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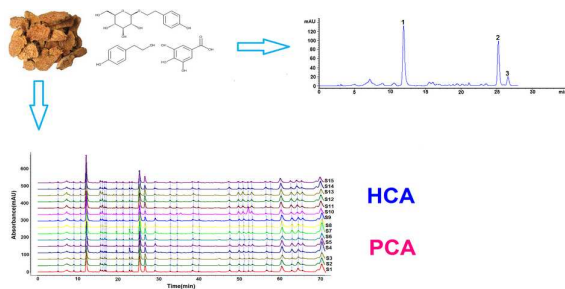
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HPLC-DAD quantitative analysis and chromatographic fingerprint analysis were developed to evaluate the quality of *Rhodiola crenulata* H. Ohba from different origins.



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Combinative method using multi-components quantitation by single reference standard and HPLC fingerprint for comprehensive evaluation of *Rhodiola crenulata* H.Ohba

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Abstract

A combinative method using HPLC-DAD quantitative analysis and fingerprint were developed to evaluate the quality of *Rhodiola crenulata* H.Ohba (*R. crenulata*) from different origins. In this study, three phenolic compounds were first simultaneously determined by single standard to determine multi-components (SSDMC) method. The chromatographic separation was performed on a Merck Purospher STAR RP-18 column (4.6×250mm, 5μm) with a gradient elution program within 28 min at 275nm wavelength. The method has achieved desired specificity, linearity ($r^2 \geq 0.9999$), precision, accuracy (95-105%), stability and robustness. Compared with the normal external standard method (ESM), this alternative method can be used for simultaneous determination of multiple indexes effectively and accurately, and can resolve the problem of lacking of chemical standards. Additionally, the herbal chromatographic fingerprint was calculated using similarity evaluation system and SPSS 19.0 software as a result of analysing all the *R. crenulata* samples. The obtained data showed good repeatability and stability of the

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3 chromatographic fingerprint. The 15 *R. crenulata* samples from different origins were
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5 classified by hierarchical clustering analysis (HCA) and principal components analysis
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7 (PCA) according to the characteristic of common peaks and the similarity values were all
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9 above 0.90. This study demonstrated that a combination of the chromatographic
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11 quantitative analysis and fingerprint offers an efficient way to quality consistency
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13 evaluation of *R. crenulata*.

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15 Keywords: Quality consistency evaluation, *Rhodiola crenulata* H. Ohba, Single standard
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17 to determine multi-components (SSDMC), Quantitative determination, HPLC fingerprint.

18 19 **1. Introduction**

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21 The plants of the genus *Rhodiola* are widely distributed in the high cold region of Asia,
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23 Europe and North America. There are approximately 90 species recorded in the world,
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25 and more than 70 species are found in China, mainly the Hengduan Mountains region
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27 including eastern Tibet, northern Yunnan and western Sichuan. In China, *Rhodiola*
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29 species have been used for maintaining body health and treating various diseases in
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31 traditional Tibetan medicines (TTM) for thousands of years.¹ Among the variety of
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33 *Rhodiola* species,²⁻⁴ *R. crenulata* is known for best quality and has been accepted by the
34
35 Pharmacopoeia of China since 2005.⁵

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37 *R. crenulata* consists of the dried roots and rhizome of *Rhodiola crenulata* (Hook. f. et
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39 Thorns.) H. Ohba (Fam. Crassulaceae). Phytochemical studies on *R. crenulata* revealed
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41 that it contained a variety of chemical constituents, mainly including phenolic compounds,
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43 organic acids, flavonoids and tannins.⁶⁻⁸ Phenolic compounds are the major active
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45 constituents for the standardization of *R. crenulata* which exhibit many biological
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47 activities such as adaptogenic, anti-oxidation, anti-hypoxia, anti-fatigue, anti-arrhythmic,
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49 anti-cancer and enhancement in learning and memory.⁹⁻¹⁷

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51 Separation-based techniques, such as capillary zone electrophoresis and high speed
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53 counter current chromatography have been used for the determination of *R. crenulata*.^{3,18}
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55 However, on the basis of satisfactory accuracy and effectiveness, a simple method is
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57 preferred to be developed for quality evaluation of *R. crenulata*. High performance liquid
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59 chromatography coupled with diode array detector (HPLC-DAD) has proved to be one of
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3 the most powerful tools for the quality control of traditional Chinese medicine (TCM)
4 because of the simplicity, speed and stability of the method.¹⁹⁻²³ Only few studies using
5 HPLC on the evaluation of *R. crenulata* have been reported.²⁴ But it is insufficient to
6 determine merely one or two markers for completely evaluating the inner quality of *R.*
7 *crenulata*. Therefore a comprehensive and systematic quality standard for quality
8 assessment of *R. crenulata* is imperative.

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10 In this study, SSDMC method, which has already applied in some TCM for resolving
11 the problem of the expensive price and the limited availability of reference substances,^{25,}
12 ²⁶ was first used to determine phenolic compounds in *R. crenulata*. The three phenolic
13 compounds salidroside, tyrosol and gallic acid which have specific pharmacological
14 activity and abundant in *R. crenulata* were simultaneously determined with salidroside as
15 reference standard. The SSDMC method was validated, and its results were compared
16 with that obtained by traditional ESM. Additionally, the same chromatographic condition
17 was applied to fingerprint analysis for the further assessment of *R. crenulata*. The
18 similarity evaluation system, HCA and PCA were performed to classify the samples from
19 different origins according to the characteristic of common peaks in fingerprint
20 chromatogram. The aim of the present study was to develop a direct, rapid and reliable
21 HPLC-DAD method for quantitative analysis and establish the chromatographic
22 fingerprint analysis for distinguishing *R. crenulata* from various locations. This
23 combinative method can evaluate the quality of *R. crenulata* more comprehensively.

2. Experimental

2.1 Materials and reagents

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25 HPLC-grade acetonitrile and methanol were purchased from Fisher Scientific (Fisher
26 Scientific, USA). Phosphoric acid for HPLC was obtained from Kemiou Chemical
27 Reagent Co. Ltd. (Tianjin, China). Redistilled water was used for the preparation of
28 two-phase mobile solvent system. All solvents and samples were filtered through 0.45 μ m
29 membrane filters before analysis.

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31 Salidroside and tyrosol were purchased from the Institute for the Control of
32 Pharmaceutical and Biological Products (Beijing, China), and gallic acid was obtained

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3 from Acros Organics (New Jersey, USA). The purity of salidroside, tyrosol and gallic
4 acid were all determined by peak area normalization method on HPLC. Through
5 calculation, the purity of the standards was all above 98.0%.
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9 Fifteen batches of *R. crenulata* from different provinces in China were collected,
10 including Tibet Linzhi (No.5), Tibet Lasa (No.1, 2, 3, 4, 13, 14), Yunnan Kunming (No.7,
11 8), Yunnan Lijiang (No.6), Sichuan Leshan (No.9), Sichuan A-bazhou (No.10), Qinghai
12 Xining(No.11, 12), Gansu (No.15). All of the samples were identified by Professor Ying
13 Jia (Department of Traditional Chinese Medicine, Shenyang Pharmaceutical University,
14 Shenyang, China)
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20 2.2 Apparatus

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22 Agilent 1100 HPLC system (Agilent Technologies, USA) consisted of a quaternary
23 solvent delivery system, an on-line degasser, an auto-sampler, a column temperature
24 controller and a diode array detector (DAD). System control and data analysis were
25 processed with Agilent ChemStation software. Additional different HPLC instrument was
26 used. Agilent 1260 HPLC system comprised a quaternary solvent delivery system, an
27 on-line degasser, an auto-sampler, and photodiode array detector coupled with an
28 analytical workstation. A KH5200b sonicated bath (HeChuang, KunShan Co. Ltd) was
29 used for sample preparation.
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37 2.3 Preparation of solutions

38 2.3.1 Preparation of standard solution

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40 Stock solutions of the reference standards (salidroside, tyrosol and gallic acid) were
41 prepared by dissolving accurately weighed standards in methanol to yield the
42 concentrations of 3.016, 1.618 and 2.020 mg/mL respectively, and stored in a 10 mL
43 volumetric flask. These solutions were stored at 4°C before use.
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49 2.3.2 Sample solutions

50 Transfer about 500mg powder of *R. crenulata*, accurately weighed to a 100mL
51 glass-stoppered conical flask. 15 mL methanol-water (6:4) was added and sonicated for
52 30 min, and then adjust to the initial weight by adding methanol-water (6:4) as needed.
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56 The extracts were filtered with a 0.45µm membrane filter prior to HPLC analysis.
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59 2.4 Chromatographic conditions

The chromatographic separation was carried out on a Merck Purospher STAR RP-18 (4.6×250mm, 5µm) with a guard column (4.6×10mm, 5µm; Merck). Agilent Zorbax SB C₁₈ (4.6×250mm, 5µm). The mobile phase consisted of water containing 0.02% phosphoric acid (A) and methanol: acetonitrile=9:1(B). The segmented gradient profile was 5-5% (B) in 0-10 min, 5-17% (B) in 10-15 min, 17-17% (B) in 15-28 min. 17-23% (B) in 28-45min, 23-25.5% (B) in 45-72min. The assay determination was stop at 28 min. The mobile phase flow rate was 1.0 mL per minute. The DAD detector was operated at 275 nm with 8 nm bandwidth and no reference wave. The column temperature was at 25°C. All injection volumes of sample and standard solutions were 10µL.

2.5 Data analysis

To calculate the RRFs, three different concentration calibration curves was prepared. The RRFs of tyrosol and gallic acid at each concentration level of calibration curve were calculated as Eq. (1). The content of reference standard solidoside was calculated by external standard method. The final RRFs of each analyte were obtained as the mean values calculated from the triplet of six gradient concentrations level.

$$\text{RRF} = \frac{A_s / C_s}{A_{sx} / C_{sx}} \quad (1)$$

In which A_s and C_s were the peak area and the concentration of solidoside obtained from standard solution; A_{sx} and C_{sx} were the the peak area and the concentration of analyte obtained from standard solution.

The relative retention time of the analyte (RRT_x) was calculated as the ratio of retention time of the analyte (t_x) and the solidoside (t_{SA}).

$$\text{RRT}_x = \frac{t_x}{t_{SA}} \quad (2)$$

A reference chromatographic fingerprint was calculated by professional software named Similarity Evaluation System for Chromatographic Fingerprint of Traditional Chinese Medicine (version 2009A). The software was to employ the correlative coefficient in evaluating the similarities of different chromatograms. The HCA and PCA were done by SPSS 19.0 software. P value was the probability and it was used to compare

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3 the differences between two groups of data by paired *t*-test. If $P < 0.05$ meant there were
4 significant difference between two groups of data.
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7 **3. Results and discussion**

8 **3.1 Optimization of extraction conditions**

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10 Sample pretreatment conditions were optimized by investigating extraction solvents,
11 extraction methods, solvent volumes, extraction time and extraction frequency on the
12 extraction efficiencies for chemical markers which used for HPLC quantification and
13 fingerprinting. In the preliminary studies for the selection of different proportions of
14 extraction solvents, methanol-water (6:4) was found to be the most effective solvent for
15 extracting phenolic compounds based on the HPLC results. Extraction methods including
16 ultrasonication, heat-reflux and maceration were then investigated for extraction
17 efficiencies for marker compounds. In consideration of the convenience and shorter
18 time-length of sonication, ultrasonication was chosen for further experiment. The
19 appropriate volume of solvent (5mL, 15mL, 30mL), extraction time (15minutes, 30
20 minutes, 45 minutes) and extraction frequency (once, twice) were also investigated, and
21 the results showed that 15mL solvent volume for extraction once in 30 minutes was the
22 appropriate condition for extraction in consideration of convenience operation and the
23 relatively higher value achieved. The optimal extraction conditions for *R. crenulata* used
24 in this study are presented in detail in Section 2.3.2.
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40 **3.2 Optimization of HPLC chromatographic conditions**

41 HPLC conditions including detection wavelength and mobile phase were investigated
42 for optimization of chromatographic separations for marker compounds. The wavelength
43 for the detection of the target compounds in the preparation was scanned in the entire UV
44 range (200-400nm) to determine the λ_{max} . The chemical structures of these reference
45 compounds were shown in Fig. 1(A). Most chemical constituents had the best responses
46 at 275nm, thus the detection wavelength was set at 275nm.
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53 In the selection of mobile phase, mobile phase system, pH of mobile phase, program
54 of mobile phase, flow rate, column temperature and injection volume were investigated to
55 achieve better separation. It was found that good resolution and symmetric peak shape
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3 were obtained when methanol: acetonitrile=9:1–water containing 0.02% phosphoric acid
4 was selected as a mobile phase with a step linear gradient. Meanwhile, the results showed
5 that column temperature maintained at 25°C, flow rate was set at 1 mL/min and the
6 injection volume was 10µL which could obtain good resolution and acceptable peak
7 parameters. Optimal HPLC condition used in this study are shown in Section 2.4.
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10 11 12 3.3 Selection of reference standard

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14 In order to generate the 'standard' RRF for each analyte to the reference analyte, it is
15 essential to choose suitable reference analyte. Selected reference analyte should be
16 abundant in sample, stable, easily accessible and has a clear pharmacological effect or is
17 related to clinical efficacy. Meanwhile, it should achieve a good separation under the
18 chromatographic conditions. Therefore, salidroside was selected as reference analyte
19 because of its low cost, high stability, high content and significant pharmacological
20 activities in *R. crenulata*.
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28 3.4 Calculation of relative reference standards and relative retention time

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30 Using salidroside as reference standard, the results of RRFs and RRT gained by six
31 concentration standard solutions are shown in Table 1. Additionally, the influence of
32 concentrations of standard solution on the RRFs was studied. It was prepared through
33 diluting the mixed standard solution in order. Triplicate experiments were performed, and
34 average RRFs were calculated based on the RRFs obtained in each experiment. The
35 results showed that the RRF of gallic acid was 0.091, and the RRF of tyrosol was 0.43.
36 The average retention time of gallic acid, salidroside and tyrosol were 12.0min, 25.1min
37 and 26.5min respectively. And the RRT of gallic acid was 0.48, the RRT of tyrosol was
38 1.05. It also proved that the RRFs and RRT were stability at different of standard
39 solution.
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48 3.5 Method validation

49 3.5.1 Specificity

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51 The specificity was estimated by comparing the consistency of the retention time of
52 each analyte between a sample and the corresponding reference standard. The integration
53 peak in the chromatogram of the sample solution was corresponding in time to the peak
54 in the chromatogram of standard solution (Fig. 1(B)).
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3.5.2 Linearity, limits of quantification and detection

Methanol stock solutions containing three phenolics were prepared and diluted into six appropriate concentrations for the construction of calibration curves. The calibration curve was constructed by plotting the peak area of each compound against the concentration of each compound. The results exhibited good linearity ($r^2 \geq 0.9999$) over the concentration range. The limits of detection (LOD) and quantification (LOQ) for each analyte under the chromatographic conditions were determined at the signal-to-noise ratio (S/N) of 3 and 10, respectively. The LOD and LOQ of the 3 analytes were 0.075-0.60 and 0.25-2.00 $\mu\text{g/mL}$, respectively (Table 1).

3.5.3 Repeatability, precision and accuracy

The repeatability test was analyzed by three replicates at three different concentrations. The RSD values of the content of the three markers were less than 1.1%. The intermediate precision including different analysts and days were also analyzed to ensure the validity of the method. The RSD values of the content of the three phenolics with different analysts and days were found in the range of 0.6-1.0% and 1.4-1.9%, respectively. Accuracy was calculated as the percentage of recovery by the assay of the known added amount of gallic acid, salidroside and tyrosol in the sample. Recovery was between 95.6% and 103.6% with RSD values of less than 3.7% for all the three marker compounds. The results obtained by the ESM and SSDMC method showed no remarkable differences using paired *t*-test ($p=0.435>0.05$).

3.5.4 Stability

The stability of sample solution was investigated. It was carried out by comparing the peak areas of gallic acid, salidroside and tyrosol in the chromatograph of the same sample solution after storing at room temperature for different time (0, 2, 4, 6, 8, 12, 24 hours). Stability was evaluated by calculating the RSD of area obtained. The analytes were found to be stable within 24 hours in sample solution ($\text{RSD}<1\%$).

3.5.5 Robustness

In order to introduce the SSDMC method into different laboratories, the robustness test was performed to investigate the significant affecting factors for this method. Through six concentration standard solutions, the robustness of RRT and RRFs was

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evaluated with different equipment and columns. The results showed that the RRT and RRFs were fluctuated in a relative narrow range. The robustness of the established method was also evaluated by examining its stability with small variations of procedural parameters, including pH of mobile phase, ratio of components in mobile phase, programs of mobile phase, wavelength, flow rate, injection volume, and column temperature. It was proved that a slight variation of ratio of mobile phase and flow rate had great influence on the RRT of gallic acid; a little variation of wavelength had a significant effect on RRFs of gallic acid(RSD>4%). Other Factors had slight influence on the RRFs and RRT and the fluctuation was proportionable (RSD<2%). The initial ratio of organic phase had significant influence on the retention time of gallic acid which was the first peak in the chromatogram, and it was also sensitive to flow rate mainly because the chromatography behavior of gallic acid, tyrosol and salidroside were very different at different flow rate. The variation of RRFs of gallic acid was great influenced may due to the UV spectrums of gallic acid, tyrosol and salidroside were different. The results showed that RRFs and RRT of gallic acid were greatly sensitive to the variation of conditions. Therefore it was recommended that the ratio of organic phase and flow rate should not be changed too much, and the wavelength should be controlled strictly. The pH of mobile phase, the time program of mobile phase, the injection volume and the column temperature can be slightly adjusted to meet the system suitability.

3.6 Quantitative determination of three phenolics in *R. crenulata* samples

The ESM and SSDMC method were employed for quantitative analysis of three phenolics in *R. crenulata* samples from different sources. All samples were analyzed by the optimized extraction procedure under the optimized HPLC conditions. Three main phenolics, salidroside, tyrosol and gallic acid were selected as analytes to evaluate the quality. The contents of three phenolics were calculated using these two methods are shown in Table 2.

It was proved that there was no significant differences between these two methods using the paired *t*-test($p=0.164>0.05$). The values of RRFs determination is complimentary and can be effectively used as an alternative method to conventional HPLC method especially in the absence of suitable reference materials for the active

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3 pharmaceutical ingredients.

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5 3.7 Establishment of chromatographic fingerprint of *R. crenulata* and similarity
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9 Fifteen batches of materials from various locations, including almost all of the
10 growing places of *R. crenulata* in China, were obtained to ensure that the reference
11 fingerprint thus developed was geographically representative and authentic. Peaks
12 existing in all chromatograms of the samples were assigned as the 'common peak'. Using
13 the software of Similarity Evaluation System for Chromatographic Fingerprint of
14 Traditional Chinese Medicine (Version 2009A), the chromatograms of *R. crenulata*
15 containing 31 common peaks within 72 min were shown in Fig. 2. It was found that these
16 samples had similar HPLC profiles. The representative standard fingerprint was generated
17 by the median method were shown in Fig. 3. Similarity analysis was conducted based on
18 the standard fingerprints, and the results are shown in Table 3. Five peaks were identified
19 comparing with standard compounds, including gallic acid (4), salidroside (13), tyrosol
20 (14), epicatechin (20), *p*-coumaric acid (26). Peak 13 (salidroside) was chosen to calculate
21 the RRT and RPA. The similarity values of all the 15 samples were more than 0.90
22 indicating that the chemical constituents of the 15 batches *R. crenulata* samples were
23 similar. Therefore, if 0.90 is set as an appropriate threshold, it is easy to identify *R.*
24 *crenulata* based on the chromatographic fingerprint.
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39 3.8 HCA of the samples
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41 In order to evaluate the variation of *R. crenulata* from various origins, HCA was used
42 to find natural cluster of samples according to RPA was performed from HPLC profiles.
43 The clustering analysis was operated in SPSS software, and the results were shown in Fig.
44 4. It was found that 15 tested samples of *R. crenulata* were divided into two main clusters
45 (I and II) containing 7 and 8 samples, respectively. The results indicated that samples 3,
46 14, 1, 2, 5, 4 and 13 which all collected from Tibet province were classified in the same
47 cluster (cluster I). Sample 3 and 14 were classified in the same group may due to their
48 similarity values were relative low in the cluster I. And samples 6 and 7 (from Yunnan
49 province, China) were classified in another group. The other samples were classified in
50 one cluster (cluster II) might be due to the constituent of *R. crenulata* from different
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3 cultivation regions were similar.

4 5 3.9 PCA of the samples

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7 PCA was implemented as a data reduction technique to generate a visual scatterplot
8 for qualitative evaluation of resemblances and differences between the studied samples.
9 The first two PCs explained more than 97.2% of total variability, and the other principal
10 components which had a minor effect on the model were discarded. Hence it met a
11 sufficient condition for studying the relation between *R. crenulata* samples in a
12 bi-dimensional plate (Fig. 5). From the scatter points, the PC1 values of samples were all
13 comparatively concentrated. However, the samples can be classified into two groups
14 according to the difference of the PC 2 values and marked as group I and II, respectively.
15 In group I, sample 3, 14, 2, 4, 5, 1 and 13 were all collected from Tibet. And the content
16 of salidroside from group I was higher than group II. The result was closely related to the
17 unique climatic ecological environment in the plateau of Tibet and the categorized data
18 were corresponded with the results of the HCA.
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21 The presented method suggested that the quality of different samples for sale was
22 not consistent, and it may be helpful for discriminate *R. crenulata* according to their
23 cultivation regions. The *R. crenulata* from Tibet occupies an important place in the *R.*
24 *crenulata* resource of the world, thus it would be significant to classify and identify them.
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4. Conclusions

The proposed HPLC quantitative analyse combined with fingerprint method is an efficient and comprehensive tool for quality control of *R. crenulata*. In this research, SSDMC method was first developed to quantify the chromatographically diverse constituents in *R. crenulata*. Comparison with accurate values obtained from external standard method proved that it was a fast, convenient, and accurate method to determine the contents of phenolics in *R. crenulata*. The fingerprinting analysis using similarity, HCA, and PCA approaches has produced the desirable results with high accuracy. This study revealed that quantitation of pharmacologically active constituents by SSDMC method combined with chromatographic fingerprints offers an efficient way for monitoring of the *R. crenulata* quality consistency.

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References

- 1 Y. C. Yang, T. N. He, S. L. Lu, R. F. Hung, Z. X. Wang, *Zang Yao Zhi*, Qinghai People's Publishing House, Xining, 1991, 34-35, 432-434.
- 2 R. P. Brown, P. L. Gerbarg, Z. Ramazanov, *HerbGram*, 2002, **56**, 40–52.
- 3 S. Y. Cui, X. L. Hu, X. G. Chen, Z. D. Hu, *Anal Bioanal Chem*, 2003, **377**, 370–374
- 4 H. B. Li, F. Chen, *J. Chromatogr., A*, 2001, **932**, 91–95.
- 5 China Pharmacopoeia Committee, *Pharmacopoeia of the People's Republic of China*, China Chemical Industry Press, Beijing, 2005, 106.
- 6 G. G. Yousef, M. H. Grace, D. M. Cheng, I. V. Belolipov, I. Raskin, M. A. Lila, *Phytochemistry*, 2006, **67**, 2380–2391.
- 7 G. Z. Ma, W. Li, D. Q. Dou, X. L. Chang, H. Bai, T. Satou, J. Li, D. J. Sun, T. G. Kang, T. Nikaido, K. Koike, *Chem. Pharm. Bull.*, 2006, **54**, 1229–1233.
- 8 M. W. Lee, Y. A. Lee, H. M. Park, S. H. Toh, E. J. Lee, H. D. Jang, Y. H. Kim, *Arch. Pharm. Res.*, 2000, **23**, 455–458.
- 9 A. A. Spasov, G. K. Wikman, V. B. Mandrikov, I. A. Mironova, V. V. Neumoin, *Phytomedicine*, 2000, **7**, 85–89.
- 10 N. N. Rege, U. M. Thatte, S. A. Dahanukar, *Phytother Res.*, 1999, **13**, 275–291.
- 11 W. Zheng, S. Wang, J. Agric, *Food Chem.*, 2001, **49**, 5165–5170.
- 12 A. Panossian, G. Wikman, J. Sarris, *Phytomedicine*, 2010, **17**, 481–493.
- 13 C. Y. Ma, J. Tang, H. X. Wang, X. H. Gu, G. J. Tao, *Chromatographia*, 2008, **67**, 383–388.
- 14 A. Tolonen, A. Hohtola, J. Jalonen, *Chromatographia*, 2003, **57**, 577-579.
- 15 A. Bharathi, Y. H. Wang, A. Zulfiqar, J. S. Troy, F. Vicky, C. Alain, T. A. John, A. K. Ikhlas, *Biomed. Chromatogr.*, 2009, **23**, 865–872.
- 16 W. Z. Fan, Y. Tezuka, K. M. Ni, S. Kadota, *Chem. Pharm. Bull.*, 2001, **49**, 396-401.
- 17 C. Carlo, V. M. Maria, V. Giovanni, R. Valentina, C. Roberto, M. Maurizio, *Hysiology & Behavior*, 2010, **101**, 555–562.
- 18 X. Han, T. Y. Zhang, Y. Wei, X. L. Cao, Y. Ito, *J. Chromatogr., A*, 2002, **971**, 237–241

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4 19 A. Escarpa, M. C. Gonzalez, *J. Chromatogr., A*, 1998, **823**, 331–337.
- 5
6 20 M. Francisco, D. A. Moreno, M. E. Cartea, F. Ferreres, C. G. Viguera, P. Velasco, *J.*
7
8 *Chromatogr., A*, 2009, **1216**, 6611–6619.
- 9 21 Q. An, N. W. Lu, Y. M. Dong, *Anal. Methods*, 2013, **5**, 5775-5784
- 10
11 22 M. L. Calabrò, V. Galtieri, P. Cutroneo, S. Tommasini, P. Ficarra, R. Ficarra, *J.*
12
13 *Pharm. Biomed. Anal.*, 2004, **35**, 349–363.
- 14
15 23 C. Y. Luo, X. L. Zuo, Y. Q. Li, C. J. Sun, Y. Jiang, Z. Y. Wu, *Food Chem.*, 2011, **127**,
16
17 314–320.
- 18
19 24 Y. Mao, Y. Li, N. Yao, *J. Pharm. Biomed. Anal.*, 2007, **45**, 510–515
- 20
21 25 J. J. Hou, W. Y. Wu, J. Da, S. Yao, H. L. Long, Z. Yang, L. Y. Cai, M. Yang, X. Liu,
22
23 B. H. Jiang, D. A. Guo, *J. Chromatogr., A*, 2011, **1218**, 5618–5627.
- 24
25 26 China Pharmacopoeia Committee, *Pharmacopoeia of the People's Republic of China*,
26
27 China Chemical Industry Press, Beijing, 2010, 285
- 28
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Table 1 The results of linearity, LOD, LOQ, RRFs and RRT. (n=2)

Analytes	Linearity equation	R ²	Range (µg/mL)	LOD	LOQ	RRFs		RRT	
				(µg/mL)	(µg/mL)	RRFs	RSD%	RRT	RSD%
Salidroside	Y=2.881x-6.406	0.9999	55.10-1102	0.60	2.00	1.0	0	1.0	0
Tyrosol	Y=6.711x-8.06	0.9999	19.42-388.4	0.29	0.97	0.43	1.0	1.05	0.1
Gallic acid	Y=31.81x-31.18	0.9999	16.16-323.2	0.075	0.25	0.091	0.5	0.47	0.4

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Table 2 Content of 15 batches of *Rhodiola crenulata* H.Ohba determined with ESM and SSDMC method (n=2)

No.	origin	ESM				SSDMC		Method	
		Total phenolics(%)	Gallic acid(%)	Tyrosol(%)	Salidroside(%)	Total phenolics(%)	Gallic acid(%)	Tyrosol(%)	Salidroside(%)
1	Tibet Lasa	2.02	0.22	0.28	1.52	2.02	0.22	0.28	1.52
2	Tibet Lasa	2.29	0.20	0.27	1.82	2.29	0.20	0.27	1.82
3	Tibet Lasa	2.54	0.11	0.23	2.20	2.54	0.11	0.23	2.20
4	Tibet Lasa	2.42	0.28	0.22	1.92	2.42	0.27	0.22	1.92
5	Tibet Linzhi	2.17	0.25	0.15	1.77	2.17	0.25	0.15	1.77
6	Yunnan	1.86	0.46	0.71	0.69	1.86	0.46	0.71	0.69
7	Yunnan	1.56	0.30	0.40	0.87	1.56	0.29	0.40	0.87
8	Yunnan	2.35	0.32	0.33	1.70	2.35	0.32	0.33	1.70
9	Sichuan	1.76	0.31	0.13	1.33	1.76	0.31	0.13	1.33
10	Sichuan	1.51	0.25	0.16	1.10	1.51	0.25	0.16	1.10
11	Qinghai	2.69	0.59	0.33	1.77	2.68	0.58	0.33	1.77
12	Qinghai	2.50	0.41	0.27	1.82	2.50	0.40	0.28	1.82
13	Tibet Lasa	1.77	0.26	0.31	1.20	1.76	0.25	0.31	1.20
14	Tibet Lasa	2.51	0.18	0.16	2.17	2.51	0.17	0.16	2.17
15	Gansu	1.93	0.28	0.28	1.37	1.93	0.28	0.28	1.37
Average		2.13	0.29	0.28	1.55	2.12	0.29	0.28	1.55

Total phenolics were the sum of the content of salidroside, tyrosol and gallic acid.

Table 3 The similarities of chromatograms of 15 samples

No.	Similarities
1	0.976
2	0.961
3	0.901
4	0.972
5	0.981
6	0.918
7	0.944
8	0.974
9	0.951
10	0.948
11	0.971
12	0.996
13	0.977
14	0.921
15	0.998

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Figure caption

Fig.1 (A) The chemical structures of the investigated compounds.

Fig.1 (B) (I) mixed standards: (1) gallic acid (2) salidroside (3) tyrosol, (II) sample of *Rhodiola crenulata* H.Ohba (Tibet Linzhi).

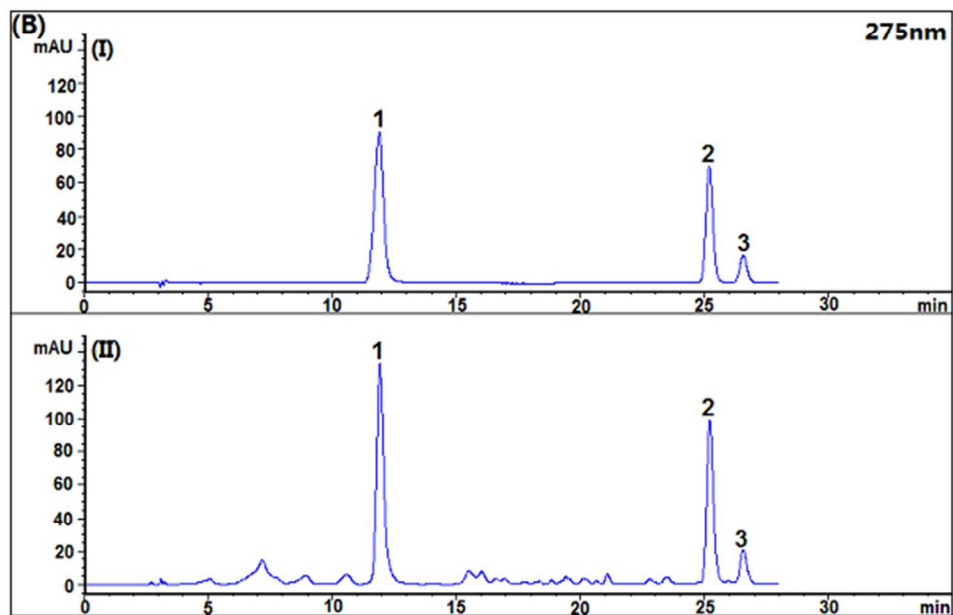
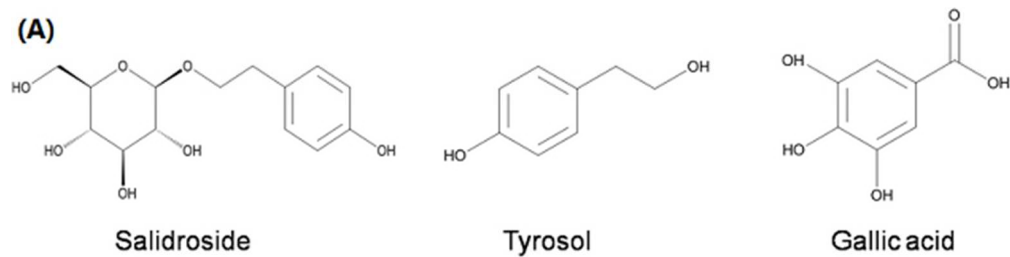
Fig.2 Chromatographic fingerprints for all the *Rhodiola crenulata* H.Ohba samples.

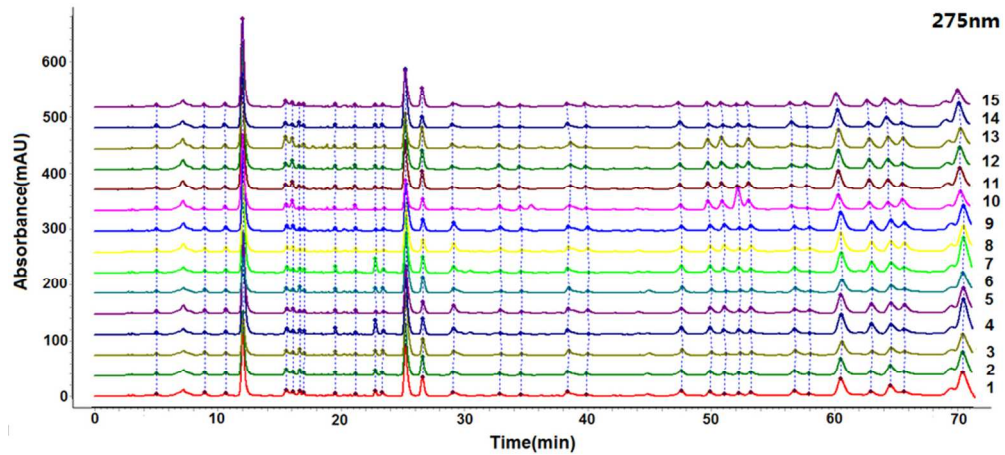
Fig.3 The representative standard fingerprint was generated by the median method.

Gallic acid (4), salidroside(13), tyrosol(14), epicatechin(20), *p*-counaricacid(26).

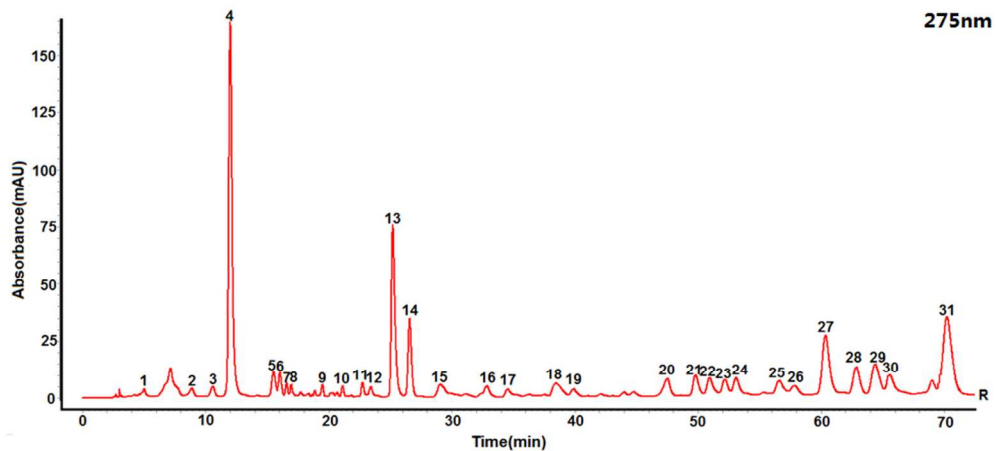
Fig.4 Dendrograms of hierarchical cluster analysis for the 15 tested samples of *Rhodiola crenulata* H.Ohba samples.

Fig.5 PCA scatterplot for all the *Rhodiola crenulata* H.Ohba samples.

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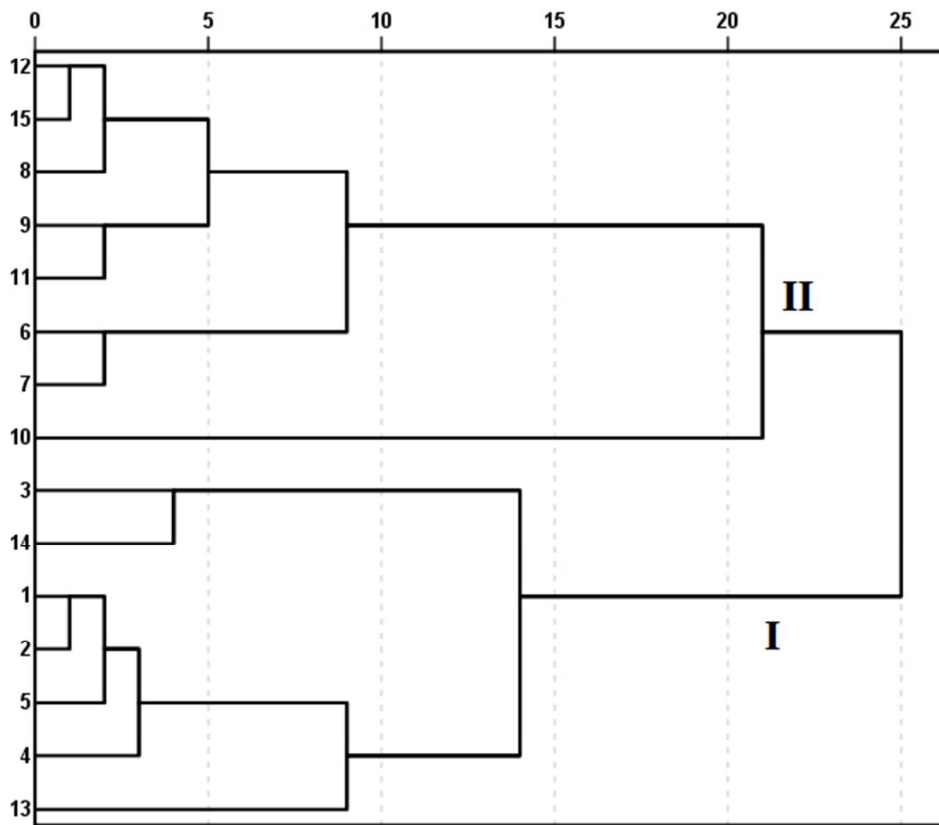
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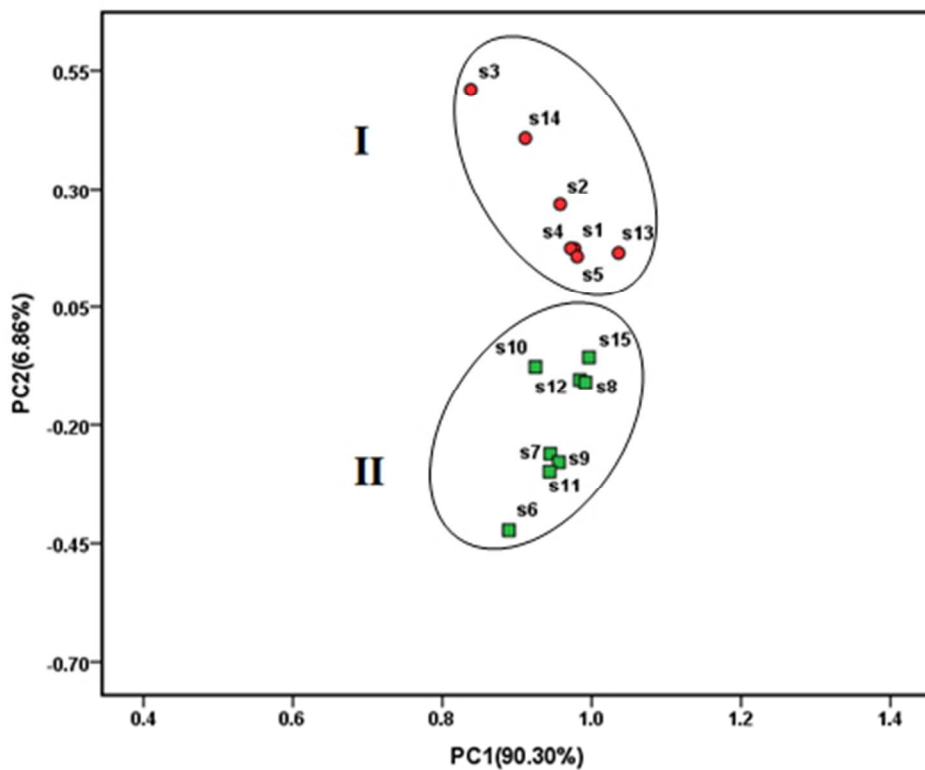
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Dendrograms using Average Linkage (Between Groups)

Rescaled Distance Cluster Combine



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