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ARTICLE

A step towards Mobile Arsenic measurement for surface waters

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C.A. de Villiers,^a M.C. Lapsley^a and E. A. H. Hall^{a*},

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Surface modified quantum dots (QDs) are studied using a bio-inspired cysteine rich ligand (glutathione, GSH) and their quenching response and selectivity to arsenic examined. As predicted from As³⁺ binding with highly crosslinked phytochelatin-(PCn)-like molecules, better arsenic selectivity is obtained for a thicker more 3-dimensional GSH surface layer, with exposed sulfhydryl groups. A detection limit of at least 10 μM can be achieved using CdSe/ZnS core-shell QDs capped with this GSH structure. The system is also demonstrated using a mobile phone camera to record the measurement, producing a detection limit of 5 μM. However, copper remains the main interferent of concern. Water-soluble CdTe QDs show little sensitivity to As³⁺ even with a GSH surface, but they remain sensitive to Cu²⁺, allowing a copper baseline to be established from the CdTe measurement. Despite anticipating that spectrally non overlapping fluorescence would be required from the two types of QDs to achieve this, a method is demonstrated using RGB channels from a mobile phone and processing the raw data for CdTe QDs, with an emission wavelength of 600nm, and CdSe/ZnS QDs, with emission maximum of 630nm. It is shown that As³⁺ measurement remains feasible at the WHO guideline value of 10 μg/L up to a copper concentration of around 0.3 μM Cu²⁺, which corresponds to the highest recorded level in a selection of large rivers worldwide.

Introduction

Many water sources, in both developed and under-developed countries, continue to contain high levels of arsenic (As), at concentration levels far exceeding the WHO drinking water guideline value of 10 μg/L.¹ The chronic exposure to arsenic through the consumption of these waters has been recognised as an environmental health disaster. Ingestion of amounts in excess of the guideline is putting 100 million people at risk of cancer and other arsenic-related diseases.² It has long been realised that the determination of the total arsenic concentration is insufficient for environmental considerations, because the bioavailability as well as the physiological and toxicological effects of arsenic depend on its chemical form.^{3,4} Consequently, knowledge of the speciation of arsenic in natural water is important.

It is generally accepted that trivalent arsenicals (As(III)) are more toxic than the corresponding pentavalent arsenicals (As(V)).^{5,6} The toxicity of trivalent arsenicals likely occurs through its interaction with sulfhydryl groups of cysteines in biomolecules. As(III) binding to a specific protein could block its normal capacity for binding its normal substrate(s), alter the protein's conformation and function as well as its interaction with other functional proteins leading to a deterioration in cellular functions.⁶

Due to the well-recognised importance of arsenic detection, an over-abundance of detection methods based on spectrometric and electrochemical methods, inductively coupled plasma-mass spectrometry, neutron activation analysis, optical spectrometric techniques, colorimetric measurement methods and biosensors have been developed, reported and reviewed.^{3,4,7,8} Most of these detection techniques obtain limits of detection below the WHO arsenic guideline value. Nevertheless, a vast number of existing methods, including atomic fluorescence spectrometry (AFS), which is considered the best option for ultra-trace arsenic in water, are suitable for laboratory conditions only.^{4,9-11}

At present, the majority of commercially available field sensing kits for arsenic are equipped with a colorimetric readout scheme.⁴ The best reported limit of detection with these kits is 2 mg/L.^{4,12} Although the colorimetric field kits are able to provide rapid results in on-site conditions, they lack sensitivity and may show weak correlation with laboratory methods.¹³ Scaling up to a global remote analytical method requires innovative analysis combined with mobile hardware solutions. Due to their ubiquity, connectivity, and increasing complexity and power, the field of mHealth (mobile health) has emerged around mobile phones¹⁴. The goal is to provide low cost, efficient, healthcare solutions to people around the world. There are almost 7 billion mobile phones worldwide, with enthusiastic uptake not just in the developed world, but also in

1 developing countries where it is predicted that mobile phone
2 subscriptions will soon account for 78% of the world's total¹⁵.
3 Technologies embedded within these devices are developing
4 fast due to high consumer demand, leading to the proliferation
5 of 'smartphones' which include powerful processors, wireless
6 connectivity, and sensors such as cameras, microphones,
7 accelerometers and GPS positioning¹⁶.

8 Modern mobile phone technology is attractive and, potentially,
9 distributed data networks allowing interpretation and
10 processing of environmental data (eg arsenic levels) within the
11 communication network itself, could enable other benefits such
12 as water-course cartography, guiding safer consumption.

13 The camera within the mobile phone, offers a clear detection
14 system. However, the intended use for mobile phone camera
15 modules is to record pictures to be viewed by the human eye.
16 Translating this to metrology, where an image is taken, then
17 requires analysis either by a person (at the scene or remotely) or
18 computationally. Bourouis *et al.* have developed an Artificial
19 Neural Network algorithm which runs on a smartphone to
20 analyse images of skin to identify cancers¹⁷, allowing
21 examinations in remote and poor areas.

22 Many of these ideas make use of the RGB capabilities of the
23 smartphone camera by providing a reasonably faithful colour
24 representation to the viewer in a complex 2-D image. In these
25 cases exact spectral information is not important, rather the
26 colours demarcate different areas of the images. However,
27 successful quantisation on a smartphone for a diagnostic has
28 also been shown for Roche's Cardiac proBNP test strip¹⁸, and
29 accurate yes/no readings have been shown using a mechanical
30 attachment test strip holder for tuberculosis, HIV and malaria
31 commercial test strips, with an app which geotags the locations
32 and sends them to a central server for disease mapping¹⁹.

33 The goal now is to take the next step that goes behind the
34 processed camera image and takes the RAW data directly from
35 the pixels of the ccd. Without the same power of laboratory
36 based spectroscopy tools, successful use of a portable mobile
37 phone analytical laboratory also needs well defined optical
38 properties for the reagents that can be matched with the phone's
39 camera output (ie RGB). In recent years, colloidal
40 semiconductor nanoparticles (NPs), also known as quantum
41 dots (QDs), have emerged as fluorescent probes for the
42 detection of analytes, including many biological, organic and
43 inorganic species.²⁰ With their broad absorption and *narrow*
44 and tunable emission spectra,²⁰⁻²³ they offer a clear opportunity
45 for compatibility with the RGB component resolution well
46 suited to a mobile phone system. As QDs are semiconductors,
47 absorption of a photon can excite an electron within the valence
48 band to the conduction band creating an electron-hole pair.^{24,25}
49 Recombination of the electron-hole pair results in fluorescence.
50 The breakthrough QD papers of Chan & Nie²⁶ and Bruchez *et*
51 *al.*²⁷ demonstrated that QDs can be made water-soluble,
52 biocompatible and functionalised via surface modification and
53 biconjugate techniques. Therefore, through the use of
54 appropriate functionalization techniques, QDs have a high
55 potential for use as highly sensitive fluorescent biomarkers,

(bio)chemical probes and inorganic ion sensors in aqueous
environments.^{20,22,25,28-30}

56 Wang *et al.*³¹ examined the reaction of As(III) with
mercaptoacetic acid (MA)-capped CdTe, MA-capped
CdTe/ZnS and glutathione (GSH)-capped CdTe QDs. Wang *et*
al. found that only the GSH-capped CdTe QDs showed a
response to As(III) and was quenched upon adding As(III)
between 375 µg/L - 1900 µg/L.³¹ These QDs did not respond
significantly to other metal ions such as Na⁺, Ca²⁺, Mg²⁺, Al³⁺,
Ni²⁺, Cr³⁺, Mn²⁺, Zn²⁺, Cu²⁺ and Fe³⁺.³¹ Using similar chemistry
to Wang *et al.*³¹, Butwong *et al.*³² took MA-capped CdS QDs,
but this required a preconcentration step which generated arsine
(AsH₃), which was then reacted with the QDs in a gas diffusion
unit in a concentration range of 0.08 - 3.20 mM.³² Arsine
generation is the most common preconcentration technique for
ultra-trace determination of arsenic, as it greatly improves
sensitivity regardless of the analytical technique used.
However, this imposes limitations in construction as a portable
device for *in situ* determination of As(III). As the maximum
concentration limit of As allowable in drinking water is 10 µg/L
(0.133 µM),¹ the selectivities and limits of detection (LOD) for
As, especially for trivalent arsenicals, reported using QDs for
detection, without the use of preconcentration, are still not
sufficiently low to be considered as a viable option for As(III)
sensing in water.

57 The research reported herein starts with the premise that
sulfhydryl groups of cysteines in peptides and proteins are
highly efficient binding ligands of As(III), central to the
toxicity process⁶, and further examines the capacity for a
sensitive As(III) determination involving a bio-inspired
cysteine-containing peptide affinity ligand linked with
responsive quantum dots (QDs). The approach is based on
examination of the glutathione ligand, as a simple affinity unit
for a QD, that could be transferred to a mobile phone based
assay and consideration of its improved As(III) binding
selectivity and sensitivity.

Materials and Methods

58 All reagents used were of analytical grade and Milli-Q water
was used for dilutions. All solvents were used as supplied. QDs
were acquired from PlasmaChem. Hydrophobic QDs were
CdSe/ZnS core-shell particles were supplied dry with a
surfactant capping of trioctylphosphine/ trioctylphosphine
oxide (TOP/TOPO). These QDs are referred to as QD630
because of their maximum emission wavelength at 630 ± 5nm.
Hydrophilic QDs were CdTe core particles with short
mercaptocarboxylic acid capping ligands. These QDs are
referred to as QD₆₀₀ and QD₅₅₀ referring to their maximum
emission wavelength. The hydrophilic QDs were suspended in
water at a concentration of 1 mg/ml.

59 3-Mercaptopropionic acid (MPA), 3-Mercaptoundecanoic acid
(MUA), N-Hydroxysuccinimide (NHS) 98%, L-glutathione,
N,N-Dimethylformamide (DMF) anhydrous 99.8%, potassium
chloride, potassium phosphate monobasic, potassium
phosphate dibasic, Ethylenediaminetetraacetic acid disodium

salt dehydrate (EDTA), aluminium chloride, calcium chloride, copper (II) chloride, magnesium chloride, manganese chloride, arsenic (III) chloride and iron (III) chloride were acquired from Sigma Aldrich. Nickel chloride, and cobalt chloride was acquired from Acros Organics. Zinc Chloride was acquired from Fisher Scientific. 1-ethyl-3-(3-(dimethylamino)propyl) carbodiimide hydrochloride (EDC) was acquired from Fluka Analytical, Amersham. Cy5 Maleimide mono-reactive dye was acquired from GE Healthcare. All reactions took place at standard laboratory conditions.

Preparation of water-soluble quantum dots

The CdTe QD₆₀₀ were used as supplied with a mercaptocarboxylic acid capping. However, the CdSe/ZnS QD₆₃₀ were hydrophobic, and consequently water-soluble ligand capping exchange procedures were performed on these QDs. The QD₆₃₀s were functionalised with different carboxy-thiol capping ligands, namely, l-glutathione (GSH), mercaptopropionic acid (MPA), mercaptoundecanoic acid (MUA), and a mixture of MPA and MUA in a 2:1 MPA:MUA molar ratio.

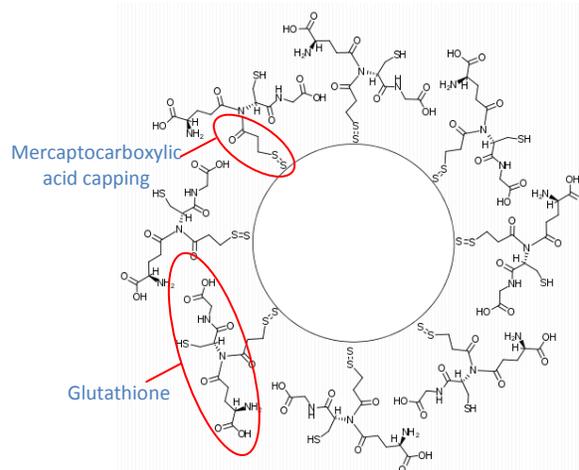
The method was adapted from Ruedas-Rama *et al.*²⁸ and Zeng *et al.*²³ with the following modifications: QD₆₃₀ were re-dispersed in chloroform at a concentration of 1 mg/mL. QD₆₃₀s in chloroform were then rendered water-soluble by ligand exchange. For GSH ligand exchange, typically, 1 mL QD₆₃₀ in chloroform (~ 2.67 nmol) was used, to which a GSH solution (800 µL from a solution containing 71 mg GSH and 20 mg NaOH in 1 mL methanol) was added. The resulting solution was stirred at room temperature for several hours or overnight. NaOH solution (1 M, 0.5 mL) was then added to extract QDs to the aqueous phase, and the chloroform layer was removed using a separation funnel. NaOH solution (1 M, 0.5 mL) was added to the extracted chloroform to wash out any remaining QDs. The aqueous QD solution was then separated into 4 eppendorfs. Excess GSH was removed by two consecutive precipitation steps, using acetone to aid precipitation followed by centrifugation. The resulting QD₆₃₀s were dissolved in 1 mL phosphate buffer (50 mM, pH 7.2) to produce a resulting concentration of 2.67 µM QD equivalent. The capped QDs were stored at 4 °C and shielded from sunlight to prevent photodegradation.

MPA, MUA, and MPA/MUA (2:1 molar ratio) capped QDs were prepared using a similar procedure. CdSe/ZnS QDs capped with MPA, MUA, GSH and MPA/MUA (2:1 molar ratio) are referred to as QD₆₃₀-MPA, QD₆₃₀-MUA, QD₆₃₀-GSH and QD₆₃₀-MPA/MUA, respectively.

Peptide Attachment to Quantum Dots

Peptide conjugation to QDs was performed through EDC/NHS activation using the method adopted from Ruedas-Rama *et al.*,²³ Wang *et al.*³², and recommended by Plasmachem³³. Briefly 0.5 mL of GSH solution (0.08 to 80 mg/mL), 0.5 mL of EDC/NHS solution containing 40 mg/mL EDC and 4 mg/mL NHS, were mixed with 1 mL of carboxylated QD₆₀₀ or QD₆₃₀ solution (1 mg/mL) all in phosphate buffer (50 mM, pH 7.2).

The EDC/NHS solution was prepared per batch and used immediately. After preparation of the reaction mixture, it was



Scheme 1. Schematic of QD₆₀₀-peptide conjugate product through EDC/NHS attachment of GSH to carboxylated QD surface.

stirred at room temperature for 2-4 hours and then stored at 4 °C overnight.

To ensure removal of excess unreacted reagents, the QD product was washed three times with phosphate buffer (50 mM, pH 7.2) using centrifugal concentrators with a 10000 MWCO membrane and centrifugation at 4000 rpm for 20 minutes per wash. The final washed QD-peptide conjugated product was resuspended in 1 mL phosphate buffer (50 mM, pH 7.2), unless stated otherwise. Scheme 1 shows a schematic of the QD₆₀₀-peptide conjugate product obtained. The CdTe QD₆₀₀-peptide conjugate obtained is referred to as QD₆₀₀-GSH in this document, while the CdSe/ZnS QD₆₃₀-peptide obtained is referred to as QD₆₃₀-GSH-GSH.

Characterisation of Successful Peptide Attachment

Different concentrations of GSH (0, 0.08, 0.8, 8 and 80 mg/mL) was conjugated to equal concentrations of QDs using the method described above and reacted with a maleimide reactive dye, which has a high affinity to form sulfur bonds with the sulfhydryl groups on the GSH. Formation of a bond between the fluorescent dye and the peptide attached to the QD will result in FRET as the dye comes in close vicinity to the QDs and the QD mixture is excited at a wavelength around 400 nm.

The Amersham Cy5 maleimide mono-reactive dye was prepared by adding 50 µL of anhydrous DMF to one pack of dye as received. The vial was flushed with nitrogen gas, capped and mixed thoroughly. After preparation of the dye, 1.5 µL of the dye solution was added to each eppendorf of QD-GSH conjugates (1 mL of ~ 1 mg/mL QD-GSH). The eppendorfs were flushed with nitrogen, capped and their contents were mixed thoroughly. The dye reaction was left to incubate at room temperature for two hours with additional mixing every 30 minutes, before being left overnight at 4 °C. The

fluorescence emission spectra of the QD-GSH-dye conjugates were scanned before washing excess dye away. The QD-GSH-dye conjugates were then separated from the excess unconjugated dye by dialysis using centrifugal concentrators with a 10000 MWCO membrane and centrifugation at 4000 rpm for 20 minutes. The centrifugation washing step was performed three times, using 1 mL phosphate buffer (50 mM, pH 7.2) for re-suspension of QDs between washing steps. The final washed product was resuspended in 1 mL phosphate buffer (50 mM, pH 7.2) and the fluorescence emission spectra scanned. Control emission spectra from solutions containing only GSH-dye, dye alone, and QD-GSH alone were also scanned at excitation wavelength of 400 nm to determine the change in the emission spectra caused by FRET.

Fluorescent Intensity Measurements

Steady-state fluorescence emission spectra were measured using a Cary-Eclipse fluorescence spectrofluorometer (Varian). The spectrofluorometer was equipped with a xenon discharge lamp (75 kV), Czerny-Turner monochromators, two detectors (sample and internal reference), and an R-928 photomultiplier tube (PMT) with manual and automatic voltage controlled using the Cary-Eclipse software. All samples, except for the samples used for characterisation of successful peptide attachment using a maleimide reactive dye, were illuminated at an excitation wavelength of 360 nm and the emission was scanned from 550 nm to 680 nm. Samples used for characterisation of successful peptide attachment were illuminated at an excitation wavelength of 400 nm and the emission was scanned from 550 nm to 720 nm. The instrument excitation and emission slits as well as the PMT detector voltage was maintained constant for each batch of QD-peptide configurations. Excitation and emission slit sizes were selected at either 10 or 20 nm for both slits and PMT values chosen were between 580 and 800 V. 96-well black microtiter plates from Nunc were used for measurement of most fluorescence emission spectra. Samples for the characterisation of successful peptide attachment were placed in a quartz microcell (Starna) with a light path length of 10 mm (160- μ m inner volume) for measurement of fluorescence emission spectra.

Effect of Cations on QD Fluorescence

The effect of various metal ions on the fluorescence intensity of QD₆₃₀-MPA, QD₆₃₀-MUA, QD₆₃₀-GSH, QD₆₃₀-MPA/MUA and QD₆₀₀ was investigated. Sets of samples were prepared by incubating 200 μ L of metal ion solution (ranged from 0.5 μ M - 1 mM) in phosphate buffer (50 mM, pH 7.2) and 100 μ L of 0.1 mg/mL QD-conjugate solution (\sim 0.132 nmol equivalent QD₆₀₀ conjugates or \sim 0.027 nmol equivalent QD₆₃₀ conjugates) in each well. The QD fluorescence emission response toward the metal ions at different concentrations was scanned and compared.

Arsenic Calibration of Carboxylated QDs and QD-peptide Conjugates

The response of the QD₆₃₀ and QD₆₀₀ fluorescence emission in the presence of various concentrations of As³⁺ was investigated to establish whether a calibration relationship between QD fluorescence response and the As³⁺ concentration exists. 200- μ L aliquots of As³⁺ ion stock solutions (ranged 5 nM-30 mM) in phosphate buffer (50 mM, pH 7.2) were added to 100 μ L of QD-peptide conjugates of QD₆₃₀ (\sim 0.026 nmol) or QD₆₀₀ (\sim 0.132 nmol) in each well. The mixtures were left to incubate for an hour before the fluorescence emission spectra were scanned. Because of the broad As³⁺ ion response range in all cases, logarithmic representations were made. The data, corresponding to the average of three determinations, were fitted by a standard least-squares treatment to determine a calibration curve.

Mobile Phone

The mobile telephone used for this study was the 2008 Nokia N85 smartphone which has an integrated 5 megapixel camera and runs the Symbian OS v9.3 operating system. This module was provided by the Nokia Research Centre, Cambridge, UK as a development handset. The development handset runs an in-house application called RAWCam which allows control of camera settings which are usually automated, and gives access to the unprocessed raw data files which are produced by the camera chip. Some black-box information has also been provided on the Carl-Zeiss camera lens

In comparison with current mid/top end mobile phone cameras the N85 is now a low spec but ubiquitous camera. The specifications are summarized below in Table 1:

| | |
|--|---|
| Resolution: | 5MP |
| CMOS sensor: | 2560x1920 pixels |
| Lens: | Carl Zeiss optics |
| Flash: | LED flash |
| Focal length: | 5.2mm |
| Aperture Diameter: | 1.82mm |
| F-number: | 2.8 |
| Diagonal FOV: | 67deg (infinity case) |
| Horizontal FOV: | 56deg |
| Pixel pitch: | 2.2 μ m |
| Sensor max image height (from centre to image corner): | 3.5mm |
| CPU: | 369 MHz ARM 11 |
| Connectivity: | Bluetooth, wireless, voice call and SMS |
| Data output: | raw, jpeg |

Table 1 N85 smartphone specifications

The camera in the Nokia phone is a 1/2.5" five megapixel autofocus module produced by Toshiba to the SMIA95 size specification. It comprises a 2560x1920 pixel CMOS sensor overlaid by a Bayer filter to provide colour information (see figure 5b). The task of processing image data is shared between the phone processor and the camera module itself. The CMOS chip contains circuitry behind each pixel to convert photon count to voltage, amplify this voltage, and then to digitise the information. The phone then performs various post processing algorithms to make the image more pleasing to the human eye and compress the image into a jpeg file. Both raw and jpeg

images are produced. Raw files contain the calculated average R, G and B values for each pixel without any further image processing. These files are uncompressed and therefore relatively large. The jpeg files are unsuitable for use as they are not only subject to lossy compression, but the images are manipulated to create a colour balance which is pleasing to the eye rather than an accurate representation of the spectral information captured. In order to run post-processing algorithms more suitable for analytical data the pre-processed raw file was used. The Nokia development application RAWcam gives access to these raw files as well as more control over the camera. The following control and set-up was used:

The LED (Forge Europa 3mm Blue LED, manufacturer number FNL-U300B22WCSL. Peak wavelength 458nm. Driven by a 5V power supply in series with a 75 Ohm metal film resistor) was positioned approx 5mm away from a well plate (transparent flat bottomed well strip, Greiner bio-one. Catalogue number 756070), illuminating from the side through the well wall. The filter (Edmund optics glass long pass filter 475nm, name: GG-475, 12.5mm Dia. Longpass Filter. catalogue number: #54-651) was attached to the camera module in front of the lens. The camera and filter were placed above the well looking down into it. Each QD mixture was left in the dark for 1hr before the image was taken.

Images used 5000s exposure, no flash, no neutral density filtering, no analogue gain, raw format images. Image analysis was performed by identifying the image area at the centre of the well selected by hand. This area is then used to analyse all images. The red/green/blue channel readings were recorded and processed separately and averaged over the defined area for each image to give the reading.

Results and Discussion

QD capping ligand

QD capping-ligand exchange using a carboxy terminated thiol,

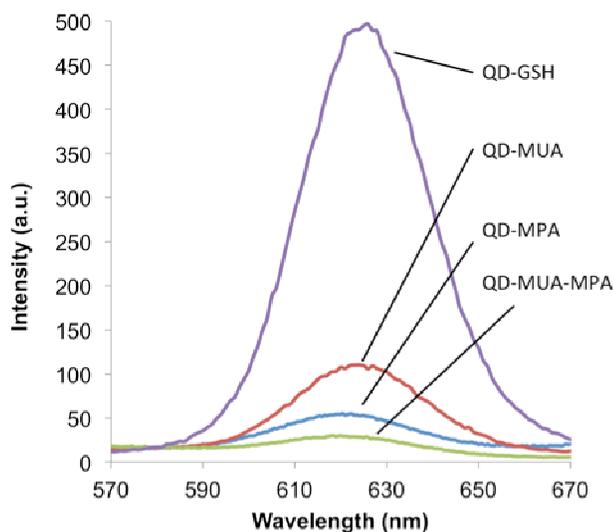


Figure 1: Fluorescence Emission Spectra of 0.09 μM CdSe/ZnS QDs after attachment of each different capping ligand (Excitation at 360nm; PMT: 800V; Excitation Slit: 20nm; Emission Slit: 20nm).

particularly a straight chain thiol like MUA or MPA^{23,28} has become a routine method for rendering water insoluble QDs both water-soluble, and displaying a surface active carboxyl, for further derivatisation. Nevertheless, this exchange is not without consequence and usually results in loss of QD emission intensity. In the case of arsenic, low detection levels are required so that loss of QD sensitivity will have an impact. Figure 1 compares the fluorescence emission spectra of QD₆₃₀-MPA, QD₆₃₀-MUA, QD₆₃₀-GSH, and QD₆₃₀-MPA/MUA. QD₆₃₀ conjugated to GSH retained the highest fluorescence intensity compared with the other capping ligands. It can also be noted that MUA and MPA caused hypsochromatic shifts (blue-shift) up to 10 nm in the wavelength of the maximum emission peak compared to that of the unfunctionalised QD₆₃₀. This, combined with the fluorescence emission intensity decrease, is consistent with core degradation as a result of photo-oxidation of the QD surface and, consequently, an increasing number of surface defects and a decreasing size of the nanocrystal.^{34,35} Surface defects contribute to radiationless recombination of the excitons, which limits the fluorescence efficiency of the QDs. In this instance, the QD₆₃₀-GSH conjugate, with a branched structure has a higher quantum yield and provides a more efficient capping ligand for the QD. This is anticipated from consideration of the QD surface curvature and the lower expected space-filling capability of a straight chain thiol versus a branched thiol. This corresponds to reports also suggesting that multidentate thiolated ligands improve stability and quantum yield.³⁶ However, by analogy with a mechanism proposed by Tehrani et al³⁷ for GSH on Au-nanoclusters, the association of GSH at the QD surface may also be associated with oxidation of the GSH to GSSG by the QD.

In contrast to the QD₆₃₀, CdTe QD₆₀₀ were supplied already rendered water-soluble with a carboxylated capping ligand, so there was no capacity to investigate the impact of the capping ligand.

In addition to its protective role, the capping ligand needs to impart selectivity towards As³⁺, appropriate for drinking water consumption. In particular this means that selectivity with respect to other metal cations. Metal binding ligands are well recognised in nature and include both macromolecular structures such as proteins, smaller oligos and chelation by organic acids, such as citrate and malate³⁸. Taking inspiration from these biological models, the simple MUA and MPA QD functionalisation may provide a suitable analogue of the organic acid chelator, whereas complexation of As³⁺ through Cys-rich peptides is a feature of metallothioneins (MTs) and the enzymatically synthesized phytochelatin (PCs)³⁹. Cadmium, lead and mercury are also known to form metal-GSH complexes, but as these ions are not commonly present in water unless there has been significant pollution ([Cd²⁺] < 0.0009 μM ; [Pb²⁺] < 0.024 μM ; [Hg²⁺] < 0.00013 μM),¹ they are not expected to be the cause of significant interference in the routine measurement of arsenic concentrations in surface water samples.^{40,41} The GSH capping ligand thus could provide the first building block towards a Cys-rich metal binding

environment, and since it is confined at the QD surface, this may offer sufficient spatial flexibility to provide additional multidentate complexation of the metal cation. Nevertheless, As^{3+} ions typically form a very stable three-coordinate trigonal-pyramidal complex by binding to three GSH thiol groups in a highly crosslinked phytochelatin-(PC_n)-like molecules.⁴² Furthermore, Xia *et al.*⁴³ have also shown highly selective coordination of As^{3+} to the three cysteine residues of PC_3 molecules in the presence of Cu^{2+} and Zn^{2+} ions. Therefore, increasing GSH on the surface through peptide bonding may offer the possibility of enhancing sensitivity.

Figure 2a shows the relative intensity of the QD_{600} -GSH fluorescence following increasing EDC/NHS coupled GSH. The EDC/NHS activation potentially also enables cross-linking of the QDs, so that at low concentrations of GSH in solution, aggregation was observed, suggesting that the preferred reaction results in QD cross-linking and QD-QD quenching. At higher concentrations of GSH, fluorescent intensity is partially recovered. Increase in GSH on the surface Yang *et al.*⁴⁴ noted the molar ratio of free GSH to QDs may be in the range of about 100 to 5000 to prevent aggregation. In this instance using 13.2 nmol QD_{600} no aggregation was observed for molar ratios between 1000:1 and 10000:1. As the greatest fluorescence intensity was obtained using 80mg/mL GSH, this amount of GSH was used during EDC/NHS modification of the QD that were used for arsenic detection. The data in Figure 2 also suggest a red shift and broadening of the emission. While the primary determinant of emission wavelength is the QD core size, it is also influenced by surface states. QDs with a large number of surface states can show weak broad emissions⁴⁵ that are often red shifted, consistent with the data recorded here. Clearly, while these QD_{630} -GSH-GSH QDs are brighter than the MUA or MPA capped QDs, the fluorescence is less intense than the QD_{630} -GSH, so that sensitivity and selectivity may require compromise.

Further insight into the GSH derivatisation can be gained using the Amersham Cy5 maleimide mono-reactive dye, which has a high affinity for the sulfhydryl groups on the GSH, a FRET pair can be established between the QD and the dye (figure 2b). As can be seen in the figure, the maximum peak intensity of the QDs decreases, while that of the dye increases for the QD-GSH-GSH-dye conjugates (figure 2c), consistent with a FRET pair. However, interestingly, the decrease in the QD fluorescence is focused on the longer wavelength emission so the residual maximum shows a blue shift. This probably suggests that the NHS/EDC coupling of GSH has resulted in a heterogeneous population, and caused additional surface states resulting from the peptide bonded GSH. Since the dye binds with the sulfhydryl groups on GSH, it will selectively react with QDs that present the GSH for reaction.

Metal ion susceptibility

Although Callan & Mulrooney⁴⁶ and Ke *et al.*⁴⁷ reported that introduction of a ZnS shell reduced the effect of metal ions on

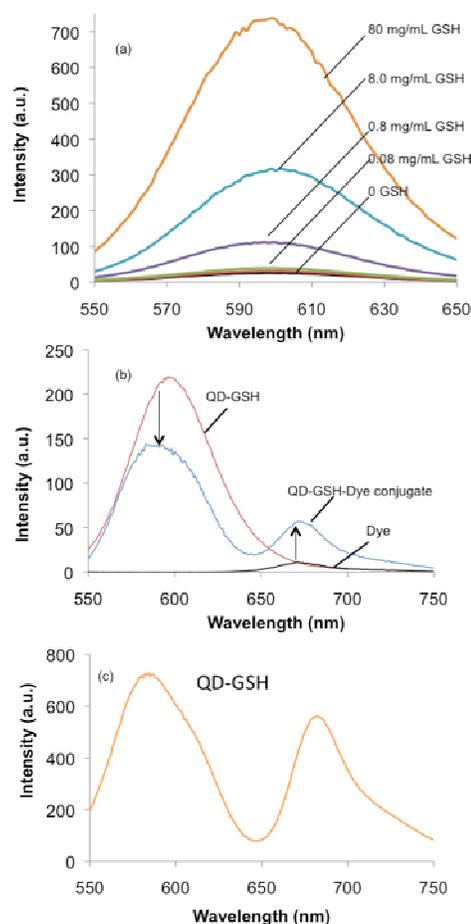


Figure 2 (a) Emission spectra of CdTe QD-GSH before and after the covalent EDC/NHS reaction with GSH in phosphate buffer (pH 7.2) (b) FRET emission spectrum of CdTe QD-GSH-Dye (Amersham Cy5 maleimide mono-reactive dye) conjugate compared to emission spectra of dye alone (and QDs alone) (c) Emission spectra of the QD-GSH-GSH-dye conjugates for 80 mg/mL GSH (added during EDC/NHS conjugation. (Excitation at 400 nm; Excitation slit size: 20 nm; Emission slit size: 20 nm; PMT: 600 V)

the fluorescence intensity of QDs, since it serves as a protective layer to the core, the ZnS layer still offers significant activity towards metal cations. It is evident from Table 2 that the CdSe/ZnS core-shell QDs were mainly more susceptible to quenching by metal ions than the CdTe QDs (except in the case of Cu(II)). While a robust shell is likely to increase quantum yield, and reduce environmental effects (if it is not chemically reactive) the carboxylate and GSH capping efficiency is also likely to play a part in the stability and reactivity, particularly as a metal complexing ligand. Furthermore, for the QDs compared here, the larger particle size of the CdSe/ZnS QDs (375 000 Da) compared to the CdTe QDs (76 000 Da), suggests a greater number of surface binding sites for interaction with cations in solution.

In terms of As^{3+} measurement, from Table 2 interfering ions, Cu^{2+} , Fe^{3+} and Ni^{3+} are of primary concern because water sources, especially those affected by acid mine drainage, may contain high concentrations, and these ions are well-known quenchers of fluorescence.⁴⁸

| Metal | Average Concentration Reported (mg/L) | WHO Drinking Water Limit (mg/L (μM)) | Concentration (μM) | | | | | | |
|---------|---------------------------------------|--------------------------------------|--------------------|-------------|-----------------|---------------------|-----------------|-----------------|---------------------|
| | | | CdTe QD | QD CdTe-GSH | CdSe/ZnS QD-GSH | CdSe/ZnS QD-GSH-GSH | CdSe/ZnS QD-MUA | CdSe/ZnS QD-MPA | CdSe/ZnS QD-MPA/MUA |
| Zn(II) | 0.01 - 5 | 0.01 (0.15) | 1000 | 100 | 2.5 | 1 | 2.5 | 10 | 10 |
| Co(II) | 0 - 0.11 | 0.004 (0.07) | 25 | 500 | 50 | 100 | 100 | 100 | 2.5 |
| Al(III) | 0.001 - 1 | 0.9 (33.36) | 1000 | 100 | 500 | 500 | 1000 | 1 | 50 |
| Fe(III) | 0.5 - 50 | 2 (35.81) | 250 | 1000 | 250 | 500 | 0.5 | 0.5 | 0.5 |
| Mn(II) | 0.001 - 10 | 0.4 (7.28) | 1000 | 500 | 250 | 500 | 50 | 25 | 50 |
| Ni(II) | 0.02 - 1 | 0.07 (1.19) | 50 | 250 | 100 | 25 | 100 | 0.5 | 0.5 |
| Mg(II) | 1 - 130 | 50 (2.06) | 250 | 250 | 1000 | 1000 | 1000 | 100 | 1000 |
| Cu(II) | 0.005 - 30 | 1.5 (22.9) | 0.5 | 0.5 | 0.5 | 1 | 0.5 | 0.5 | 0.5 |

Table 2 Highest cation concentration tested without effect on QD luminescence (phosphate buffer pH 7.2) for CdSe/ZnS QD₆₃₀-GSH, QD₆₃₀-GSH-GSH, QD₆₃₀-MUA, QD₆₃₀-MPA, QD₆₃₀-MPA/MUA and CdTe QD₆₀₀ conjugates. WHO metal cation drinking water guidelines and top limit of average worldwide cation concentration ranges in water sources are also listed (WHO, 2008)

Since transition metal ions, such as Fe³⁺ and Ni²⁺, can engage in d-d energy level transitions, the quenching observed is expected to originate from electron transfer between metal ions and QDs, resulting in nonradiative recombination.⁴⁹ As seen in Table 2, Cu²⁺ is the most significant quencher at WHO drinking water concentrations, but Cu²⁺ sensitivity is reduced for QD₆₃₀-GSH-GSH, compared with the other QD₆₃₀S.

The severe quenching effect of Cu²⁺ on QDs is expected and has been widely reported.^{46,50-54} Cu²⁺ ions are effective electron scavengers, preventing QD electron-hole recombination resulting in fluorescence quenching. Furthermore, at high enough concentrations, previous reports have suggested that Cu²⁺ can penetrate the capping layer and diffuse into the CdSe or CdTe core,^{23,55} where, due to the low solubility of CuSe and CuTe, the Cd²⁺ in the QD lattice could be displaced by Cu²⁺. This is consistent with the observation for the CdTe QDs at Cu²⁺ > 5 μM, where QD emission was quenched completely and irreversibly. CdSe/ZnS QD₆₃₀ is also susceptible to quenching at around the same concentration, and the response curve overlays that for CdTe, except at higher concentrations where ~20% fluorescence remains. Furthermore, with the additional GSH-GSH modification, overall Cu²⁺ susceptibility was reduced. This is consistent with the copper becoming complexed by the GSH layer, instead of penetrating into the core as described above. In an electrochemical analogue using GSH modified Au-nanoparticles, Guo *et al.*⁵⁶ have proposed that the position of the LUMO for a metal-GSH dimer is dependent on the metal, so that electron scavenging of the QD by the metal will be influenced by the relative positions of the LUMO of the GSH-metal complex and conduction band of the QD, as well as the GSH-metal affinity kinetics. This will depend on both the metal and the GSH binding. In Singh's review⁴⁰ of glutathione-metal binding it is reported that both the thiol group of the cysteine residue and the amino group of the glutamic acid are involved in complexation of Cu²⁺, forming octahedral complexes, with higher involvement of the amino group indicated with increase in Cu²⁺ concentration. This Cu²⁺-GSH complex defines the LUMO for scavenging of electrons

from the QD. This may be consistent with the contrasting findings in the literature, that L-cysteine (rather than glutathione) capped QDs were not susceptible to Cu²⁺ quenching,⁴⁹ because the metal complex LUMO will not facilitate electron scavenging. Singh⁴⁰ points out that Cu²⁺ oxidation occurs rapidly with GSH, producing GSSG, but the reoxidation of GSSG is slow, so that reversibility is compromised.

In contrast, Fe³⁺ binding to GSH is proposed to involve thiolate complexation, resulting in rapid reduction to Fe²⁺ and dimerisation to GSSG. This may be followed by high spin Fe²⁺ binding, involving 5 and 6 coordinate complexes that will catalyse thiol oxidation in the presence of oxygen. The lower sensitivity towards Fe³⁺ observed, suggests that this metal complexation is either less favoured kinetically, or produces a LUMO that is less efficient as an electron scavenger. Overall, the likely involvement of the different metal-GSH complex LUMOs in the QD quenching mechanism will impact the degree of quenching and, dependent on the GSH structure on the QD surface, also drive the metal ion selectivity since different coordination geometry and chemistry is required for each ion.

In the case of Ni²⁺, Singh concluded that complexation involved the amine and carboxyl group of the glutamic acid branch. The decrease in Ni²⁺ quenching observed for QD₆₃₀-GSH-GSH (Table 2) compared to QD₆₃₀-GSH and, for QD CdTe-GSH compared with QD CdTe, may also be consistent with a raised LUMO for GSH-Ni²⁺ complexes with respect to the QD conduction band, thereby modulating charge separation. From Table 2 it is evident that at 50 μM, Ni²⁺ interference of As³⁺ could occur. However, as Ni²⁺ is not present in surface waters at concentrations greater than 17 μM on average (Table 2), interference of Ni²⁺ on QD fluorescence is expected to be negligible.

Table 2 also identifies a strong susceptibility to Zn²⁺. In this instance the result is a fluorescence enhancement. It has been well reported that GSH has a high selectivity towards complex formation with Zn²⁺.⁴⁰ According to the idea of LUMO versus conduction band positioning, this might suggest that the LUMO

energy level is above the conduction band, providing a protective rather than quenching role. However, an alternative interpretation of this enhancement has been proposed by Chen and Rosenzweig, who found that Zn^{2+} could form bridging complexes between adjacent cysteine modified QDs that form small clusters (about 3-4QDs). This resulted in an observed fluorescence increase; these explanations are not necessarily contradictory.

Thus, examination of the principles of metal-ion complex interactions with QDs, can rationalise different reports of metal ion selectivity and quenching and shows that GSH capping has

capacity to modulate metal ion selectivity and sensitivity depending on its surface presentation. Based on these foundations, the response of As^{3+} to GSH and GSH-GSH modified QDs was examined.

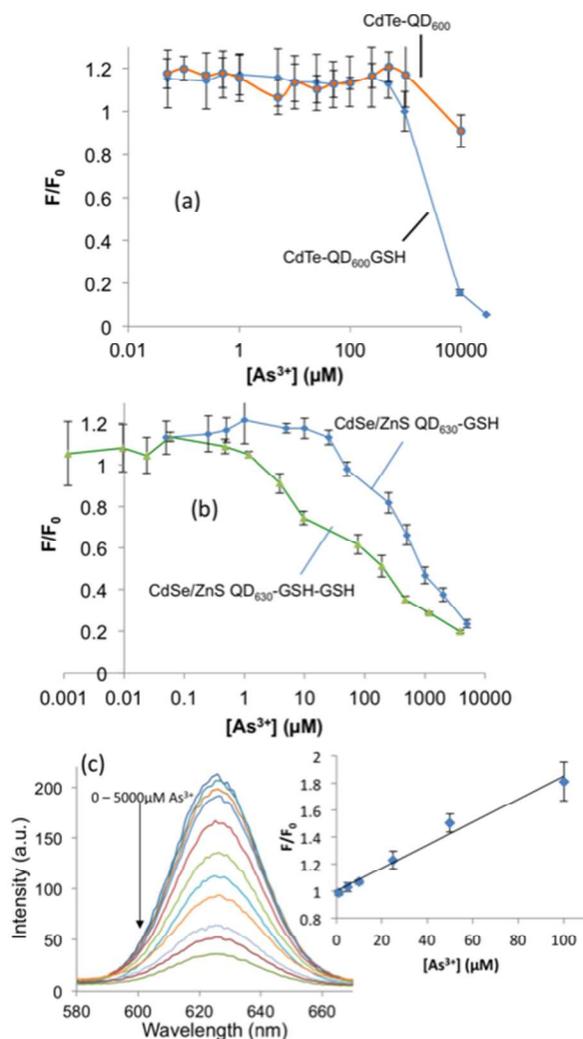


Figure 3: The normalised fluorescence maxima response of (a) carboxylated CdTe QDs compared with functionalised CdTe QD₆₀₀-GSH to different concentrations of As^{3+} ; (b) CdSe/ZnS QD₆₃₀-GSH and functionalised CdSe/ZnS QD₆₃₀-GSH-GSH to different concentrations of As^{3+} . (c) Fluorescence emission spectra for CdSe/ZnS QD₆₃₀-GSH-GSH showing progressive quenching for concentrations of As^{3+} from 0 – 5000 μM As^{3+} . Insert: Stern Volmer plot for the data. (phosphate buffer pH 7.2, Excitation at 360 nm; Excitation slit size: 20 nm; Emission slit size: 20 nm; PMT: 800 V).

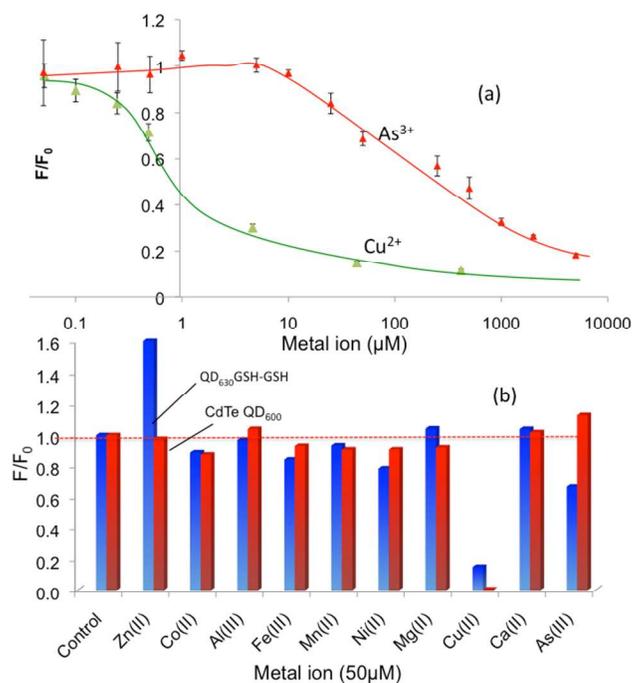


Figure 4: (a) The normalised fluorescence maxima of CdSe/ZnS QD₆₃₀-GSH-GSH: comparison of the response by 90nM CdSe/ZnS QD₆₃₀-GSH-GSH to As^{3+} and Cu^{2+} (b) Effect on the normalised fluorescence for a concentration of 50 μM of different ions for 90 nM CdSe/ZnS QD₆₃₀-GSH-GSH and 440 nM CdTe QD₆₀₀ (n = 3) (Excitation at 360 nm; Excitation slit size: 20 nm; Emission slit size: 20 nm; PMT: 800 V

Arsenic response

Figure 3a compares the response for CdTe QDs with and without GSH and demonstrates that only slight As^{3+} sensitivity can be seen in the functionalised QD₆₀₀-GSH. The limit of detection (LOD) ($S/N > 3$) lies around $F/F_0 = 0.78-0.80$, at a concentration of circa 1mM for the GSH modified QD and >10mM for the unmodified CdTe QD. It is evident that the CdTe core QDs are not responsive to arsenic, although GSH functionalised CdTe QDs showed some improvement in response >1mM. In terms of surface water concentrations, this renders the CdTe QD unsuitable for As^{3+} detection.

In contrast, figure 3b compares the As^{3+} response for CdSe/ZnS QD₆₃₀-GSH and the functionalised QD₆₃₀-GSH-GSH. Although a clear linear quenching trend was visible, starting at an As^{3+} concentration of 1 μM , the limit of detection lies around $F/F_0 = 0.88-0.87$ and a concentration of ~150 μM in the case of ~90 nM CdSe/ZnS QD₆₃₀-GSH and ~10 μM in the presence of ~90 nM CdSe/ZnS QD₆₃₀-GSH-GSH. This suggests an order of magnitude improvement compared with current field-testing systems. The GSH-GSH surface modification was tested for concentrations >80 $\mu g/mL$ and optimised at 80 mg/mL solution concentration during the surface reaction with the linear response to As^{3+} improving from 250 μM to 10 μM in this range.

In mammalian metabolism, As^{3+} generally has a high propensity to react with cysteine thiols, particularly glutathione as the most abundant endogenous thiol, and is known to form a tridentate complex with As^{3+} . However, unlike the endogenous glutathione, the GSH's cysteine groups are probably bonded to the QD ZnS surface and potentially not available for As^{3+} complexation. This leaves the As^{3+} in competition at the ZnS QD surface. This is in contrast with the QD_{630} -GSH-GSH, where the outer coating of GSH is attached via a peptide bond, leaving thiol and amide chemistry for metal complexation. Furthermore the thicker GSH layer may achieve better phytochelatin (PC) mimicry for As^{3+} binding.⁵⁷ Consequently, there are GSH sulphhydryl groups on the surface of the QD which are expected to have a high affinity towards As^{3+} binding.^{6,43} According to Shen *et al.*⁶, trivalent arsenic binds to three or four cysteine residues far more stably and selectively than one or two residues. There are various possible mechanisms that may be connected with the quenching. Unlike Cu^{2+} , As^{3+} is not expected to be a good oxidant of GSH. Indeed, As^{3+} affinity is reversible, whereas with Cu^{2+} it is mostly irreversible. This suggests that there is not a resultant redox electron exchange with the QD. However, the As^{3+} complex formed with the GSH thiol and amide may place the LUMO of a GSH(n)- As^{3+} complex below the QD conduction band to allow electron transfer and thus quenching. In the design of an affinity ligand modified QD for metal-ion detection, both the kinetics of the affinity ligand and the LUMO of the resulting complex need to be taken into account. In this context, other peptide sequences might reveal greater sensitivity to other ions in figure 4b, even though they do not show a response with GSH surface modification.

Depending on the balance between metal-GSH affinity kinetics and the quenching mechanism, the behaviour will be different. The quenching of the QD excited state may be limited by collision with a quencher (the dynamic mechanism) or by a static mechanism arising from charge transfer (or electron tunnelling) or the overlap of molecular orbitals. Metal ion quenching typically involves electron transfer to the metal/metal complex, which might be expected to result in a non-linear Stern Volmer plot, as seen with many other semiconductor nanoparticles²⁸⁻³¹. However, for the "metalloid", arsenic, we consider this unlikely in this instance because of the overlay of GSH ligand binding kinetics. Indeed, a characteristic Stern-Volmer plot emerges (figure 3c). The 'dynamic' behaviour seen here may be indicative of the equilibrium binding between the GSH and As^{3+} limiting the response rather than static electron transfer between QD and metal complex. The sensitivity of QD_{630} -GSH-GSH to As^{3+} is indicated by a Stern-Volmer coefficient of $K = 8.5 \times 10^3 \text{ M}^{-1}$.

Capacity for Arsenic detection in water

LABORATORY DETECTION

Figure 4a compares the response of the QD_{630} -GSH-GSH for Cu^{2+} and As^{3+} . As expected this highlights the issue of copper interference. Despite the generally good selectivity toward As^{3+} with this GSH layer, zinc and copper are also clear responders

(figure 4b). In nature, binding of copper and arsenic regularly occur at similar sites, so that total selectivity is unlikely. In the case of zinc, the water levels anticipated do not present a problem. Copper is an essential dietary mineral, with recommended intake 2-3mg/day and maximum 10mg/day, so it is not a particularly critical problem in drinking water. According to WHO, EPA and the Bangladesh standard, the maximum permissible limit for drinking water is 1500, 1300 and 1000 $\mu\text{g/L}$, respectively. In western drinking water, the prime source of copper arises from copper pipes, but various natural sources in different parts of the world do have measurable copper levels. For example, Balu River Bangladesh 3.81 - 19.99 $\mu\text{g/L}$; Mississippi River, 21.5 $\mu\text{g/L}$; Yangtze River, 7.31 $\mu\text{g/L}$; Indus River, Pakistan >10 $\mu\text{g/L}$ ⁵⁸; all are much lower than the permissible limit. Thus, assuming a typical maximum level in the region of $\sim 20 \mu\text{g/L}$ (0.3 μM), then from figure 4, this corresponds to up to 20% quenching by Cu^{2+} .

Figure 4b also shows the same array of metal ion responses for the CdTe QDs, where excellent selectivity for copper is demonstrated. Thus, used in conjunction with the QD_{630} -GSH-GSH, the CdTe QD provides an option for a copper "baseline", as a way to validate the integrity of the arsenic assay in the presence of copper cross reactivity, so long as the arsenic determination remains sufficiently sensitive to achieve the WHO recommended levels. This is explored further in a mobile phone assay system.

MOBILE ARSENIC

As discussed earlier, the majority of commercially available field sensing kits for arsenic are equipped with a colorimetric readout scheme⁴. This makes the methodology for these, as well as the QD method described here, ideally suited for smart phone camera technology. Although the camera sensor only gives RGB component readings for each pixel rather than a detailed spectral readout, spatial encoding can instead be used for wavelength spectroscopy.

Figure 5b shows the transmittance profile for the red, blue green and infra-red filters in the Nokia N85 phone camera. The RGB output of the phone cameras can be predicted and calibrated using a spectrum recorded from incident light and the data in figure 5b. Figure 5a shows the spectrally predicted RGB signals compared with the rawcam image data taken from the N85 camera. A reasonable match can be seen, at least at these wavelengths.

From knowledge of the camera filter properties, we can design the QD wavelength to give output in a particular RGB channel. Ideally, as with a normal spectrophotometric assay, maximum wavelength separation is desirable. Figure 5c shows a mixture of the QD_{630} -GSH-GSH and CdTe QD_{560} , where good wavelength separation is indicated. Reference to figure 5b, suggests that overall, this should lead to a CdTe QD_{560} dominated green channel, a QD_{630} -GSH-GSH dominated red channel and a blue referencing channel. Unfortunately, the CdTe QD_{560} were unstable and the signal intensity diminished rapidly with time. In contrast, both CdTe QD_{600} and QD_{630} -GSH-GSH gave viable output but without anticipated good

spectral separation. Nevertheless, figure 5e shows the QD₆₃₀-

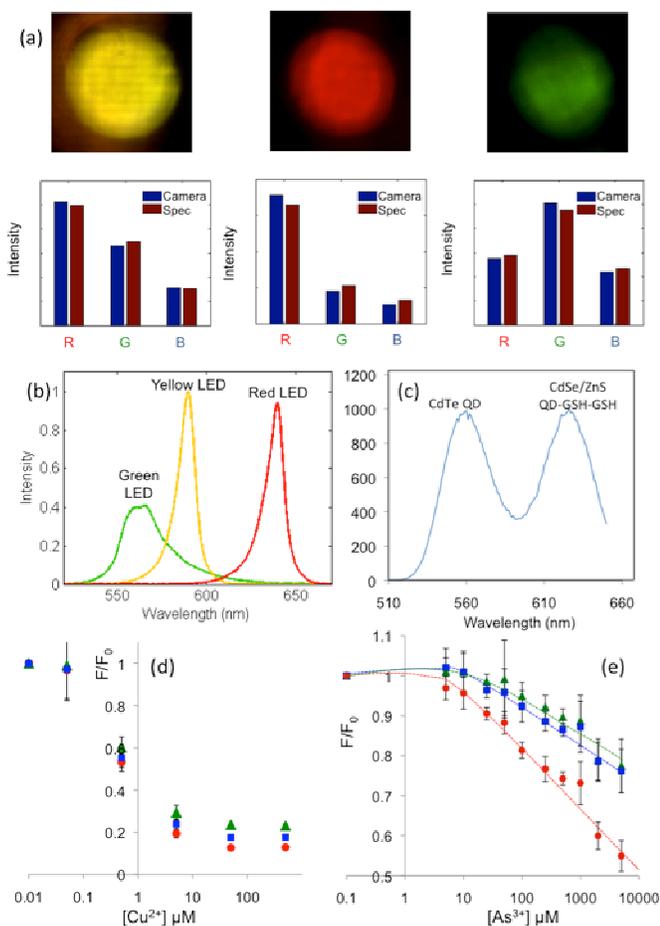


Figure 5: Data from the Nokia N85 mobile phone camera. (a) Camera output for different spectral images and the RGB raw output, together with predicted output based on filter transmission data; (b) Spectra for the green, red and yellow LEDs. (c) Emission spectrum for mixture of CdTe QD₆₀₀ and QD₆₃₀-GSH-GSH showing spectral separation. (d) RGB response from CdTe QD₆₀₀ for [Cu²⁺]. (e) RGB response from QD₆₃₀-GSH-GSH for [As³⁺].

GSH-GSH RGB response to arsenic, is consistent with the data collected with a fluorometer (figures 3 and 4) and broadly confirms the design expectation on RGB output, from examination of the output from the yellow and red LEDs (figure 5b). It can also be seen that [As³⁺] can be measured either directly from the absolute output from the red channel or as the difference between the red and green or blue channels. Of special note, compared with the fluorometer data, the lowest detection level is $\leq 5 \mu\text{M}$, a slight improvement compared with the lab-based fluorometer method.

The detection integrity in the presence of copper interference can be checked from the CdTe QD₆₀₀ (figure 5d) data, which also mimics the fluorometer data in figure 4d. These data show almost the same output from all RGB channels. This was predicted for red and green, but the coincident output from the blue channel takes into account of the additional longpass filter used, and suggests a higher relative transmittance in the blue channel, for this camera at this wavelength. However, these

data also demonstrate the significant potential to allow QDs, even with closely aligned emission wavelength to be separated according to their relative and absolute RGB output.

From the experimental mobile phone data obtained in figure 5c and 5d arsenic measurement can be computed against a copper corrected baseline (figure 6). The figure predicts that using the CdTe QD₆₀₀ / QD₆₃₀-GSH-GSH combination and a mobile phone camera, arsenic can be measured in the presence of copper, even at a typical maximum river water Cu²⁺ level in the region of $\sim 20 \mu\text{g/L}$ ($0.3 \mu\text{M}$).

Conclusion

According to British Geological Survey⁵⁹, 35 million Bangladeshi people use drinking water with arsenic concentration exceeding the national standard ($50 \mu\text{g/L}$) and 57 million people are exposed to concentrations exceeding the World Health Organization (WHO) guideline value of $10 \mu\text{g/L}$ ⁶⁰. Although arsenic estimation in the laboratory achieves levels of detection capable of measurement of arsenic at $10 \mu\text{g/L}$, on site portable measurement is typically an order of magnitude less sensitive, or requires preconcentration. Furthermore, potentially portable assay methods often show cross reactivity, particularly with copper, which is also an essential dietary element. Some of these methods have adopted cysteine-rich ligands for As³⁺ binding, but have found that the same ligands can bind other metals and thus reduce the selectivity. Similar, effects are found in nature where subtle spatial presentation of the affinity groups can play a significant role in the outcome. For example, As³⁺ ions typically form a very stable three-coordinate trigonal-pyramidal complex by binding to three GSH thiol groups in a highly crosslinked phytochelatin-(PC_n)-like molecules.⁴² However, GSH can bind

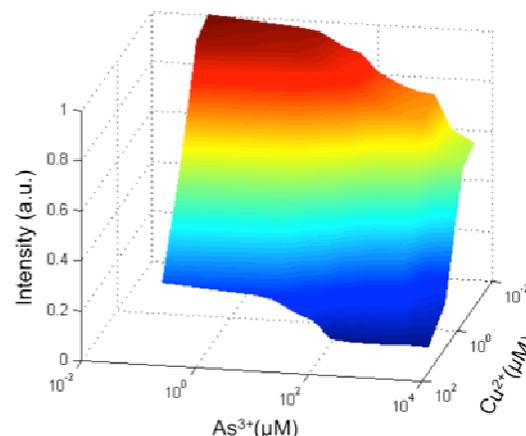


Figure 6: QD₆₃₀-GSH-GSH response to [As³⁺] and [Cu²⁺]. Data from CdTe QD₆₀₀ and QD₆₃₀-GSH-GSH are used to compute the QD₆₃₀-GSH-GSH response to [As³⁺] in the presence of [Cu²⁺]. The Cu²⁺-selective response from CdTe QD₆₀₀ (R,G or B channel) provides the input to calculate the red channel response to Cu²⁺ on QD₆₃₀-GSH-GSH. This provides the baseline for the [As³⁺] measurement.

metals through the amine and carboxyl groups as well as the

thiol and thus the presentation of these groups can alter the affinity for a particular method. By paying attention to the presentation of free sulfhydryl on the surface of a CdSe/ZnS QD, this work has demonstrated better As³⁺ selectivity and sensitivity than reported previously. Furthermore, CdTe QDs could be prepared which showed no As³⁺ sensitivity, but were selective towards Cu²⁺, thereby offering the possibility of a baseline measurement for copper. Using the QDs as the analytical element had the benefit of a narrow emission fluorescence, which could be resolved using a reasonably low specification mobile phone camera, by direct processing of the raw pixel data. Surprisingly, by using separate processing for R, G and B channels, even QDs as close in emission maximum as 630 and 600, showed distinct intensity data. It could be demonstrated that taking copper specific data from the CdTe QDs produced a background 'copper-signal' for the GSH-GSH modified CdSe/ZnS QDs, that, in principle would enable arsenic measurement to be made on a mobile phone, even in the presence of copper at the highest recorded levels in some large rivers around the world. With the significant increase in resolution now available in smart phone cameras, it is possible to consider that such portable analysis is now feasible.

Acknowledgements

Nokia Research Centre, Cambridge, UK provided the 2008 Nokia N85 smartphone as a development handset.

Notes and references

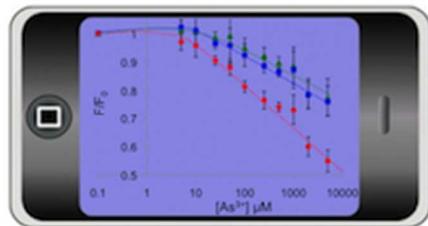
^a Institute of Biotechnology, Department of Chemical Engineering and Biotechnology, University of Cambridge, Tennis Court Road, Cambridge, CB2 1QT, UK.

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GSH modified QDs are used for selective As^{3+} measurement for river waters in a mobile phone camera-linked unit.

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