

# Analytical Methods

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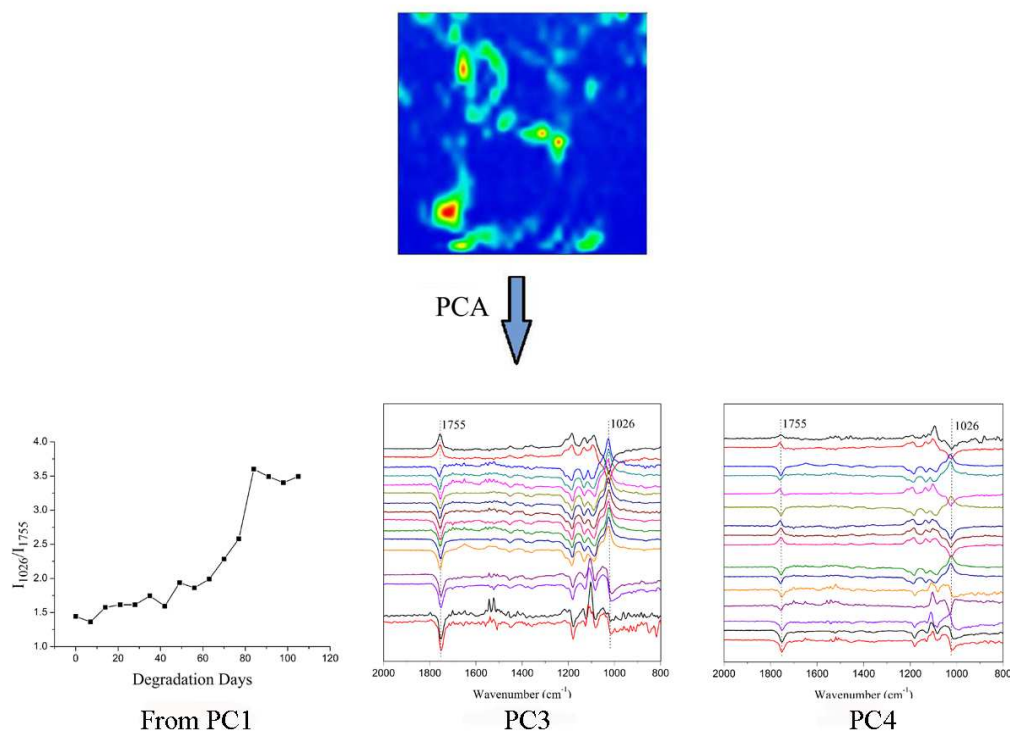
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## Table of Contents Entry



This paper proposes coupling ATR/FTIR mapping with principal component analysis for the biomimetic degradation of porous poly(L-lactide)/hydroxyapatite (PLLA/HA) composite material.

1  
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4 1 **Attenuated Total Reflectance/Fourier Transform Infrared**  
5  
6 2 **(ATR/FTIR) Mapping Coupled with Principal Component**  
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8 3 **Analysis for the Study of *In Vitro* Degradation of Porous**  
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10 4 **Poly lactide/Hydroxyapatite Composite Material**

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16 6 Nan Jing, Xiaoting Jiang, Qian Wang, Yongjiao Tang and Pudun Zhang\*  
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21 8 **Abstract:**

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24 9 Attenuated total reflectance/Fourier transform infrared (ATR/FTIR) mapping was  
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26 10 used to characterize the degradation of porous polylactide/hydroxyapatite (PLLA/HA)  
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28 11 composite material. The ATR/FTIR images were acquired using a Continuum XL  
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30 12 FTIR imaging microscope coupled with a Slide-On Si ATR accessory at a resolution  
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32 13 of  $8\text{ cm}^{-1}$  with 8 co-added scans in the wavenumber range of  $4000\text{--}650\text{ cm}^{-1}$ .  
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34 14 Principal component analysis (PCA) method was used to analyze the FTIR image data.  
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36 15 Four principal components (PCs) were identified by analyzing both the scree plot and  
37  
38 16 the loading spectra. The degradation at varied days was clearly described by the curve  
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40 17 of the intensity ratio of the band at  $1026\text{ cm}^{-1}$  to the one at  $1755\text{ cm}^{-1}$  ( $I_{1026}/I_{1755}$ ) in the  
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42 18 first PC's (PC1's) loading plot versus the days. The shape and position of the second  
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44 19 PC (PC2) is similar to the IR spectrum of water, implying that it comes from the  
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46 20 contribution of water absorbed on the porous sample. The variations of the bands at  
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48 21  $1755\text{ cm}^{-1}$  and  $1026\text{ cm}^{-1}$  in PC3's loading plots reflected the changes of PLLA and

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4 1 HA in different days, respectively, while the fourth PC (PC4) suggested a transition  
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6 2 from the abstract signal to the noise. These results demonstrate that ATR/FTIR  
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8 3 mapping coupled with PCA could effectively characterize the degradation process of  
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10 4 porous PLLA/ HA composite material.  
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## 16 **1 Introduction**

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19 7 Fourier transform infrared (FTIR) spectroscopic imaging is a chemical imaging  
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21 8 technique developed in recent twenty years<sup>1</sup> for detecting the distribution of chemical  
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23 9 components of a sample in a micro-region. It expands traditional FTIR technique from  
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26 10 “point” analysis to “plane”, even “stereoscopic” analysis. This technique can be  
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28 11 achieved by mapping or imaging on the basis of the adopted detector.<sup>2</sup> As with  
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30 12 traditional FTIR method, three collecting modes, *i.e.* transmission, reflection and  
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32 13 attenuated total reflectance (ATR), are provided for FTIR mapping or imaging. FTIR  
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34 14 imaging has been applied in life sciences,<sup>3-6</sup> biomedical science,<sup>7-9</sup> polymers,<sup>10-13</sup>  
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36 15 artworks and archeology,<sup>14-16</sup> and forensic science,<sup>17,18</sup> etc.  
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42 16 Univariate analysis is often used in exploring FTIR images, in which one or two  
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44 17 characteristic absorption bands of a particular component are selected to build visual  
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46 18 chemical image and further to obtain the composition and distribution information of  
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48 19 components in a sample.<sup>19</sup> Though this method is simple and easy-to-understand, it  
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50 20 cannot accurately reflect the compositions and distribution of a multi-component  
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52 21 sample or a sample with complex structure because of the spectral overlapping. More  
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54 22 importantly, since only one or two absorption bands are used, much of the information  
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4 1 is sacrificed and thus it can not represent the whole spectral and spatial distribution of  
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6 2 the components in some cases. Principal component analysis (PCA) is a commonly  
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8 3 used multivariate analysis method.<sup>20-22</sup> When being used in processing FTIR images,  
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10 4 it regards each spectrum (pixel) as a whole rather than just considering individual  
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12 5 absorption band in a spectrum. PCA decomposes the spectral data matrix of an image  
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14 6 into the product of loading and score of a series of principal components (PC), which  
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16 7 represent the spectrum and the concentration distribution of each abstract pure  
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18 8 component, respectively.

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21 9 Hydroxyapatite (HA) is an important biocompatible inorganic component of bone  
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23 10 and tooth of vertebrates.<sup>23</sup> Poly(L-lactide) (PLLA) is a nontoxic polymer and it can be  
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25 11 degraded into CO<sub>2</sub> and H<sub>2</sub>O under wet conditions, which are not harmful to human.  
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27 12 Just because of their excellent biocompatibility, PLLA/HA porous composite materials  
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29 13 are often used as scaffolds for bone tissue engineering.<sup>24</sup> However, to guarantee a  
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31 14 healthy growth of bone, the degradation rate of scaffold must be adapted to the growth  
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33 15 of tissue during the bone-repair process. Due to the great limitations and the strict and  
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35 16 severe conditions for *in vivo* study, most of the degradation was *in vitro* investigated  
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37 17 under biomimetic condition.<sup>25,26</sup> The *in vivo* degradation of the composite material  
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39 18 can be well predicted from the *in vitro* results. In this work, ATR/FTIR mapping  
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41 19 coupled with PCA was used to study the *in vitro* degradation of PLLA/HA composite  
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43 20 material and to further analyze the changes of chemical composition during the  
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45 21 degradation process. Different from our previous work,<sup>27</sup> this presented manuscript  
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47 22 systematically explored the application of PCA in analysis of the FTIR images. PCA  
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4 1 can highlight the subtle but important changes of the spectra that are often difficult to  
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6 2 be observed using univariate methods. Therefore it is an expectable approach to  
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8 3 characterize the degradation of PLLA/HA composites. Our result demonstrates that  
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10 4 some interesting and meaningful results were obtained by using this method.  
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## 16 **2 Experimental**

### 17 **2.1 Instruments, materials and reagents**

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20 8 A Thermo Nexus 8700 FTIR spectrometer coupled with a Continuum XL FTIR  
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22 9 imaging microscope (Thermo Electron, WI, USA) was used to collect the FTIR  
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24 10 images. The microscope was allocated with a Slide-on Si ATR accessory (Thermo  
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26 11 Electron, USA). The biomimetic degradation was conducted in a DK-420S three-way  
27  
28 12 water bath (Shanghai Jinghong, China). Poly(L-lactide) with an inherent viscosity ( $\eta$ )  
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30 13 of 1.22 dl/g and a weight-average molecular weight ( $M_w$ ) of about 121,000 g/mol was  
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32 14 purchased from Jinan Daigang Biomaterials Co. Ltd. (Jinan, China). The rod-like  
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34 15 nano-HA powder with an average length of 150 nm and width of 20 nm was obtained  
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36 16 from Nanjing Emperor Nano Materials Co. Ltd (Nanjing, China). 1,4-Dioxane,  
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38 17 AgNO<sub>3</sub>, NaCl, KCl, Na<sub>2</sub>HPO<sub>4</sub> and KH<sub>2</sub>PO<sub>4</sub> were of analytical grade.  
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### 46 **2.2 Preparation of PLLA/HA composite material**

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49 19 Solvent casting/salt leaching method<sup>27,28</sup> was used to prepare the porous PLLA/HA  
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51 20 composite material. PLLA and HA were first ultrasonically dispersed in two same  
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53 21 volume of dioxane, the two solutions were then mixed together under vigorous  
54  
55 22 magnetic stirring. The ground NaCl powder with particle size of 150-200  $\mu$ m (80-100  
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4 1 mesh) was added as the porogen into the mixed suspension and continued to stir till  
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6 2 the suspension became homogeneous. The as-prepared mixture was cast onto the petri  
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8 3 dishes to a thickness of about 1 mm and then dried at room temperature. The dried  
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10 4 samples were soaked into deionized water to remove the salt. The water was replaced  
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12 5 by the fresh one every 12 hours till the residual chloride ion was no longer detected.  
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14 6 Finally, the obtained composite materials were dried in a vacuum oven at 45 °C for 48  
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16 7 h.  
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### 21 8 **2.3 *In vitro* degradation of PLLA/HA composite material**

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23  
24 9 PLLA/HA composite materials were immersed in phosphate buffered saline (PBS)  
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26 10 solution (pH=7.4) at 37 °C for *in vitro* degradation. A few pieces of sample were taken  
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28 11 out every 7 days and were rinsed with deionized water to remove the residual PBS  
29  
30 12 salts. The buffer solution was simultaneously replaced by the fresh one. The as-treated  
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32 13 materials were dried in a vacuum oven at 45 °C for 24 h.  
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### 36 14 **2.4 ATR/FTIR mapping**

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39 15 The dried PLLA/HA samples were cut into sheets with a size of 1 cm × 1 cm and  
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41 16 was fixed onto a glass slide using a double-sided adhesive tape, which was placed on  
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43 17 the stage of FTIR imaging system and was detected using a liquid nitrogen-cooled  
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45 18 mercury cadmium telluride (MCT) detector. A Slide-on Si ATR accessory was  
46  
47 19 mounted on a 15 × IR objective (N.A.=0.58) for collecting ATR/FTIR images. Before  
48  
49 20 collection, the Si ATR element was pulled out in order that the beams were focused on  
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51 21 the sample. The mapping area with a size of 300 μm × 300 μm was selected in the  
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53 22 bright field of view and then the ATR element was pushed in. The ATR/FTIR images  
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4 were acquired with a pixel size of 10  $\mu\text{m}$  at a resolution of 8  $\text{cm}^{-1}$  with 8 co-added  
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6 scans in the wavenumber range of 4000–650  $\text{cm}^{-1}$ , implying that 961 spectra (31  $\times$  31)  
7  
8 was involved in an ATR/FTIR image. The scanning step was triggered once the Si  
9  
10 element was contacted with the sample surface. The acquiring, the processing and the  
11  
12 analysis of the ATR/FTIR images were implemented using the Atlus software of  
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14 Omnic 7.2 (Thermo Electron). It was stressed that the mapping areas are not the same  
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16 at each time because all the FTIR images were off-line collected. The collection of  
17  
18 ATR/FTIR images at different area/location is better than at only one micro-region to  
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20 characterize the degradation behavior.  
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### 29 **3 Results and discussion**

#### 30 **3.1 ATR/FTIR spectra of PLLA and HA**

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33 Fig.1 shows the ATR/FTIR spectra of PLLA, HA and their mixture. The bands at  
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35 1755  $\text{cm}^{-1}$ , 1184  $\text{cm}^{-1}$  and 1088  $\text{cm}^{-1}$  in PLLA's spectrum arising from the stretch  
36  
37 vibration of the C=O group ( $\nu_{\text{C=O}}$ ), the asymmetric and symmetric stretch vibrations  
38  
39 of the C-O-C group ( $\nu_{\text{as(C-O-C)}}$  and  $\nu_{\text{s(C-O-C)}}$ ), respectively; while the band at 1026  $\text{cm}^{-1}$   
40  
41 attributes to the stretch vibration of the P-O group of HA. It is observed that this band  
42  
43 is overlapped with the one at 1044  $\text{cm}^{-1}$  ( $\nu_{\text{C-CH}_3}$ ) of PLLA in the spectrum of their  
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45 mixture.  
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21 **Fig. 1**



### 3.2 Interpretation of ATR/FTIR image

Fig.2 (a) shows an ATR/FTIR image of non-degrading PLLA/HA composite. Since degradation always occurred on PLLA and HA remained unchanged throughout the degradation process, all the FTIR images involved in this paper are generated based on the absorption band of the carbonyl group of PLLA at  $1755\text{ cm}^{-1}$ . Fig.2 (b) shows the color scale in which the absorbance sequentially increases from blue to red.

An FTIR image contains large amounts of spectral information in which each pixel corresponds to an infrared spectrum. Fig.2(c) shows the infrared spectra in different pixels in Fig.2(a). The blue area has no any absorption signal other than noise, representing the pore region of the composite; while other color areas show the spectra of the mixture of PLLA and HA, suggesting the pore walls of the composite.

**Fig. 2**

### 3.3 The principle of PCA for FTIR image

FTIR image data usually consist of four dimensions, *i.e.*  $x$ - and  $y$ - dimensions (defining the spatial position), a spectral dimension (describing the chemical information of components) and an intensity dimension (corresponding to the concentration of component). These raw data cannot be completely displayed on three-dimensional images, thus a big challenge in analysis of FTIR image is how to effectively extract information from these hyperspectral images.

The spectrum of a pixel in FTIR image is essentially the weighted sums of the

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4 1 product of each pure spectrum by the concentration of the pure component in this  
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6 2 pixel based on the Beer-Lambert law. Although the concentration of each pure  
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8 3 component varied with pixel, the spectra are identical. FTIR image data can be  
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10 4 expressed using a matrix  $\mathbf{D}=\mathbf{C}\mathbf{S}^T+\mathbf{E}$ , where  $\mathbf{D}$  is the original experimental data matrix  
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12 5 with a size that equals the number of pixel  $\times$  number of wavenumbers,  $\mathbf{C}$  the  
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14 6 concentration matrix of the components with a size that equals the number of pixel  $\times$   
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16 7 number of components,  $\mathbf{S}^T$  the spectral matrix consisting of the spectra of pure  
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18 8 components with a size that equals number of components  $\times$  number of wavenumbers,  
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20 9 and  $\mathbf{E}$  is the error matrix. All of the image data are mean-centered by spectrum before  
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22 10 PCA and are then linearly decomposed into a few uncorrelated new variables (*i.e.*  
23  
24 11 PCs). The new variables are ranked by decreasing order of contribution to total  
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26 12 variance, meaning that the first PC (PC1) covers the largest variance and PC2 covers  
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28 13 the second largest variance and so forth. After PCA, the FTIR image data are  
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30 14 expressed as matrix  $\mathbf{D}=\mathbf{T}\mathbf{P}^T+\mathbf{E}$ , where  $\mathbf{T}$  is the score matrix, in which the column  
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32 15 vectors can be folded back to form the score maps, representing the abstract  
33  
34 16 concentration distribution; while  $\mathbf{P}^T$  is the loading matrix, in which the row vectors  
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36 17 represent the abstract pure spectra.<sup>29</sup> The loading plots are usually associated with  
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38 18 infrared spectra,<sup>30,31</sup> in which the positive or negative bands indicate that certain  
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40 19 characteristics are appearing or missing in the corresponding PCs.  
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### 51 3.4 Determination of the number of PCs

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54 21 In principle, the number of PCs is no larger than the number of wavenumbers.  
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56 22 However, since experimental errors always exist when acquiring FTIR image, only  
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1 the first few PCs can describe the spectral variations relating to the chemical  
2 composition, others are the contribution of noise. Therefore, how to determine the  
3 number of PCs is an important issue for analyzing FTIR image. Scree plot is one of  
4 the often used methods,<sup>20</sup> in which a plot of the eigenvalue (or the fraction of total  
5 variance) of each PC versus the serial number of PC is created. Generally only the  
6 first few PCs has large eigenvalues and thus show a steep slope in the scree plot, other  
7 PCs forms a “scree-like” platform due to their small eigenvalues. The PCs before  
8 inflexion in the scree plot are usually selected for PCA. Fig. 3 shows the scree plot of  
9 PLLA/HA composite at day 84, which indicates that only PC1 is the valuable  
10 component. Similar results were also achieved by analyzing the composites in other  
11 degradation days. This method had been successfully applied in our previous work.<sup>13</sup>

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13 **Fig. 3**

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15 Nevertheless, when observing the loading plots (Fig. 4), we find that they do not  
16 display the noise until PC5; while for PC2-PC4, some “characteristic bands” are still  
17 shown in the loadings even though their eigenvalues are small. These results imply  
18 that determination of the number of PCs using scree plot method may sometimes  
19 leads to the loss of some valuable PCs. A solution to this problem is to compare the  
20 loadings with the infrared spectra of the real components and retain the PCs that show  
21 the “characteristic bands” in loadings.<sup>20</sup> On the basis of this principle, PC1 to PC4  
22 may be retained, however, the shape and position of the loading plot of PC2 is similar

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4 1 to the IR spectrum of water, implying that it might be derived from the adsorption of  
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6 2 water on the porous material during the collection.  
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11 **Fig.4**  
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16 **3.5 Principal component analysis for the degradation of PLLA/HA composite**  
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19 **material**  
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21 8 For samples at every degradation day, the PCA results show that PC1 covers more  
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23 9 than 90% of the total variance and the loading plot (Fig.5(a)) contains nearly the  
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25 10 entire characteristics of the spectrum of the mixture of PLLA and HA, which  
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27 11 represents the average spectrum of the image. Correspondingly, the score map of PC1  
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29 12 (figure not shown) also reflects the distribution of the predominant component, which  
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31 13 is almost consistent with the ATR/FTIR image based on a characteristic band (image  
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33 14 not shown). Further, a degradation curve of the intensity ratio of the band at  $1026\text{ cm}^{-1}$   
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35 15 to the one at  $1755\text{ cm}^{-1}$  in the loadings of PC1 ( $I_{1026}/I_{1755}$ ) versus the degradation days  
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37 16 is achieved (Fig.5(b)). It indicates that this ratio increases slowly before day 56,  
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39 17 suggesting a slow degradation is occurred in this stage. During the period 56-84 day,  
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41 18 this ratio increases rapidly, suggesting the degradation is accelerated. The acceleration  
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43 19 of the degradation might result from the hydrolysis of the polymer which can  
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45 20 auto-catalyze the degradation.<sup>32</sup> These ratios become stable after day 84, suggesting  
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47 21 that some of the HA might be peeled off from the pore walls.  
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**Fig.5**

The loading plots of PC3 of the FTIR images at different degradation days are shown in Fig. 6. The band at  $1755\text{ cm}^{-1}$  corresponds to the characteristic absorption of C=O group in PLLA and is positive at day 0 and 7, indicating that the degradation has not yet been initiated. From the 14th day, the band becomes negative and gradually increases with the days, suggesting that the content of PLLA is reduced with the increase of degradation days. Compared with our previous work that the change of morphology cannot be observed until day 28,<sup>28</sup> ATR/FTIR mapping coupled with PCA can reflect the degradation of composite more effectively. Since the significant degradation of PLLA in the late period resulted in a fragmentation of the sample, it is difficult to collect an ATR/FTIR image after day 105. In addition, the band at  $1026\text{ cm}^{-1}$  in the loadings of PC3 is caused by HA and is nearly unchanged during the period 14-77 day. However, it changes to negative band after day 84, suggesting a decrease of HA's content. This result is identical to the one drawn from the degradation curve (Fig. 5(b)).

**Fig. 6**

Fig.7 shows the loading plots of PC4 at different degradation days. All the bands at  $1755\text{ cm}^{-1}$  show negative values till the materials are degraded more than 63 days, which is consistent with our previous study on the degradation half-life.<sup>28</sup> Since the

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4 1 eigenvalues of all the PC4 are low (<1.6%), it is essentially a minor factor that transits  
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6 2 from the abstract signal to the noise. the variations of the bands (positive or negative)  
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9 3 before day 63 may be caused by the big noise during the collection of FTIR images.  
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11 4 These small eigenvalues can also be applied to interpret the variations of the band at  
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13 5 1026 cm<sup>-1</sup> in PC4.  
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7 **Fig. 7**

#### 9 **4. Conclusion**

10 ATR/FTIR mapping coupled with PCA was used to study the degradation of porous  
11 PLLA/HA composite material. Four PCs were identified by analyzing both the scree  
12 plot and the loadings of the images. The loading plot of PC1 represents the average  
13 spectrum of an image. The degradation curve based on the ratio of the band at 1026  
14 cm<sup>-1</sup> to the one at 1755 cm<sup>-1</sup> ( $I_{1025}/I_{1755}$ ) in the loading plots of PC1 versus the days  
15 indicates that the degradation is slow in the first 56 days and is accelerated during the  
16 period 56-84 day. However, the degradation becomes stable after day 84. PC2  
17 attributes to the water absorbed on the sample. Both PC3 and PC4 show the variations  
18 of the characteristic bands of PLLA and HA during the degradation process. The  
19 loadings of PC3 suggest that the degradation of the composite starts from day 14. In  
20 addition, PC4 reflects a transition from the abstract signal to the noise and the  
21 variations of the bands (positive or negative) in PC4 are caused by the experimental  
22 noise. All other PCs (e.g. PC5 to PC8) only reflect the experimental noise. Our results

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4 1 indicate that ATR/FTIR mapping coupled with PCA is an effective method for  
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6 2 characterizing the degradation of PLLA/HA composite material. Combining further  
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9 3 with some quantitative approaches, it is possible to quantitatively describe the  
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11 4 degradation process and establish a degradation model.  
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## 16 **References**

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4 **1 Figure Captions**  
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3 **Fig.1** FTIR spectra of PLLA, HA and their mixture.  
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5 **Fig.2** FTIR image of non-degrading PLLA/HA composite material  
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6 a) FTIR image based on the absorption band at  $1755\text{cm}^{-1}$ ; b) the color scale; and c)  
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7 FTIR spectra of different pixels in the image.  
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9 **Fig.3** The scree plot of the FTIR image of the PLLA/HA composite at day 84.  
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11 **Fig.4** Loading plots of the first eight PCs of the FTIR image of PLLA/HA composite  
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12 materials at day 84.  
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14 **Fig.5** (a) The loading plots of PC1 of the ATR/FTIR images at day 0, 7, 14, 21, 28, 35,  
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15 42, 49, 56, 63, 70, 77, 84, 91, 98 and 105 (from top to bottom). (b) Degradation curve  
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16 of  $I_{1026}/I_{1755}$  versus the degradation days.  
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18 **Fig.6** The loading plots of PC3 of the FTIR images at day 0, 7, 14, 21, 28, 35, 42, 49,  
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19 56, 63, 70, 77, 84, 91, 98 and 105 (from top to bottom).  
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21 **Fig.7** The loading plots of PC4 of the FTIR images at day 0, 7, 14, 21, 28, 35, 42, 49,  
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22 56, 63, 70, 77, 84, 91, 98 and 105 (from top to bottom)  
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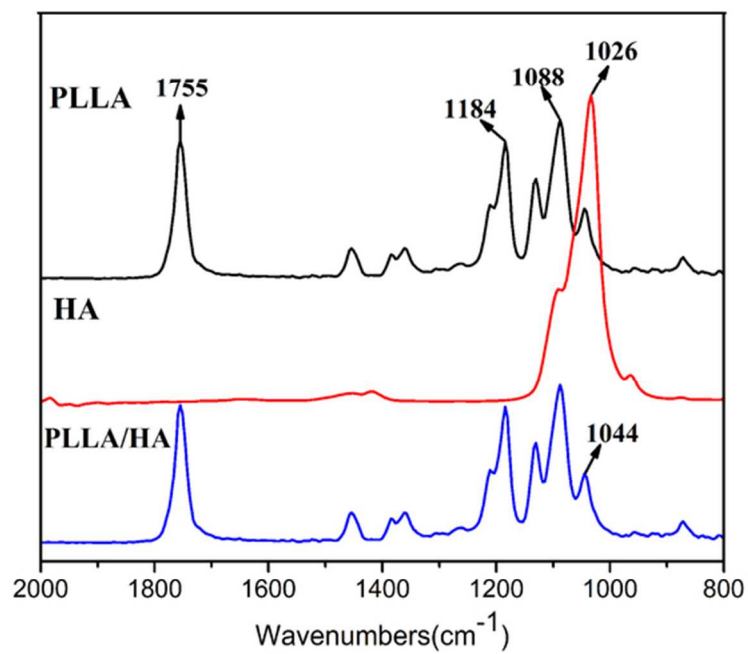


Fig.1  
63x44mm (300 x 300 DPI)

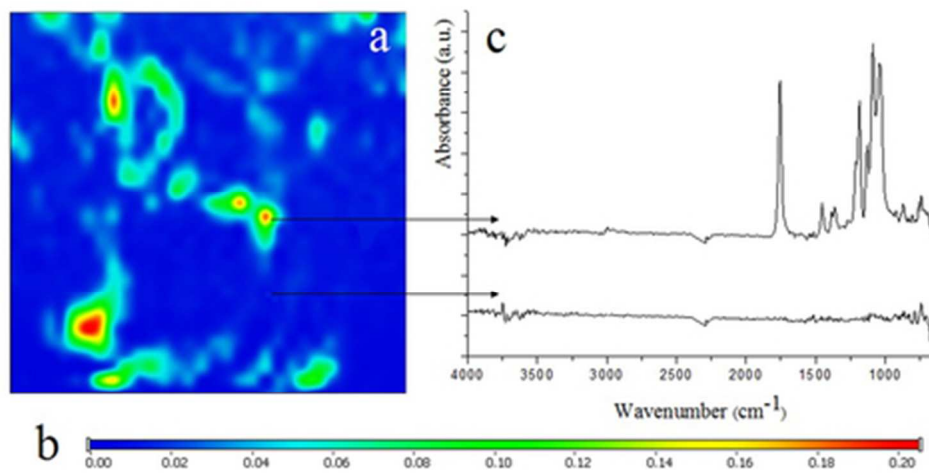


Fig.2  
42x21mm (300 x 300 DPI)

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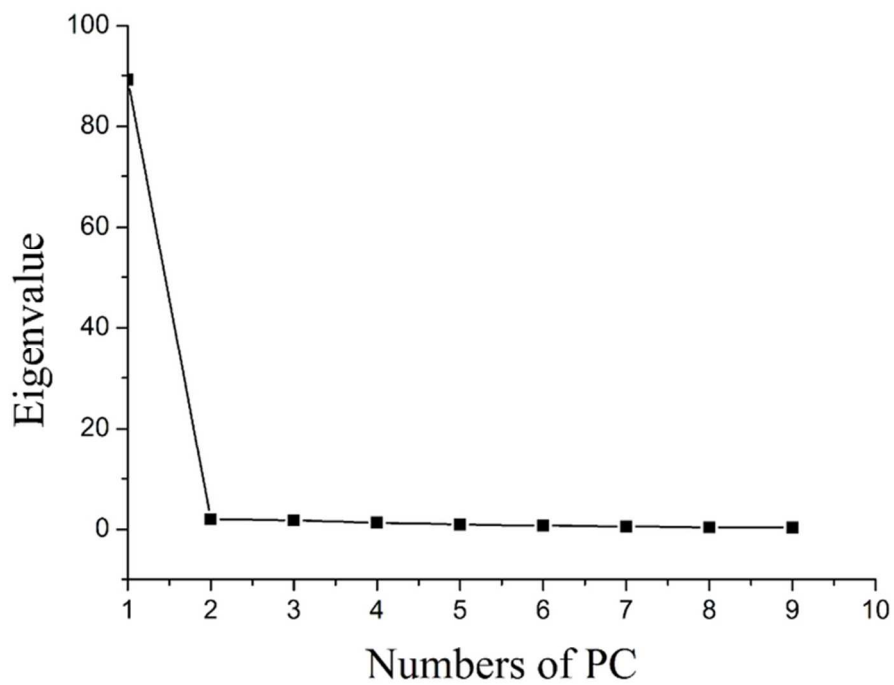


Fig.3  
67x53mm (300 x 300 DPI)

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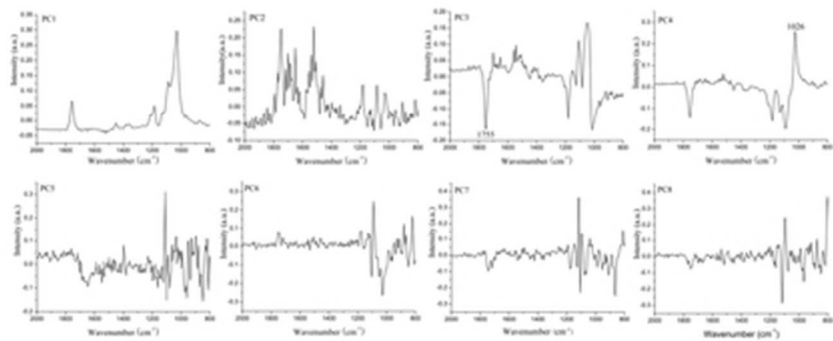


Fig.4  
35x15mm (300 x 300 DPI)

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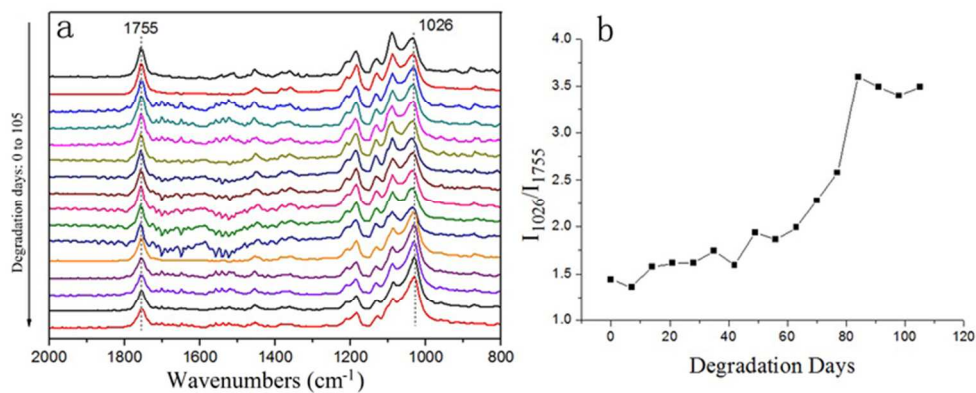


Fig.5  
68x27mm (300 x 300 DPI)

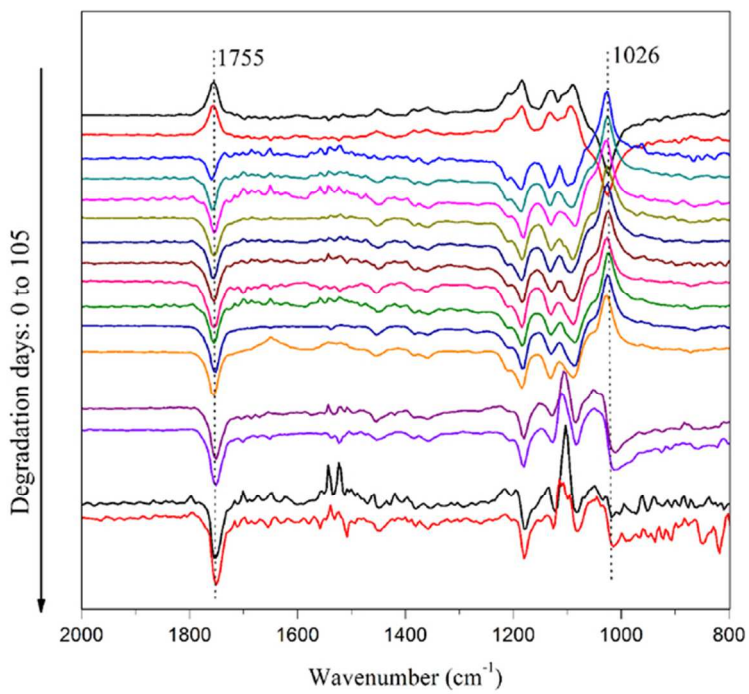


Fig.6  
68x48mm (300 x 300 DPI)

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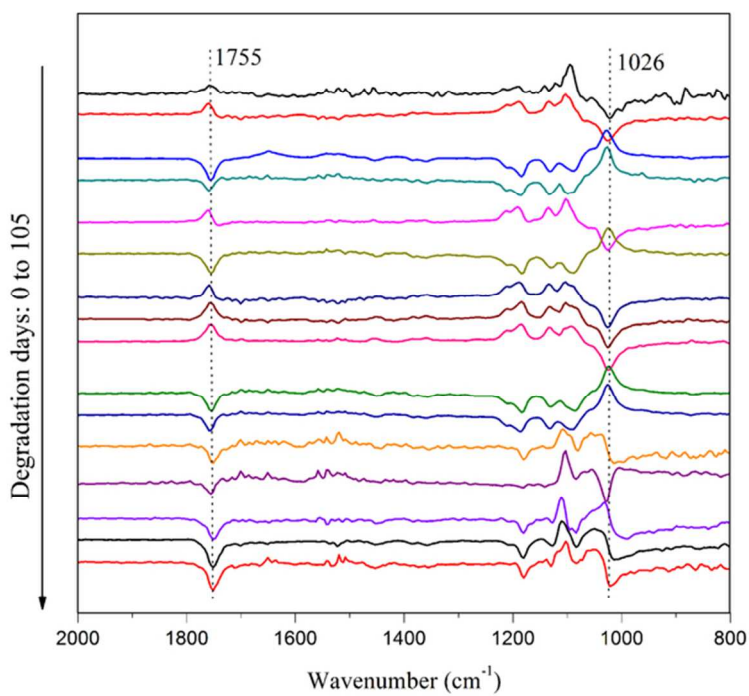


Fig.7  
68x48mm (300 x 300 DPI)