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## COMMUNICATION

## Mn-Salen Catalysed Benzylic C-H Activation for the Synthesis of Aryl [<sup>18</sup>F]CF<sub>3</sub>-containing PET probes

Cite this: DOI: 10.1039/x0xx00000x

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Accepted 00th January 2012

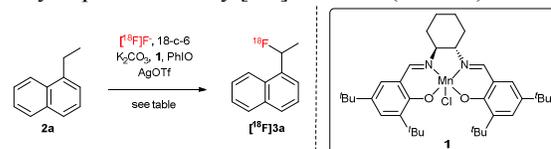
DOI: 10.1039/x0xx00000x

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**The development of a Mn-salen complex catalysed oxidative benzylic fluorination of non-activated C-H bonds using [<sup>18</sup>F]fluoride is described for installation of [<sup>18</sup>F]CHRF, [<sup>18</sup>F]CR<sub>2</sub>F and particularly [<sup>18</sup>F]CF<sub>3</sub> containing groups in the presence of other functional groups.**

PET is a non-invasive molecular imaging modality which allows the *in vivo* imaging of a variety of biological processes in patients. Of the leading PET radioisotopes currently available, fluorine-18 (<sup>18</sup>F) is one of the most important, as evidenced by the numerous clinically-deployed <sup>18</sup>F radiotracers. These tracers include 2-[<sup>18</sup>F]fluorodeoxyglucose ([<sup>18</sup>F]FDG), 3'-deoxy-3'-[<sup>18</sup>F]fluorothymidine ([<sup>18</sup>F]FLT) and [<sup>18</sup>F]fluorodopa, which are widely used in the fields of oncology and neurology.<sup>[1-4]</sup> While there has been an increase in the number of <sup>18</sup>F-containing pre-clinical radiotracers and new methods for the attachment of <sup>18</sup>F-containing prosthetic groups to target molecules, the initial incorporation of [<sup>18</sup>F]fluoride is in most cases still achieved by classical nucleophilic substitution reactions (*i.e.*, via S<sub>N</sub>2 or S<sub>N</sub>Ar mechanisms). Recently however, protocols for the preparation of [<sup>18</sup>F] aryl fluorides from aryl iodonium salts,<sup>[5],[6]</sup> and by the use of Pd(IV) and Ni(II) complexes,<sup>[7]</sup> [<sup>18</sup>F]allylic fluorides by Pd-catalysed substitution, [<sup>18</sup>F]4-fluorophenols by oxidative fluorination,<sup>[8]</sup> and [<sup>18</sup>F]trifluoromethyl aromatics catalysed by copper,<sup>[9]</sup> have been described that bypass these traditional radiofluorination paradigms. Moreover, Groves *et al.* have very recently disclosed an appealing method for direct oxidative [<sup>18</sup>F]fluorination at benzylic methylene C-H positions using [<sup>18</sup>F]fluoride using a Mn(salen)OTs pre-catalyst.<sup>[10-12]</sup> This report has prompted us to disclose a related procedure that we have developed and applied with particular success for the fluorination of aryl difluoromethyl compounds as an efficient approach to [<sup>18</sup>F]labelled trifluoro aryl PET probes.

Selecting to work with Jacobsen-type pre-catalyst (±)-**1** and with iodobenzene as stoichiometric oxidant due to their commercial availability, our initial objective was to define optimal conditions for the benzylic oxidative [<sup>18</sup>F] fluorination of 1-ethylnaphthalene **2a** by [<sup>18</sup>F] fluoride (Table 1).<sup>[13]</sup>



Entry	Solvent	PhIO <sup>[c]</sup>	T/°C	Time	%RCI <sup>[d]</sup>
1	MeCN <sup>[a]</sup>	P	60	15	27
2	MeCN/MeOH <sup>[a]</sup>	P	60	30	trace
3	MeCN/ <i>t</i> -BuOH <sup>[a]</sup>	P	60	15	25
4	MeCN/DMF <sup>[a]</sup>	P	60	15	2
5	MeCN <sup>[b]</sup>	P	60	30	72
6	MeCN <sup>[b]</sup>	S	60	30	32
7	MeCN <sup>[b]</sup>	S	90	30	28
8 <sup>[c]</sup>	MeCN <sup>[b]</sup>	P	60	30	70

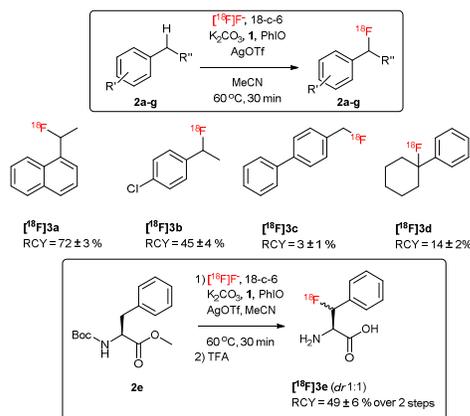
<sup>[a]</sup>[substrate] = 0.3 M; <sup>[b]</sup>[substrate] = 0.1 M; <sup>[c]</sup>Method of PhIO addition, either P = in equal portions at 5 min intervals, N = None used or S = Single addition; <sup>[d]</sup>RCI = Radiochemical Incorporation, calculated from radio-HPLC analysis, n = 3 in each example; <sup>[e]</sup> Kryptofix 2.2.2. (K222) used instead of 18-c-6.

**Table 1** – Optimisation of the oxidative [<sup>18</sup>F] fluorination of 1-ethylnaphthalene **2a**.

A combination of different solvents, temperatures durations and protocols for addition of the PhIO were explored. Portion-wise addition of solid PhIO gave a promising radiochemical incorporation of 27% within 15 minutes (n = 3; entry 1). As PhIO is only sparingly soluble in most organic solvents, and full dissolution is critical for automated radiosynthesis, we explored the use of a co-solvent. Use of either MeOH or DMF as co-solvents (1:1 v/v) allowed complete dissolution of the

oxidant, but suppressed product formation (entries 2 and 4) whereas *t*-BuOH was ineffective as a co-solvent and gave identical levels of incorporation to when using MeCN alone (entry 3 *cf.* entry 1). Reverting to the use of just MeCN as solvent but with increased dilution (0.1 M *cf.* 0.3 M) we were pleased to observe that after a reaction time of 30 minutes 72% radiochemical incorporation could be achieved (entry 5). At this dilution, addition of the PhIO solution as a single portion gave 32% RCI after 30 min, which we considered adequate for compound labelling (entry 6). Increasing the temperature to 90 °C gave no discernible improvement in incorporation (entry 7) and replacement of 18-c-6 with Kryptofix<sup>®</sup> 2.2.2 (K222) had no significant influence on the level of RCI (entry 8).

Application of this reaction to several simple substrates was undertaken to delineate the scope of the method (Figure 1).

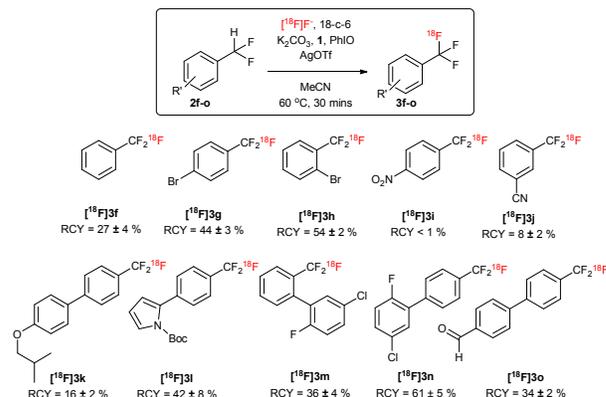


**Figure 1** - Oxidative benzylic [<sup>18</sup>F] fluorination of several different compounds using optimised labelling conditions, n = 4.

Successful labelling of compounds was achieved with moderate to good levels of incorporation in most cases. In general, secondary fluorides were introduced efficiently, whereas introduction at a primary position gave less than 5% RCY ([<sup>18</sup>F]3c) and at a tertiary position gave a more promising 14% RCY ([<sup>18</sup>F]3d). Site-selective labelling to give phenylalanine derivative [<sup>18</sup>F]3e gave 49% RCY although no diastereoselectivity was observed.

Notwithstanding the attraction of installing a single fluorine atom site-selectively within a target molecule, labelling of CF<sub>3</sub> groups was of particular interest due to their occurrence in a number of pharmaceuticals and we anticipated that <sup>18</sup>F-labelling of a difluoromethyl group could be an attractive entry to labelled CF<sub>3</sub> units.<sup>14</sup> The benzylic difluoromethyl containing precursors required for this approach are readily prepared by fluorodeoxygenation of benzaldehydes or by copper-catalysed difluoromethylation of aryl iodides using TMS-CF<sub>2</sub>CO<sub>2</sub>Et (or TMS-CF<sub>2</sub>H).<sup>[15-17]</sup>

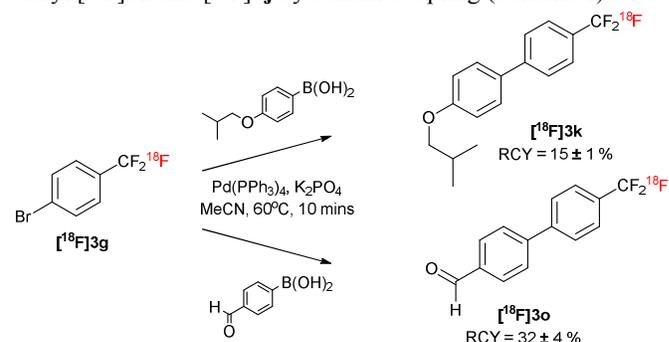
Ten benzylic difluoromethyl substrates also containing other potentially reactive functional groups (*i.e.* ether, aryl halide and carbamate groups) were investigated (Figure 2).



**Figure 2** - <sup>18</sup>F Oxidative benzylic fluorination to give CF<sub>3</sub>-containing compounds, n = 3.

[<sup>18</sup>F]Trifluoromethylbenzene ([<sup>18</sup>F]3f) and its 4- and 2-brominated derivatives [<sup>18</sup>F]3g and [<sup>18</sup>F]3h were prepared in 27%, 44% and 54% RCYs respectively whereas the more electron deficient 4-nitro and 3-cyano derivatives [<sup>18</sup>F]3i and [<sup>18</sup>F]3j were much less efficiently labelled (1% and 8% RCY respectively). It appears therefore that although strongly electron withdrawing substituents deter [<sup>18</sup>F]fluorine trapping of the presumed radical intermediate, bulky 2-substituents are tolerated. This is consistent with the successful formation of isomeric 2- and 4-substituted biaryls [<sup>18</sup>F]3m and [<sup>18</sup>F]3n and of 4-substituted biaryls [<sup>18</sup>F]3k, [<sup>18</sup>F]3l and [<sup>18</sup>F]3o in 36%, 61%, 16%, 42% and 34% RCYs respectively.

Labelled bromophenyls [<sup>18</sup>F]3g and [<sup>18</sup>F]3h can also be deployed as prosthetic groups as illustrated by the preparation of biaryl [<sup>18</sup>F]3k and [<sup>18</sup>F]3l by Suzuki coupling (Scheme 1).



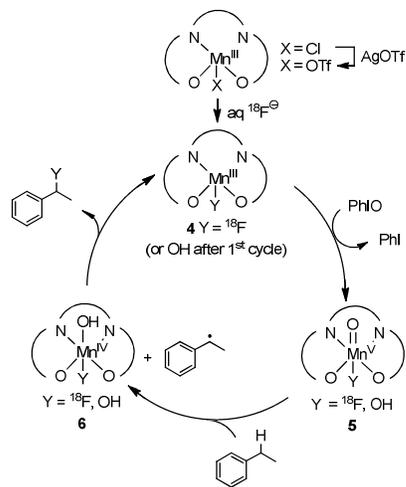
**Scheme 1** – Synthesis of biaryls [<sup>18</sup>F] 3k and 3o from [<sup>18</sup>F]3g by Suzuki coupling, n = 3.

The specific radioactivity of a labelled probe, quantifies the amount of radioactivity relative to non-labelled material present, and constitutes an important property of the probe. For the synthesis of [<sup>18</sup>F]3g, the specific activity was ~4 GBq/μmol, starting from 190 MBq of starting [<sup>18</sup>F]fluoride. This level of specific activity is ~10 fold greater than that reported by Huiban *et al.* for their [<sup>18</sup>F]CF<sub>3</sub> labelling.<sup>[9]</sup>

We also performed ICP-MS analysis on a purified sample of [<sup>18</sup>F]3a which had been allowed to decay, and no manganese was detected (<0.01 mg/l). Unlike Pd, which has a non-dietary limit of 100 μg/day for a 50 Kg adult, Mn is essential for

various functions in the human body and a daily intake of 2.5 mg/day is recommended for a 50 Kg adult.<sup>[18]</sup>

Mechanistically, we anticipate that the [<sup>18</sup>F]labelling reaction proceeds *via* essentially the same catalytic cycle as proposed by Liu and Groves for their <sup>18</sup>F labelling using Mn(salen)OTs (Figure 5).<sup>[11]</sup>



**Figure 5** – Proposed catalytic cycle for the <sup>18</sup>F-labelling reaction, and side-reaction to form alcohol by-products.

Thus, following initial Cl to OTf ligand exchange at Mn(III) effected by the AgOTf, [<sup>18</sup>F]fluoride is introduced *via* ligand exchange ( $\rightarrow$  4). Oxidation by PhIO then gives Mn(V) oxo species 5 which effects hydrogen atom abstraction from a benzylic position on the substrate concomitantly forming fluorohydroxy-manganese(IV) species 6 (X = <sup>18</sup>F). This species then ‘rebounds’ the [<sup>18</sup>F]fluorine atom to the benzylic carbon centred radical to give the labelled product. Due to the low concentration of [<sup>18</sup>F]fluoride, hydroxide can presumably compete as a ligand at Mn giving *e.g.* the dihydroxymanganese(IV) intermediate 6 (X = OH), hydroxide radical ‘rebound’ from which would give alcohol side-products, as are observed in these reactions.

## Conclusions

We have described a new method for radiolabelling *via* site-selective oxidative fluorination of benzylic difluoromethyl groups using [<sup>18</sup>F]fluoride to give aryl [<sup>18</sup>F]CF<sub>3</sub> compounds. The labelling reactions can be achieved without extensive investment in complex precursors and the straightforward experimental protocol holds promise for adaption to automated clinical PET probe production.

## Acknowledgements

We thank Imanova Ltd. for [<sup>18</sup>F]fluoride and use of their radiochemistry facilities, Dr. Neil Bramall (University of Sheffield) for ICP analysis, and CRUK/EPSRC/MRC/DoH (England) for funding (grant C2536/A10337).

## Notes and references

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