

Analytical Methods

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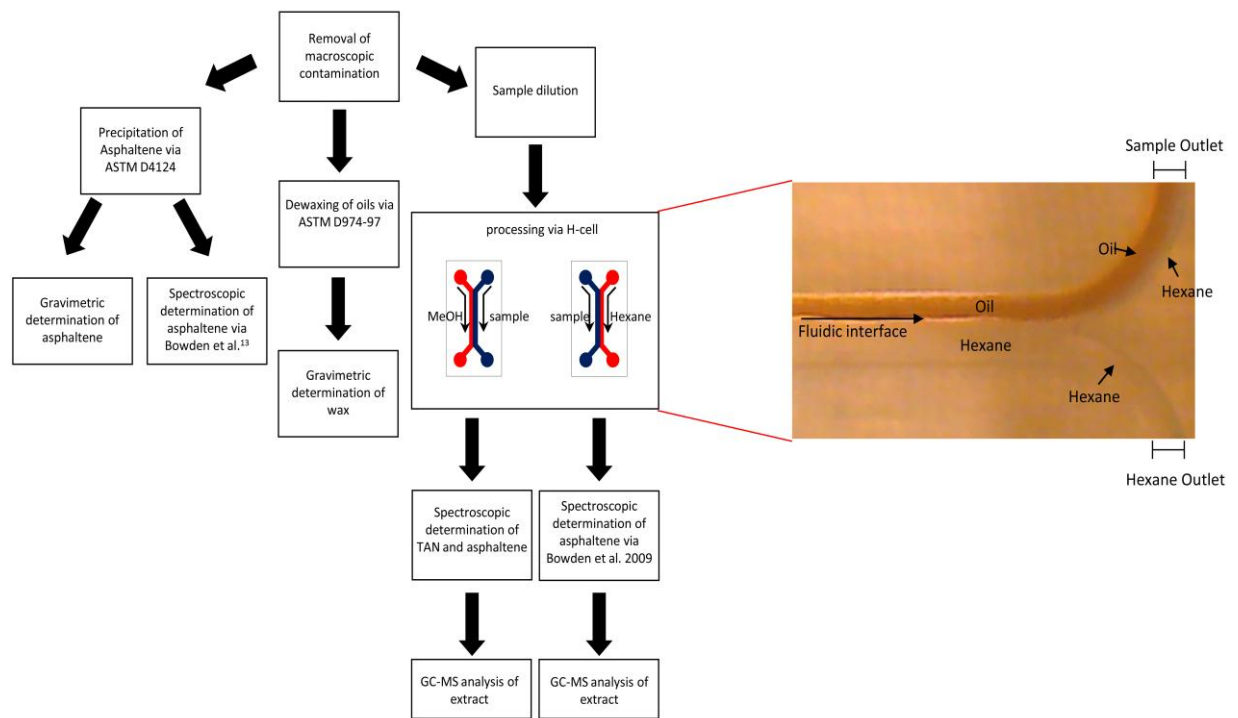
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TABLE OF CONTENT ENTRY



Analytical scheme for H-cell capable of performing rapid and accurate asphaltene and TAN assays.

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4 18 Simultaneous and rapid asphaltene and TAN determination for heavy
5 19 petroleum using an H-cell
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9 21 Oluwarotimi O. Alabi, Stephen A. Bowden, John Parnell

10 22 Dept Geology and Petroleum Geology, University of Aberdeen, Aberdeen,
11 23 AB24 3UE
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15
16 25 Abstract

17 26 Characterising the asphaltene and carboxylic acid (naphthenic acid) content
18 27 of crude oil is important for petroleum production, transport, storage and
19 28 environmental science. This is because, the proportion of asphaltene and the
20 29 concentration of acidic compounds in petroleum can be used to characterise
21 30 viscosity (e.g. producibility), refining potential (e.g. its value) and chemical
22 31 recalcitrance and thus behaviour as a contaminant. Here we present an assay
23 32 for determining the proportion of asphaltene and total acid number (TAN) of
24 33 petroleum. The method utilises a microfluidic component called an H-cell and
25 34 produces an asphaltene-free fraction, either hydrocarbon or methanol-soluble,
26 35 that can be forwarded for further advanced analysis and used to determine
27 36 asphaltene content and TAN value. The H-cell method depends on a
28 37 diffusion-based separation that is only practical when a sample is manipulated
29 38 at a microscale and thus is fundamentally different to previous methods for
30 39 assaying these parameters that utilise solubility- or chromatography-based
31 40 methods. Comparisons of asphaltene and TAN measurements derived from
32 41 the H-cell based assay have very high correlations with the ASTM D4124 and
33 42 ASTM D974-97 methods. Therefore rapid and simultaneous determination of
34 43 asphaltene content and TAN value can be achieved by an H-cell based
35 44 format. While this format is suited to miniaturisation and point of need
36 45 analysis, the main benefit of the H-cell method might be its capacity to provide
37 46 new analytical windows.
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3 48 1.0 Introduction
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5 49 The fractionation of petroleum into its constituent parts falls into two basic
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7 50 categories; thermally driven distillation processes applied in refining and
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9 51 trading petroleum as a commodity and chemical methods applied to evaluate
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11 52 oil and gain crucial technical information that can answer problems
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13 53 encountered during oil exploration, production, transportation and
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15 54 environmental remediation¹. Of the chemical assays reported for asphaltene
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17 55 analysis, the SARA (Saturates, Aromatics, Resins and Asphaltenes) scheme
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19 56 is currently at the fore¹⁻⁴. Standard methods for asphaltene analysis define
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21 57 asphaltene as the component of petroleum that is insoluble when diluted with
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23 58 an excess of *n*-alkane solvent; within ASTM D4124² this is hexane although
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25 59 other solvents may be chosen¹.
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27 60
28 61 A standard method for assaying the acidity of crude oil (ASTM 974-975) is the
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30 62 Total Acid Number (TAN) – number of milligrams of potassium hydroxide
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32 63 required to neutralise the acidity in a gram of petroleum. Particularly when
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34 64 hydrogen sulphide is absent, it is the carboxylic acids (naphthenic acids)
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36 65 present in petroleum that are often responsible for its acidic behaviour and
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38 66 thus corrosion during petroleum transport, storage and refining⁶. Furthermore,
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40 67 the carboxylic acids within petroleum may combine with salts to create
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42 68 surfactants and emulsifying agents that are important for determining the
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44 69 interaction of petroleum with aqueous phases⁷. Carboxylic and naphthenic
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46 70 acids can be extracted from petroleum by ion exchange methods^{8,9}, and the
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48 71 yields obtained generally, although not always, correlate with TAN number.
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50 72
51 73 Complexity in handling and assaying petroleum derives from its chemical
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53 74 heterogeneity, varying physical properties (e.g. viscosity and density) and the
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55 75 sensitivity of methods to a wide range interferences (some of these factors are
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57 76 described from the perspective of an oil field operative in the opening chapter
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59 77 Mullins⁴). The latter can include non-petroleum and non-liquid materials, such
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61 78 as inorganic minerals and compounds that become entrained within petroleum
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63 79 during its production, refining and transport. The consequences of these are
64
65 80 that asphaltene data appears to have considerable noise and this can

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4 81 produce notable variation in the application of methods between laboratories
5 82 – particularly with regard to TAN values⁹. The effect of this is the widespread
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7 83 and prolific modification of methods to create a range of propriety-assays and
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9 84 discipline specific methods. Examples of this include alternatives to the TAN
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11 85 assay (Horvath-Gumulka test – Naphthenic Acid Number and Carboxylic Acid
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13 86 Number¹), utilisation of different solvents for analytical handling of asphaltene
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15 87 e.g. toluene or dichloromethane⁴, and the widespread use of heptane over
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17 88 hexane by academic laboratories in preparing asphaltene-free fractions for
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19 89 biomarker analysis¹⁰. Thus while assaying petroleum for sale as a commodity
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21 90 mandates standardisation of assay formats (e.g. the use of ASTM methods)
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23 91 day to day problem solving and research drives considerable analytical
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25 92 development for what would generally be considered basic assays.

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28 94 In this paper we investigate the application of an H-cell based method to
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30 95 determine asphaltene and carboxylic acid content for a range of petroleum-
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32 96 types. Within an H-cell, two fluids are flown in hard contact with each other,
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34 97 and analytes are permitted to diffuse from one fluid to the other¹¹. To extract
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36 98 specific fractions from petroleum previous work has used hexane¹² and
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38 99 methanol¹³, but only for a limited number of samples. The asphaltene
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40 100 assaying method used here^{12,13} is fundamentally novel in that it does not
41
42 101 strictly fall into either the thermal (evaporative) or chemical (chromatography
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44 102 or solubility) based methods¹ for fractionating petroleum, instead it depends
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46 103 on the varying rates of diffusion of different petroleum components within
47
48 104 liquids to achieve separation¹¹⁻¹³. For a range of sample types we have used
49
50 105 two assay formats; 1) one using hexane to determine asphaltene content and
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52 106 2) another using methanol to simultaneously determine asphaltene content
53
54 107 and TAN number. Despite the potential of H-cell based methods, given the
55
56 108 inherently complex nature of petroleum and potential interferences, a key
57
58 109 need is to evaluate the robustness of the H-Cell methods when applied to a
59
60 110 range of heavy petroleum - both naturally occurring and anthropogenically
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112 111 extracted from the subsurface. There is also a need to scope and document
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114 112 the analytical window provided by H-cell methods with regard to petroleum,
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116 113 and heavy petroleum.

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5 115 2.0 Method

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7 116 Analytical procedures are summarised in Fig. 1, and the methods employed at
8
9 117 each stage are described in the following sections.

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12 119 *2.1 Samples*

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14 120 The petroleum samples used in this study represent a range of physical types
15
16 121 from tar through to conventional oil (Fig. 2). Twelve petroleum samples were
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18 122 chosen for analysis (Table 1). Produced oils were analysed dead (they were
19
20 123 degassed by exposure to ambient conditions) and taken from a stock
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22 124 collection at the University of Aberdeen. Seep samples and naturally occurring
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24 125 bitumen were analysed in the condition in which they were collected, except
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26 126 where obvious physical interferences were present; e.g. dead insects,
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28 127 sediment etc.

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31 129 Two samples (Boa A & B) possessed high wax content. The wax component
32
33 130 of petroleum is known to be a significant interference during asphaltene
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35 131 determination. To provide an assessment of this interference the sample was
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37 132 split, and the aliquot dewaxed prior to further analysis. Wax was removed
38
39 133 from the waxy oils using the method detailed in ASTM D721-06¹⁴. Briefly; an
40
41 134 oil sample is dissolved in a 3:1 Methyl Ethyl Ketone/Toluene mix and cooled
42
43 135 to -32 °C. The precipitated wax is recovered by filtering the solution. The
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45 136 percentage of wax filtrate retained on the filter was determined gravimetrically,
46
47 137 after it had been dried in a desiccator for 24 hours.

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50 139 *2.2 Asphaltene and SARA Analysis*

51 140 The % asphaltene content of samples and SARA composition was obtained
52
53 141 using ASTM D4124². The oils were separated into component constituents of
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55 142 asphaltene and maltene using excess hexane in a 1:40 oil/ hexane ratio.
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57 143 Silica gel column chromatography was then used to separate the maltene
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59 144 fraction into saturate, aromatics, and resin components. The amount of each
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145 fraction, including the asphaltene fraction, was deduced gravimetrically.

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147 *2.3 TAN analysis*

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4 148 The ASTM D974-97⁵ method was used to perform total acid number
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6 149 measurements. In summary, about 2 gram of oil is dissolved in 250ml conical
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8 150 flask using 100ml solution of titration solvent (toluene 500ml, water 5ml and
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10 151 propan-2-ol 495ml). P-Naptholbenzein solution was used as an indicator. The
11
12 152 mixture (a yellow-orange coloration) was titrated with potassium hydroxide
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14 153 solution in small increments until the end point was indicated by a colour
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16 154 change. A blank titration was performed and the Total Acid Number calculated
17
18 155 using the prescribed formula.
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20 157 *2.4 Petroleum Acid Fraction Extraction - Ion Exchange Solid-Phase Extraction*

21 158 Carboxylic acids contained in the acid fraction of 9 crude oils were extracted
22
23 159 using the Ion Exchange Chromatography (IEC). The Solid-Phase Extraction
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25 160 (SPE) method described in Jones et al.⁸ was used on oil samples. In
26
27 161 summary, a SAX quaternary amine SPE ion exchange column was
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29 162 conditioned with 40ml of *n*-hexane. One gram of oil, spiked with 75 μ g of 1-
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31 163 adamantanecarboylic acid and 50 μ g 5 β -cholanic acid (recovery standards),
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33 164 was pipetted onto the column and allowed to adsorb. After eluting non-acid
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35 165 fractions with *n*-hexane and DCM, an acid fraction was eluted with a mixture
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37 166 of diethyl ether and 2% formic acid. The acid fraction was reduced to dryness
38
39 167 in a rotor-evaporator and the recovered acid-fraction re-dissolved in methanol
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41 168 for spectroscopy and then redissolved in DCM prior to derivatisation with N,O-
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43 169 bis(trimethylsilyl)trifluoroacetamide (BSTFA) to convert alkenoic acids to their
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45 170 silylated ethers and esters for GC-MS analysis.
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47 172 *2.5 Gas Chromatography-Mass Spectrometry*

48 173 GC-MS analysis was performed using an Agilent 6890N GC fitted with a J&W
49
50 174 DB-5 phase 50 m length column (0.25 mm id, 0.25 μ m film thickness)
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52 175 connected to a 5975 MSD and a quadruple mass spectrometer operating in
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54 176 SIM mode (dwell time 0.1 s/ion and ionisation energy 70 eV). Samples were
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56 177 injected manually using a split/splitless injector operating in splitless mode
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58 178 (purge 40 ml min⁻¹ for 2 min). The temperature program for the GC oven was
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60 179 80 – 295 °C, holding at 80 °C for two minutes, rising to 10 °C min⁻¹ for 8 min
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180 and then 3 °C min⁻¹ and finally holding the maximum temperature for 10 min.

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3 181 Compounds were identified by comparing retention times to well-
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5 182 characterised materials that served as reference samples.
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8 9 184 *2.6 Microfluidic Separation*

10 185 Microfluidic separation followed the method presented in Bowden et al.¹³
11
12 186 using an H-cell with the following channel dimensions: channel length 20 mm,
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14 187 width 260 μm and 60 μm depth. Microfluidic chips were fabricated by Dolomite
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16 188 microfluidics from sodalime glass. The H-cell was held in a Mitos chip
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18 189 interface fitted with a multiflux 4-way linear connector. Heavy oils with API's <
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20 190 23° were diluted with hexane to lower the viscosity and improve sample
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22 191 manipulation (typical dilution factor of 1:5). Lighter oils (API's > 34°) were
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24 192 analysed with limited dilution (1:1 to 1:0). Methanol and Hexane were used as
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26 193 the extracting solvents and were pumped through the device with samples
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28 194 introduced as slugs for discrete batch analysis and processing. Solvents were
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30 195 pumped continuously through the microfluidic chip to establish optimal wetting
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32 196 characteristics within the channel and limit interaction between the oil and the
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34 197 sides of the channel (interface can occur via viscous effects caused by the
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36 198 adsorption of asphaltene and wax precipitates on the sides of channels).
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38 199 Three residence times were investigated; 2.8, 5.6 and 11.3 seconds –
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40 200 residence time is a key operational parameter within a Y- or H-cell and
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42 201 represents the maximum time that a particle will spend at the interface
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44 202 between two fluid streams¹⁵. The ratio of the flow rates between petroleum
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46 203 and extracting solvent was in the order of 1:6, this parameter governs the
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48 204 separation of the fluid streams at the downstream-end of the H-cell. The
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50 205 stability of the interface between the fluids was visually monitored at the down
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52 206 stream end of the device during experiments (Fig. 2).
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51 208 *2.7 Determination of asphaltene content by UV-Vis absorption spectra*

52 209 UV-Vis absorption spectra were obtained for the products of SARA fractions
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54 210 and off-line for microfluidic chip effluents using a USB4000 Ocean Optics
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56 211 spectrometer measuring within the wavelength range of 178-890 nm, in effect,
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58 212 operating the spectrometer as a single-beam spectrometer. Integration time
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60 213 was 500 ms and four scans were averaged to produce a single spectrum.
214 214 Samples were diluted in solvent to increase volume for ease of sample

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4 215 manipulation. Spectra were normalised, smoothed and then cropped (290nm
5 216 to 410 nm range) and the difference between whole oil and maltene spectra
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7 217 used to obtain the proportion of asphaltene in a sample as described in
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9 218 Bowden et al.¹³. The same spectral acquisition parameters were utilised for
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11 219 the determination of methanol extractables, except in this instance the
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13 220 absolute units of absorption used were measured at 205.4 nm and a
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15 221 calibration curve obtained using the acid fraction obtained by ion exchange
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17 222 chromatography.
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19 223

20 224 3.0 Results

21 225 *3.1 Comparison of spectroscopic and gravimetric determination of Asphaltene*

22 226 The H-cell based assay utilises differences in adsorption in the 290 to 410 nm
23
24 227 range for whole and asphaltene-free oils to determine asphaltene content ¹³.
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26 228 This element of the assay was investigated separately from H-cell
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28 229 parameters. It was found that a relatively high correlation ($r = 0.95$, $n = 13$
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30 230 which is significant with an alpha value greater than 0.001) could be obtained
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32 231 between the gravimetric and spectroscopic analysis of the products of ASTM
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34 232 D4124² (Data shown in supplementary information 1). However, a notable
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36 233 under-prediction occurred and the limit of detection was 4 %, e.g. when
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38 234 gravimetric analysis returned 4 % asphaltene the spectroscopic method
39
40 235 presented in Bowden et al.¹³ detected no asphaltene. This is significant
41
42 236 because petroleum with an asphaltene content of 5 % would be considered
43
44 237 asphaltic.

45 238

46 239 The limit of detection was improved by aggregating the asphaltene spectra of
47
48 240 all twelve samples to produce an “averaged asphaltene spectra” in the range
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50 241 290 to 410 nm. Results for this approach are shown in Fig. 3 and data
51
52 242 approach a 1:1 gradient, whilst the intercept of a straight line fitted to the data
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54 243 suggests a minimum detection limit of less than 1 %. Variance in the data is
55
56 244 also well explained using this approach ($r = 0.97$, $n = 13$, which is significant
57
58 245 with an alpha value greater than > 0.001). At present we have not determined
59
60 246 the exact cause for this improvement, but it is likely that the improvement
247 achieved by using averaged asphaltene spectra reflects the difficulties
248 inherent to chemically separating and characterising a pure asphaltene

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3 249 fraction of oil by ASTM D4124². Numerous studies have shown that
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5 250 asphaltene subfractions can be highly variable and can contain many non-
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7 251 macromolecular compounds that have little similarity to classic models of
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9 252 asphaltenes but that can precipitate with an asphaltene fraction (e.g. non-
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11 253 macromolecular heteroatomic compounds that are insoluble in hexane¹⁶).
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13 254 Asphaltic oils would be expected to yield purer asphaltene, e.g. relatively
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15 255 speaking they are less affected by interfering compounds. Averaged
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17 256 asphaltene spectra were therefore used when analysing the products of H-cell
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19 257 separations.
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21 258

21 259 *3.2 Comparison of H-cell based measurement of asphaltene with ASTM* 22 23 260 *D4124*

24
25 261 Predictions of asphaltene content produced by the H-cell processing of oils
26
27 262 were compared to results obtained from ASTM D4124². The correlation
28
29 263 between the results of the standard and H-cell method, for all three H-cell
30
31 264 residence times using either hexane or methanol as the solvent, were
32
33 265 significant indicating that the different methods are at least comparable
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35 266 (significant for an alpha value of 0.001, $r = 0.99$ and $n = 9$). Thus a calibration
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37 267 of the standard ASTM method to the H-cell method can be achieved. Based
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39 268 on the intercepts of straight-lines fitted to the raw data (Figure 4), the best
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41 269 detection limits were obtained for residence times of 5.6 seconds; the limit of
42
43 270 asphaltene-detection when hexane was used was less than 1 % and when
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45 271 methanol was used was less than 0.3%. (Data for other residence times is
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47 272 shown in supplementary information 2 and 3 and results listed Table 1).
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50
51 274 Asphaltenes are not the only high molecular weight and poorly soluble
52
53 275 component of petroleum that forms a solid precipitate. The wax components
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55 276 of oil can variably co-precipitate with asphaltene during ASTM D4124² to
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57 277 cause an erroneous assay. Waxes are held to be hydrocarbon compounds
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59 278 with high molecular weights – typically saturated compounds such as *n*-
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279 alkanes or structurally similar compounds. Asphaltenes are held to be
280 heteroatom-containing compounds, with an aromatic nucleus and a disputed
281 high molecular weight, (in excess of 550 amu⁴. The difference between the
282 two compounds classes is important because the factors that increase the

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3 283 stability of one compound type may destabilise the other (for example
4 284 blending a waxy oil with lower molecular hydrocarbon compounds may help
5 285 solubilise wax but could destabilise asphaltene). Two waxy-oils were
6 286 processed (Boa A, 9.3% and Bob B, 13% wax). For a residence time of 5.6
7 287 seconds, a comparison of dewaxed and pristine samples (that still have their
8 288 wax content and have not been dewaxed via ASTM D721-06¹⁴) suggests that
9 289 waxy samples assay with 1.5 to 1 % more asphaltene than would be expected
10 290 (Table 1).

11 291

12 292 *3.3 Hexane extracts analysis*

13 293 In addition to permitting an analysis of asphaltene content the hexane extract
14 294 obtained by diffusive separation within an H-cell is effectively asphaltene-free
15 295 and readily amenable for GC-MS analysis¹². Ion chromatograms of the
16 296 hexane extracts and the saturate fraction (obtained via the SARA-scheme and
17 297 ASTM D4124²) of sample Brid E are compared in Figure 5. Superficially they
18 298 appear similar, expect that *n*-alkanes with a carbon number greater than
19 299 twenty nine are not prominent on the 85 *m/z* ion chromatograms of H-cell
20 300 extracts. Additionally, H-cell fractions with the shortest residence times have
21 301 the lowest abundance of higher carbon number *n*-alkanes. This could be of
22 302 concern when making use of petroleum-biomarker proxies and parameters
23 303 that utilise homologous series of compounds, as these parameters are
24 304 sensitive to differences of a single carbon number. Little variation in the
25 305 relative proportions of the acyclic isoprenoids pristane and phytane is observed
26 306 as a function of residence time, and similarly the proportion of these
27 307 isoprenoids relative to their neighbouring *n*-alkanes also varies little (Figure 5).
28 308 Thus when preparing a sample for GC-analysis, longer residence times would
29 309 be needed if the final focus of analysis was on higher carbon number
30 310 biomarkers such as hopanes or similar terpanes.

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32 312 *3.6 Determination of TAN value using a methanol extract*

33 313 To evaluate the feasibility of predicting TAN value from an H-cell extract, the
34 314 yield of methanol was first compared to the concentration of naphthenic acid
35 315 obtained by Ion Exchange Chromatography (IEC). The greatest yields of
36 316 methanol-extractables were obtained for H-cell residence times of 5.6

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3 317 seconds (Fig. 6a – see Supplementary Information 4 for other residence
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5 318 times), but a significant correlation (alpha value greater than 0.01) between H-
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7 319 cell and IEC yields was observed for residence times of both 5.6 and 11.3
8
9 320 seconds. Detection limits (e.g. the value at which IEC yields an acid fraction
10
11 321 but the H-cell method would not) decreased from 4.6 mg/g through 4.4 mg/g
12
13 322 to 3.4 mg/g for residence times of 2.8, 5.6 and 11.3 seconds.

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15
16 324 GC-MS analysis of BSTFA derivatised H-cell and IEC fractions revealed that
17
18 325 both fractions are similar (both contain *n*-alkanoic acids and *n*-alkanols), but
19
20 326 that the H-cell extract contained far greater proportions of *n*-alkanols (Fig. 7).
21
22 327 This is a reasonable finding as methanol would not be expected to offer much
23
24 328 selectivity in terms of preferably solubilising alkanols over alkanolic acids, and
25
26 329 the two compound classes would have similar diffusivities. The presence of
27
28 330 other compound-types such as alcohols in addition to naphthenic acids
29
30 331 explains the relatively high yields obtained for the H-cell extracts in
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32 332 comparison to acid fractions obtained by ion exchange chromatography.

33 333
34 334 Previous work linking concentrations of naphthenic acids in petroleum to TAN
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36 335 values^{9,17} has used correlations between IEC yields and TAN value rather
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38 336 than concentrations of specific compounds. This is because the identification
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40 337 of compounds types that contribute most to crude oil acidity has been
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42 338 inconclusive – e.g. *n*-alkanoic acids have been shown to contribute little to the
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44 339 proton donating ability of crude oil and therefore have little influence on TAN
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46 340 value¹⁷. They were utilised in this study because of the highly diagnostic M-15
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48 341 ions produced by BSTFA-derivatised alkanolic acids during electron impact
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50 342 ionisation mass spectrometry. To convert methanol extract yields to a TAN
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52 343 equivalent a straight-line equation derived from IEC-acid fraction yields and
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54 344 TAN values was used^{8,9,17}. The TAN values obtained via ASTM D974-97⁵ are
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56 345 compared to H-cell predictions of TAN value in Fig. 6b - see Supplementary
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58 346 Information 5 for other residence times. There are notable outliers, but
59
60 347 generally, data fall in the same sequence as those determined via ASTM
348 D974-97⁵ (e.g. the most acidic sample determined via ASTM D974-97⁵ is the
349 most acidic sample according to the H-cell method).

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3 351 4.0 Discussion
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5 352 *4.1 Precision and sensitivity of H-cell method*
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7 353 A residence time of 5.6 seconds has the most repeatable measurement (%
8 354 relative standard deviation of 19% compared to 25% for both 2.8 and 11.3
9 355 seconds and 34% for the SARA method) (Table 2). Low repeatability for
10 356 ASTM D4124² likely derives from difficulty in weighing small amounts of
11 357 asphaltene in asphaltene poor samples. Measurement accuracy of the H-cell
12 358 method is also indicated by limits of detection (Table 2), where as low as
13 359 0.05% asphaltene in oil can be detected depending on the solvent used.
14 360 Sensitivity for asphaltene prediction is higher for methanol than for hexane.
15 361 Repeatability of TAN data is best at 11.3 seconds compared to other
16 362 residence times (although reasonable for 5.6 seconds) (table 2) but low when
17 363 compared to ASTM D974-97⁵. H-cell method appears sensitive at detecting
18 364 very low acid concentrations in oils, typically detecting acid content which can
19 365 be equivalent to as low as ~0.001mg of potassium hydroxide per gram of oil
20 366 however; this is a combined assay for both acids and alcohols.
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34 368 *4.2 Other considerations*
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36 369 For conventional asphaltene determination, the wax content of crude oil is a
37 370 known potential interference because of the formation of microcrystalites of
38 371 wax¹⁸. Previous work suggested that the formation of large asphaltene
39 372 aggregates that, in common with wax crystallites are a solid phase, are larger
40 373 and slower diffusing than their constituent molecules, did not impinge on the
41 374 production of an asphaltene-free fraction and thus asphaltene determination
42 375 via an H-cell¹². The results presented here suggest that waxes within oils
43 376 interfere slightly in the determination of asphaltene content; a 1 to 1.5 % over
44 377 estimation of asphaltene content was found for waxy oils. This over estimation
45 378 is relatively minor; by way of example, this is less than the error introduced by
46 379 various oil field sampling methods¹⁹, but significant within the context of the
47 380 generally high accuracy seen in Fig. 4 and table 2. The main cause of over
48 381 estimation is that wax compounds, as straight chain *n*-alkanes, contribute to
49 382 the spectra of the whole oil in a similar way to the saturate fraction of a non-
50 383 waxy oil (Fig. 8). Fig. 9 presents diffusive mixing times for *n*-alkanes within the
51 384 H-cell, and from this it could be concluded that both waxes and particularly
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4 385 wax-precipitates would not be expected in the extracting phase. As waxes will
5 386 not have defused to the asphaltene-free extract, their contribution to the final
6 387 spectra is missing and the proportion of asphaltene over estimated (the
7 388 amount of maltene fraction is underestimated). From an applications
8 389 perspective it would be beneficial to have an *a priori* knowledge of wax
9 390 content prior to sample analysis.
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15 392 Unlike ion exchange chromatography, an H-cell, using methanol as an
16 393 extracting solvent, does not separate *n*-alkanoic acids from mixtures
17 394 containing *n*-alkanols and *n*-alkanoic acids (Fig. 7). The effects created by the
18 395 presence of *n*-alkanols within methanol extracts can be adjusted by using the
19 396 straight line relationship shown in Fig. 6a to recalibrate results. Thus from the
20 397 perspective of producing a proxy for TAN, the presence of alcohols in
21 398 methanol-extracts is not a significant interference. However alcoholic
22 399 compounds such as phenols despite being classed as corrosive have not
23 400 been shown to be significant contributors to TAN value in the same way as
24 401 carboxylic acid species¹⁷. Compounds such as phenols, although not detected
25 402 in the small volumes analysed by GC-MS in this instance, would be expected
26 403 to partition to the methanol phase of an H-cell extract. Longer term, this
27 404 aspect of the H-cell separation procedure could be developed to try and
28 405 obtain a broader spectrum assay for polar compounds in petroleum.
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44 408 *4.3 Applications*

45 409 The asphaltene content and acidity of petroleum is used to inform decision
46 410 making for oilfield (often termed the upstream sector of the petroleum
47 411 industry), refinery (the downstream sector) and natural or environmental
48 412 science applications. Conventional methods typically yield this information
49 413 subsequent to laboratory analysis. Recent work has sought to develop
50 414 methods to provide this information at point of need and hopefully more
51 415 rapidly^{13,20}. In the case of oilfield applications varying asphaltene contents can
52 416 be used to predict variation in the physical properties of petroleum in the
53 417 subsurface; an example of this includes subsurface intervals within oil
54 418 reservoirs that contain exceptionally viscous or even solid petroleum (tar-

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4 419 mats). These intervals, aside from containing immobile bitumen may even
5 420 constitute barriers to flow, can be identified by a simple chemical assay²¹.
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7 421 Aside from this special case, it is also not uncommon for subsurface
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9 422 accumulations of petroleum to be the product of complex filling histories – e.g.
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11 423 oil of different types migrated to its current position at varying times, and was
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13 424 then variably altered in different regions of the subsurface²²). Chemical
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15 425 assays (notably asphaltene content) are often a viable and far less costly
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17 426 proxy for identifying and characterising variation than physical measurements.
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19 427 More recently the role of organic acids within petroleum in influencing wetting
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21 428 behaviour and the final level of oil recovery has been highlighted by laboratory
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23 429 studies²³. Therefore the simultaneous and rapid measurements of both
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25 430 asphaltene and acid content would be extremely useful for appraising oilfield
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27 431 potential and in particular what fraction is ultimately recoverable.
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31 433 Further into the life of an oilfield, the actions (workovers) initiated by operators
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33 434 to improve oil production often result in transient changes in the composition
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35 435 of produced fluids e.g. increased concentrations of corrosive surfacting agents
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37 436 or asphaltene as a consequence of the removal of blockages⁶. Chemical
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39 437 measurements performed at point of need could help engineers identify when
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41 438 the deleterious effects of such interventions had abated. Similar benefits could
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43 439 also be envisioned for point of need assays performed for the purposes flow
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45 440 assurance in refineries.
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49 442 For natural and environmental science applications, the asphaltene and TAN
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51 443 number are useful because changes in both parameters can be linked to oil
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53 444 degradation. Asphaltene is the recalcitrant proportion of petroleum and thus
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55 445 within spilled petroleum asphaltene concentrations rise as other components
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57 446 are degraded²⁴. Organic acid concentrations initially rise as oil is degraded,
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59 447 likely because of a contribution from hydrocarbon-metabolites formed by
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448 microbial activity. In the terminal stages of petroleum degradation the
449 concentration of organic acids has been observed to decrease²⁵. Repeated
450 simultaneous measurements of asphaltene and TAN values via an H-cell
451 would therefore record the attenuation of oil-spills and possibly help
452 differentiate fresh from degraded petroleum. Measuring the asphaltene and

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3 453 TAN content of petroleum thus provides a rapid method of characterisation –
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5 454 e.g. helping to distinguish fresh from weathered petroleum.
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8 456 5.0 Conclusion

9 457 An H-cell based method for separating heavy petroleum can be optimised to
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11 458 yield proxy measurements of asphaltene and the carboxylic acid content of
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13 459 petroleum (which can be expressed as a TAN value). Detailed analysis of the
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15 460 extracts produced by the H-cell demonstrates that the method, because it
16
17 461 utilises microscaled diffusion, provides an analytical window that is inherently
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19 462 different to distillation and chemical based methods for assaying heavy
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21 463 petroleum, The H-cell method gives better accuracy for measuring asphaltene
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23 464 when compared to a gravimetric-based methods like ASTM D4124². The
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25 465 capability of this technique to provide rapid simultaneous TAN and asphaltene
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27 466 proxy measurements has the potential to impact both petroleum and
28
29 467 environmental science.
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32 469 6.0 References

33 470 1 J. G. Speight, *The Chemistry and Technology of Petroleum*, 5th edn, Taylor
34
35 471 and Francis group, Florida, 2014.
36
37 472

38 473 2 ASTM D4124 Standard Test Method for Separation of Asphalt into Four
39
40 474 Fractions, 2000, American Society for Testing and Materials, West
41
42 475 Conshohocken, PA.
43
44 476

45 477 3 K.E. Peters, C.C. Walters, and J.M. Moldowan, *The Biomarker Guide: Biomarkers and Isotopes in the Environment and Human History*, Cambridge
46
47 478 University Press, Cambridge, vol. 1, 2nd edn, 2005.
48
49 479
50
51 480

52 481 4 O.C. Mullins, E. Y. Sheu, A. Hammami, and A. G. Marshall, *Asphaltenes, Heavy Oils and Petroleomics*, Springer, New York, 2007.
53
54 482
55
56 483

57 484 5 ASTM D974-97 Standard Test Method for Acid and Base Numbers by
58
59 485 Colour Indicator Titration, 2013, American Society for Testing and Materials,
60
486 West Conshohocken, PA, 2013.

- 1
2
3 487
4
5 488 6 W.A. Derungs, *Corrosion*, 1956, **12**, 41- 46
6
7 489
8
9 490 7 Z. Mendez, R.E. Anton, and J.L.J. Salager, *Dispersion Sci., Technol.* 1999,
10 491 **20**, 883.
11
12 492
13
14 493 8 D.M. Jones, J.S. Watson, W. Meridith, M. Chen, and B. Bennett, *Anal.*
15 494 *Chem.*, 2001, **73**, 703.
16
17 495
18
19 496 9 A.E. Borgund, and K. Erstad, T. Barth *Energy & Fuels*, 2007, **21**, 2816.
20
21 497
22
23 498 10 K. Gurgey, *Org. Geochem.*, **29**, 1139.
24
25 499
26 500 11 P. B. Brody and P. Yager, *Sens. Actuators, A*, 1997, **58**, 13.
27
28 501
29
30 502 12 S. A. Bowden, P. B. Monaghan, R. Willson, J. Parnell and J. M. Cooper,
31 503 *Lab Chip*, 2006, **6**, 740.
32
33 504
34
35 505 13 S. A. Bowden, R. Wilson, J. Parnell and J. M. Cooper, *Lab Chip*, 2009, **9**,
36 506 828.
37
38 507
39
40 508 14 ASTM D721-06 Standard Test Method for Oil Content of Petroleum Waxes
41 509 2011. American Society for Testing and Materials, West Conshohocken, PA.
42
43 510
44
45 511 15 N.T. Nguyen and S.T. Wereley, *Fundamentals and Applications of*
46 512 *Microfluidics*, Artech House Inc, Massachusetts, 2002.
47
48 513
49
50 514 16 J.S. Sinninghe Damsté, W. Irene, C. Rijpstra, J.W. de Leeuw, and P.A.
51 515 Schenck, *Org. Geochem.*, 1988, **13**, 593.
52
53 516
54
55 517 17 W. Merideth, S. J. Kelland and D. M. Jones, *Org. Geochem.*, 2000, **31**,
56 518 1059.
57
58 519
59
60

1
2
3 520 18 E.D. Burger, T.K. Perkins, and J.H. Striegler, J. Petrol. Tech. 1981, **33**,
4 1075.
5 521
6 522

7
8 523 19 S.A. Bayliss, S.A. Org. Geochem., 1998, **29**, 463.
9 524

10
11 525 20 M.H. Schneider, V.J. Sieben, A.M. Kharrat, and F. Mostowfi, F. Anal.
12 526 Chem., 2013, **85**, 5153.
13 527

14
15 528 21 A. Wilhelms, and S.R. Larter, *The Geochemistry of Reservoirs* (e.d Cubitt,
16 529 J.M. and England W.A.), Geological Society Special Publication No. 86, 87-
17 530 101.
18 531

19
20 532 22 K.E. Peters, J.M. Moldowan, A.R. Driscoll, and G.J. Demaison, The
21 533 American Association of Petroleum Geologists Bulletin, 1989, **73**, 454.
22 534

23
24 535 23 Y. Tanino, and M.J. Blunt, Water Resour., 2013, **49**, 4311.
25 536

26
27 537 24 J. Connan, Biodegradation of Crude oils in Reservoirs in *Advances in*
28 538 *Petroleum Geochemistry*, ed. J. Brooks & D. Welte, Academic Press, London,
29 539 1984, pp.299-235.
30 540

31
32 541 25 J.S. Watson, D.M. Jones, R.P.J. Swanell, and A.C.T. van Duin, Org.
33 542 Geochem., 2002, **33**, 1153.
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36 544 Figure captions

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38 545 Figure 1: Schematic representation of the analytical scheme used during this
39 546 study, illustrating the alternative analysis methods employed for heavy
40 547 petroleum.
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43 549 Figure 2: LHS - Variation in the sample type used for this study that ranges
44 550 from solid through to viscous liquid petroleum. RHS - Image of H-cell device in
45 551 operation. Labelled is the interface between the two fluids and the fluids
46 552 exiting the device. Note that the relative pressure of the two fluid streams is
47 553 set so that some of the extracting phase exits through the sample outlet. This

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3 554 represents a factor of safety to ensure that none of the asphaltene containing
4 555 sample unintentionally exits with the sample phase needs to be asphaltene-
5 556 free.
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10 558 Figure 3: Graph comparing spectroscopic and gravimetric determination of
11 559 asphaltene. Raw data (filled black circles) and data recalibrated using straight-
12 560 line fitted to raw data (Hollow circles).
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16 562 Figure 4: Comparison of percentage asphaltene predicted by the H-Cell
17 563 method with ASTM D4124². LHS – analyses using hexane as the extracting
18 564 solvent and for a residence time of 5.6 seconds; RHS – analyses using
19 565 methanol as the extracting solvent and for a residence time of 5.6 seconds.
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26 568 Figure 5: m/z 85 Ion chromatograms of the H-cell hexane extracts and
27 569 hydrocarbon fraction yielded by ASTM D4124². Data are shown for sample
28 570 Bride E for the H-cell residence times indicated. Carbon numbers correspond
29 571 to those of the n-alkane homologous series.
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34 573 Figure 6: a) Comparison of the yield of methanol extractables at 5.6 seconds
35 574 with acid-fraction yields obtained by ion exchange chromatography⁸. b) Acid
36 575 fraction yields at 5.6 seconds expressed as TAN values using approach
37 576 presented in Borgund, et al.⁹, and Meridith, et al.¹⁷ with TAN values obtained
38 577 by ASTM D974-97⁵.
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47 579 Figure 7: m/z Ion chromatograms for methanol extracts and acid fractions
48 580 obtained by IEC. Plotted ions in the range m/z 285 to 341 correspond to the
49 581 M-15 ions for BSTFA derivatised n-alkanols and n-alkanoic acids.
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54 583 Figure 8: Absorption spectra of various fractions of oil in the 280-410 nm
55 584 wavelength range. Note: Wax spectra are similar to that of maltene fraction
56 585 (de-asphaltene oil) causing over estimation of asphaltene content.
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4 587 Figure 9: Scatter plot of the relative abundance of *n*-alkanes in H-cell extracts
5 588 (relative to the abundance of the compound in the fraction produced by
6 column chromatography) plotted by carbon number. Also shown is the
7 589 calculated diffusive mixing times¹⁵ shown for key *n*-alkanes.
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598 Table 1: Sample Descriptions and Analysis Details

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Sample*	Type [†]	API ^{††}	TAN mg KOH/g	Wax %	Asph ₁ [‡] %	ICE acid fraction* mg/g oil	H-cell analyses [°] (%)									
							Dilute [°]	RT: 2.8			RT: 5.6			RT: 11.3		
								Hex	MeOH	Hex	MeOH	Hex	MeOH			
Beatrice Oilfield, Moray Firth, UK	Boa A	Stock Oil	38°	n.d.	9.3	4	1	n.d.	1:1	4(4.5) ^w	n.d.	2(3) ^w	n.d.	4(5) ^w	n.d.	
	Bob B	Stock Oil	38°	n.d.	13	5.5	7	n.d.	1:1	5.5(5) ^w	5	5(5) ^w	6	4.5 (4.5) ^w	n.d.	
Siljan, Sweden	Sil C	Stock Oil	15-20°	5.3	n.d.	10	12	9.51	1:3	10	10	10	10	10	n.d.	
Wytch farm, Dorset	Sher D	Stock Oil	37°	5.17	n.d.	18	25	6.89	1:2	17	18	18	18	18	n.d.	
	Brid E	Stock Oil	38°	0.55	n.d.	6.9	8	4.28	1:1	6.9	6	7	6.5	6	n.d.	
	From F	Stock Oil	38°	1.08	n.d.	7	1	4.77	1:2	n.d.	7	n.d.	6	n.d.	n.d.	
Bengal deep fore-	Syl G	Stock Oil	28.3°	0.84	n.d.	5.7	5	4.17	1:½	n.d.	5.7	n.d.	6	n.d.	n.d.	
	BM J	Stock Oil	n.a.	0.84	n.d.	6	2	5.15	1:2	n.d.	n.d.	n.d.	n.d.	n.d.	7	
Murchison Field, North Sea	Oryx L	Stock Oil	38°	0.55	n.d.	6.3	6	4.5	1:½	n.d.	7	n.d.	6.5	n.d.	5.5	
	Oryx M	Stock Oil	38°	0.28	n.d.	7	10	4.1	3:1	7	6.3	8	7	7	5	
Thurso, Sutherland, UK	Cait	Fresh, Seep	n.d.	n.d.	n.d.	16	12	n.d.	1:5	16	n.d.	22	n.d.	18	n.d.	
Pitchford Bridge, Shropshire, UK	Pit Br	Fresh, Seep	n.d.	6.57	n.d.	30	37	12.13	1:10	30	n.d.	29	n.d.	38	n.d.	

600 *Sample name and code referred to in text. †Stock samples are taken from a stock collection held at the University of Aberdeen; Fresh Seep samples were collected during fieldwork. ††API
601 values were taken from the information listed with samples in the collection, except for seep samples. ‡The first asphaltene value was obtained using ASTM D4124, the second as described in
602 text. ●Acid fraction obtained by ion exchange chromatography (solid phase extraction). ° Dilution factor of samples to solvent (v/v), RT = residence time of particle in H-cell, hex = hexane used
603 as extracting solvent, MeOH = methanol used as extracting solvent, n.d. = not done, ()^w = Asphaltene data for waxy samples.

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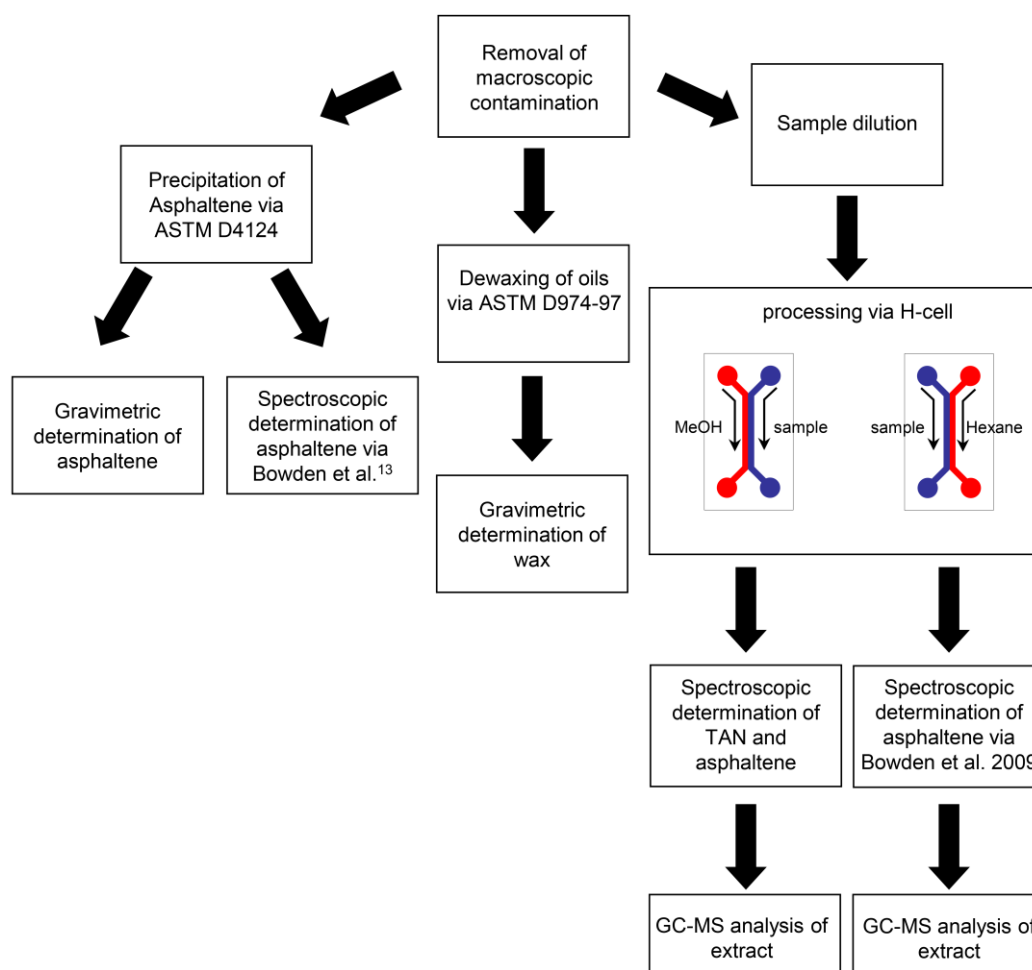
Table 2: Precision and Sensitivity

Method	RT(s)	RSD (%)		LoD		
		Asph	TAN	Asph%	Methanol	TAN
				Hexane		mgKOH/g
H-cell	2.8s	25	13	0.5	0.3	0.9
	5.6s	19	11	0.1	0.05	~0.001
	11.3	25	7	0.9	0.1	~0.001
ASTM D4214		34				
ASTM D974-9726			4			

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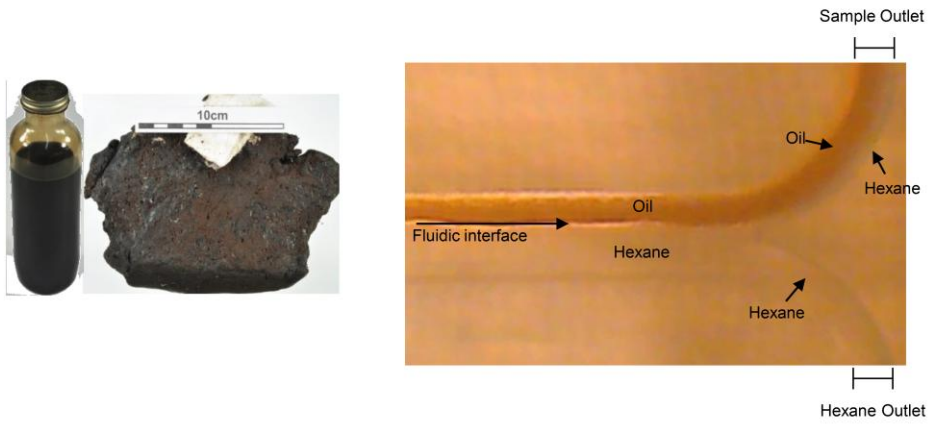
RT = residence time of particle in H-cell, Asph = Asphaltene, TAN = Total Acid number, RSD (%) = Relative standard deviation expressed in percentage, LoD = Limit of Detection.

Fig 1.



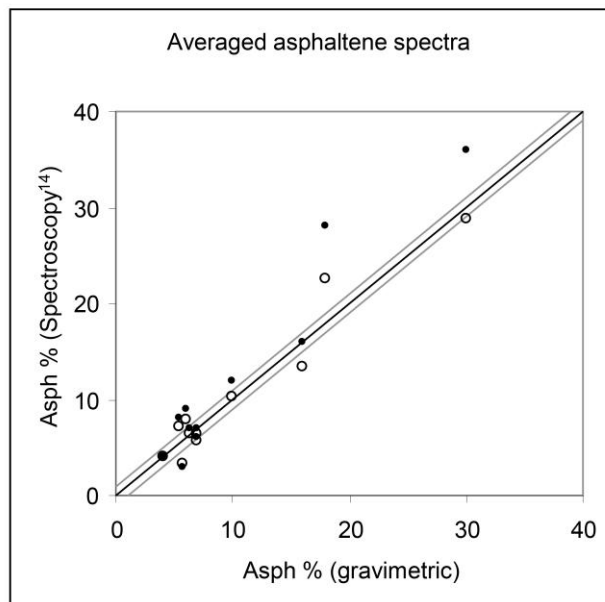
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Fig 2.



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Fig 3.



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Fig 4.

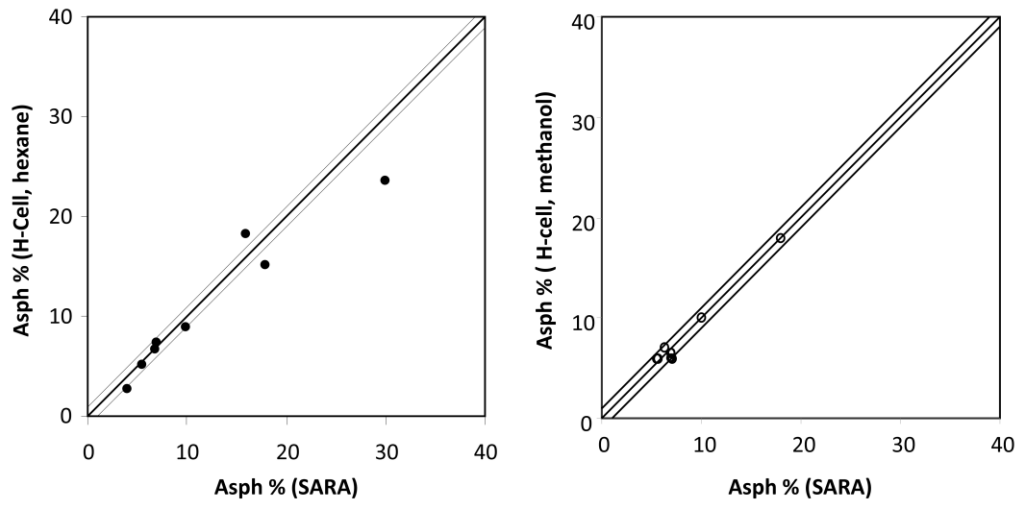
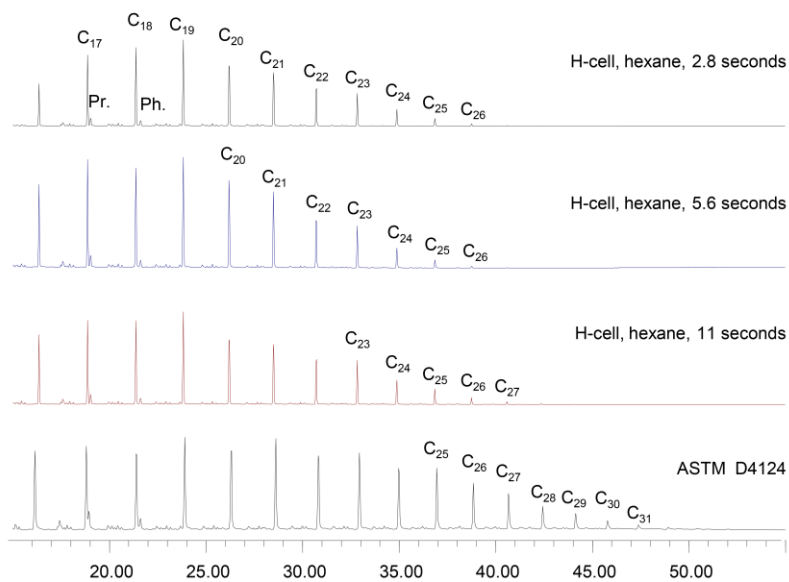
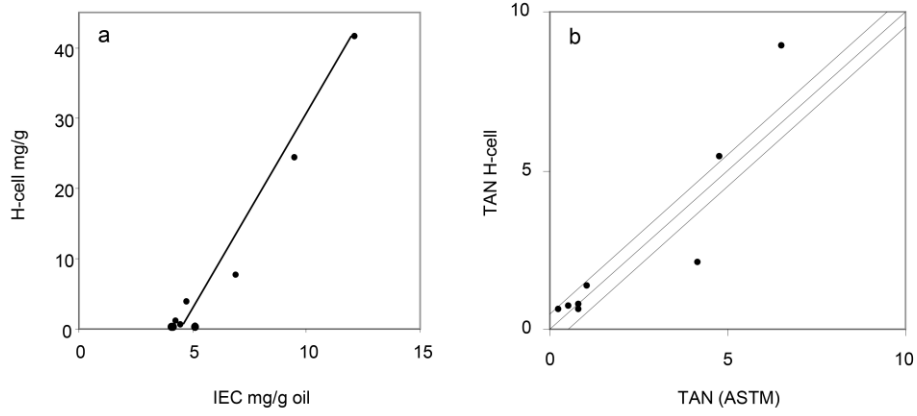


Fig 5.



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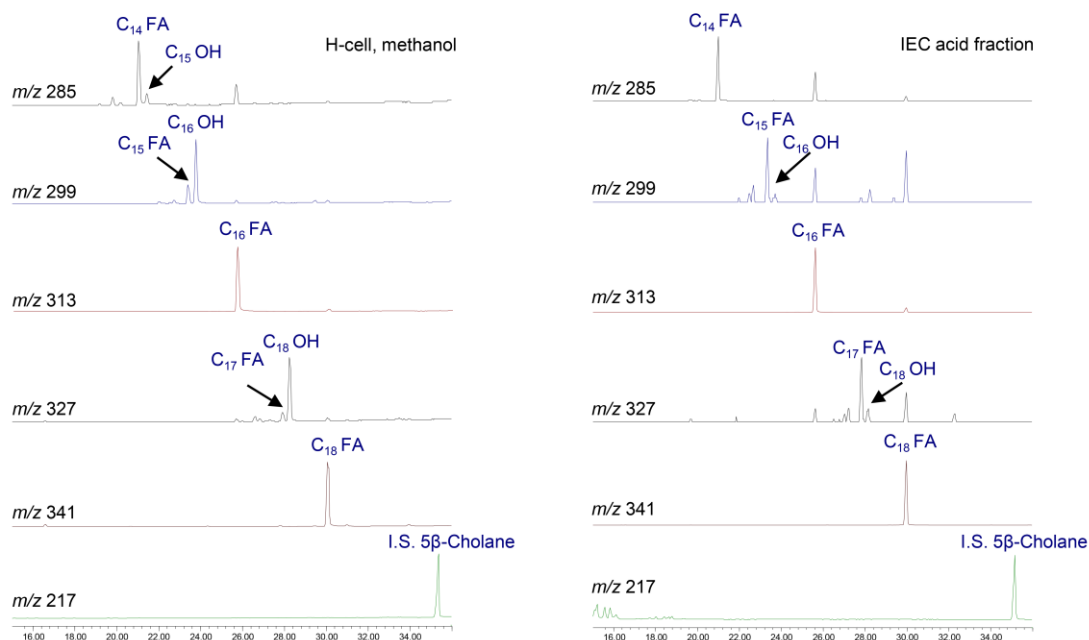
Fig 6.



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Fig 7.



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Fig 8.

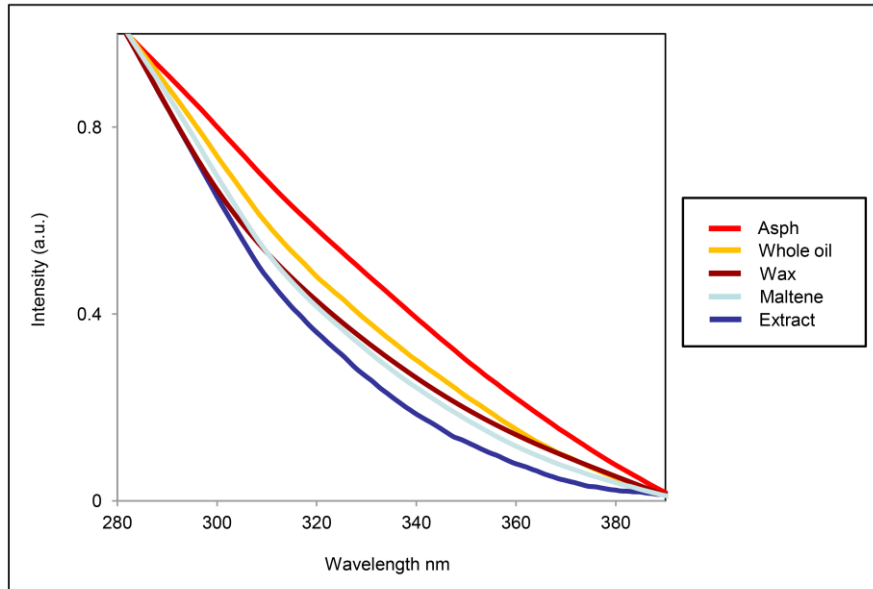
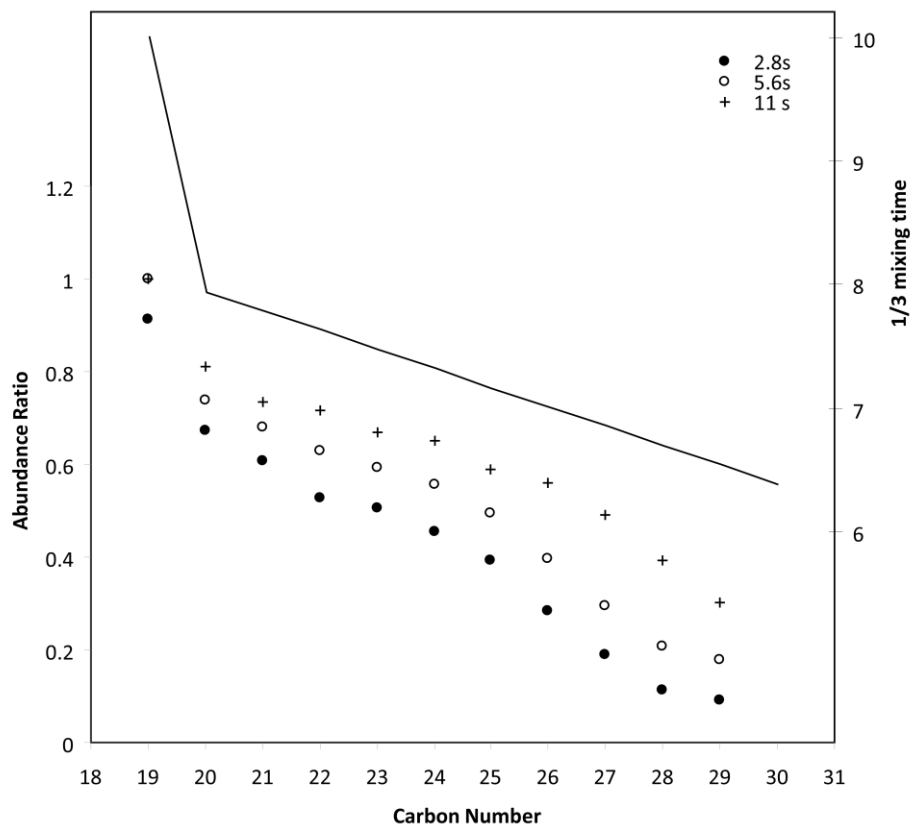


Fig 9.



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