTrp, derived from 1-octanol/water partitioning experiments, is significantly higher than that of native Trp. However, systematic studies towards structure-guiding effects of fluorinated amino acids have mostly focused on α-helical systems and only a limited number of approaches for fluorination of specific β-sheet positions have been reported, so far. Finally, since both peptides AzoChig1 and AzoChig2 showed high solubility only in polar solvents like methanol and acetonitrile, AzoChig3 (GYDP-GTWG) equipped with an additional, N-terminal triethylene glycol residue, was prepared to furnish a photoswitchable and water-soluble chignolin derivative.

Amino acid building blocks for the solid-phase peptide synthesis (SPPS) of photocontrolled chignolin-derived β-hairpins were prepared, as follows (ESI†): Fmoc-protected AMPP derivative was synthesized in seven steps according to the known strategy of Renner et al. Thus, (9H-fluoren-9-yl)methyl-(3-aminobenzyl)carbamate, derived from Fmoc-protected 3-(aminomethyl)aniline, was reacted with 2-(3-nitroso phenyl)acetic acid under Mills condition to afford the AMPP building block in 59% yield.

The chignolin-derived peptides were assembled in an automated SPPS of photocontrolled chignolin-derived β-hairpins were prepared, as follows (ESI†): Fmoc-protected AMPP derivative was synthesized in seven steps according to the known strategy of Renner et al. Thus, (9H-fluoren-9-yl)methyl-(3-aminobenzyl)carbamate, derived from Fmoc-protected 3-(aminomethyl)aniline, was reacted with 2-(3-nitroso phenyl)acetic acid under Mills condition to afford the AMPP building block in 59% yield.

Fmoc-protected of commercially available 5FTrp yielded the requisite fluorinated building block, while Fmoc-TEG-derived was synthesized in five steps starting from triethylene glycol. The chignolin-derived peptides were assembled in an automated microwave-assisted CEM Liberty 1 peptide synthesizer on a preloaded Fmoc-Gly-Wang LL resin (Novabiochem).
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After Fmoc-deprotection by piperidine in N-methylpyrrolidone (NMP), the standard amino acid couplings were performed using HBTU/HOBt and diisopropylethylamine (DIPEA) in DMF for activation. In contrast, incorporation of the non-standard building blocks required modified coupling procedures and the use of the more reactive HATU/HOAt/N-methylmorpholine cocktail. Release from the resin with simultaneous deprotection using TFA/H$_2$O mixtures (95:5) followed by preparative RP-HPLC, finally provided the targeted chignolin-derived peptidomimetics with yields of 10-33%.

Photomodulation of β-hairpins with azobenzene derivatives as backbone elements mostly relies on an ultrafast trans ⇆ cis isomerization, which can be induced by light of different wavelengths and involves large changes in geometry and dipole moments. For instance, cis-AMPP-TrpZip peptides forming a β-hairpin-like structure can rapidly unfold within 1 ns by photo-isomerization of AMPP to its trans-isomeric state. In chignonin, the β-hairpin conformation is stabilized by H-bonds of Asp3:N-Thr8:O, Gly7:N-Asp3:O$^\delta$, Thr8:N-Asp3:O, Glu5:N-Asp3:O$^\delta$ and Thr6:N-Asp3:O, as well as by hydrophobic interactions between Tyr2:Pro4 and Tyr2:Trp9. The newly synthesized AzoChig derivatives should retain most of the aforementioned stabilizing interactions, and in particular AzoChig2 comprising the 5FTrp residue should reveal enhanced hydrophobic interactions between Tyr2 and Trp9 (vide supra). RP-HPLC analyses and NMR spectroscopy demonstrate the structural integrity and purity of the assembled AzoChig1-3 mimetics (see ESI†). To elucidate the three-dimensional solution structures of cis/trans-AzoChig1-3, 1H and 13C, as well as correlation (COSY), hetero nuclear single quantum (HSQC) and hetero nuclear multiple bond (HMBC) NMR spectra were recorded in 10 mM MeOH-d$_4$ solutions at 400 and 600 MHz. Unfortunately though, AzoChig3 aggregates strongly in solution, which leads to heavy signal broadening and precludes any precise signal assignments. In contrast, sharp 1H NMR signals, unequivocally assignable were obtained for cis/trans-AzoChig1 and 2, enabling their application to total correlation (TOCSY) and rotating-frame nuclear Overhauser enhancement (ROESY) spectroscopy at 400 MHz in MeOH-d$_4$ (Fig. 2C-F). All samples were kept in the dark for two days before measurement, yielding trans-configured peptides.

The 1H-NMR spectra of AzoChig1 and AzoChig2 peptides show distinct peak offsets for the aromatic residues Tyr2, AMPP5/6 and Trp9 (8.0 – 6.5 ppm) upon cis ⇆ trans isomerizations. Moreover, NOE cross-peaks detected between Tyr2:Trp9, Pro4:Gly7, AMPP5/6:Gly7 and AMPP5/6:Trp9 in the ROESY spectra of cis-AzoChig1 and cis-AzoChig2, depict the expected formation of folded structures for both derivatives (Fig. 2D-F),
with NOE signals between aromatic Tyr2, AMPP5/6 and Trp9 moiety, pinpointing the presence of turns stabilized by hydrophobic interactions. The $^{19}$F-NMR spectra of cis- and trans-AzoChig2 finally reveal distinct downfield shifts of the fluorine signals (Fig. 2B), indicating significant changes in the electronic structure and interactions upon isomerization. To substantiate the presence of unfolded trans conformations and their conversions into the desired $\beta$-hairpin structures upon azobenzene trans-to-cis isomerizations, UV/Vis, FT-IR, and CD spectroscopic measurements were performed. Therefore, 1 mM stock solutions of AzoChig1-3 peptides in MeOH were prepared and stored in the dark at room temperature for two days, furnishing trans-configured peptides. Using these stock solutions, peptide samples with concentrations of $c = 76 \mu M$ (AzoChig1), $77 \mu M$ (AzoChig2) and $78 \mu M$ (AzoChig3) were prepared through dilution with MeOH to record stationary UV/Vis absorption spectra before and after illumination at appropriate wavelengths (Fig. 3A). Irradiation of the trans-peptides at $\lambda = 350 \text{ nm}$ for 180 s led to reversible formation of photostationary states comprising in each case 84% of the trans-form of the peptides. This feature, similar to the one observed previously in ATZ12,27,28 points to a change of the peptide structure with an open trans-form to a compact cis form. Latter thereby shows additional, presumably inter-strand H bonds pointing to a $\beta$-hairpin-like structure.

CD spectra in the 195-250 nm range were determined for trans-AzoChig1-3 peptides at concentrations of $c = 76-78 \mu M$ in MeOH at 5 °C (vide supra). Solvent dependent CD spectra of AzoChig1 and AzoChig2 were recorded at concentrations of $c = 82-112 \mu M$ in MeOH/H$_2$O mixtures at 5 °C. For the cis-azo isomers, CD spectra were recorded after irradiation at 350 nm for 180 s, i.e., at the cis-photostationary state (Fig. 3D). All three cis-azo peptides feature distinct maxima at 231 nm — assigned to stacking of Trp9 to Tyr2 — as well as minima at 212 nm and 200 nm, and pronounced positive signals below 195 nm. Hence, they strongly resemble the spectra of native chignolin suggesting a folded, hairpin-like structure. The enhanced hydrophobic interactions between Tyr2 and 5FTrp9 in AzoChig2 become visible by the increase of the CD-signal in the 231 nm range. Temperature-dependent CD spectra of cis-AzoChig1 and cis-AzoChig2 in the 5-60 °C range (data not shown; see ESI†) reveal a thermal unfolding process to be working at higher temperatures, similar to native chignolin. The CD spectra of trans-AzoChig1-3 display small maxima at 231 nm and minima at 213 nm but deviate in the 200 nm range. The deep minimum found for AzoChig2 at 200 nm can be tentatively interpreted in terms of a better ordering of the peptide part due to the increased hydrophobic interaction of 5FTrp9 with Tyr2.
Moreover, solvent-dependent CD measurements were performed using mixtures of MeOH/H$_2$O (Fig 3E and 3F, see ESI$^+$), with ratios from 10-100% methanol. The MeOH/H$_2$O spectra of the cis-trans-AzoChig1 and cis-trans-AzoChig2 peptidomimetics show increasing values for the maximum around 230 nm and deeper minima at 200 nm with increasing water amounts. Presumably, the hydrophobic turn region of the cis-peptides becomes more stabilized by hydrophobic interactions between Tyr2:Trp9 in aqueous surroundings, which leads to the observed increase in molar ellipticity at 230 nm. Furthermore, interstrand interactions are stabilized by water-mediated hydrogen bonding as reflected by the decreased ellipticity around 200 nm. Finally and at higher water ratios, the observed CD signals of AzoChig1 and AzoChig2 approximate nicely the reported molar ellipticity characteristics of the parent peptide chignolin.$^{14}$

Conclusions

We have presented a novel class of photoswitchable β-hairpin model peptides derived from the designer mini protein chignolin by substitution of two central amino acids from the turn sequence by the known azobenzene chromophore AMPP. The resulting AzoChig1-3 peptidomimetics were assembled by SPPS and carefully characterized at both photosisomeric states using UV/VIS, IR, CD, and NMR spectroscopy. In the trans-state of AMPP, the peptides mostly exhibit a disordered structure, while trans → cis photoisomerization of the azobenzene chromophore induces folded β-hairpin-like structures.

Notes and references

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