# ChemComm

## Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemcomm

Cite this: DOI: 10.1039/coxx00000x

www.rsc.org/xxxxx

## COMMUNICATION

### Femtogram Level Competitive Immunoassay of Mercury (II) Based on Surface-enhanced Raman Spectroscopy

Yuzhen Wang<sup>*a,b,c*</sup>, Shuai Chen<sup>*a*</sup>, Chao Wei<sup>*a*</sup>, Minmin Xu<sup>*a*</sup>, Jianlin Yao<sup>\**a*</sup>, Yuan Li<sup>*b*</sup>, Anping Deng<sup>\**a*</sup> and Renao Gu<sup>a</sup>

s Received (in XXX, XXX) Xth XXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

A femtogram level and specific surface enhanced Raman spectroscopy (SERS) based competitive immunoassay was developed to detect Hg(II) in aqueous solution for the first 10 time. This novel approach provides an alternative, ultrasensitive and specific analytical method for the detection of Hg(II).<sup>1</sup>

Mercury is a toxic and widespread pollutant that originates from oceanic and volcanic emissions, and human activities.<sup>1</sup> 15 Mercuric ions [Hg(II)] is the most stable form of inorganic mercury, which will damage the brain, nervous system, kidneys, and endocrine system.<sup>2</sup> Furthermore, microorganisms are able to biomethylate Hg(II) to methylmercury [CH<sub>3</sub>Hg(I)]. Such process allows bioaccumulation and biomagnifications to occur and 20 consequently be of concern for public health.<sup>1,3</sup> Therefore, it is necessary to develop mercury detectors for both environmental and biological samples.<sup>4</sup> Various methodologies such as sensors based on fluorophores, chromophores, foldamers, ampter, silver nanoparticles. conjugated polymers, gold nanoparticles, 25 electroanalytical methods such as capillary electrophoresis and voltammetry, pyridylporphyrin–DNA, peptides, proteins. enzymes, oligonucleotides, and DNAzyme have been reported in

the past few years.<sup>5</sup> Nevertheless, the above methodologies will be interfered by chemically-related metals, delayed response to 30 Hg(II), and/or insufficient sensitivity.

Surface-enhanced Raman spectroscopy (SERS) is a general and powerful spectroscopy technique that has been generally used by lots of surface science scientists in the past few years. It provides non-destructive and ultrasensitive characterization down

<sup>35</sup> to single molecular level.<sup>6</sup> SERS has been applied to various areas such as chemical sensing, medical diagnostics, and the studies of living cells and bacteria.<sup>7</sup> Very recently, SERS can also be employed to determine the Hg(II) ions, mainly by the nanoparticle-based chemical approaches.<sup>8</sup> For example, Zhang 40 selected appropriate organic ligands for the femtomolar detection

of  $Hg^{2+}$  by a SERS chip with silver-coated gold nanoparticles.

E-mail: jlyao@suda.edu.cn, denganping@suda.edu.cn

These SERS chips possess many advantages, such as excellent anti-interference ability, recycling usage, long-term stability, portable feasibility, and simple operation, it exhibited high 45 sensitivity and specificity.<sup>8</sup>

The introduction of immunoassay in SERS measurements can integrate their advantages of specificity and sensitivity.<sup>9</sup> For instance, the strong and specific binding of an antibody to its antigen has been widely exploited in biochemical studies, clinical <sup>50</sup> diagnostics, sensor design, and environmental monitoring.<sup>10</sup> In past years, many different approaches such as scintillation counting, fluorescence, chemiluminescence, electrochemical detection, enzymatic,<sup>11</sup> and surface plasmon resonance (SPR)<sup>12</sup> have been developed for a direct measurement of antigen-55 antibody binding. The expose of SERS first appeared in the enzyme immunoassay at the end of 1980s, in which Cotton et al. employed the SERRS effect to detect human thyroid simulating hormone (TSH) antigen.13 Nevertheless, the SERS based immunoassay was usually used for macromolecule proteins using 60 a sandwiched pattern<sup>14</sup>. In our previous studies, we reported the high sensitive detection and separation of antigen based on magnetic SERS immunoassay<sup>15</sup>. Herein, we report a femtogram level and specific competitive immunoassay for the detection of Hg(II) in aqueous solution.

Using hybridoma technique we successfully produced 65 monoclonal antibody (mAb) against Hg(II), which was then characterized by an indirect competitive enzyme linked immunosorbent assay (ELISA).<sup>16</sup> The IC<sub>50</sub> value of the ELISA for Hg(II) was 1.12 ng/mL (i.e. 5.6 nM) and the limit of detection 70 (LOD) at 3 times of signal/noise was 0.08 ng/mL (i.e. 0.4 nM). Low values of IC<sub>50</sub> and LOD indicated that the sensitivity of the ELISA is high. The cross-reactivity values of the mAb with Hg(II), CH<sub>3</sub>Hg(I), 6-mercaptonicotinic acid (MNA, the ligand selected for the preparation of immunogen) and CH<sub>3</sub>Hg-MNA 75 were 100%, 1.97%, 0.72% and 1.16%, respectively. There was no cross-reactivity of the mAb with Cu2+, Cr3+, Sn2+, Ni2+, Mn2+,  $Pb^{2+}$ ,  $Zn^{2+}$ ,  $Cd^{2+}$ ,  $Fe^{3+}$ ,  $Co^{2+}$  and  $Mg^{2+}$ . The values clearly demonstrate that the mAb has highly specific recognition with Ho(II)

<sup>&</sup>lt;sup>a</sup> College of Chemistry, Chemical Engineering and Materials Science, Soochow University, Suzhou 215123, China

Fax: +86 512 65880089; Tel:+86 512 65880359

<sup>&</sup>lt;sup>b</sup> College of Chemistry, Sichuan University, Chengdu 610041, China <sup>c</sup> College of Chemistry and Environmental Engineering, Datong

University, Datong 037009, China

Combining the highly specific mAb against Hg(II) with the highly sensitive analytical method SERS, we expect that the SERS based immunoassay is able to detect Hg(II) with ultrasensitivity and specificity. In general, the antibody is immobilized on the surface of the substrate and the target analyte

is packed by the immobilized Ab and another Ab labeled with SERS reporter, so called as sandwich immunoassay for detecting macromolecules. However, the competitive immunoassay is utilized to detect the small species that is unable to react with two s antibodies simultaneously, such as Hg(II). The principle and

- processes of the SERS based competitive immunoassay for Hg(II) were illustrated in Scheme 1. The coating antigen (i.e. antigenprotein conjugate) is firstly immobilized on the surface of substrate, it competes the limited binding sites of the mAb with
- <sup>10</sup> the Hg(II) in the solution. The weak SERS signal is due to the less amount of the mAb-gold nanoparticles-4-mercaptobenzoic acid (MBA) conjugate bound on the substrate which is caused by the competition of the high concentration of Hg(II) in the solution.
- To establish a SERS based competitive immunoassay, 15 besides mAb and coating antigen, another requirement is to prepare qualified MBA labeled immunogold. Firstly, we synthesized gold nanoparticles with the size of 30 nm. Then, MBA was attached onto the surface of Au nanoparticles with full coverage, and an appropriate amount of mAb was immobilized.
- <sup>20</sup> Finally, excessive bovine serum albumin (BSA) was added to the MBA labeled immunogold to completely block nonspecific binding sites.



35 Scheme 1 the principle and processes of the SERS based competitive immunoassay for Hg(II)

In order to achieve high sensitivity of the assay, there are several criteria, such as maintaining consistent concentration of coating antigen (500 ng/mL) and diluting ratio of MBA labeled <sup>40</sup> immunogold, have to be fulfilled. On the other hand, the possibility of SERS background signals from the coating antigen (i.e. antigen-protein conjugate) should be clarified and eliminated

- because some amino-acid residues are Raman active species. Fortunately, it was confirmed that there was no any background <sup>45</sup> SERS signals raised from coating antigen, and all detected signals
- in the SERS based immunoassay were generated from MBA bound on immunogold.

Under optimal assay conditions, the standard curve for Hg(II) was constructed within the range of  $10^{-5}$ -100 ng/mL. The

- <sup>50</sup> procedures of the SERS based immunoassay were as followed: 25 ng of coating antigen, diluted by phosphate-buffered saline (PBS), was dropped onto the substrate. After being placed in a chamber with a relative humidity of 65%-75% overnight, the substrate was dried at room temperature for 30 minutes. It was
- ss then incubated in 5% BSA for 1 hour to block nonspecific active sites followed with the treatment by Millii-Q water and dry nitrogen gas. Then, it was exposed to a mixture of 20  $\mu$ L of different concentrations of Hg(II) and 150  $\mu$ L of MBA labeled immunogold in borate buffer solution for 2 hours at room

<sup>60</sup> temperature. It was lastly treated in the order of TBS, 0.1% Tween, TBS, ultrapure water, and dry nitrogen gas. The SERS signal of the MBA labeled immunogold immobilized on the substrate was read out by a confocal microprobe Raman system. The SERS intensity at 1077 cm<sup>-1</sup> arose from Raman reporter <sup>65</sup> (MBA) was used for quantitative analysis.



**Fig 1** The SERS signals and calibration curve of the SERS based immunoassay for Hg(II) with different standard concentrations (a-i). The inserted figure is the calibration curve, where *B* and  $B_0$  is the SERS signal of the analyte at the standard point and at zero 70 concentration of the analyte, respectively.

The spectra and the calibration curve of the SERS based immunoassay for Hg(II) were shown in Figure 1. It was found that the SERS signal steadily decreased (a-i) as the concentration of Hg(II) increased,. The IC<sub>50</sub> value was found to be 0.06 ng/mL <sup>75</sup> (i.e. 0.3 nM) and the LOD was  $8 \times 10^{-5}$  ng/mL (i.e. 0.4 pM, in this case, the analyte volume for detection was 20 µL, thus the quantitative amount was 1.6 fg). The values of IC<sub>50</sub> and LOD obtained in this approach are 18.7 and 1000 times respectively lower than those in ELISA using the same mAb,<sup>16</sup> indicating the series was integrated. Comparing to other reports for detection of Hg(II), the proposed SERS based immunoassay is the most sensitive analytical method for Hg(II) achieved (see Table S1).

The specificity of the SERS based immunoassay was also verified. Six species including Pb(II), Cd(II), Ag(I), MNA, CH<sub>3</sub>Hg(I) and CH<sub>3</sub>HgMNA were selected to test the crossreactivity (CR). The standard solutions of cross-reacting species were prepared in the concentration range of  $10^{-5}$ -100 ng/mL and applied to the procedures of the assay. CR was expressed as percent IC<sub>50</sub> value based on the response of Hg(II), e.g. CR(%) = [IC<sub>50</sub> for Hg(II)]/[IC<sub>50</sub> for competing chemicals] × 100%. The cross-reactivity values of the mAb with Hg(II) and six species were shown in Figure 2. The original spectra were presented in Figure S4. One could conclude that no cross reactivity with Pb<sup>2+</sup>, S Cd<sup>2+</sup>, Ag<sup>+</sup>, CH<sub>3</sub>Hg(I), MNA and CH<sub>3</sub>HgMNA was detected. It clearly demonstrated that SERS based immunoassay exhibited high specificity in detecting Hg(II).

Canal water samples collected from Suzhou city was filtrated by a nylon membrane filter (0.45  $\mu$ m). The filtrates were adjusted to pH 7.0 and were analyzed by the proposed method. Then, the SERS based assay was applied to detect the filtrates which were spiked with Hg(II) at the concentrations of 0.1 ng/mL

and 0.5 ng/mL. Acceptable recovery rates of 83.5-95.7% were obtained, which suggested that the method could be used for the detection of Hg(II) in real aqueous samples.



**Fig 2** The cross-reactivity values of the mAb with Hg(II) and s other species tested by the proposed SERS based competitive immunoassay.

#### Conclusions

- In summary, a femtogram level and specific SERS based competitive immunoassay for the detection of Hg(II) in aqueous <sup>10</sup> solution was developed successfully. The labeled immunogold was synthesized by linking MBA and mAb to the surface of gold nanoparticles. The LOD value of the assay for Hg(II) was found to be 8×10<sup>-5</sup> ng/mL (i.e. 0.4 pM or 1.6 fg in 20 µL analyte solution), which was much lower compared to that in normal <sup>15</sup> ELISA approach. No cross reactivity with Pb<sup>2+</sup>, Cd<sup>2+</sup>, Ag<sup>+</sup>, MNA,
- <sup>15</sup> ELISA approach. No cross reactivity with Pb<sup>-</sup>, Cd<sup>-</sup>, Ag<sup>-</sup>, MNA,  $CH_3Hg^+$  and  $CH_3Hg$ -MNA revealed high specificity of the assay. The proposed method was used for the detection of Hg(II) in spiking water samples with acceptable recovery rate. This novel approach not only provides an alternative, ultrasensitive and
- <sup>20</sup> specific analytical method for the detection of Hg(II), but also can be extended as an useful model for the detection of other small molecular compounds in biological, food and environmental areas.

This work was supported by National Natural Science <sup>25</sup> Foundation of China (NSFC, Nos. 20835003, 21175097 21033007 and 21173155), the National Instrumentation Program (2011YQ031240402), and Priority Academic Program Development of Jiangsu Higher Education Institutions.

Yuzhen Wang and Shuai Chen contributed equally to this 30 work.

#### Notes and references

- (a) H. H. Harris, I. J. Pickering and G. N. George, *Science* 2003, 301, 1203-1203; (b) N. Givelet, F. Roos-Barraclough and W. Shotyk, *J. Environ. Monit.* 2003, 5, 935 – 949.
- <sup>35</sup> 2 N. Givelet, F. Roos-Barraclough and W. Shotyk, *J. Environ. Monit.*, 2003, **5**, 935 – 949.
  - 3 T. W. Clarkson, L. Magos and G. J. Myers, N. Engl. J. Med. 2003, 349, 1731-1737.
  - 4 (a) A. B. Descalzo, R. MartNnez-Mopez, R. Radeglia, K. Rurack
- and J. Soto, J. Am. Chem. Soc. 2003, 125, 3418-3419; (b) J. S. Lee, M. S. Han and C. A. Mirkin, Angew. Chem. Int. Ed. 2007, 46, 4093 4096.
- 5 (a) A. B. Othman, J.W. Lee, J.-S. Wu, J. S. Kim, R. Abidi, P.ThuQry, J. M. Strub, A. V. Dorssleaer and J. Vicens, *J. Org. Chem.* 2007, **72**, 7634 – 7640; (b) E. M. Nolan and S. J. Lippard,

*J. Am. Chem. Soc.* 2007, **129**, 5910 – 5918; (c) S. Yoon, E. W. Miller, Q. He, P. H. Do and C. J. Chang, *Angew. Chem. Int. Ed.* 2007, **46**, 6658 –6661; (d) M. K. Nazeeruddin, D. D. Censo, R. Humphry-Baker and M. GrItzel, *Adv. Funct. Mater.* 2006, **16**, 189

- 194; (e) Y. Zhao and Z. Zhong, J. Am. Chem. Soc. 2006, 128, 9988 9989; (f) Q. Wei, R. Nagi, K. Sadeghi, S. Feng, E. Yan, S. J. Ki, R. Caire, D. Tseng and A. Ozcan. ACS Nano 2014, 8, 1121-1129; (g) L. Chen, N. Qi, X. Wang, L. Chen, H. You and J. Li. RSC Adv., 2014, 4, 15055–15060; (h) I. B. Kim and U. H. F.
- <sup>55</sup> Bunz, J. Am. Chem. Soc. 2006, **128**, 2818 –2819; i) M. Rex, F. E. Hernandez and A. D. Campiglia, Anal. Chem. 2006, **78**, 445–451; (j) P. Salaun and C. M. G. van den Berg, Anal. Chem. 2006, **78**, 5052–5060; (k) J. K. Choi, G. Sargsyan, A. M. Olive and M. Balaz. Chem. Eur. J. 2013, **19**, 2515–2522; (l) S. V. Wegner, A.
- Okesli, P. Chen and C. He, J. Am. Chem. Soc. 2007, 129, 3474 3475; (m) D. Li, A. Wieckowska and I. Willner Angew. Chem. Int. Ed. 2008, 47, 3927 –3931; (n) M. Hollenstein, C. Hipolito, C. Lam, D. Dietrich and D. M. Perrin, Angew. Chem. Int. Ed. 2008, 47, 4346 –4350
- <sup>65</sup> 6 (a) S. Nie and S. R. Emory, *Science*, 1997, **275**, 1102–1106; (b) K. Kneipp, Y. Wang, H. Kneipp, L. T. Perelman, I. Itzkan, R. R. Dasari and M. S. Feld, *Phys. Rev. Lett.* 1997, **78**, 1667–1670; (c) J. F. Li, Y. F. Huang, Y. Ding, Z. L. Yang, S. B. Li, X. S. Zhou, F. R. Fan, W. Zhang, Z. Y. Zhou, D. Y. Wu, B. Ren, Z. L. Wang and Z. Q. Tian, *Nature* 2010, **464**, 392–395.
- 7 (a) A. L. Alak and T. Vo-Dinh, *Anal. Chem.* 1987, **59**, 2149–2153;
  (b) J. M. Bello, D. L. Stokes and T. Vo-Dinh, *Anal. Chem.* 1989, **61**,1779–1783.
- 8 (a) Y. X. Du, R. Y. Liu, B. H. Liu, S. H. Wang, M. Y. Han and Z. P. Zhang, *Anal. Chem.* 2013, **85**, 3160–3165; (b) W. Ma, M. Z. Sun,
- Lidaig, Indi. Chem. 2019, 65, 9160–9165, (f) W. Ma, M. Z. Sdil,
   L. G. Xu, L. B. Wang, H. Kuang and C. Xu, *Chem. Commun.*,
   2013, **49**, 4989 -4991; (c) W. Ji, L. Chen, X. X. Xue, Z. N. Guo, Z.
   Yu, B. Zhao and Y. Ozaki, *Chem. Commun.*, 2013, **49**, 7334 7336; (d) L. X. Chen, N. Qi, X. K. Wang, L. Chen, H. Y. You and
- 80 J. H. Li, *RSC Adv.* 2014, 4, 15055-15060; (e) X. F. Ding, L. T. Kong, J. Wang, F. Fang, D. D. Li and J. H. Liu, *ACS Appl. Mater. Interfaces* 2013, 5, 7072-7078.
- 9 V. Kanda, J. K. Kariuki, D. J. Harrison and M. T. McDermott, *Anal. Chem.* 2004, **76**, 7257–7262.
- <sup>85</sup> 10 (a) Y. Wang, D. Wei, H. Yang, Y. Yang, W. Xing, Y. Li and A. Deng *Talanta* 2009, **77**, 1783-1789; (b) Y. Li, Y. Wang, H. Yang, Y. Gao, H. Zhao and A. Deng *J. Chromatog. A* 2010, 1217, 7840-7817; (c) D. Li, S. Wei, H. Yang, Y. Li and A. Deng *Biosens. Bioelectron.* 2009, **24**, 2277-2280.
- <sup>90</sup> 11 (a) S. Gutcho and L. Mansbach, *Clin. Chem.* 1977, *23*, 1609-1614;
   (b) M. Bruchez, M. Moronne, P. Gin, S. Weiss and A. P. Alivisatos, *Science* 1998, **281**, 2013-2016;
   (c) J. Yakovleva, R. Davidsson, A. Lobanova, M. Bengtsson, S. Eremin, T. Laurell and J. Emneus, *Anal. Chem.* 2002, **74**, 2994-3004;
   (d) C. Duan and M. E. Meyerhoff, E. *Anal. Chem.* 1994, **66**, 1369-13777.
- 12 B. K. Oh,; Y. K. Kim, W. Lee, Y. M. Bae, W. H. Lee and J. W. Choi, *Biosens. Bioelectron*, 2003, **18**, 605-611.
- 13 T. E. Rohr, T. Cotton, N. Fan and P. J. Tarcha, Anal. Biochem. 1989, 182, 388-398.
- 100 14 (a) S. P. Xu, X. H. Ji, W. Q. Xu, X. L. Li, L.Y. Wang, Y. B. Bai, B. Zhao and Y. Ozaki, *Analyst*, 2004, **129**, 63-68; (b) N. Jing, M. H. Harpster; H. Zhang; J. O. Mechamc; W. C. Wilsonc and P. A. Johnson, *Biosens. Bioelectron.*, 2010, *26*, 1009-1015; (c) J. Ni, R. J. Lipert, H. Y. Park and M. D. Porter, *Anal. Chem.*, 1999, **71**, 4903-4908.
  - 15 (a) F. Bao, J. L. Yao, R. A. Gu Langmuir 2009, 25, 10782– 10787; (b) S. Chen, Y. X. Yuan, J. L. Yao, S. Y. Han, R. A. Gu, *Chem. Commun.*, 2011, 47, 4225 -4227.
- 16 Y. Z. Wang, H. Yang, M. Pschenitza, R. Niessner, Y. Li, D.
   Knopp and A. P. Deng, *Anal. Bioanal. Chem.* 2012, **403**, 2519-2528.
  - <sup>†</sup> Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/b00000x/