



## Electrochemical detection of guaiacol in bamboo juice based on the enhancement effect of RGO nanosheets

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4 Electrochemical detection of guaiacol in bamboo juice based on the  
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6 enhancement effect of RGO nanosheets  
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28 Reduced graphene oxide (RGO) nanosheets with high quality were chemically  
29 synthesized by hydrothermal reduction of well-dispersed graphene oxide (GO)  
30 suspension. The electrochemical behavior of guaiacol was studied on different carbon  
31 materials surface. Compared with the bare glassy carbon, graphite, and GO, RGO  
32 nanosheets exhibited strong enhancement effect and the electrochemical oxidation  
33 response of guaiacol was remarkably improved. The influence of the pH value,  
34 amount of RGO, scan rate and accumulation time on the oxidation signal of guaiacol  
35 were investigated. Based on the enhancement effect of RGO, a simple, fast and  
36 sensitive electrochemical method was developed for the detection of guaiacol.  
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38 The linear range was from 0.5  $\mu\text{M}$  to 500  $\mu\text{M}$  with a correlation coefficient of  
39 0.998, and the limit of detection was as low as 0.2  $\mu\text{M}$  (S/N = 3). Finally, it was used  
40 to determine guaiacol in bamboo juice sample, and the recovery was over the range  
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4 between 98.9% and 105.8%.  
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9 **Keywords:** RGO nanosheets; Guaiacol; Bamboo juice; Electrochemical detection.  
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## 11 12 13 **Introduction** 14

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16 Graphene, a two dimensional monolayer of  $sp^2$  carbon in a honeycomb-like network,  
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18 has attracted a great deal of scientific interest in these years.<sup>1-3</sup> Due to its unique  
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20 electric, thermal and mechanical characteristics, graphene has been extensively  
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22 applied in many technological fields such as transparent conducting films,<sup>4</sup> sensors,<sup>5</sup>  
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24 supercapacitors,<sup>6</sup> and batteries.<sup>7</sup> Up to now, various approaches have been developed  
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26 for the synthesis of graphene, including mechanical exfoliation,<sup>8</sup> epitaxial growth,<sup>9</sup>  
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28 chemical vapor deposition,<sup>10</sup> unzipping carbon nanotubes,<sup>11</sup> liquid phase exfoliation  
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30 of graphite,<sup>12</sup> chemical reduction of GO,<sup>13</sup> etc. According to data, graphene prepared  
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32 through chemical reduction of GO possesses many structural defects, which are  
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34 advantageous for the electrochemical sensing, for example, the response signals and  
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36 detection sensitivity of glucose,<sup>14</sup>  $H_2O_2$ ,<sup>15</sup> and  $NO_2$ <sup>16</sup> were enhanced remarkably on  
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38 the surface of RGO nanosheets.  
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47 Guaiacol is a kind of natural organic matter which is the major ingredient of  
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49 creosote. Because of the special pharmacological effects such as strong anti-microbial  
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51 action, anti-inflammation action and anti-nociceptive action, guaiacol was widely  
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53 used in the field of medicine.<sup>17,18</sup> Therefore, the detection of guaiacol is quite  
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55 important and interesting. Liquid chromatography-mass spectrometry<sup>19</sup> and gas  
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3 chromatography-mass spectrometry<sup>20</sup> are the most common approaches for the  
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6 detection of guaiacol. Although electrochemical method possesses the advantages  
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8 such as high sensitivity, short analysis time, low cost and handling convenience, the  
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11 electrochemical detection of guaiacol is limited. As shown in Fig. 1, the structural  
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14 formula of guaiacol contains phenolic hydroxyl group and should be electrochemical  
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17 active, even so, electrochemical determination of guaiacol using RGO nanosheets has  
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20 not been reported.

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22 The main objective of this work is to study the electrochemical response of  
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24 guaiacol on the surface of RGO and then develop a simple, rapid and sensitive  
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27 electrochemical method for the detection of guaiacol utilizing the unique property of  
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30 RGO nanosheets. RGO nanosheets were synthesized by using  $\text{Na}_2\text{S}_2\text{O}_3$  as the  
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33 reducing reagent from graphene oxide through hydrothermal method. The obtained  
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36 RGO was well-dispersed in water, resulting in a stable and homogeneous suspension.  
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39 Compared with the bare GCE, GO and the RGO modified electrodes, RGO greatly  
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42 improves the oxidation signal of guaiacol, indicating that RGO exhibits remarkable  
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45 surface enhancement effect toward the oxidation of guaiacol. Based on this, a  
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48 sensitive and convenient electrochemical method was proposed for the determination  
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51 of guaiacol, which was successfully demonstrated with traditional Chinese medicines  
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54 of bamboo juice.

## 55 **Experimental section**

### 56 **Reagents**

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4 All chemicals were of analytical grade and used as received. Guaiacol was obtained  
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6 from Guangfu fine chemical insititute (Tianjin, China).  $\text{KMnO}_4$ , sulphuric acid (98%),  
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8 graphite powder (99.99% purity, ~100 mesh), and  $\text{Na}_2\text{S}_2\text{O}_3 \cdot 5\text{H}_2\text{O}$  were purchased  
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10 from Zhongtian chemical reagent company (Wuhan, China). Absolute ethyl alcohol  
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12 and absolute ether were obtained from Fuchen chemical reagent company (Tianjin,  
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### **Instruments**

The electrochemical measurements were performed using 660D electrochemical analyzer (CH Instruments, USA) with a conventional three-electrode system. The working electrode was RGO modified glassy carbon electrode, the reference electrode an Ag/AgCl with saturated KCl, and the counter electrode was a Pt wire. Scanning electron microscopy (SEM) was performed with a JEOL microscope (JSM-6510LV, Japan). Transmission electron microscopy (TEM) images were measured using a Tecnai G220 microscope (FEI Company, Netherlands). Raman spectra were carried out on a LabRAM HR800 confocal Raman microscopy using 532 nm laser (Horiba JobinYvon, France). Fourier Transform Infrared spectroscopy (FTIR) were obtained with a Avatar 360 spectrometer (Nicolet, America) with a KBr plate. X-ray diffraction (XRD) patterns were measured using a diffractometer (Shimadzu XRD-7000, Japan). The ultraviolet spectrum measurement was carried out with TU-1901 (Persee corporation of beijing, China).

### **Synthesis of GO and RGO**

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4 Graphene oxide, was synthesized by Hummer's method.<sup>21</sup> RGO used in this work was  
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6 prepared by the chemical reduction of GO with  $\text{Na}_2\text{S}_2\text{O}_3 \cdot 5\text{H}_2\text{O}$ . In brief, 50 mg GO  
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8 was first ultrasonically dispersed in 80 mL  $\text{H}_2\text{O}$  for 1.5 h to get graphene oxide  
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10 suspension, then 0.5 g  $\text{Na}_2\text{S}_2\text{O}_3 \cdot 5\text{H}_2\text{O}$  was added into the GO suspension. The mixture  
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12 was then sealed into a Teflon-lined stainless steel autoclave and maintained at 180 °C  
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14 for 24 h. After it was cooled down to the room temperature, the resulting product was  
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16 separated by centrifugation, washed with deionized water and ethanol for several  
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18 times, and dried at 60 °C under vacuum overnight.  
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#### 26 **Fabrication of RGO modified GCE**

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28 RGO nanosheets (10.0 mg) were added into doubly distilled water (10.0 mL), and  
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30 then sonicated in a KQ3200DE ultrasonicator for 1 h, giving a stable and black  
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32 graphene suspension. Before modification with RGO, the GCE with a diameter of 3  
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34 mm was polished with 0.05 mm alumina slurry, and then sonicated in doubly distilled  
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36 water for 2 min. After drying, the GCE surface was coated with 10  $\mu\text{L}$  RGO  
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38 suspension, and the water was evaporated under an infrared lamp in air. The RGO  
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40 nanosheets modied GCE was prepared. For the comparasion, 1 mg/mL graphite and  
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42 GO suspension was also prepared to modify the glassy carbon electrode, respectively.  
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#### 51 **Sample preparation**

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53 The bamboo juice (pharmaceutical co., LTD of tongyuan Sichuan, China) used in this  
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55 study was purchased from a local pharmacy. The extraction process of guaiacol was  
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3 performed as described like this. Briefly, the pH of 60 mL bamboo juice was adjusted  
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5 close to 1~2. 20 mL ether was added into the above solution and the extraction  
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7 process parallely undergoes 3 times, then the ether extraction solution was  
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9 mixed together. The mixture of the ether extraction solution was washed with 50 mL  
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11 5% NaHCO<sub>3</sub> for 3 times, the layer of NaHCO<sub>3</sub> solution was abandoned and the final  
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13 ether extraction solution was gathered. The ether in the final extraction solution was  
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15 evaporated at room temperature. Finally, the remaining residue was dissolved in 25  
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17 ml ethanol for measurement.  
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### 26 **Analytical procedure**

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28 Unless otherwise stated, 0.1 M, pH 6.5 phosphate buffer (PBS) was used as the  
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30 supporting electrolyte for the detection of guaiacol. After 30 s accumulation, the  
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32 differential pulse voltammetry (DPV) curves were recorded from 0.1 to 1.0 V, and the  
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34 oxidation peak current at 0.60 V was measured for guaiacol. The pulse amplitude is  
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36 50 mV, pulse width is 40 ms, and the scan rate is 40 mV s<sup>-1</sup>.  
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## 43 **Results and discussion**

### 44 **AFM Characterization of GO and RGO**

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46 RGO together with the original GO were first examined by AFM. As shown in Fig. 2,  
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48 the mean thickness of GO is about 1.24 nm (a) and RGO is 1.41 nm (b), the small  
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50 distinction in thickness between GO and RGO, suggesting the original GO and the  
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52 obtained RGO possess the same layers. The theoretical thickness for a perfectly flat,  
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4 single layer  $sp^2$ -carbon atom network is about 0.3 nm, so it can be concluded that the  
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6 number of layers of the synthesized RGO is about 4.  
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9 The morphologies of different carbon materials were further characterized by  
10 SEM and TEM. As shown in Fig. 4A, the unmodified GCE surface was smooth and  
11 virtually featureless. On the other hand, the pristine graphite was made of large bulk  
12 particles in micron meter as displayed in Fig. 4B. After modification with GO  
13 nanosheets as illustrated in Fig. 4C, wrinkled sheets were observed. In addition, the  
14 RGO film demonstrated a curly and corrugated appearance consisting of a wrinkling  
15 paper-like structure, which is significantly different from the flat and characterless  
16 GCE surface. To preferably check the microstructure, TEM test was also performed  
17 for GO and RGO. Fig. 3E and 4F are TEM images of GO and RGO. It was clear that  
18 both of GO and RGO exhibited wrinkled flake-like shapes. The wrinkled nature of  
19 graphene is highly beneficial in maintaining a high surface area, which is  
20 advantageous to improve the accumulation efficiency of the target analyte in  
21 electrochemical sensing.  
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#### 44 **Enhancement effect of RGO nanosheets**

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46 The electrochemical behavior of guaiacol was studied on the unmodified,  
47 graphite-modified, GO-modified and RGO-modified GCEs using cyclic voltammetry  
48 (CV). Fig. 4 shows the electrochemical response of 50  $\mu$ M guaiacol on different  
49 electrode surface in 0.1 M (pH 6.5) phosphate buffer solution. Successive cyclic  
50 sweep was conducted from 0 to 1.0 V, the oxidation peak current was recorded as the  
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4 analytical signal. The electrochemical signals on the un-modified GCE (black line)  
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6 and graphite modified GCE (red line) were negligible, indicating the oxidation  
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8 activity of guaiacol on unmodified and graphite-modified GCEs was very low. When  
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10 using GO-modified GCE (blue line), a small oxidation peak appeared at about 0.60 V,  
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12 suggesting that GO is more active to the electrochemical detection compared with the  
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14 unmodified and graphite-modified GCEs. What interesting is that when  
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16 RGO-modified GCE was used, a well-defined oxidation peak at about 0.63 V was  
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18 observed (green line), and the peak current increased greatly relative to GO-modified  
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20 GCE. The notable peak current enhancement indicates that RGO nanosheets can  
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22 greatly accelerate the electron transfer rate. The strong enhancement effect toward the  
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24 oxidation of guaiacol may attribute to the superior electroconductivity, huge specific  
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26 area and strong accumulation efficiency of RGO.  
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34 To explain the different activities of GO and RGO toward the oxidation of  
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36 guaiacol, the surface functional group of pristine graphite, GO and RGO was  
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38 investigated by FTIR. Fig. 5 shows the FTIR spectra of graphite (curve a), RGO  
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40 (curve b) and GO (curve c). After chemical oxidation of pristine graphite, numerous  
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42 oxygen-containing functional group was introduced on the surface of GO (curve c),  
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44 such as O-H ( $3428\text{ cm}^{-1}$ ), C=O ( $1724\text{ cm}^{-1}$ ), C=C ( $1635\text{ cm}^{-1}$ ) and alkoxy C-O ( $1045$   
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46  $\text{cm}^{-1}$ ). Compared with GO, the peak at  $1724\text{ cm}^{-1}$  (C=O),  $1250\text{ cm}^{-1}$  (epoxy C-O) and  
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48  $1400\text{ cm}^{-1}$  (carboxy C-O) significantly decreased on RGO surface, indicating the  
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50 effective reduction of the GO. Of course, part of oxygen-containing functional group  
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52 still existed on the surface of RGO though most of the functional groups in GO were  
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4 removed, which was similar to the other report.<sup>22</sup>  
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6 The element contents of graphite, GO and RGO were analyzed using XPS. Fig. 6  
7 displays the XPS spectra of graphite, GO and RGO. For graphite, the content of  
8 oxygen was as low as 2.60% and the content of carbon was as high as 97.40%. On the  
9 other hand, the content of oxygen remarkably increased to 32.57% for GO. However,  
10 the oxygen content decreased greatly to 15.44% for RGO compared with GO,  
11 suggesting considerable deoxygenation by the hydrothermal reduction process.  
12 Moreover, no extra elements than oxygen and carbon were detected, suggesting the  
13 purity of graphite, GO and RGO was very high. From the FTIR and XPS spectra, we  
14 know that RGO was successfully reduced from GO through the hydrothermal process.  
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29 To obtain further information on the structure of RGO, Raman spectra was  
30 carried out. Fig. 7 shows the Raman spectra of pristine graphite (a), GO (b) and RGO  
31 (c). As shown in Fig. 7a, two obvious peaks at  $1580\text{ cm}^{-1}$  and  $2700\text{ cm}^{-1}$  were  
32 observed for the pristine graphite, which was assigned to the G-band and 2D-band of  
33 the natural graphite.<sup>23</sup> Additionally, a small peak at  $1350\text{ cm}^{-1}$  was also examined  
34 which was attributed to the disorder-related D-band. According to data, the D band is  
35 due to the existence of defects in basal plane and edge of carbon material, and its  
36 intensity is linked with the amount of disorder.<sup>24,25</sup> The tiny D-band intensity ( $I_D$ )  
37 suggests that few defects existed in pristine graphite. However, the D-band became  
38 well-defined and the intensity increased obviously in the spectra of GO and RGO. The  
39 greatly-increased D-band indicates great deal of defects existed in basal plane and edge  
40 of graphene than in graphite. On the other hand, the relative intensity ratio of the D  
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3 and G ( $I_D/I_G$  ratio) is proportional to the number of defect sites in graphite carbon.<sup>24</sup>  
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6 Compared with GO, an increased  $I_D/I_G$  intensity ratio can be observed for RGO,  
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8 indicating there are more defective sites existing on the surface of RGO, which may  
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10 be the reason of higher activity toward the oxidation of guaiacol relative to GO and  
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12 the pristine graphite.  
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16 Fig. 8 shows the X-ray diffraction (XRD) patterns of graphite, GO and RGO  
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18 nanosheets. A very sharp peak was observed at  $2\theta = 26.5^\circ$  for graphite (curve a),  
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20 which attributes to the diffraction of (002) plane, and the interlayer distance obtained  
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22 from the (002) plane was 3.48 Å. A typical oxidation leads to the appearance of  
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24 diffraction peak of the GO at  $2\theta = 10.8^\circ$  without the peak of  $d_{002}$  visible in graphite  
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26 due to the introduction of oxygen-containing groups to the graphite surface,<sup>26</sup> and the  
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28 corresponding interlayer distance was calculated to be 8.37 Å. In the case of RGO,  
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30 only a broad peak at  $2\theta = 23.4^\circ$  can be observed, and the interlayer distance was 3.63  
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32 Å, indicating a sufficient reduction process of the original graphene oxide.<sup>26</sup>  
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#### 41 **Effect of pH and scan rate on the oxidation of guaiacol at RGO/GCE**

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43 In order to optimize the response of guaiacol at RGO/GCE, some different supporting  
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45 electrolytes such as phosphate buffer solution, Tris buffer solution, acetate buffer  
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47 solution, B-R buffer solution, HCl and HClO<sub>4</sub> solution was discussed by cyclic  
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49 voltammetry. The results indicate that the peak current of guaiacol in 0.1 M phosphate  
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51 buffer solution is larger than in the others (data not shown), so phosphate buffer  
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53 solution was chosen as the supporting electrolyte. The effect of phosphate buffer  
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4 solution with different pH values ranging from 5.7 to 8.0 on the oxidation response of  
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6 10  $\mu\text{M}$  guaiacol was investigated. It was found that  $E_{\text{pa}}$  was depended on the solution  
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8 pH. A good linear relationship was obtained between the  $E_{\text{pa}}$  and pH. The liner  
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10 regression equation is  $E_{\text{pa}}/\text{V} = 1.037 - 0.0624\text{pH}$ , and the correlation coefficient is  
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12 0.995. According to the relationship between the peak potential and pH, the slope of  
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14  $-62.4 \text{ mV/pH}$  indicated that the electron transfer was accompanied by an equal  
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16 number of protons in the electrode reaction process.<sup>27</sup> In addition, the maximum  
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18 current response of guaiacol was obtained at pH 6.5 phosphate buffer solution.  
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20 Therefore, pH 6.5 phosphate buffer solution was chosen for the subsequent analytical  
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22 experiments.  
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29 The influence of scan rate ( $\nu$ ) on the oxidation of guaiacol at the RGO/GCE was  
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31 investigated by linear sweep voltammetry (LSV). In pH 6.5 phosphate buffer solution,  
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33 the anode peak current of guaiacol increased continuously with the increase of scan  
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35 rate ( $\nu$ ) over the range from 0.05 to 0.25  $\text{V s}^{-1}$ . A good linear relationship between the  
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37 peak current and square root of the scan rate ( $\nu^{1/2}$ ) was obtained. The linear regression  
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39 equation is  $i_{\text{pa}}(\mu\text{A}) = -0.2739 + 30.3315\nu^{1/2}$  ( $\nu$  in  $\text{V s}^{-1}$ ), and the correlation coefficient  
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41 is 0.995, indicating the oxidation of guaiacol was controlled by diffusion. Moreover,  
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43 according to Fig. 4, only an oxidation peak was observed, indicating that the oxidation  
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45 of guaiacol was a totally irreversible electrode process.  
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52 Additionally, the  $E_{\text{pa}}$  of guaiacol shifted positively with increasing the scan rate.  
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54 The dependence of  $E_{\text{pa}}$  with  $\ln \nu$  can be expressed as  $E_{\text{pa}}(\text{V}) = 0.74926 + 0.06548 \ln \nu$   
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56 ( $\text{V s}^{-1}$ ), and the correlation coefficient is 0.995. As for an irreversible electrode  
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4 process, according to Laviron,<sup>28</sup>  $E_{pa}$  is defined by the following equation:  $E_p =$   
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6  $E^0 - (RT/\alpha nF) \ln(\alpha nF/RTk_s) - (RT/\alpha nF) \ln v$ , where  $\alpha$  is the electron-transfer coefficient,  $n$   
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8 is the electron-transfer number,  $k_s$  is the rate constant of the reaction,  $v$  is the scan rate  
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10 and  $E^0$  is the formal redox potential. So the value of  $n$  can be easily calculated from  
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12 the slope of  $E_p$  versus  $\ln v$ . Here, taking  $R = 8.314$ ,  $T = 298$  and  $F = 96500$ , generally,  
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14  $\alpha$  is assumed to be 0.5 in a totally irreversible electrode process, so it can be  
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16 calculated that one electron and one proton was involved in the oxidation of guaiacol.  
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#### 24 **Detection of guaiacol using RGO/GCE**

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26 Fig. 9 shows the influence of the amount of RGO suspension on the oxidation peak  
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28 current of 5  $\mu\text{M}$  guaiacol using differential pulse voltammetry (DPV). After 30 s  
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30 accumulation under open circuit, the oxidation peak current was recorded as the  
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32 analytical signal. The oxidation peak current of guaiacol increased remarkably with  
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34 the volume of RGO suspension over the range from 0 to 10  $\mu\text{L}$ . During this period,  
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36 the accumulation efficiency of RGO/GCE obviously enhanced. However, the  
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38 oxidation current gradually decreased when further improving the amount of RGO,  
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40 which may be due to the block effect of RGO for the decreased electric conductivity.  
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42 In order to shorten the time of solvent evaporation and to achieve high sensitivity, 10  
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44  $\mu\text{L}$  RGO suspension was used to modify the GCE surface.  
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52 Fig. 10 displays the effect of accumulation time on the oxidation peak current of  
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54 50  $\mu\text{M}$  guaiacol. By extending the accumulation time from 0 to 30 s, the oxidation  
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56 peak current increased greatly, revealing that accumulation is efficient to improve the  
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4 detection sensitivity. It is a surprise that longer accumulation time than 30 s does not  
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6 enhance the oxidation peak current, suggesting that the amount of guaiacol on the  
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8 surface of RGO tends to a limiting value. Considering sensitivity and analysis time,  
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10 30 s accumulation was employed.  
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14 The RGO modified GCE was used for single determination in this work. The  
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16 reproducibility for multiple modified GCEs was estimated by measuring the peak  
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18 current of 5  $\mu\text{M}$  guaiacol. For 5 RGO modified GCEs, the value of the relative  
19  
20 standard deviation (RSD) was 3.2%, suggesting excellent fabrication reproducibility  
21  
22 and detection precision.  
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26 The potential interferences for the determination of guaiacol were examined.  
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28 Under the optimized conditions, the oxidation peak current of 5  $\mu\text{M}$  guaiacol was  
29  
30 individually measured in the presence of different concentrations of interferents, and  
31  
32 then the peak current change was checked. No influence on the detection of guaiacol  
33  
34 was observed after the addition of 500-fold amounts of  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Pb}^{2+}$ ,  
35  
36 100-fold amounts of the pyrogallol, resorcinol, phenole (peak current change was  
37  
38 below 5%).  
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44 The linear range and limit of detection were tested using DPV under the  
45  
46 optimized conditions. As displayed in Fig. 11, the oxidation peak current of guaiacol  
47  
48 ( $i_p$ ,  $\mu\text{A}$ ) was linear with its concentration ( $C$ ,  $\mu\text{M}$ ) over the range from 0.5 to 500  $\mu\text{M}$ ,  
49  
50 obeying the following linear equation:  $i_p = 0.06898 C + 4.725$ . The correlation  
51  
52 coefficient was 0.998, suggesting good linearity. After 30 s accumulation under open  
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54 circuit, the limit of detection was evaluated to be 0.2  $\mu\text{M}$  based on three signal to  
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4 noise ratio.  
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### 8 9 **Analytical application**

10  
11 To demonstrate its applicability of the proposed method for real sample analysis,  
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13 bamboo juice sample was used for the quantitative analysis. Each sample solution  
14  
15 undergoes five parallel detections, and the RSD was below 5%. The concentration of  
16  
17 guaiacol was obtained by the standard addition method, and the results are listed in  
18  
19 Table 1. In order to testify the accuracy of this method, the content of guaiacol was  
20  
21 also analyzed using ultraviolet spectrum. The obtained results are in good agreement,  
22  
23 revealing that this method is satisfactory. In addition, a known amount of guaiacol  
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25 standard was spiked in the sample, and then analyzed according to the same procedure.  
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31 The value of recovery was in the range from 98.9% to 105.8%, also indicating that  
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determination of guaiacol using RGO modified GCE is accurate and feasible.

### 39 **Conclusion**

40  
41 RGO nanosheets with high quality were chemically synthesized by hydrothermal  
42  
43 reduction of well-dispersed GO suspension. Compared with glassy carbon, graphite  
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45 and GO, RGO remarkably increased the peak current toward the oxidation of  
46  
47 guaiacol. Based on the great enhancement effect of RGO nanosheets, a rapid,  
48  
49 sensitive and convenient electrochemical method was developed for the detection of  
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51 guaiacol. This method was successfully used in bamboo juice sample, and revealed  
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promising application.

## Acknowledgements

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## References

- 1 J.L. Achtyl, I.V. Vlassioux, P.F. Fulvio, S.M. Mahurin, S. Dai, F.M. Geiger, *J. Am. Chem. Soc.*, 2013, **135**, 979-981.
- 2 B. Zhang, L.X. Fan, H.W. Zhong, Y.W. Liu, S.L. Chen, *J. Am. Chem. Soc.*, 2013, **135**, 10073-10080.
- 3 A.K. Geim, K.S. Novoselov, *Nat. Mater.*, 2007, **7**, 6183-6191.
- 4 X. Wang, L.J. Zhi, K. Müllen, *Nano Lett.*, 2008, **8**, 323-327.
- 5 Z.G. Cheng, Q. Li, Z.J. Li, Q.Y. Zhou, Y. Fang, *Nano. Lett.*, 2010, **10**, 1864-1868.
- 6 H.L. Wang, Q.L. Hao, X.J. Yang, L.D. Lu, X. Wan, *Electrochem. Commun.*, 2009, **11**, 1158-1161.
- 7 C.Y. Wang, D. Li, C.O. Too, *Chem. Mater.*, 2009, **21**, 2604-2606.
- 8 S.P. Pang, J.M. Englert, H.N. Tsao, Y. Hernandez, A. Hirsch, X.L. Feng, K. Müllen, *Adv. Mater.*, 2010, **22**, 5374-5377.
- 9 H. Huang, W. Chen, S. Chen, A.T.S. Wee, *ACS Nano*, 2008, **2**, 2513-2518.
- 10 A. Reina, X.T. Jia, J. Ho, D. Nezich, H. Son, V. Bulovic, M.S. Dresselhaus, J.



- 1  
2  
3  
4 Kong, *Nano. Lett.*, 2009, **9**, 30-35.  
5  
6 11 D.B. Shinde, J. Debgupta, A. Kushwaha, M. Aslam, V.K. Pillai, *J. Am. Chem. Soc.*,  
7  
8 2011, **133**, 4168-4171.  
9  
10 12 G.X. Wang, B. Wang, J. Park, Y. Wang, B. Sun, J. Yao, *Carbon*, 2009, **47**,  
11  
12 3242-3246.  
13  
14 13 M. Zhou, Y.M. Zhai, S.J. Dong, *Anal. Chem.*, 2009, **81**, 5603-5613.  
15  
16 14 X. Chen, H. Ye, W. Wang, B. Qiu, Z. Lin, G. Chen, *Electroanalysis*, 2010, **22**,  
17  
18 2347-2352.  
19  
20 15 Q. Lu, X. Dong, L.J. Li, X. Hu, *Talanta.*, 2010, **82**, 1344-1348.  
21  
22 16 J.D. Fowler, M.J. Allen, V.C. Tung, Y. Yang, R.B. Kaner, B.H. Weiller, *ACS Nano.*,  
23  
24 2009, **3**, 301-306.  
25  
26 17 T. Ohkubo, M. Shibata, *J. Dent. Res.*, 1997, **76**, 848-851.  
27  
28 18 H.F. Liu, B. Lepoittevin, C. Roddier, V. Guerineau, L. Bech, J.M. Herry, M.N.  
29  
30 Bellon-Fontaine, P. Roger, *Polymer*, 2011, **52**, 1908-1916.  
31  
32 19 M. Takahashi, S. Sakamaki, A. Fujita, *Biosci. Biotechnol. Biochem.*, 2013, **77**,  
33  
34 595-600.  
35  
36 20 P.R. Pérez-Cacho, M.D. Danyluk, R. Rouseff, *Food Chem.*, 2011, **129**, 45-50.  
37  
38 21 W.S. Hummers, R.E. Offeman, *J. Am. Chem. Soc.*, 1958, **80**, 1339-1340.  
39  
40 22 C. Wu, D. Sun, Q. Li, K.B. Wu, *Sens. Actuator B-Chem.*, 2012, **168**, 178-184.  
41  
42 23 A.C. Ferrari, J. Robertson, *Phil. Trans. Roy. Soc. A.*, 2004, **362**, 2269-2270.  
43  
44 24 F. Tuinstra, J.L. Koenig, *J. Chem. Phys.*, 1970, **53**, 1126-1130.  
45  
46 25 A.C. Ferrari, J. Robertson, *Physical Review B*, 2000, **61**, 14095-14107.  
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4 26 C. Hontoria-Lucas, A.J. López-Peinado, J.D.D. López-González, M.L.  
5  
6 Rojas-Cervantes, R.M. Martín-Aranda, *Carbon*, 1995, **33**, 1585-1592.  
7  
8  
9 27 F.Y. Zhang, Z.H. Wang, Y.Z. Zhang, Z.X. Zheng, C.M. Wang, Y.L. Du, W.C. Ye,  
10  
11 *Talanta*, 2012, 93, 320-325.  
12  
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14 28 E. Laviron, *J. Electroanal. Chem.*, 1974, **52**, 355-393.  
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**Captions for figures and table**

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9 Fig. 1 Structural formula of guaiacol.

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14 Fig. 2 AFM images of GO (a) and RGO nanosheets (b).

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19 Fig. 3 SEM images of GCE (A), graphite (B), GO (C) and RGO (D); TEM images of  
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21 GO (E) and RGO (F).

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26 Fig. 4 CV curves of 50  $\mu\text{M}$  guaiacol on the unmodified GCE (black line),  
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28 graphite-modified GCE (red line), GO-modified (blue line) and RGO-modified (green  
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30 line) GCEs. Scan rate: 100  $\text{mV s}^{-1}$ .

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36 Fig. 5 FTIR spectra of graphite (a), RGO (b) and GO (c).

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41 Fig. 6 XPS spectra of graphite, GO and RGO.

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46 Fig. 7 Raman spectra of Graphite, GO and RGO.

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51 Fig. 8 The picture of XRD: graphite (a), GO(b) and RGO (c).

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56 Fig. 9 Influence of amount of RGO suspension on the oxidation peak current of 5  $\mu\text{M}$   
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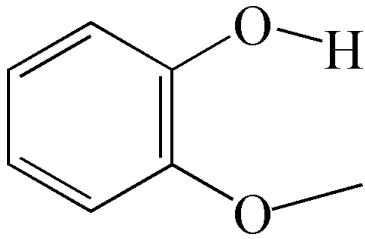
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4 guaiacol.  
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9 Fig. 10 Influence of accumulation time on the oxidation peak current of 50 $\mu$ M  
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11 guaiacol.  
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16 Fig. 11 DPV curves of guaiacol with different concentrations on RGO modified GCE.  
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18 (a) 0  $\mu$ M, (b) 2  $\mu$ M, (c) 5  $\mu$ M, (d) 10  $\mu$ M, (e) 50  $\mu$ M, (f) 105  $\mu$ M and (g) 200  $\mu$ M.  
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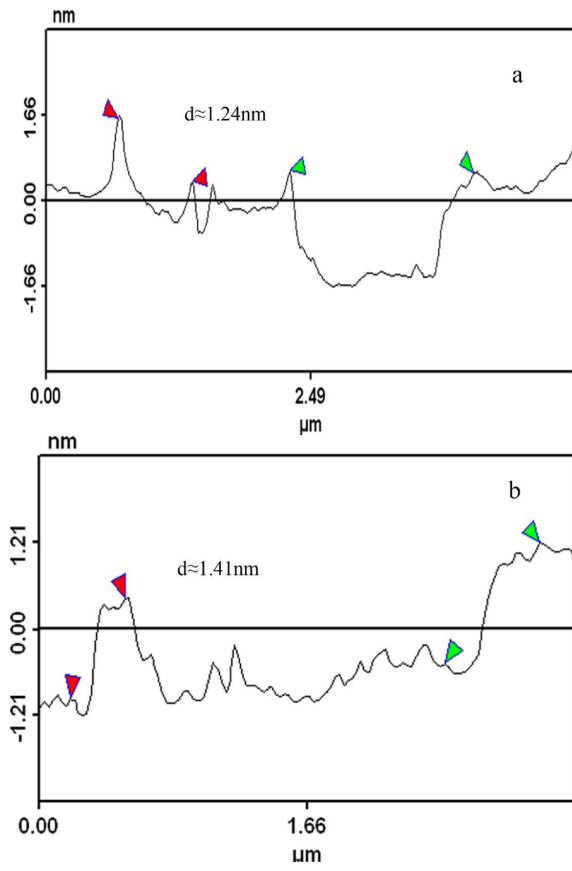
23  
24 Table 1 Determination of guaiacol in bamboo juice.  
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Fig. 1



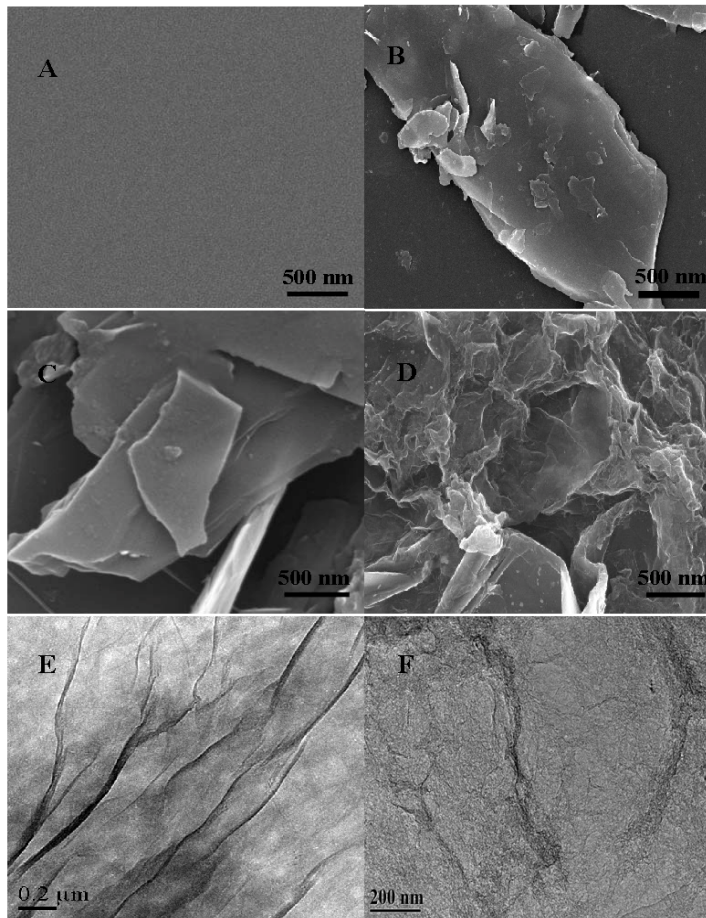
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Fig. 2



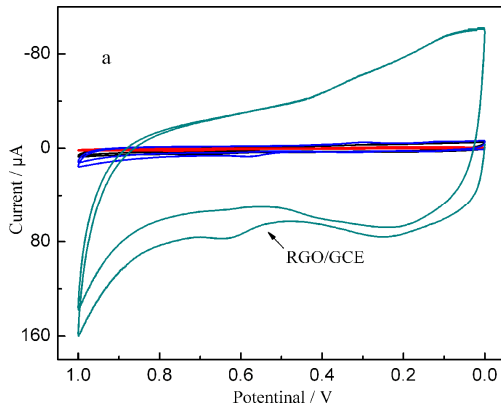
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Fig. 3



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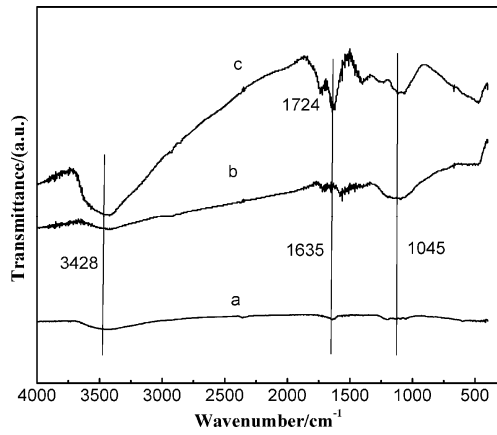
Fig. 4



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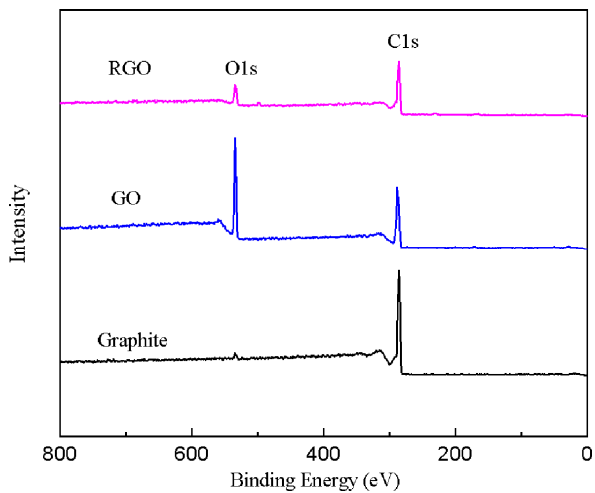


Fig. 5



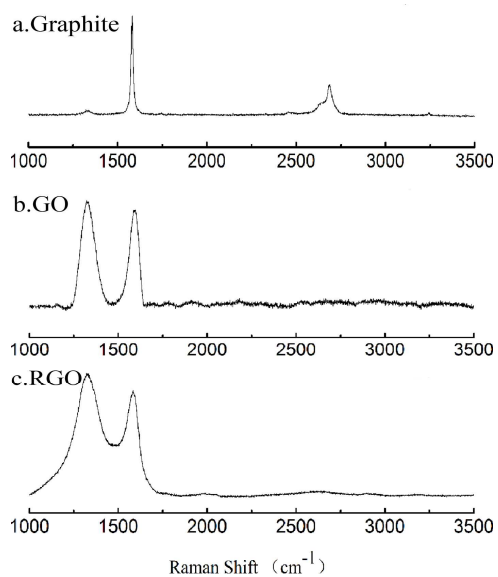
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Fig. 6



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Fig. 7



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Fig. 8

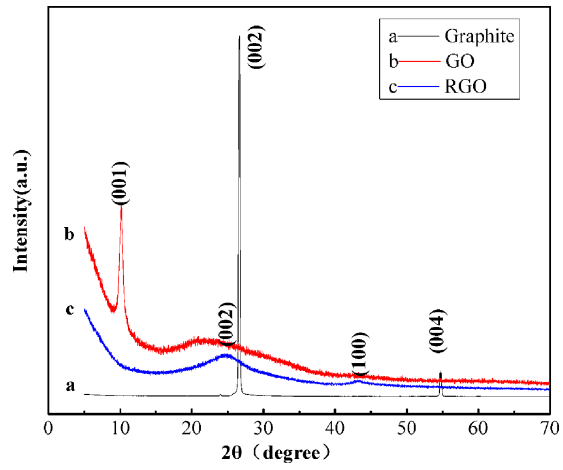
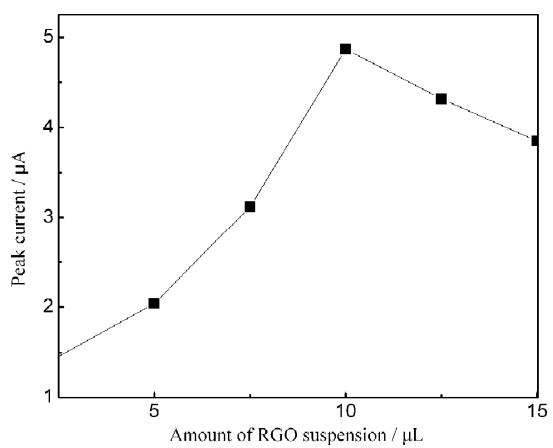


Fig. 9



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Fig. 10

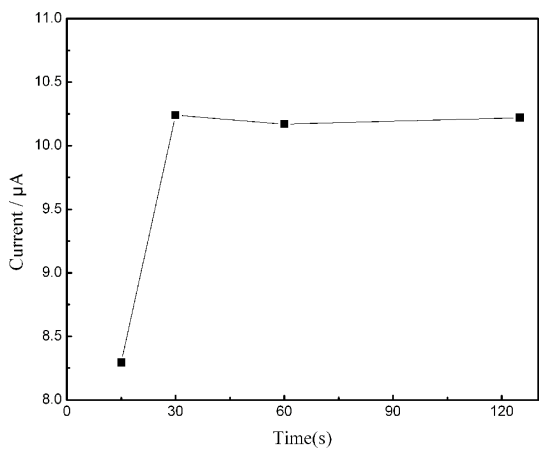
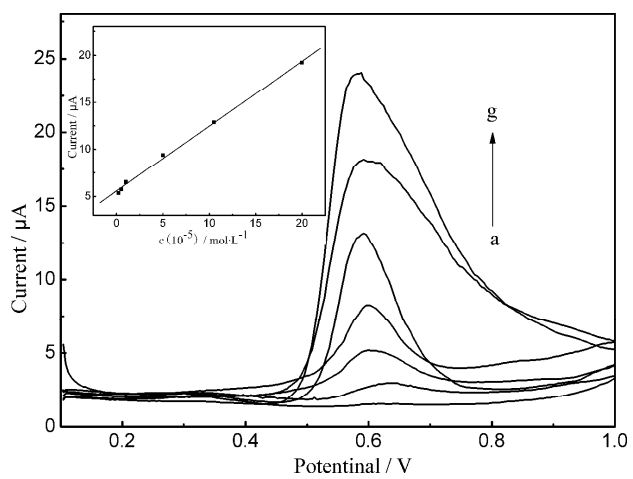


Fig. 11



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Table 1 Determination of guaiacol in bamboo juice.

| Sample | UV<br>(M)             | This method<br>(M)    | Recovery<br>(%) | RSD<br>(%,n=5) |
|--------|-----------------------|-----------------------|-----------------|----------------|
| A      | $5.07 \times 10^{-6}$ | $5.29 \times 10^{-6}$ | 105.8           | 1.2            |
| B      | $6.72 \times 10^{-5}$ | $6.96 \times 10^{-5}$ | 99.4            | 4.5            |
| C      | $2.59 \times 10^{-4}$ | $2.67 \times 10^{-4}$ | 98.9            | 3.6            |

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4 Reduced graphene oxide (RGO) nanosheets with high quality were chemically  
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6 synthesized by hydrothermal reduction of well-dispersed graphene oxide (GO)  
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8 suspension. The electrochemical behavior of guaiacol was studied on different carbon  
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10 materials surface. Compared with the bare glassy carbon, graphite, and GO, RGO  
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12 nanosheets exhibited strong enhancement effect and the electrochemical oxidation  
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14 response of guaiacol was remarkably improved.  
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