ChemComm

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemcomm

ChemComm

ARTICLE TYPE

Cite this: DOI: 10.1039/c0xx00000x

www.rsc.org/xxxxx

Dynamic Kinetic Resolution of Dehydrocoronamic Acid

David A. Chaplin,^a Martin E. Fox^{*a} and Sebastian H. B. Kroll^a

Received (in XXX, XXX) Xth XXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

⁵ Dehydrocoronamic acid can be racemised by dehydration of an *N*-acyl derivative to an azlactone, which undergoes facile racemisation. For the *N*-trifluoroacetyl derivative, the racemisation process was combined with an enzymatic resolution, to achieve a dynamic kinetic resolution process by 10 which the racemate can be converted to either enantiomer.

A family of Hepatitis C NS3 NS4A protease inhibitors which contain the common chiral motif (1R, 2S)-dehydrocoronamic acid **1** (Figure 1) comprises several antiviral drugs and

developmental drug candidates (Figure 2).¹⁻⁹



Figure 1 (1R, 2S)-Dehydrocoronamic acid.

Efficient synthesis of this densely functionalised quaternary



Figure 2. Examples of Hepatitis C NS3 NS4A protease inhibitors.

amino-acid as a single enantiomer is challenging. However, a concise route to the racemate has been developed,^{10,11} and routes to the single enantiomer based on resolution of both the racemate¹⁰ and a malonate precursor¹² have been described. ⁴⁵ Synthesis from the chiral pool material butane-1,2,4-triol¹³ and asymmetric routes employing asymmetric alkylation under chiral phase-transfer catalysis¹¹ and palladium-catalysed asymmetric allylic alkylation¹⁴ have also been published. We reasoned that if a method for racemisation of the off-isomer could be found, this ⁵⁰ could be combined with the established racemic synthesis and a method of resolution to provide an overall high-yielding synthesis

- of this increasingly commercially important compound. On dehydration with DCC, as an entry to subsequent coupling reactions, amide derivatives of dehydrocoronamic acid **5** cleanly
- ss give azlactones 6 (Scheme 1). Subsequent methanolysis of 6 gives the corresponding methyl esters 7. To our surprise, we discovered the enantiomeric excess of the product ester was lower than that of the starting acid.



Scheme 1. Racemisation of dehydrocoronamic acid azlactones.

The rate of racemisation was strongly influenced by the nature of the amide group. Thus, the single enantiomer methyl azlactone **6a** racemised[‡] very slowly at room temperature in acetonitrile, and with $t_{1/2}$ of 60 minutes at 80 °C. Azlactone **6b** 5 with an electron-withdrawing 3-trifluoromethyl substituent

- racemised more readily, with $t_{1/2}$ of 7.5 h at 15 °C in acetonitrile. In addition to racemisation, slower *cis/trans* isomerisation (Schemes 1 and 4) took place. This was detectable both by NMR and by chiral GC after methanolysis, which showed the *trans*-
- ¹⁰ isomer **8** formed to be racemic. Rates of *cis/trans* isomerisation of **6a** and **6b** were almost identical at 80 °C in acetonitrile. Hence the relative rates of racemisation to *cis/trans* isomerisation were lower for **6a** (8.5:1 at 80 °C in acetonitrile) than **6b** (165:1 at 15 °C). After *in-situ* formation of **6a** from (1*R*)-**5a**, heating for
- ¹⁵ 4 h at 80 °C, then methanolysis at ambient temperature, racemic **7a+8a** (6:1) was isolated in quantitative yield. For (*1R*)-**6b**, after 1 day at 15 °C, near-racemic (12% ee) **7b** + **8b** (189:1) was isolated in 70% yield. This behaviour was in marked contrast to the thermal isomerisation of other dehydrocoronamic acid
- ²⁰ derivatives, for example, the benzaldehyde imine 9,^{9,10} where racemisation was much slower than *cis/trans* isomerisation (Scheme 2). For this compound, after 120 minutes at 80 °C in toluene, the enantiomeric excess was 98.1%, but the ratio of imine 9 to dihydroazepine 11, formed by [3,3]-signatropic isomerisation of the trans isomer $10^{10,11}$ was 51 (66% da)





Scheme 2. Isomerisation of dehydrocoronamic ester imine.

⁴⁰ Likewise, the *N*-BOC, *cis*-ethyl ester **12** underwent more rapid *cis/trans* isomerisation than racemisation, reaching 90% ee and 38:62 *cis/trans* after 16 h in *o*-dichlorobenzene at 150 °C.



⁵⁰ Scheme 3. Isomerisation of *cis-N*-BOC dehydrocoronamic acid ethyl ester.

These observations can be rationialised if a *trans*-selective thermal isomerisation mechanism is possible for all ⁵⁵ dehydrocoronamic acid derivatives, but a different and *cis*-stereospecific isomerisation is possible for azlactones **6** (Scheme 4). A diradical mechanism¹⁵⁻¹⁷ is widely accepted for cyclopropane thermal isomerisation. For the intermediate **14**,

C-C bond rotation is possible, hence this pathway would give the thermodynamically more stable *trans* isomer **15** as the main product. For the *cis*-stereospecific rearrangement, we propose a mechanism which proceeds by concerted [3,3]-sigmatropic oxadivinylcyclopropane rearrangement to an achiral dihydrooxepine intermediate **16**, for which C-C bond rotation is ⁶⁵ not possible. This mechanism should be energetically more favoured for azlactone **6** compared to other dehydrocoronamic acid derivatives owing to the stabilisation provided by the simultaneous formation of an aromatic oxazole heterocycle. Increasing the electron-withdrawing power of the azlactone ⁷⁰ substituent R¹ withdraws electron density from the C=O bond and destribuilings the generation to the output



Scheme 4. Proposed mechanisms of azlactone isomerisation.

Evidence for this mechanism is provided by the ¹H NMR ⁹⁰ spectrum of **6b** (Figure 3), where in addition to the major signals which can be assigned to the cyclopropane structure, a minor set of four signals [$\delta_{\rm H}$ (400 MHz, CDCl₃; Me₄Si) 6.18, (1 H, dt, J =11.2, 5.2 Hz, OCH₂CH), 5.97 (1 H, dtt. J = 11.2, 5.9, 2.2 Hz, OCH₂CHCH), 4.72 (2H, d, J = 5.9 Hz, OCH₂) and 3.47(2 H, dd, 95 J = 5.2, 2.2 Hz, OCH₂CHCHCH₂)] is observed in a 30:1 ratio which can be assigned to dihydrooxepine **16**, present as an equilibrium component.



¹¹⁰ Figure 3. ¹H NMR spectrum of CF₃-azlactone 6b.

Given the ready racemisation of trifluoromethyl azlactone **6b** at ambient temperature and the utility of trifluoroacetyl as a removable nitrogen protecting group,¹⁸⁻²¹ we were able to apply ¹¹⁵ this discovery to our objective of an asymmetric synthesis of dehydrocoronamic acid. The racemisation could be applied in a

rac-**6** cis

65

resolution-based approach by recycle of the off-isomer of acid 5 to racemic ester 7 or acid 5. The desired (1R)-ester was obtained by resolution of 7b by enantioselective hydrolysis of the ester group of the (1S)-enantiomer with Subtilisin A, with an E value

- ⁵ of around 100, sufficiently high for an efficient resolution. However, this enzyme also cleaved the trifluoroacetyl group to a significant extent, reducing the possible efficiency of the recycle of (1*S*)-**5b**. Dynamic kinetic resolution of azlactone **6** provides a more direct and attractive approach than resolution/recycle of
- ¹⁰ ester **7**. Dynamic kinetic resolution of azlactones bearing a single 5-substituent, which can tautomerise to an enol form, by ringopening with nucleophiles in the presence of a chiral catalyst is well-established.²²⁻²⁶ Application of this concept to reaction of the 5,5-disubstituted azlactone **6b** with alcohols mediated by
- ¹⁵ enzymes proved feasible (Scheme 5). On a 7.5 g scale, the isolated yield of rac-**6b** from rac-**5b** was 89%. In the dynamic kinetic resolution, ethanol gave the best combination of reactivity and selectivity. With Novozym-435 (immobilised *Candida antarctica* Lipase B) under optimised conditions, (100:1 w/w
- ²⁰ MTBE-ethanol at 25 °C) on a 1 g scale, (1*S*)-**7c** was obtained in 95% ee and 97% isolated yield after 11 days (E~40). For (1*R*)-**7c**, the isomer required for the antiviral compounds, the highest enantioselectivity was achieved with immobilised Lipase PS. On a 1 g scale, under the same conditions, 75% ee and 96% isolated

25 yield was obtained after 18 days (E~7).



40 Scheme 5. Dynamic kinetic resolution of azlactone 6b.

Conclusions

We have discovered a method for racemisation of dehydrocoronamic acid and applied this to the asymmetric synthesis of either enantiomer of this commercially significant ⁴⁵ molecule by enzyme mediated dynamic asymmetric kinetic resolution. A concerted [3,3]-sigmatropic mechanism is proposed for this isomerisation.

Notes and references

^a Chirotech Technology Centre, Dr Reddy's Laboratories EU Ltd., Unit

50 410 Cambridge Science Park, Milton Road, Cambridge, CB4 0PE, UK.

Fax: 44(0)1223 506710; Tel: 44(0)1223 728010; E-mail: mfox@drreddys.com

† Electronic Supplementary Information (ESI) available: Experimental

procedures for preparation and isomerisation of **6a**, **6b** and **10**, ⁵⁵ isomerisation of **12**, resolution of **7b** and dynamic kinetic resolution of

6b. GC analytical methods and chromatograms for 7a-c, 12 and 13.

¹H NMR spectra of **1**, **5a**, **5b**, **6a**, **6b**, **7a**, **7b**, **7c**, **9**, **11**, **12**, **13**, **15a**, **15b**. See DOI: 10.1039/b00000x/

 \ddagger Enantiomeric excesses of **6** determined by methanolysis to ester **7** then $_{60}$ chiral GC.

- 1 For a review see L. Chatel-Chaix, M. Baril and D. Lamarre, *Viruses*, 2010, **2**, 1752.
- 2 Simeprevir: M.D. Cummings, J. Lindberg, T.-I. Lin, H. de Kock, O.L.E. Lenz, S. Fellander, V. Baraznenok, S. Nystrom, M. Nilsson, L. Vrang, M. Edlund, A. Rosenquist, B. Samuelsson, P. Raboisson
- and K. Simmen, Angew. Chem., Int. Edn., 2010, 49, 1652.
 Simeprevir: M. Nilsson, A.K. Belfrage, S. Lindstroem, H. Waehling, C. Lindquist, S. Ayesa, P. Kahnberg, M. Pelcman, K. Benkestock, T. Agback, L. Vrang, L. Y.Terelius, K. Wikstroem, E. Hamelink, C.
- 70 Rydergard, M. Edlund, A. Eneroth, P. Raboisson, T.-I. Lin, H. de Kock, P. Wigerinck, K. Simmen, B. Samuelsson and A. Rosenquist, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 4004.
- Asunaprevir: X.A. Wang, L.Q.-Sun, S.-Y. Sit, N. Sin, P.M. Scola, P. Hewawasam, A.C. Good, Y. Chen and J.A. Campbell, PCT Patent
 Application WO 2003099274 A1.
- Faldaprevir: M. Llinas-Brunet, M.D. Bailey, N. Goudreau, P.K. Bhardwaj, J. Bordeleau, M. Bos, Y. Bousquet, M.G. Cordingley, J. Duan, P. Forgione, M. Garneau, E. Ghiro, V. Gorys, S. Goulet, T. Halmos, T. S.H. Kawai, J. Naud, M.-A. Poupart and P.W. White, J. Med. Chem., 2010, 53, 6466.
- Danoprevir: S.D. Seiwert, S.W. Andrews, Y. Jiang, V. Serebryany, H. Tan, K. Kossen, P.T.R. Rajagopalan, S. Misialek, S.K. Stevens, A. Stoycheva, J. Hong, S.R. Lim, X. Qin, R. Rieger, K.R. Condroski, H. Zhang, M.G. Do, C. Lemieux, C, G.P. Hingorani, D.P. Hartley, J.A. Josey, L. Pan, L. Beigelman and L.M. Blatt, *Antimicrobial*
- Agents and Chemotherapy, 2008, 52, 4432.
 Neceprevir: A. Phadke, X. Wang, G. Pais, A. Hashimoto, V. Gadhachanda, D. Chen, A. Agarwal, S. Zhang, C. Liu, S. Li and M. Deshpande, Milind, PCT Patent Application WO 2010068761 A2.
- 90 8 Sovaprevir: A. Phadke, X. Wang and S. Zhang, S. PCT Patent application WO 2008008502 A1.
- 9 Grazoprevir: S. Harper., J.A. McCauley, M.T. Rudd, M. Ferrara, M. DiFilippo, B. Crescenzi, U. Koch, A. Petrocchi, M.K. Holloway, J.W. Butcher, J.J. Romano, K.J. Bush, K.F. Gilbert, C.J. McIntyre,
- ⁹⁵ K.T. Nguyen, E. Nizi, S.S. Carroll, S.W. Ludmerer, C. Burlein, J. DiMuzio, D.J. Graham, C.M. McHale, M.W. Stahlhut, D.B. Olsen, E. Monteagudo, S. Cianetti, C. Giuliano, V. Pucci, N. Trainor, C.M. Fandozzi, M. Rowley, P.J. Coleman, J.P. Vacca, V. Summa and N.J. Liverton, ACS Med. Chem. Lett., 2012, 3, 332.
- ¹⁰⁰ 10 P.L. Beaulieu, J. Gillard, M.D. Bailey, C. Boucher, J.-S. Duceppe, B. Simoneau, X.-J. Wang, L. Zhang, K. Grozinger, I. Houpis, V. Farina, H. Heimroth, T. Krueger and J. Schnaubelt, *J. Org. Chem.*, 2005, **70**, 5869.
- K. M. Belyk, B. Xiang, P.G. Bulger, W.R. Leonard, J. Balsells, J.
 Yin, C.-Y. Chen, *Org. Process. Res. Dev.*, 2010, 14, 692-700.
 - 12 C. Fliche, J. Braun, and F. Le Goffic, Synth. Commun., 1994, 24, 2873.
 - 13 W. Tang, X. Wei, N.K. Yee, N. Patel, H. Lee, J. Savoie and C.H. Senanayake, *Org. Process Res. Dev.*, 2011, **15**, 1207.
- ¹¹⁰ 14 M.E. Fox, I.C. Lennon and V. Farina, *Tetrahedron Lett.*, 2007, **48**, 945.
 - 15 P.H. Mazzocchi and H.J. Tamburin, J. Henry, J. Am. Chem. Soc., 1975, **97**, 555.
 - 16 W.E. Doering, and E.A. Barsa, Tetrahedron Lett., 1978, 19, 2495.
- 115 17 C. Isborn, D.A. Hrovat, and W.T. Borden and W. Thatcher, J. Phys. Chem. A, 2004, 108, 3024.
 - 18 H. Tanaka, Y. Yoshimura, N.J. Dovichi, M.M. Palcic and O. Hindsgaul, *Tetrahedron Lett.*, 2012, 53, 1812.
- F. Formaggio, M. Crisma, G. Ballano, C. Peggion, M. Venanzi and
 C. Toniolo, *Org. Biomol. Chem.*, 2012, **10**, 2413.
 - 20 B.J. Wolfe, F. Ghomashchi, T. Kim, C.A. Abam, M. Sadilek, R. Jack, J.N. Thompson, R.C. Scott, M.H. Gelb and F. Turecek, *Bioconjugate Chemistry*, 2012, 23, 557.
- 21 D. Rejman, R. Pohl and M. Dracinsky, *Eur. J. Org. Chem.*, 2011, 2172.
 - 22 N.J Turner, J.R. Winterman, R. McCague, J.S. Parratt, and S.J.C. Taylor, *Tetrahedron Lett.*, 1995, 36, 1113.

- 23 M.D. Truppo and G. Hughes, Org. Process Res. Dev., 2011, 15, 1033.
- 24 A. Peschiulli, C. Quigley, S. Tallon, Y.K. Gun'ko and S.J. Connon, J. Org. Chem. 2008, 73, 6409.
- 5 25 A. Berkessel, S. Mukherjee, F. Cleemann, T.N. Mueller and J. Lex, *Chem. Commun.*, 2005, 1898.
- 26 A. Berkessel, F. Cleemann, S. Mukherjee, T.N. Mueller and J. Lex, *Angew. Chemie, Int. Edn.*, 2005, **44**, 807.