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1 Rapid and nondestructive evaluation of fish Freshness by near infrared 2 reflectance spectroscopy combined with chemometrics analysis ¹Ran Ding, ¹Xingyi Huang^{*}, ¹Fangkai Han, ¹Huang Dai, ^{1, 2}Ernest Teve, ¹Fubin Xu 3 ¹School of Food and Biological Engineering, Jiangsu University, 4 5 Xuefu Road 301, Zhenjiang 212013, Jiangsu, P. R. China ²School of Agriculture, Department of Agricultural Engineering, 6 7 University of Cape Coast, Cape Coast, Ghana 8 *Corresponding author: Xingyi Huang, Email: h xingyi@163.com 9 Tel: +86-51188792368, Fax: +86-51188797308 10 Abstract 11 Rapid and nondestructive measurement of freshness is essential for control of fish 12 and its products' quality and safety. In this study, K value was measured by high 13 performance liquid chromatography (HPLC) and employed as an index of fish 14 freshness. The prediction models of the silver chub freshness were developed using 15 Fourier Transform Near Infrared Reflectance Spectroscopy (FT-NIRS) with Several 16 Partial Least Squares (PLS, i-PLS, Si-PLS), Support Vector Machines Regression 17 (SVMR) and Synergy interval plus Support vector machine regression leading to 18 Si-SVMR. By comparison, the performance of Si-SVMR model was superior to the others for the prediction of K value, where RMSECV = 0.027095 and Rc = 95.59%19 20 for calibration set, while RMSEP = 0.036525 and Rp = 93.74% for prediction set. The 21 results indicated that FT-NIR spectroscopy together with Si-SVMR model could be a 22 reliable method for detection of fish freshness.

23 Keywords: NIRS, fish freshness, K value, SVMR

24 **1.0 Introduction**

It is well known that freshness is the primary characteristic of the quality of freshwater fish and a positive property well evaluated by consumers. Loss of freshness and spoilage of fish are complicated processes and various factors such as fishing, handling, bleeding, storage temperature, the kind of fish species, the amount of food in the guts influence the spoilage pattern. The traditional detection methods of fish freshness mainly include sensory evaluation, microbiological, chemical, and

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physical testing methods. Sensory evaluation methods based on significant appearance parameters such as, skin, slime, eves, gills, belly and odor are conducted by experienced and well-trained humans(Nollet and Toldra 2010). Although these methods can be done without damage to the fish, these are subjective due to the subjective opinion of human judges and difficult to make quantitative analysis(Pons-Sánchez-Cascado, Vidal-Carou et al. 2006). At present microbiological methods based on total viable counts, chemical and biochemical methods (total volatile basic nitrogen, trimethylamine, pH, ATP, etc.), and physical testing methods play a key role in industrial fish quality and safety evaluation and inspection. Even certain of them have been used as regulation methods and gold standards serving scientific researches with their reliability and accuracy (Watanabe, Tamada et al. 2005, Alimelli, Pennazza et al. 2007, Zaragozá, Ribes et al. 2012). However, these methods are normally expensive, time-consuming, laborious, destructive and difficult for the application of the real-time detection. Recently emerging sensor array technologies, such as electronic tongue, electronic nose and olfaction visualization technology, were also used for the study of freshness detection. These methods have their advantages. Nevertheless, gas gathering, complicated testing procedures and moisture sensibility weaken the overall performance for electronic nose technology. Electronic tongue technology is not suitable for online testing due to the troublesome liquid sample preparation. As for olfaction visualization technology, the colorimetric sensor array preparation is a time-consuming effort and expensive metalloporphyrins directly result in the increase of detection cost (Gil, Barat et al. 2008, Huang, Xin et al. 2011, Ruiz-Rico, Fuentes et al. 2013). Therefore, it is necessary to develop a simple, fast, non-destructive, objective, economical and quantitative method for the detection of fish freshness (Wei and Wu 2007, Dowlati, de la Guardia et al. 2012).

Near infrared reflectance spectroscopy (NIRS) is the high-tech analytical techniques in the field of analytical chemistry. It is the integration of multi-disciplinary knowledge, such as spectroscopy, chemometrics, and computer multi-disciplinary knowledge and others. This method is simple, fast, green, low cost and has a good reproducibility. The root of modern near infrared analytical techniques

began earlier in the 1960s by the American Scientist Norris for Agriculture product quality measurement. This technology was usually used for a measurement of agricultural moisture, fat, protein and other substances(Khodabux, L'Omelette et al. 2007, Tito, Rodemann et al. 2012). With the rapid development of computer technology and the stoichiometry, the technology has been applied to various walks of life, such as pharmaceutical analysis(Porfire, Rus et al. 2012), chemical engineering(Ribeiro, Raja et al. 2014), environmental science (Buerck, Roth et al. 2001), agricultural products quality researches (Nicolaï, Beullens et al. 2007).

The previous research objects of the detection of fish freshness with NIRS mostly were with fish fillets and surimi (Bøknæs, Jensen et al. 2002, Xu, Huang et al. 2012), with the fish body broken. Recently the back of the entire fish was also studied (Zhang, Xu et al. 2011, Li, Wang et al. 2013). Remarkably, the back of fish is covered by fish scales which are mainly composed of CaO, P_2O_5 and Collagen(Atta 2013). Normally, Inorganic Chemical (CaO, P_2O_5) has reflection interference on NIRS. Besides, NIRS may be also interfered with the different distribution of organs and fishbone in the abdominal cavity of fish. NIRS detection technology of large yellow croaker by removing fish scales based on PLS methods was proposed (Zhang, Xu et al. 2011). The result is ideal, but the method is also destructive. In contrast, the structure of fish eye is sample and easy to locate(Dowlati, Mohtasebi et al. 2013), which bring about a good test repeatability. Fish eye gradually becomes cloudy from the clear, until the corruption and collapse the storage duration, which is a sound determination indicator of fish freshness for consumers and fish merchants. Hence, NIR detection method based on fish eye could be rapid, simple, nondestructive and easy to locate in theory.

Recently, a new perspective that K value is a more accurate index for the early freshness of fish than TVBN (Total Volatile Basic Nitrogen) was put forward. In the view, the freshness of fish depends primarily on its own biochemical reactions. That a series of biochemical reactions occurred after death is the essence of the change of fish freshness and has nothing to do with the microbial decomposition. The microbial decomposition occurred in the late stage of fish deterioration. The decomposition of

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ATP in fish meat sets in after death, and its compounds are subsequently produced according to the following sequence.

 $ATP \rightarrow ADP \rightarrow AMP \rightarrow IMP \rightarrow HxR(inosine) \rightarrow Hx(hypoxanthine)$

The K value represents the proportion (%) of the total amount of inosine and hypoxanthine relative to that of ATP-related compounds. The higher the K value, the less fresh the fish. ATP had a rapid degradation from the dead to rigor mortis, leading to a sharp increase of K value. Hence, K value is a more accurate index for the early freshness of fish than TVBN(Etsuo Watanabe 2005). The researches on horse mackerel(Losada, Piñeiro et al. 2005), trout (Bizri, Bouhours et al. 1985), yellow fin tuna(Kamalakanth, Ginson et al. 2011) and other during iced storage(Lougovois, Kyranas et al. 2003) showed that K value could be used as an important chemical indicator for the evaluation of fish freshness.

Freshwater fish is popular with consumers because of its delicacy and nutrition. Silver carp (Hypophthalmichthys molitrix) is a widespread species of freshwater fish in China. In this study, NIRS was attempted for the detection of K value in silver carp during storage and preservation. In addition, HPLC was employed for K value measurement of silver carps. Multiple spectral preprocessing methods were compared. PCA algorithm was applied for dimensionality reduction. Several partial least square techniques were applied (PLS, i-PLS, Si-PLS). Support Vector Machine Regression and Synergy interval plus Support vector machine regression leading to Si-SVMR were also acquired for the quantitative prediction of K value of silver carp.

2.0 Materials and methods

2.1Silver carp samples

114 180 fresh silver carps (*Hypophthalmichthys molitrix*) were purchased from 115 Zhenjiang fish market with an average weight of 950 g and average body length of 40 116 cm. The fish were washed with running water and then put into food preservation kits 117 with labels (1~180). Then all fish were stored in a refrigerator at about 4°C. 15 fish 118 were selected randomly for the spectral collection every day and then they were 119 slaughtered for the measurement of K values. The test was conducted for 12 days.

2.2. NIR instrumentation and data collection

The spectrum of each sample was collected in the reflectance mode using the Antaris II Near Infrared Spectrophotometer (Thermo Electron Company, USA) with an integrating sphere. Built-in background was as a reference. Each eye of the fish was scanned 5 times. The average of 10 data was used as the final raw data of the fish. The spectra of 15 fish were collected one by one every day. Each time a fish was taken out of the fridge for the spectral collection. After the spectral collection of the fish was finished, it was transferred to the next laboratory in an ice box for the measurement of K value. The whole experiment was conducted at an ambient temperature of $25 \pm 1^{\circ}$ C and the air humidity was kept at steady state. Each spectrum was an average of 16 scans with a spectral range of 10,000–4000 cm⁻¹, and the raw data set were measured in the interval of 8 cm⁻¹, resulting in 1557 variables. The raw NIRS profile of a silver carp is shown in Fig.1.

2.3. Determination of K value

K value is an indicator of the fish freshness. The researches on horse mackerel(Losada, Piñeiro et al. 2005), trout(Bizri, Bouhours et al. 1985), yellow fin tuna(Kamalakanth, Ginson et al. 2011) and other during iced storage(Lougovois, Kyranas et al. 2003) showed that K value could be used as an important chemical indicator for the evaluation of fish freshness. It equals to the percentage of the sum of inosine and hypoxanthine and the total of adenosine triphosphoric (ATP) and its decomposition product. Expressed as(Losada, Piñeiro et al. 2005, Kimiya, Sivertsen et al. 2013):

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$$K(\%) = \frac{100 \times (HxR + Hx)}{ATP + ADP + AMP + IMP + HxR + Hx}$$
(1)

- 143 ATP—adenosine triphosphate,
- 144 ADP—adenosine diphosphate ,
- 145 AMP—adenosine monophosphate,
- 146 IMP—inosine monophosphate,
- 147 HxR—inosine,
- 148 Hx—hypoxanthine .
- 149 All concentration unit were ' μ mol / g'.

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 The fish was immediately slaughtered for the measurement of K values after its near-infrared spectral collected. Specific steps are described below: The flesh of fish back without scales was ground into minced fish with the mincing machine. 1g of minced fish was homogenized with 10mL of purified water for 1 min. 20mL of 15% PCA(perchloric acid solution) was added to the mixture, keeping on stirring for 5 min. Then the sample was centrifuged and supernatant was poured into a beaker (100mL). The sediment was washed by 5% PCA and further was centrifuged. Afterwards, the first and the second supernatant were merged. Repeat the above twice. The supernatant was neutralized (pH 6.8) by adding 5M KOH, standing in an ice bath for 30 min. Then the solution was centrifuged and the supernatant was poured into 100-mL volumetric flask. The sediment was washed by 5% PCA-KOH. The first and the second supernatant were merged and transferred into the above volumetric flask. Repeat the above twice. Finally, the solution in the 100-mL volumetric flask was diluted with purified water to 100 mL and agitated, standing in the an ice bath in a short time. Centrifugation conditions: 9000r/min, 10°C, 10min. Liquid chromatography conditions: 1) high performance liquid chromatography (Ultimate 3000 DIONEX company in Germany), C18 reverse phase column (Shimadzu inertsustain C18 4.6 × 150 mm, 5µm). 2) mobile phase: 0.05M disodium hydrogen phosphate-the phosphodiesterase sodium hydroxide buffer (pH 6.8), 3) detection time : 30 min, 4) detection wavelength: 254 nm, 5) flow rate: 0.8mL/min, 6) column temperature: 2600° C. The types and concentration of the single standard and mixed standard solution of ATP, ADP, AMP, IMP, HxR and Hx were measured under the same conditions. The standard curve was drawed. Fig.2. shows High Performance Liquid Chromatogram (HPLC) of the standard ATP-related compounds. The types and concentration of the standard samples were determined by comparing the retention time and peak area of samples with standards. K value was calculated according to the method described elsewhere(Kimiya, Sivertsen et al. 2013). The variation trend of K value of silver chub during the cold storage was shown in Fig. 3. 2.4 Chemometric methods and evaluation index All computations, chemometric analysis and graphics were executed with programs developed in Matlab 2008b (The Mathworks, Inc., Natick, MA, USA). The SVMR models were fitted using the functions provided in the libsvm-mat-2.89-3 toolbox. The chemometric techniques was used in the research, including Savizky-Golay

190 polynomial curve smoothing (SG), 1st derivation(D1), Principal Component Analysis

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(PCA), Support Vector Machine Regression (SVMR) and multifarious Partial Least
Squares (PLS, i-PLS, Si-PLS).

The application of SG-D1 is beneficial not only for the removal of noise but also for extending the differences of characteristics(Xu, Xie et al. 2011, Chen, Ding et al. 2012). Compared with Partial Least Squares (PLS), Synergy Interval Partial Least Squares (Si-PLS) needs the less number of spectra variables but the result is not less than the result from PLS model in general (Chen, Zhao et al. 2007, Godoy, Vega et al. 2014, Teye, Huang et al. 2014). Support Vector Machine (SVM) determines the appropriate trade-off between learning ability of limited samples and the learn accuracy of specific samples for the best generalization performance (Devos, Ruckebusch et al. 2009, Alves and Poppi 2013).

Root mean square error of cross-validation (RMSECV), Root mean squared error of prediction (RMSEP) and Correlation coefficient (R) were employed as index for the performance of the achieved model. The smaller RMSECV and RMSEP were, the stronger the predictive ability of the achieved model was. The higher the similarity of measurements and prediction values will make the R output move further towards 1, leading to the stronger predictive ability of the achieved model(Chen, Zhao et al. 2009, Teye, Huang et al. 2013).

2.5 Calibration and prediction set

On the first day, due to the difference of the silver carp vitality, the time of their death was different after they were placed into the refrigerator. That brought large differences on K value of the first day. Some fish stayed alive corresponding to K value but others did not. Considering that the freshness of dead fish was studied in the paper, the data from the next day to the twelfth day (a total of 11 days) were used for modeling. The samples were divided into two sets, namely calibration set and prediction set. The calibration set was made up of 110 samples and these samples were used to model. 55 samples were used as the prediction set, which was used to test the reliability and stability of the model. To minimize the error and avoid bias in the division of the subsets, the data is rearranged according to the size of K value from low to high order. Then in every 3 samples, two were selected as calibration set

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and 1 as the prediction set.

2.6 Quantification models

Multifarious Partial Least Squares (PLS, i-PLS, Si-PLS), SVMR and Si-SVMR models were applied for the prediction of K value of silver carps during the cold storage. Firstly, multiple pretreatment methods and combination between them were compared, including Savitzky-Golay (SG), First Derivative (D1), Second Derivative (D2), Standard Normal Variate (SNV), Multiplicative Scatter Correction (MSC) and Mean Centre (MC) as well as the combination of SG and D1. The selection principle was based on maximal correlation coefficient (R). Then PLS with the full spectrum, i-PLS and Si-PLS quantitative models were respectively achieved. And the efficient joint intervals were also generated by Si-PLS model. Next, Principal Component Analysis (PCA) was used for dimensionality reduction and then SVMR model with the full spectra and the optimal combined subintervals from Si-PLS model were respectively obtained.

3.0 Results and discussion

3.1K value analysis

ATP in the fish flesh would be degraded as: $ATP \rightarrow ADP \rightarrow AMP \rightarrow IMP \rightarrow$ $HxR \rightarrow Hx$ after the death of fish. K value is more accurate to indicate the changes in the early stage (freshness) of fish than TVBN (Total Volatile Basic Nitrogen). Because the rapid ATP decomposition leads to the soar of K value. However, the rate of protein degradation is slow from the death of fish to final decay. The lower the K value, the more fresh the fish. Inversely, the higher the K value, the less fresh the fish. There are many studies on the relationship between K value and fish freshness (Lougovois, Kyranas et al. 2003, Losada, Piñeiro et al. 2005, Kamalakanth, Ginson et al. 2011). It is generally acknowledged that K value of live fish is not more than 10%; K value of the secondary freshness range from 20% to 40%; the fish is inedible/harmful when the K value is more than 40% (Khodabux, L'Omelette et al. 2007).

In this work, the behavior of K value during cold storage is shown in Fig.1. K value is on the rise with the storage duration, with a sharp increase especially in the first three days. The silver carps were fresh food with an average K value of 10.64%

on the first day. K values of some fish were less than 10%, indicating that the fish were still dying on the first day. And K value of the death on the first day ranged from 10% to 20%, suggesting that they were fresh. On the second day silver carps were also fresh with average K value of 19.67%. The silver carps were the secondary freshness with an average K value of 20%~40% from the third to seventh days. The fish is inedible starting from the eighth day. The perception of spoilage happened on the twelfth day(Kimiya, Sivertsen et al. 2013).

3.2 NIR examination

Many factors have an effect on spectral response values in the process of measurement, including high frequency random noise, baseline drift, signal to background, uneven concentration, light scattering, or the optical path change and others. Spectra preprocessing is very important in near infrared spectroscopy analysis for the decrease of error. Multiple preprocessing methods were investigated combining with Si-PLS model, including Savizky-Golay polynomial curve smoothing (SG), 1st derivation (D1), 2nd derivation (D2), Standard Normal Variate (SNV), Multiplicative Scatter Correction (MSC) and Mean centering (MC) as well as SG-D1. A SG filter with five smoothing points was employed to smooth and remove random noise from NIR spectra. D1 and D2 of the spectra were also used to increase the spectral resolution and to solve problems of baseline shift and linear tilt. MSC and SNV were used to correct both multiplicative and additive effects of the spectra due to scattering, light scattering, particle size, and the change of optical path. In MSC, each spectrum was linearized to the average spectrum. Moreover, SNV was conducted by normalizing each individual spectrum to zero mean and unit variance (Jamshidi, Minaei et al. 2012). The arbitrary origin of interval scale variables can be eliminated by MC. From Tab.1, the result of SG-D1 was better than others with maximum correlation coefficient (R). So SG-D1 method was applied to the spectrum preprocessing in the study. The profile of the raw NIRS and corresponding SG-D1 of a fish is shown in Fig.2. It could be found that the raw spectra were smoothed and weak peaks were enhanced after the application of SG-D1 preprocessing methods.

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3.3. Different PLS models

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281	The full spectrum range from 4000 to 10000 cm ⁻¹ was used. PLS model with the
282	full spectrum and iPLS model were obtained in sequence. The results are respectively
283	shown in the Fig.4 and Fig.5, where R=0.8087 and 0.8017 respectively in the
284	prediction sets. The performances of the two models were unsatisfactory, so Si-PLS
285	model was further used. Si-PLS was divided into 15, 16, 17,, 21 intervals combined
286	with 5 to 9 subintervals as seen in Table 2. The optimal combination of intervals and
287	the number of PLS factor was optimized by cross validation. The best performance
288	was chosen according to the lowest root mean square error (RMSE). The best results
289	obtained by Si-PLS are shown in bold and italic characters from Table 2. Scatter plots
290	of Si-PLS model are shown in Fig.6. Seven spectra subintervals from the spectral
291	regions were selected by Si-PLS model as seen from Fig.7, including 5142.9-5428.6
292	$\rm cm^{-1}$, 6571.3-7142.7 $\rm cm^{-1}$, 7428.4-7999.8 $\rm cm^{-1}$ and 8856.9-9428.3 $\rm cm^{-1}.$ All the
293	ranges have 523 variables out of the 1557 variables. The selection was in accordance
294	with the group active spectral region in general. The spectra between (5142.9-5428.6
295	cm ⁻¹) showed peak at 5169 cm ⁻¹ was associated with O–H stretching and combination,
296	which indicates the absorption peak of the water, at 5421 cm^{-1} associated with C-H
297	first overtone. The spectra (6571.3-7142.7 cm ⁻¹) were obtained at 6861 cm ⁻¹ and 6852
298	cm ⁻¹ both associated with C-H stretching, at 7030 cm ⁻¹ associated with O-H first
299	overtone(Hunter, Kourtellis et al. 2011, Teye, Huang et al. 2014) , at $8000 \sim 8800 \text{ cm}^{-1}$
300	associated with C-H second overtone(Atta 2013). These spectral regions are typical
301	for fat, fatty acid and water in the fish(Atta 2013, Teye, Huang et al. 2014), which
302	have a close relationship with the freshness.

303 3.4. SVMR models

SVMR model with synergy intervals partial least squares (to form Si-SVMR model) and SVMR model with the full spectra were also applied in this work. The top priority was the choice of kernel function for modeling SVMR. Because mapping the original data X nonlinearly into a higher dimensional feature space is implemented by a kernel function(Chauchard, Cogdill et al. 2004). RBF kernel was used in the study referring to the previous work (Gunasekaran, Paulsen et al. 1985). In order to get a better performance, the penalty parameter C and kernel parameter γ have to be

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optimized. The optimal performance of SVMR models was superior with penalty parameter C = 9.2674 and kernel parameter $\gamma = 0.01$ (setting by γ in kernel function RBF) in the work. PCA was used for data dimension reduction. Then 3 PCs of both the full spectra and the joint intervals were as the input of the models. The result of SVMR model with the full spectra is shown in Fig.8. SVMR model had a correlation coefficient of 0.9410 in the calibration set and 0.9192 in prediction set, with RMSECV=0.03177 and RMSEP= 0.03656 respectively, which displayed a good performance but under fitted in the prediction model. Besides, the result of Si-SVMR model was shown in Fig.9. From Fig.9, R =0.9595 for the calibration set and 0.9374 for the prediction set, with RMSECV =0.02710 and RMSEP = 0.03653 respectively, which indicated an excellent performance of the prediction model. The predictive values of Si-SVMR model and the measured K values from the prediction set are shown in Table 3. The average relative error is 0.084. The model has a relatively large error for the prediction of K value. But for the freshness discrimination the error is still relatively reasonable. The first fresh sample was predicted as the secondary freshness with K=0.2335. The 30th secondary fresh sample was estimated as the inedible object with K=0.4232. And 5 inedible samples were predicted as the secondary freshness (20% <K<40%). Then Si-SVMR model has an accuracy of 87% for the freshness discrimination. The prediction accuracy of K value near the critical value has a great influence for the freshness discrimination. The prediction accuracy of model still needs to be optimized.

As shown in Table 4. Si-SVMR was compared with different PLS models and SVMR model with the full spectra. It was obvious that i-PLS model had the worst generalization performance among the six. It is possible that some important information were left out with the single interval. In the contrast, PLS model with the full spectra which included all the spectra had more noise and redundant information, which deprived the performance of the model. Si-PLS model merged together the advantages of i-PLS and PLS, where as much as useful information remained and noises were eliminated. So Si-PLS model was better than i-PLS and PLS models. Even so, the performance of PLS models are inferior to SVMR models. One of

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reasons is that two SVMR models belong to the non-linear recognition model while Si-PLS model used here belong to the linear recognition model (Godoy, Vega et al. 2014). Generally, nonlinear models have significantly stronger self-learning ability in the training process than the linear model. In fact there are also some potential nonlinear relationship between spectral data and chemical measurements. So the performance of the two SVMR models was better than the three PLS models. Si-SVMR model had the best generalization performance of the six. It is possible that the full spectra region contains more noise and redundant information, which weaken the predictive performance of the model. Conversely Si-SVMR selected multiple spectral sub-intervals that are informative and irrelevant information eliminated that could deprive the performance of the model. Besides, the Si-SVMR model with the less wave numbers is more efficient than SVMR model. When all the above factors are taken into account, NIRS combined with Si-SVMR model was more suitable for the detection of K value of fish.

4.0 Conclusions

This study shows that FT-NIRS coupled with chemometrics techniques could be used for the quantitative prediction of K value of silver chub during the cold storage. The eye of the silver chub is a feasible location for the detection by NIR spectroscopy, due to the simple structure of fish eyes, the constant location and close relationship with the fish freshness during the cold storage. The chemometrics treatment by (Si-PLS, SVMR and Si-SVMR) of spectral data was faster, simpler and more convenient than HPLC. Si-SVMR model had the best performance with a correlation coefficient of 95.59% in the calibration set and 93.74% in prediction set, which showed that NIRS together with Si-SVMR model has a high potential for the prediction of K value in silver chubs during the cold storage. Although the performance of Si-SVMR model needs to be further improved, the study could provide a reference for the quantitative analysis of the fish freshness by near infrared spectroscopy.

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8 9	374	References
10 11	375	Alimelli, A., G. Pennazza, M. Santonico, R. Paolesse, D. Filippini, A. D'Amico, I.
12 13	376	Lundström and C. Di Natale (2007). "Fish freshness detection by a computer screen
14	377	photoassisted based gas sensor array." Analytica chimica acta 582 (2): 320-328.
16	378	Alves J C L and R J Poppi (2013) "Biodiesel content determination in diesel fuel
17 18	379	blends using near infrared (NIR) spectroscopy and support vector machines (SVM) "
19 20	220	Talanta 104: 155 161
21 22	380	$\frac{1}{1} \frac{1}{1} \frac{1}$
23	381	Atta, K. (2013). "Morphological, anatomical and histological studies on the olfactory
24 25	382	organs and eyes of teleost fish:< i> Anguilla anguilla in relation to its feeding
26	383	habits." The Journal of Basic & Applied Zoology 66(3): 101-108.
27 28	384	Bøknæs, N., K. N. Jensen, C. M. Andersen and H. Martens (2002). "Freshness
29 30	385	assessment of thawed and chilled cod fillets packed in modified atmosphere using
31 32	386	near-infrared spectroscopy." LWT-Food Science and Technology 35(7): 628-634.
33	387	Bizri, M., JF. Bouhours, D. Garin and R. Got (1985). "Effect of acclimation
34 35	388	temperature on $\langle i \rangle K \langle i \rangle \langle sub \rangle m \langle sub \rangle$ values towards GDP-mannose of the
36 37	389	microsomal dolichol phosphate mannosyltransferase activity of trout liver (< i>
38 39	390	Salmo gairdnerii) " Comparative Biochemistry and Physiology Part B:
40 41	201	Comparative Dischamistry 80 (4): 602 606
42	391	<u>Comparative Biochemistry</u> 80(4). 693-696.
43	392	Buerck, J., S. Roth, K. Kraemer, M. Scholz and N. Klaas (2001). "Application of a
45	393	fiber-optic NIR-EFA sensor system for in situ monitoring of aromatic hydrocarbons in
46 47	394	contaminated groundwater." Journal of hazardous materials 83(1): 11-28.
48	395	Chauchard, F., R. Cogdill, S. Roussel, J. Roger and V. Bellon-Maurel (2004).
50	396	"Application of LS-SVM to non-linear phenomena in NIR spectroscopy: development
52	397	of a robust and portable sensor for acidity prediction in grapes." Chemometrics and
53 54	398	Intelligent Laboratory Systems 71(2): 141-150
55 56	570	
50 57	399	Chen, Q., J. Ding, J. Cai and J. Zhao (2012). "Rapid measurement of total acid
58 59	400	content (TAC) in vinegar using near infrared spectroscopy based on efficient variables
60		

Analytical Methods Accepted Manuscript

selection algorithm and nonlinear regression tools." Food chemistry 135(2): 590-595. Chen, Q., J. Zhao, S. Chaitep and Z. Guo (2009). "Simultaneous analysis of main catechins contents in green tea (< i> Camellia sinensis</i>(L.)) by Fourier transform near infrared reflectance (FT-NIR) spectroscopy." Food Chemistry 113(4): 1272-1277. Chen, Q., J. Zhao, C. Fang and D. Wang (2007). "Feasibility study on identification of green, black and Oolong teas using near-infrared reflectance spectroscopy based on support vector machine (SVM)." Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 66(3): 568-574. Devos, O., C. Ruckebusch, A. Durand, L. Duponchel and J.-P. Huvenne (2009). "Support vector machines (SVM) in near infrared (NIR) spectroscopy: Focus on parameters optimization and model interpretation." Chemometrics and Intelligent Laboratory Systems **96**(1): 27-33. Dowlati, M., M. de la Guardia, M. Dowlati and S. S. Mohtasebi (2012). "Application of machine-vision techniques to fish-quality assessment." TrAC Trends in Analytical Chemistry **40**: 168-179. Dowlati, M., S. S. Mohtasebi, M. Omid, S. H. Razavi, M. Jamzad and M. de la Guardia (2013). "Freshness assessment of gilthead sea bream ($\langle i \rangle$ Sparus aurata $\langle i \rangle$) by machine vision based on gill and eye color changes." Journal of Food Engineering (2): 277-287. FuBin, X., H. XingYi, D. Ran, G. HaiYang, Y. LiYa and D. Huang (2012). "Freshness evaluation model of Pseudosciaena crocea based on near-infrared spectra." Journal of Food Safety and Quality **3**(6): 644-648. Gil, L., J. M. Barat, E. Garcia-Breijo, J. Ibañez, R. Martínez-Máñez, J. Soto, E. Llobet, J. Brezmes, M. Aristoy and F. Toldrá (2008). "Fish freshness analysis using metallic potentiometric electrodes." Sensors and Actuators B: Chemical 131(2): 362-370. Godoy, J. L., J. R. Vega and J. L. Marchetti (2014). "Relationships between PCA and PLS-regression." Chemometrics and Intelligent Laboratory Systems 130: 182-191. Gunasekaran, S., M. Paulsen and G. Shove (1985). "Optical methods for nondestructive quality evaluation of agricultural and biological materials." Journal of Agricultural Engineering Research 32(3): 209-241.

Analytical Methods

431	Huang, X., J. Xin and J. Zhao (2011). "A novel technique for rapid evaluation of fish
432	freshness using colorimetric sensor array." Journal of Food Engineering 105(4):
433	632-637.
434	Hunter, M. T., A. G. Kourtellis, C. D. Ziomek and W. B. Mikhael (2011).
435	"Fundamentals of modern spectral analysis." Instrumentation & Measurement
436	<u>Magazine, IEEE</u> 14(4): 12-16.
437	Jamshidi, B., S. Minaei, E. Mohajerani and H. Ghassemian (2012). "Reflectance
438	Vis/NIR spectroscopy for nondestructive taste characterization of Valencia oranges."
439	Computers and Electronics in Agriculture 85: 64-69.
440	Kamalakanth, C., J. Ginson, J. Bindu, R. Venkateswarlu, S. Das, O. Chauhan and T.
441	Gopal (2011). "Effect of high pressure on K-value, microbial and sensory
442	characteristics of yellowfin tuna (< i> Thunnus albacares) chunks in EVOH films
443	during chill storage." <u>Innovative Food Science & Emerging Technologies</u> 12(4):
444	451-455.
445	Khodabux, K., M. S. S. L'Omelette, S. Jhaumeer-Laulloo, P. Ramasami and P.
446	Rondeau (2007). "Chemical and near-infrared determination of moisture, fat and
447	protein in tuna fishes." Food chemistry 102(3): 669-675.
448	Kimiya, T., A. H. Sivertsen and K. Heia (2013). "VIS/NIR spectroscopy for
449	non-destructive freshness assessment of Atlantic salmon (< i> Salmo salar L.)
450	fillets." Journal of Food Engineering 116(3): 758-764.
451	Li, J., L. Wang, X. Zhang and H. Li (2013). "Rapid Assessment of the Freshness of
452	Large Yellow Croaker by Near Infrared Spectroscopy Combined with PLS Methods."
453	Journal of Chinese Institute of Food Science and Technology 13(6): 209-215.
454	Losada, V., C. Piñeiro, J. Barros-Velázquez and S. P. Aubourg (2005). "Inhibition of
455	chemical changes related to freshness loss during storage of horse mackerel (< i>
456	Trachurus trachurus) in slurry ice." <u>Food Chemistry</u> 93 (4): 619-625.
457	Lougovois, V. P., E. R. Kyranas and V. R. Kyrana (2003). "Comparison of selected
458	methods of assessing freshness quality and remaining storage life of iced gilthead sea
459	bream (< i> Sparus aurata)." <u>Food Research International</u> 36 (6): 551-560.
460	Nicolaï, B. M., K. Beullens, E. Bobelyn, A. Peirs, W. Saeys, K. I. Theron and J.

Analytical Methods Accepted Manuscript

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333333344444444445555555555555555555555	23456789012345678901234567890

1

Lammertyn (2007). "Nondestructive measurement of fruit and vegetable quality by
means of NIR spectroscopy: A review." <u>Postharvest Biology and Technology</u> 46(2):
99-118.
Nollet, L. and F. Toldra (2010). Seafood and seafood product analysis, CRC Press,
Boca Raton, FL, USA.
Pons-Sánchez-Cascado, S., M. Vidal-Carou, M. Nunes and M. Veciana-Nogues

467 (2006). "Sensory analysis to assess the freshness of Mediterranean anchovies (< i>
468 Engraulis encrasicholus</i>) stored in ice." <u>Food Control</u> 17(7): 564-569.

469 Porfire, A., L. Rus, A. L. Vonica and I. Tomuta (2012). "High-throughput
470 NIR-chemometric methods for determination of drug content and pharmaceutical
471 properties of indapamide powder blends for tabletting." Journal of pharmaceutical and
472 biomedical analysis 70: 301-309.

473 Ribeiro, T., S. Raja, A. S. Rodrigues, F. Fernandes, C. Baleizão and J. P. S. Farinha
474 (2014). "NIR and visible perylenediimide-silica nanoparticles for laser scanning
475 bioimaging." <u>Dyes and Pigments</u>.

476 Ruiz-Rico, M., A. Fuentes, R. Masot, M. Alcañiz, I. Fernández-Segovia and J. M.
477 Barat (2013). "Use of the voltammetric tongue in fresh cod (< i> Gadus morhua</i>)
478 quality assessment." Innovative Food Science & Emerging Technologies 18: 256-263.

Teye, E., X.-y. Huang, W. Lei and H. Dai (2014). "Feasibility study on the use of
Fourier transform near-infrared spectroscopy together with chemometrics to
discriminate and quantify adulteration in cocoa beans." <u>Food Research International</u>
55: 288-293.

Teye, E., X. Huang, H. Dai and Q. Chen (2013). "Rapid differentiation of Ghana
cocoa beans by FT-NIR spectroscopy coupled with multivariate classification."
<u>Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy</u> 114: 183-189.
Tito, N., T. Rodemann and S. Powell (2012). "Use of near infrared spectroscopy to

487 predict microbial numbers on Atlantic salmon." Food microbiology 32(2): 431-436.

488 Watanabe, E., Y. Tamada and N. Hamada-Sato (2005). "Development of quality

489 evaluation sensor for fish freshness control based on < i> K</i>< sub> I</sub> value."

490 <u>Biosensors and Bioelectronics</u> **21**(3): 534-538.

491	Wei, Z. J. L. X. W. and X. Wu (2007). "Measurment of Bio-impedance Characteristic
492	of Freshwater Fish Based on Virtual Instrument [J]." Transactions of the Chinese
493	Society for Agricultural Machinery 9: 028.
494	Xu, L., D. Xie and F. Fan (2011). "Effects of Pretreatment Methods and Bands
495	Selection on Soil Nutrient Hyperspectral Evaluation." Procedia Environmental
496	<u>Sciences</u> 10: 2420-2425.
497	Zaragozá, P., S. Ribes, A. Fuentes, JL. Vivancos, I. Fernández-Segovia, J. V. Ros-Lis
498	J. M. Barat and R. Martínez-Máñez (2012). "Fish freshness decay measurement with
499	a colorimetric array." Procedia Engineering 47: 1362-1365.
500	Zhang, F., S. Xu and Z. Wang (2011). "Pre-treatment optimization and properties of
501	gelatin from freshwater fish scales." Food and Bioproducts Processing 89(3):
502	185-193.
503	

Tables

505 Table 1 The performance of Si-PLS model with different preprocessing methods

Methods	R _{cal}	R _{pre}	PCs
SG	0.8669	0.8675	8
SNV	0.8338	0.8117	8
D1	0.8753	0.8779	8
D2	0.8274	0.7731	8
MSC	0.8322	0.8156	8
Centralization	0.8669	0.8675	8
SG-D1	0.8880	0.8776	8

507 Table 2 Calibration results by Si-PLS model with different spectral range selection

Number of	PCs Selected intervals		RMSECV	RMSEP	
intervals					
15	8	1,4,9,10,12,13,5,7	0.0446	0.0504	
16	8	4,8,9,10,13,14,1,7,11	0.0453	0.0512	
17	8	4,8,9,11,14	0.0458	0.0489	
18	8	4,5,8,9,7,15	0.0454	0.0481	
19	8	5,10,9,13,12,17,16	0.0454	0.0499	

20	8	5,9,13,11,16,17	0.0474	0.0464
21	8	5,10,11,14,15,16,17	0.0421	0.0443
22	8	5,6,1,12,10,13,14,18	0.0463	0.0515
23	8	6,11,12,14,19,23	0.0456	0.0503
24	8	6,11,13,16,20	0.0467	0.0496
25	8	6,12,13,16,21,1,5	0.0432	0.0553

Table 3 The error analysis of measured and predictive K values in Si-SVMR

prediction set					
Sample	Measured	Predictive	Absolute	Relative	
number	K value	value	error	error	
1	0.1910	0.2335	0.0425	0.2223	
2	0.2026	0.2089	0.0063	0.0312	
3	0.2081	0.2150	0.0069	0.0331	
4	0.2117	0.2699	0.0582	0.2750	
5	0.2250	0.2010	0.0240	0.1067	
6	0.2300	0.2151	0.0149	0.0648	
7	0.2364	0.2843	0.0479	0.2028	
8	0.2409	0.2609	0.0200	0.0830	
9	0.2568	0.2686	0.0118	0.0461	
10	0.2641	0.2723	0.0082	0.0312	
11	0.2690	0.3255	0.0565	0.2101	
12	0.2739	0.3316	0.0578	0.2109	
13	0.2799	0.3335	0.0536	0.1915	
14	0.2861	0.2968	0.0107	0.0375	
15	0.2906	0.3053	0.0147	0.0505	
16	0.2946	0.2975	0.0029	0.0099	
17	0.2993	0.3470	0.0477	0.1593	
18	0.3025	0.3148	0.0123	0.0408	
19	0.3135	0.3547	0.0412	0.1313	

 $\begin{array}{c} 11 \\ 12 \\ 13 \\ 14 \\ 15 \\ 16 \\ 17 \\ 18 \\ 19 \\ 20 \\ 21 \\ 22 \\ 23 \\ 24 \\ 25 \end{array}$

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20	0.3204	0.3272	0.0068	0.0212
21	0.3306	0.3341	0.0035	0.0107
22	0.3349	0.3526	0.0177	0.0528
23	0.3395	0.3504	0.0109	0.0321
24	0.3434	0.3736	0.0302	0.0878
25	0.3491	0.3714	0.0223	0.0640
26	0.3526	0.3887	0.0360	0.1022
27	0.3689	0.3958	0.0269	0.0730
28	0.3712	0.2989	0.0723	0.1948
29	0.3750	0.3701	0.0049	0.0131
30	0.3791	0.4232	0.0441	0.1164
31	0.3914	0.3610	0.0304	0.0777
32	0.3971	0.3798	0.0173	0.0436
33	0.4055	0.3663	0.0392	0.0966
34	0.4075	0.3861	0.0214	0.0524
35	0.4170	0.4133	0.0037	0.0088
36	0.4214	0.3909	0.0305	0.0724
37	0.4292	0.3925	0.0367	0.0855
38	0.4333	0.3950	0.0382	0.0882
39	0.4385	0.4342	0.0042	0.0097
40	0.4405	0.4116	0.0289	0.0656
41	0.4503	0.4095	0.0408	0.0905
42	0.4539	0.4727	0.0188	0.0414
43	0.4566	0.4077	0.0488	0.1070
44	0.4586	0.4266	0.0320	0.0698
45	0.4603	0.4242	0.0361	0.0784
46	0.4634	0.4071	0.0563	0.1215
47	0.4774	0.4409	0.0364	0.0763
48	0.4809	0.4573	0.0237	0.0492

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49	0.4850	0.4797	0.0053	0.0109
50	0.4892	0.4103	0.0789	0.1612
51	0.4923	0.4757	0.0166	0.0337
52	0.4972	0.4176	0.0796	0.1600
53	0.5100	0.5203	0.0103	0.0202
54	0.5189	0.4813	0.0376	0.0724
55	0.5201	0.5104	0.0097	0.0187
Average	0.3661	0.3635	0.0290	0.0840
value				

515 Table 4 Results of training and predicting based on SVMR and Bi-PLS model

Re	Regression model	number	Principal	The training set		The prediction set	
10		of	component	r	RMSECV	r	RMSEP
		variable	factors				
	PLS	1557	8	0.8289	0.0512	0.8087	0.0553
	iPLS	92	5	0.8096	0.0532	0.8017	0.0554
	Si-PLS	523	8	0.8880	0.0421	0.8776	0.0443
5	SVMR	1557	3	0.94104	0.031770	0.91919	0.036561
Si	i-SVMR	523	3	0.95947	0.027095	0.93743	0.036525





Fig.4. Scatter plots of PLS model with NIR estimation and experimental K value



Fig.5. Scatter plots of i-PLS model with NIR estimation and experimental K value



Fig.6. Scatter plots of Si-PLS model with NIR estimation and experimental K value

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