

# Analyst

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1  
2  
3  
4 **Low voltage electrically stimulated lab-on-a-chip device followed by Red-**  
5  
6  
7 **Green-Blue analysis: A simple and efficient design for complicated matrices**  
8  
9

10  
11  
12  
13  
14 Shahram Seidi<sup>a,\*</sup>, Maryam Rezazadeh<sup>b</sup>, Yadollah Yamini<sup>b</sup>, Niki Zamani<sup>c</sup>, Sara Esmaili<sup>c</sup>  
15

16  
17 <sup>a</sup>*Department of Analytical Chemistry, Faculty of Chemistry, K.N. Toosi University of Technology, Tehran, Iran*

18 <sup>b</sup>*Department of Chemistry, Tarbiat Modares University, Tehran, Iran*

19 <sup>c</sup>*Farzanegan 1 Educational Center, National Organization for Development of Exceptional Talents, Tehran, Iran*  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54

---

55  
56 \*Corresponding author at: K.N. Toosi University of Technology, Faculty of Chemistry, Department of Analytical  
57 Chemistry, P.O. Box 16315-1618, Tehran, Iran. Tel.: +98(21)23064228; Fax: +98(21)22853650.  
58 E-mail address: [s.seidi@kntu.ac.ir](mailto:s.seidi@kntu.ac.ir) (S. Seidi).  
59  
60

## Abstract

In the present work, a simple and portable analysis device was designed for the first time for determination of lead ion as the model analyte. The basis of lead analysis is its extraction and preconcentration in an acceptor droplet via application of electrical field. The acceptor droplet is a KI solution and therefore, formation of yellow precipitation of  $\text{PbI}_2$  is the sign of presence of lead ion in the solution. Following that, digital picture of the final acceptor droplet was analyzed by investigating its Red-Green-Blue (RGB) components. The results show that RGB intensities of the acceptor phase is proportionate to lead concentration in the sample solution. Also, a 9.0-V battery was used to apply the electrical field and other effective parameters such as the type of organic liquid membrane, pH of the sample solution, and the extraction time were considered to reach the optimal conditions. The model analyte was determined by extracting from 100  $\mu\text{L}$  sample solution across a thin layer of 1-octanol, immobilized in the pores of a polypropylene membrane sheet, and into the acceptor droplet via applying a 9.0-V electrical potential for 20 min. The device is capable of determining the lead ion down to 20.0  $\text{ng mL}^{-1}$  with admissible repeatability and reproducibility (intra- and inter-assay precision ranged between 3.8-7.0% and 9.8-11.9%, respectively). Also, calculated Error% for the model analyte in the range of -8.5 to +4.5 depicts that the chip offers acceptable accuracy for analysis of the lead ion. The linearity was studied in the range of 50.0-1500  $\text{ng mL}^{-1}$  and the correlation coefficient was 0.9994. Ultimately, the device designed was employed for analysis of lead in real samples.

*Keywords:* Electromembrane extraction; RGB analysis; Lab-on-a-chip; Lead.

## 1. Introduction

The *in vitro* analysis of different compounds is an attractive topic in analytical chemistry. In recent years, new designs for miniaturized devices, named lab-on-a-chip (LOC) systems, were performed to attain the goal<sup>1</sup>. The main advantages of LOC systems are small sample requirements, short analysis time, portability, low cost, and low consumption of power. Thus, downsized installations or LOC systems were developed to perform laboratory analyses of chemistry, biology, and medicine over a miniaturized chip<sup>2-6</sup>. Also, electrophoretic techniques are perfectly suited for downscaling due to their smaller separation lengths, following by increase in electrical fields.

On the other hand, reduction of matrix effect, removal of interferences, and analyte preconcentration are important and necessary steps prior to analysis of target compound in a complicated medium. To this end, some extraction or sample preparation steps are necessary. Sample preparation is an important issue in analytical chemistry, and is often a bottleneck in chemical analysis. As a consequence, during the last decade new modern sample preparation methods were developed that can generally be classified as liquid based, solid based and membrane based techniques.

Among the present sample preparation methods, hollow fiber based liquid phase microextraction (HF-LPME) is known by its high sample cleanup ability, which makes it a suitable technique for analysis of dirty samples<sup>7, 8</sup>. However, relatively long process time due to its passive diffusion mechanism is the main drawback of this microextraction method. Therefore, electrical driving force was employed in HF-LPME to increase the efficiency and to reduce the extraction time for ionizable compounds<sup>9</sup>. This electrical stimulated technique was called electromembrane extraction (EME) by Pedersen-Bjergaard et al. and is based on the fact that

1  
2  
3 ionizable species can migrate in an electrical field. Thus, two electrodes were utilized in donor  
4 and acceptor phases, respectively to lead the target analytes from sample solution across the  
5 organic liquid membrane (immobilized in the HF wall pores) and into the final acceptor solution,  
6 which was placed in the lumen of HF<sup>9</sup>. EME as a new and powerful method was developed  
7 rapidly during the past years<sup>10-23</sup>. One of the most interesting applications of EME is designing  
8 the LOC systems<sup>15-17</sup>. This new concept of EME technique was first introduced in 2009<sup>15</sup> and  
9 improved during the past years<sup>16, 17</sup>. However, except for an attempt to electromembrane chip  
10 coupling to a detector (UV and mass spectrometer) for online analysis<sup>17</sup>, there is no report for  
11 designing a chip device with both extraction and detection abilities.  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24

25 In the present work, an electrical field-induced LOC design followed by RGB analysis is  
26 introduced for the first time to develop a portable device for analysis of lead ion in aqueous  
27 samples. Heavy metals, especially Pb, could cause serious threat to the environment and human  
28 health. Since the toxic metals tend to concentrate in all environmental matrices; they belong to  
29 the most deleterious pollutant group. Therefore, great attention must be focused on their  
30 concentration level. Until now, different microextraction techniques have been reported for  
31 determination of lead from various matrices<sup>24-29</sup>.  
32  
33  
34  
35  
36  
37  
38  
39  
40

41 For RGB analysis, an optical image of the acceptor droplet is decomposed into three  
42 components (red, green, and blue) using image processing software and is utilized as the  
43 concentration-dependent signal<sup>30</sup>. To this end, target ions were first extracted from few  
44 microliters of the sample solution through the organic liquid membrane, which was sustained in  
45 the pores of a sheet membrane into a droplet of the acceptor phase containing excess amounts of  
46 KI as the indicator. As the lead ions transfer into the KI solution, a yellow precipitation forms;  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 indicating the presence of the target analyte. Following that, the acceptor solution was online  
4  
5 analyzed regarding the intensities of RGB components.  
6  
7

8         Herein, a simple and fast analysis technique based on the RGB image processing was  
9  
10 employed for quantification of lead iodide in the acceptor solution. Finally, the designed LOC  
11  
12 system was used for determination and quantification of analyte of interest in the samples of  
13  
14 wastewater and street plant leaves.  
15  
16

## 17 18 **2. Experimental section** 19

### 20 21 *2.1. Apparatus* 22 23

24         The equipment used for the extraction procedure is shown in Fig. 1. A small hole with a  
25  
26 volume of 100  $\mu\text{L}$  was formed in a Plexiglass wafer and used as the sample compartment.  
27  
28 Afterwards, the hole was coated utilizing a  $1 \times 5$ -cm piece of aluminum foil with 15  $\mu\text{m}$  thickness  
29  
30 (Merck, Darmstadt, Germany) providing the electrical field. The platinum electrode used in this  
31  
32 work, with the diameters of 0.25 mm, was obtained from Pars Pelatine (Tehran, Iran). The  
33  
34 electrode and the aluminum foil were coupled to a common 9.0-V battery (GIL SUN, China) as  
35  
36 the power supply. A 40-kHz and 0.138-kW (Tecno-Gaz SpA, Italy) ultrasonic water bath with  
37  
38 temperature control and a Sepand Teb Azma centrifuge (Tehran, Iran) were used for real sample  
39  
40 pretreatment steps.  
41  
42  
43  
44

45         An atomic absorption spectrometer (GBC 932 plus, Australia) equipped with a deuterium  
46  
47 arc background corrector was used. A Pb hollow-cathode lamp was utilized as the radiation  
48  
49 source at 217.0 nm. The optimum operating parameters were adjusted according to the  
50  
51 manufacturer's recommendations.  
52  
53  
54  
55  
56  
57  
58  
59  
60

## 2.2. Chemicals and materials

Pb (NO<sub>3</sub>)<sub>2</sub>, KI, 1-heptanol, 1-octanol, and 1-decanol were obtained from Merck (Darmstadt, Germany). All the chemicals used were of analytical reagent grades. The Accurel 2E HF (R/P) polypropylene membrane sheet (157 μm thickness, 0.2 μm pore size) was supplied by Membrana (Wuppertal, Germany). Ultrapure water was obtained from a Young Lin 370 series aquaMAX purification instrument (Kyounggi-do, Korea).

## 2.3. Standard solutions and real samples

A stock solution containing 1 mg mL<sup>-1</sup> of lead ion was prepared in pure water and stored at 4 °C protected from light. Working standard solutions were prepared by dilution of the stock solution in pure water.

Wastewater samples were filtered through 0.45-μm pore size cellulose acetate membrane (Millipore, Billerica, MA, USA) filters prior to extraction.

Also, plant leaves were analyzed as the real sample. For analysis of the target heavy metal in the plant tissue, leaves were collected from city streets. The whole plant leaves were completely blended. Then, 10 mL of a 100 mM HCl solution was added to 50 g of each blended sample and it was immersed into the ultrasonic water bath. After 2 min of sonication (at 25±3 °C), the sample was centrifuged at 2500 rpm for 10 min to separate the phases. The final liquid phase was diluted (1:9) with pure water and after pH adjustment (the pH was adjusted at 6.5 via addition of proper amounts of sodium hydroxide solution), it was transferred into the EME sample hole for analysis.

## 2.4. RGB analysis

Two detection modes (offline and online) were applied for analysis of the acceptor droplet. In offline mode, the droplet acceptor was collected after completion of the extraction

1  
2  
3 process and transferred into a scanning vial. The vial was scanned by a flatbed scanner (HP  
4 Photosmart C4283, Malaysia) and the variable factor of each component (red, green, and blue)  
5 was determined using Photoshop<sup>®</sup> software (Adobe systems, USA) afterwards. The final signal  
6 was calculated by additive subtraction of RGB intensities from white.  
7  
8  
9  
10  
11

12 Also, the online mode was examined to design a portable device for lead analysis  
13 utilizing a smartphone (iPhone, USA) as the detector. In this mode, a smartphone was placed in  
14 10 cm at the top of the LOC system. When the extraction was completed, a digital photo was  
15 obtained from the phone camera and analyzed using the iDropper tool, which is available for iOS  
16 smartphones. Again, the concentration-related signal was calculated by subtraction of RGB  
17 intensities from white.  
18  
19  
20  
21  
22  
23  
24  
25

### 26 2.5. Electromembrane chip procedure

27  
28  
29  
30 One hundred microliters of the sample solution containing the model analyte in pure water  
31 was transferred into the sample vial, which was first coated by the aluminum foil. To impregnate  
32 the organic liquid membrane in the pores of the polypropylene sheet, a 1× 1-cm piece of the  
33 sheet membrane was cut out and dipped in the 1-octanol for 5 s. Also, 1 M of the KI solution was  
34 used as the acceptor droplet located at the top of the membrane sheet. Finally, the platinum  
35 electrode (the cathode) was directly introduced into the 15 μL of acceptor droplet. The electrodes  
36 (the aluminum foil and the platinum electrode) were subsequently coupled to the 9.0-V battery  
37 and the extraction unit was fixed for beginning of the process. When the extraction was  
38 completed, the final acceptor droplet ( $10 \pm 1 \mu\text{L}$ ) was analyzed by offline and online modes.  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60



## 2.6. Data analysis

RGB analysis were performed using Adobe Photoshop<sup>®</sup> CS6 software and iDropper tool version 1.1, which is available for iOS smartphones.

## 3. Results and discussion

Considering previous experiences<sup>9-23</sup>, parameters effective on efficiency of electromembrane systems are composition of organic liquid membrane, donor and acceptor phases, applied voltage, and extraction time. In this work, a common battery was used for electric field formation. Hence, applied voltage was excluded from effective variables and 9.0 V was utilized for all the experiments. Also, different signals related to red, green, and blue components were used to measure the final concentration of PbI<sub>2</sub> in the acceptor droplet. Fig. 2A confirms that maximum sensitivity was gained employing the blue color and therefore this RGB component was used in all the experiments.

### 3.1. Selection of organic liquid membrane

An adequate liquid membrane for extraction of a target analyte should have some properties. The supported liquid membrane (SLM) should have a suitable viscosity and polarity and the analyte should have an acceptable solubility in the SLM. Also, the main characteristic of the SLM is its immiscibility in water. Therefore, long-chain alcohols were studied as the organic solvent. To this end, 1-heptanol, 1-octanol, and 1-decanol were tested for extraction of lead ion from the aqueous medium. The results obtained showed that 1-octanol is the best choice for this purpose. Better behavior of 1-octanol in comparison with 1-heptanol may be justified by its higher electrical resistance that decreases the electrical current and increases the stability of the chip system. Generally, the current level through SLM in EME is a key parameter which should

1  
2  
3 be kept at relatively low levels during extraction. This issue is provided by organic solvents with  
4 proper electrical resistance. For linear alcohols, the polarity is increased and the viscosity is  
5 decreased via decreasing the number of carbon atoms. On the other hand, the electrical resistance  
6 is decreased and the electrical current is increased by increasing the polarity of alcohols as SLM.  
7  
8 Therefore, more repeatable results were obtained by 1-octanol. Moreover, high viscosity of  
9  
10 1-decanol as well as its tendency for solidification due to its low boiling temperature makes  
11 transportation of the analyte into the organic phase difficult and decreases the extraction  
12 efficiency. Thus, 1-octanol was selected as the SLM for rest of the experiments.  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22

### 23 *3.2. Investigation of extraction time*

24  
25 EME is a non-exhaustive extraction method. Therefore, extraction time is one of the most  
26 important influential parameters, which should be investigated. The RGB analysis was carried  
27 out after extraction of the model analyte for various durations in the range of 120 min. Results in  
28 Fig. 3A illustrate that the best extraction efficiency was obtained by performing the EME process  
29 for 20 min. Since stagnant conditions were applied for lead analysis (neither donor phase nor  
30 acceptor phase were agitated), relatively long extraction time is required for best extractability  
31 achievement and RGB signals were improved by increasing the extraction time. More increase in  
32 the extraction time decreases the repeatability due to evaporation of the acceptor droplet and  
33 change of its volume. It can be seen from Fig. 1 that the setup used has no shielding for the  
34 acceptor phase and it could gradually evaporate. This is while SLM evaporation may occur by  
35 increasing of the extraction time, which disturbed the EME process. Since SLM is the main  
36 electrical resistance of the system, local loosing of the liquid membrane as a result of its  
37 evaporation leads to rising of electrical current level, increase in the electrolysis reactions, and  
38 bubble formation. Therefore, 20 min would be the optimal extraction time.  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

### 3.3. Investigation of composition of donor and acceptor phases

Since formation of  $\text{PbI}_2$  precipitate is the indicator for lead existence, 1 M of the KI solution was used as the acceptor droplet. Considering the donor phase, it should have a suitable pH; such that the analyte of interest carries a net positive charge and migrate toward the cathode in the electrical field. So, the effect of HCl concentration was investigated in the range of 0-100  $\text{mmol L}^{-1}$ . Previous studies proved that the flux of analyte in EME system has a relationship with the ion balance, which is defined as ratio of the total ionic concentration in the sample solution to that in the acceptor phase<sup>31</sup>. Also, it was shown that the extractability would reduce by increasing the ion balance. Therefore, it was anticipated that the final signal of lead analysis will decrease by increasing the concentration of  $\text{H}^+$  or decreasing the pH value in the sample solution. Results in Fig. 3B confirm this and as can be seen, the best recoveries were obtained by extracting the lead ion from pure water with  $\text{pH} = 6.5$  and into the KI solution.

### 3.4. Evaluation of method performance

Figures of merit of the designed system were considered in pure water to evaluate the practical applicability of electromembrane chip for quantification of lead ion. Thus, optimal conditions were applied and calibration curves were plotted. The results summarized in Table 1 show that the proposed device is capable of determining the lead ion down to  $20 \text{ ng mL}^{-1}$  and the limit of quantification (LOQ) was  $50 \text{ ng mL}^{-1}$ . Linear range between  $50\text{-}1500 \text{ ng mL}^{-1}$  and acceptable correlation coefficient ( $R^2 = 0.9994$ ) were obtained. Precision, defined as the relative standard deviation (RSD%), was determined by intra- and inter-assays by five replicate measurements at three concentrations ( $70, 500, \text{ and } 1000 \text{ ng mL}^{-1}$ ). Regarding the RSD% in low, middle, and high concentrations, intra- and inter-assay precision in the range of 3.8-7.0% and 9.8-11.9%, respectively, were achieved (Table 2).

1  
2  
3 To validate the obtained results by the proposed methods, the acceptor phase was  
4 analyzed by both RGB and electrothermal atomic absorption spectrometry (ETAAS) after EME.  
5 For this purpose, a same sample solution spiked at the concentration of  $100 \text{ ng mL}^{-1}$  was  
6 extracted using EME-based LOC system. In the case of ETAAS, the acceptor droplet was diluted  
7 50-fold with ultrapure water since the dynamic linear range of ETAAS system was in the range  
8 of  $0\text{-}40 \text{ ng L}^{-1}$ . Determination of Pb by each detection technique was replicated three times and  
9 the calculated averages were compared by means of t-test at 95% confidence limit. The results in  
10 Table 3 indicated no significant differences between calculated concentrations using both  
11 detection techniques ( $t_{(4, 0.05)} = 1.626$ ,  $t_{\text{Table}} = 2.776$ ).  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23

24 Comparison of the designed electromembrane chip with other existing methods for  
25 determining of the lead in different plant tissues is provided in Table 1. The aim of this work is  
26 to introduce a simple, inexpensive and portable system to analyze Pb in complicated matrices  
27 due to its ability to providing sample preconcentration and cleanup using a membrane sheet. The  
28 proposed LOC system demonstrated wide linearity range, high sensitivity, and acceptable  
29 repeatability and reproducibility in a relatively short time.  
30  
31  
32  
33  
34  
35  
36  
37  
38

39 The limit of Pb in drinking water is  $50 \text{ } \mu\text{g L}^{-1}$  that is higher than the obtained LOD by the  
40 proposed method<sup>43</sup>. On the other hand, the provided LOD is comparable with some traditional  
41 methods such as conventional EME followed by CE-UV<sup>39</sup>.  
42  
43  
44  
45

46 Lower detection limits can be obtained by more sensitive detection techniques such as  
47 ETAAS, inductively coupled plasma optical emission spectrometry (ICP-OES) or mass  
48 spectrometry (ICP-MS) and x-ray fluorescence spectrometry (XRF), that are very expensive, not  
49 applicable in field analysis and also some of them are not available in common laboratories.  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Application of a smartphone as the detector not only eliminates the need for expensive  
4 and unavailable analytical instruments, but also provides a simple, inexpensive, fast, and portable  
5 analysis device. Furthermore, the LOC system offers a ligandless analysis method required only  
6 100  $\mu\text{L}$  of sample to provide the sensitivity for Pb analysis down to 20  $\text{ng mL}^{-1}$ .  
7  
8  
9

10  
11  
12 It should be noticed that other metal ions can be extracted from yellow precipitates or  
13 color solution. However, the present work is the first attempt to design a portable analysis device  
14 via coupling an electromembrane chip to a smartphone and Pb was selected as the model ion to  
15 introduce the set-up. Moreover, leaves of street plants were selected as real samples due to the  
16 highest probability of lead existence in this matrix in comparison with other metal ions reacting  
17 with I, which is attributed to fuel consumption by vehicles. A lot of reports can be found in the  
18 literature related to lead accumulation in street plants due to petrol-fueled vehicles<sup>44, 45</sup>.  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28

29 Moreover, one of the interesting points that can be considered for the introduced method  
30 is simultaneous extraction of different metal ions and analysis by multivariate curve resolution  
31 techniques. Additional experiments are undergoing and the results will be published in the  
32 future.  
33  
34  
35  
36  
37  
38

### 39 *3.5. Analysis of real samples*

40  
41  
42 The designed electromembrane chip was employed for analysis of a model heavy metal in  
43 different real samples including wastewater samples and some plant tissues to consider its  
44 practical applicability. Wastewater samples were filtered via a cellulose acetate membrane filter  
45 and their pHs were adjusted to 6.5 prior to analysis. The whole plant leaves collected around the  
46 city were blended, sonicated, and centrifuged as it was described in section 2.3. The pH of the  
47 final 10-fold diluted liquid extract was adjusted to 6.5 via addition of proper amounts of sodium  
48 hydroxide solution and the optimal conditions of EME process were applied for quantitative  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 analysis. Finally, the iDropper tool of an iOS smartphone was utilized for analysis of the final  
4 droplet. The RGB analyses of the acceptor solution, which were obtained from the wastewater  
5 sample is shown in Fig. 4. Since some real samples (plant tissues) were transformed into miry  
6 complicated solutions, some experiments were necessary to determine whether the calibration  
7 curves could be directly used for analysis of real samples or not. A matrix effect is the direct or  
8 indirect alteration or interference in response due to the presence of unintended analytes or other  
9 interfering substances in the sample. Thus, accuracy (Error%) was determined by addition of 50  
10 ng mL<sup>-1</sup> of each analyte into real samples and applying EME afterwards. The relative recovery  
11 (*RR%*) and accuracy (Error%) were calculated by the following equations:  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23

$$24 \quad RR\% = \frac{C_{\text{found}} - C_{\text{real}}}{C_{\text{added}}} \times 100 \quad (1)$$

$$25 \quad Error\% = RR\% - 100 \quad (2)$$

26  
27 where  $C_{\text{found}}$ ,  $C_{\text{real}}$ , and  $C_{\text{added}}$  are the concentrations (ng mL<sup>-1</sup>) of analyte after addition of known  
28 amount of standard into the real sample, the concentration of analyte in real sample, and the  
29 concentration of known amount of standard spiked into the real sample, respectively.  
30  
31  
32  
33  
34  
35  
36

37 The results in Table 4 show that no significant matrix effect was observed for the real  
38 samples studied and admissible precision values were obtained. Therefore, in analysis of real  
39 samples, calibration curves could be used directly.  
40  
41  
42  
43  
44  
45

#### 46 4. Conclusions

47  
48 An EME-based LOC system followed by RGB analysis was introduced for the first time  
49 for analysis of lead as the model analyte in different matrices. It was shown that the intensity of  
50 red, green, and blue components of the final acceptor droplet is proportionate to the lead  
51 concentration. Therefore, RGB analysis of the digital picture taken from the acceptor solution  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 could be used for quantification of the target analyte. Combination of EME-based LOC system  
4  
5 and a smartphone with the ability of RGB decomposition constructed a portable device for lead  
6  
7 analysis in different matrices. However, although there are some reports for designing  
8  
9 electromembrane chips, further studies are required to gain a fully portable and practical device.  
10  
11

## 12 13 14 15 **Acknowledgements**

16  
17 The authors gratefully acknowledge K.N. Toosi University of Technology and gracious  
18  
19 help of Mrs. Mirhadi, Mrs. Fakhr, Mrs. Halvaiy, Mrs. Abedini and Miss Jalaian from Farzanegan  
20  
21 Educational Center (Tehran, Iran).  
22  
23

## 24 25 **References:**

- 26  
27  
28 1 E. Verpoorte, *Electrophoresis*, 2002, **23**, 677.  
29  
30 2 I. Ali, H.Y. Aboul-Enein, V.K. Gupta, *Nano Chromatography and Capillary Electrophoresis:*  
31  
32 *Pharmaceutical and Environmental Analyses*, Wiley & Sons, Hoboken, USA, 2009.  
33  
34 3 I. Ali, Z.A. Al-Othman, A. Al-Warthan, H.Y. Aboul-Enein, *Curr. Chromatogr.*, 2014, doi:  
35  
36 10.2174/2213240601666140301001948.  
37  
38 4 Z.A. AL-Othmana, I. Ali, *J. Liq. Chromatogr. Rel. Technol.*, 2011, **34**,1295.  
39  
40 5 I. Ali, H.Y. Aboul-Enein, *Current Pharm. Anal.*, 2009, **5**, 367.  
41  
42 6 Z.A. AL-Othman, I. Ali, *Chromatographia*, 2009, **69**, S13.  
43  
44 7 S. T.S. Ho, S. Pedersen-Bjergaarda, K.E. Rasmussen, *Analyst*, 2002, **127**, 608.  
45  
46 8 C. Rosting, S. Pedersen-Bjergaard, S. Honoré Hansen, C. Janfelt, *Analyst*, 2013, **138**, 5965.  
47  
48 9 S. Pedersen-Bjergaard, K.E. Rasmussen, *J. Chromatogr. A*, 2006, **1109**, 183.  
49  
50 10 S. Seidi, Y. Yamini, M. Rezazadeh, A. Esrafil, *J. Chromatogr. A*, 2012, **1243**, 6.  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

- 1  
2  
3 11 C. Basheer, J. Lee, S. Pedersen-Bjergaard, K.E. Rasmussen, H.K. Lee, *J. Chromatogr. A*,  
4  
5 2010, **1217**, 6661.  
6  
7  
8 12 L. Guo, H.K. Lee, *J. Chromatogr. A*, 2012, **1243**, 14.  
9  
10 13 S. Seidi, Y. Yamini, M. Rezazadeh, *J. Chromatogr. B*, 2013, **913–914**, 138.  
11  
12 14 M. Rezazadeh, Y. Yamini, S. Seidi, B. Ebrahimpor, *J. Chromatogr. A*, 2013, **1280**, 16.  
13  
14 15 N.J. Petersen, H. Jensen, S.H. Hansen, K.E. Rasmussen, S. Pedersen-Bjergaard, *J.*  
15  
16 *Chromatogr. A*, 2009, **1216**, 1496.  
17  
18 16 N.J. Petersen, J.S. Pedersen, N.N. Poulsen, H. Jensen, C. Skonberg, S.H. Hansen, S.  
19  
20 Pedersen-Bjergaard, *Analyst*, 2012, **137**, 3321.  
21  
22 17 N.J. Petersen, S.T. Foss, H. Jensen, S.H. Hansen, C. Skonberg, D. Snakenborg, J.P. Kutter, S.  
23  
24 Pedersen-Bjergaard, *Anal. Chem.*, 2011, **83**, 44.  
25  
26 18 A. Šlampová, P. Kubáň, P. Boček, *J. Chromatogr. A*, 2012, **1234**, 32.  
27  
28 19 M. Rezazadeh, Y. Yamini, S. Seidi, A. Esrafil, *J. Chromatogr. A*, 2012, **1262**, 214.  
29  
30 20 M. Rezazadeh, Y. Yamini, S. Seidi, A. Esrafil, *Anal. Chim. Acta*, 2012, **773**, 52.  
31  
32 21 T.M. Middelthon-Bruer, A. Gjelstad, K.E. Rasmussen, S. Pedersen-Bjergaard, *J. Sep. Sci.*,  
33  
34 2008, **31**, 753.  
35  
36 22 M. Balchen, A. Gjelstad, K.E. Rasmussen, S. Pedersen-Bjergaard, *J. Chromatogr. A*, 2007,  
37  
38 **1152**, 220.  
39  
40 23 M. Rezazadeh, Y. Yamini, S. Seidi, *J. Chromatogr. A*, 2014, **1324**, 21.  
41  
42 24 I. López-García, Y. Vicente-Martínez, M. Hernández-Córdoba, *Talanta*, 2014, **124**, 106.  
43  
44 25 E. Teju, B. Tadesse, N. Megersa, *J. Environ. Sci. Health: Part A*, 2014, **49**, 833.  
45  
46 26 Z. Li, J. Chen, M. Liu, Y. Yang, *Anal. Methods*, 2014, **6**, 2294.  
47  
48 27 M. Shamsipur, N. Fattahi, M. Sadeghi, M. Pirsahab, *J. Iran. Chem. Soc.*, 2014, **11**, 249.  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60



- 1  
2  
3 28 S.Z. Mohammadi, D. Afzali, Z. Fallahi, *Anal. Chem.*, 2014, **94**, 765.  
4  
5  
6 29 P. Liang, J. Yu, E. Yang, L. Peng, *Atom. Spectrosc.*, 2014, **35**, 85.  
7  
8 30 F. Sponar, A. Catalin Mot, C. Sarbu, *J. Chromatogr. A*, 2008, **1188**, 295.  
9  
10 31 A. Gjelstad, K.E. Rasmussen, S. Pedersen-Bjergaard, *J. Chromatogr. A*, 2007, **1174**, 104.  
11  
12 32 H. Altundag, M. Tuzen, *Food Chem. Toxicol.*, 2011, **49**, 2800.  
13  
14 33 M.-L. Lin, S.-J. Jiang, *Food Chem.*, 2013, **141**, 2158.  
15  
16 34 W. Zhong, C. Zhang, Q. Gao, H. Li, *Microchim. Acta*, 2012, **176**, 101.  
17  
18 35 C. Locatelli, D. Melucci, *Cent. Eur. J. Chem.*, 2013, **11**, 790.  
19  
20 36 I. De La Calle, M. Costas, N. Cabaleiro, I. Lavilla, C. Bendicho, *Food Chem.*, 2013, **138**, 234.  
21  
22 37 F. Shah, E. Yilmaz, T.G. Kazi, H.I. Afridi, M. Soylak, *Anal. Methods*, 2012, **4**, 4091.  
23  
24 38 P. Liang, E. Yang, J. Yu, L. Wen, *Anal. Methods*, 2014, DOI: 10.1039/c4ay00019f.  
25  
26 39 C. Basheer, S.H. Tan, H.K. Lee, *J. Chromatogr. A*, 2008, **1213**, 14.  
27  
28 40 E. Yilmaz, M. Soylak, *Talanta*, 2013, **116**, 882.  
29  
30 41 Z.A. Alothman, E. Yilmaz, M. Habila, A. Shabaka, M. Soylak, *Microchim. Acta*, 2013, **180**,  
31  
32 669.  
33  
34 42 P.N. Nomngongo, J.C. Ngila, *Spectrochim. Acta Part B*, 2014, **98**, 54.  
35  
36 43 M. Kumar, A. Puri, *Indian J. Occup. Environ. Med.*, 2012, **16**, 40.  
37  
38 44 C.S. Mendoza, J. Hipe, *S. Pac. Stud.*, 2008, **28**, 43.  
39  
40 45 C. Aydinalp, S. Marinova, *Pol. J. Environ. Stud.*, 2004, **13**, 233.  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**Figure captions**

**Fig. 1.** The equipment used for the electrically induced LOC system (A) and detection of yellow particulate with RGB analysis (B).

**Fig. 2.** (A) Comparison of sensitivities of different red, green, and blue components for lead quantification in the acceptor phase. RGB analysis of (B) acceptor droplet for extraction from sample contains  $500 \text{ ng mL}^{-1}$  lead ion and (C) white obtained by Photoshop<sup>®</sup> software.

**Fig. 3.** Optimizations of (A) extraction time and (B) composition of donor phase. Lead ions were extracted from sample solution across 1-octanol as the SLM and into 1M KI solution by application of 9.0 V electrical field.

**Fig. 4.** EME of wastewater sample contains  $87.5 \text{ } \mu\text{g L}^{-1}$  of lead ion followed by RGB analysis using iDropper tool of an iOS smartphone.

**Table 1**Comparison of the proposed method with other analytical techniques for determination of Pb<sup>2+</sup> in different samples.

| Analytical technique <sup>a</sup> | Sample                         | Extraction time | Linear range                   | R <sup>2</sup> | LOD                      | RSD%     | Ref.      |
|-----------------------------------|--------------------------------|-----------------|--------------------------------|----------------|--------------------------|----------|-----------|
| MW-ICP-OES                        | Dried fruit                    | > 31 min        | 10-320 µg L <sup>-1</sup>      | 0.99994        | -                        | < 11.4   | 32        |
| USS-ETV-ICP-MS                    | Herbs                          | > 10 min        | -                              | > 0.9992       | 0.2 ng g <sup>-1</sup>   | <4.0     | 33        |
| Dried digestion-CdTe QDs          | Spinach and citrus leaves      | > 7-8 h         | 1.96-25.9 nmol L <sup>-1</sup> | 0.9960         | 4.7 nmol L <sup>-1</sup> | <4.8     | 34        |
| AD-VM                             | Vegetables                     | > 2 h           | -                              | > 0.9989       | 43-51 ng g <sup>-1</sup> | < 5.0    | 35        |
| USAD-XRF                          | Anatomical part of plants      | > 20 min        | -                              | -              | 2.9 µg g <sup>-1</sup>   | < 9.0    | 36        |
| VLLME-FAAS                        | Hair and urine                 | > 10 min        | 5–200 µg L <sup>-1</sup>       | 0.992          | 0.307 µg L <sup>-1</sup> | 4.09     | 37        |
| SM-DLLME-SFOD-ETAAS               | Food and water                 | > 5 min         | 0.1–30 µg L <sup>-1</sup>      | 0.9995         | 27 ng L <sup>-1</sup>    | > 2.5    | 38        |
| EME-CE-UV                         | Biological fluids and lipstick | > 15 min        | 0.1–10 mg L <sup>-1</sup>      | 0.9935         | 19 µg L <sup>-1</sup>    | 4.9-15.6 | 39        |
| DLLME-D-µ-SPE-FAAS                | Water, plant and hair          | > 2.5 min       | 1.9-? µg L <sup>-1</sup>       | 0.991          | 0.57 µg L <sup>-1</sup>  | < 7.5    | 40        |
| TC-IL-ME-FAAS                     | Hair                           | > 14 min        | 19.3-? µg L <sup>-1</sup>      | -              | 5.8 µg L <sup>-1</sup>   | -        | 41        |
| HF-SPME-ICP-MS                    | Gasoline and diesel samples    | > 40 min        | 0.8–100 µg L <sup>-1</sup>     | 0.9967         | 0.3 µg L <sup>-1</sup>   | 2.5      | 42        |
| LOC-RGB                           | Plant leaf                     | > 22 min        | 50-1500 ng mL <sup>-1</sup>    | > 0.9994       | 20 µg L <sup>-1</sup>    | < 7.0    | This work |

<sup>a</sup> Microwave (MW), Ultrasonic slurry sampling (USS), Electrothermal vaporization (ETV), Quantum dots (QDs), Acidic digestion-Voltammetric method (AD-VM), Ultrasound assisted digestion (USAD), Vortex-assisted liquid-liquid microextraction (VLLME), Flame atomic absorption spectrometry (FAAS), Supramolecular solvent (SM), Dispersive liquid-liquid microextraction (DLLME), Solidification of floating drop (SFOD), Electromembrane extraction (EME), Capillary electrophoresis (CE), Dispersive micro solid-phase extraction (D-µ-SPE), Temperature controlled ionic liquid based microextraction (TC-IL-ME), Hollow fiber-solid phase microextraction (HF-SPME).

**Table 2**

Accuracy, precision, and relative recovery of the designed LOC system for determination of lead ion in pure-water.

| Conc.<br>(ng mL <sup>-1</sup> ) | Accuracy (Error %)     |                        | Precision (RSD %)      |                        | RR%   |
|---------------------------------|------------------------|------------------------|------------------------|------------------------|-------|
|                                 | Intra-assay<br>(n = 3) | Inter-assay<br>(n = 3) | Intra-assay<br>(n = 5) | Inter-assay<br>(n = 3) |       |
| 70                              | +2.6                   | -1.9                   | 5.7                    | 10.2                   | 102.6 |
| 500                             | -2.7                   | -8.5                   | 3.8                    | 11.9                   | 97.3  |
| 1000                            | -3.0                   | +4.5                   | 7.0                    | 9.8                    | 97.0  |

**Table 3**  
The t-test results to validate LOC-RGB method.

|                                 | Detection technique |   |
|---------------------------------|---------------------|---|
|                                 | RGB                 | ETAAS                                       |
|                                 | 95.2                | 92.1  |
| Pb Conc. (ng mL <sup>-1</sup> ) | 104.2               | 89.0  |
|                                 | 108.5               | 102.7                                       |
| Average                         | 102.633             | 94.6  |
| Standard deviation              | 6.787               | 7.184                                       |
| S <sub>pooled</sub>             | 6.988               |   |
| t <sub>(4, 0.05)</sub>          | 1.626               | t <sub>(4, 0.05)</sub> < t <sub>Table</sub> |
| t <sub>Table</sub>              | 2.776               | Not significant                             |

**Table 4**  
Lead determination in real samples.

| Sample       | $C_{real}$<br>( $\mu\text{g L}^{-1}$ ) | $C_{added}$<br>( $\mu\text{g L}^{-1}$ ) | $C_{found}$<br>( $\mu\text{g L}^{-1}$ ) | RSD% | Error% |
|--------------|--|---|---|------|--------|
| Wastewater 1 | <LOQ                                   | 50.0                                    | 52.1                                    | 5.9  | +4.2   |
| Wastewater 2 | 87.5                                   | 50.0                                    | 135.8                                   | 8.3  | -3.4   |
| Plant 1      | <LOQ                                   | 50.0                                    | 47.6                                    | 6.5  | -4.8   |
| Plant 2      | 345.4 (0.69 $\mu\text{g g}^{-1}$ )     | 50.0                                    | 391.6                                   | 7.8  | -7.6   |
| Plant 3      | 148.2 (0.30 $\mu\text{g g}^{-1}$ )     | 50.0                                    | 202.9                                   | 6.4  | +9.4   |

Fig. 1

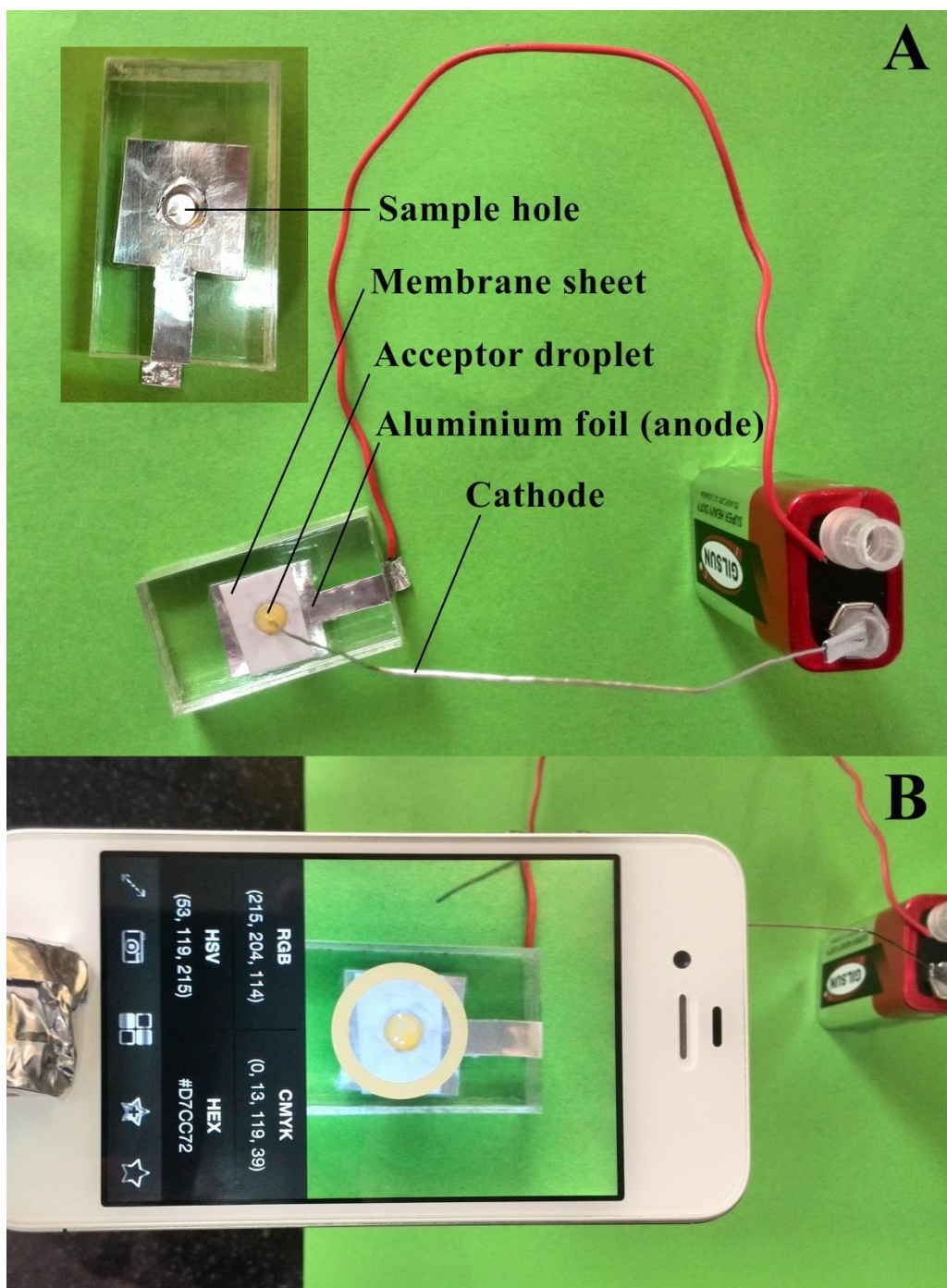


Fig. 2

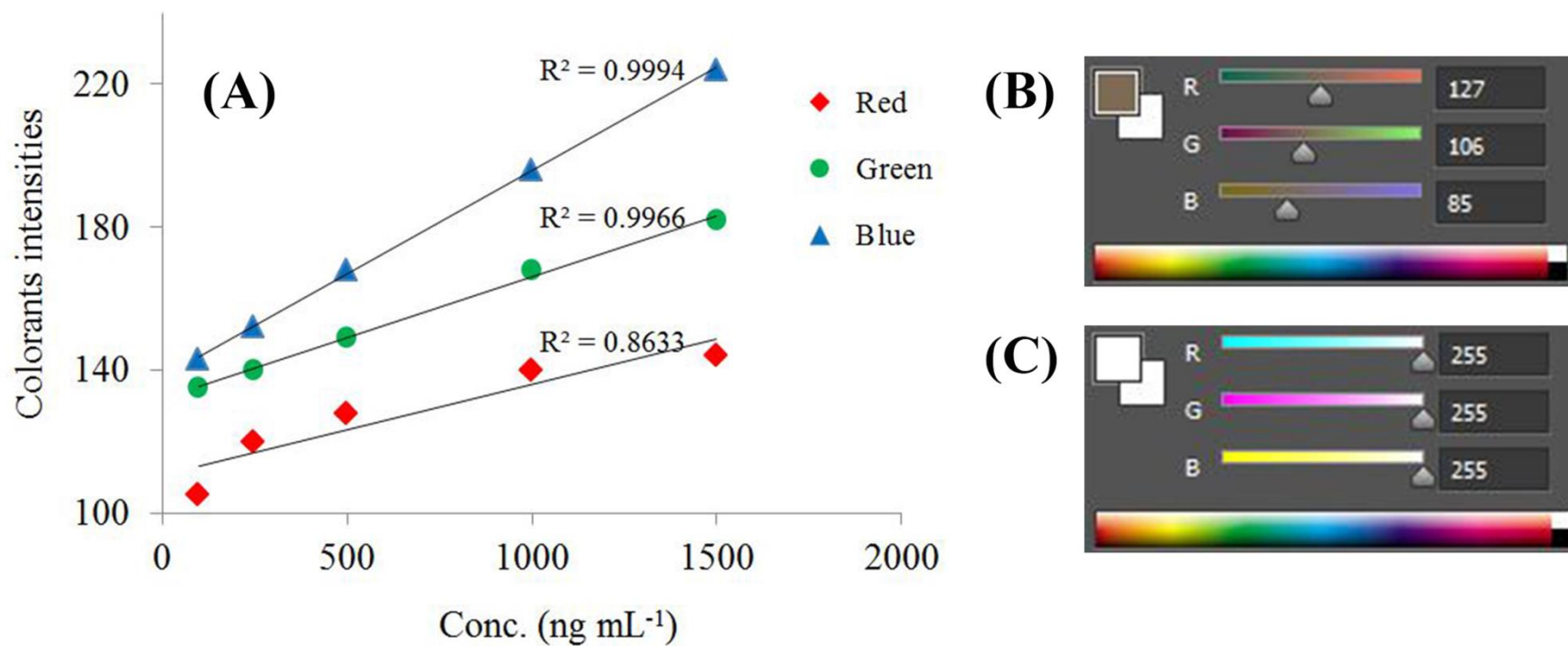
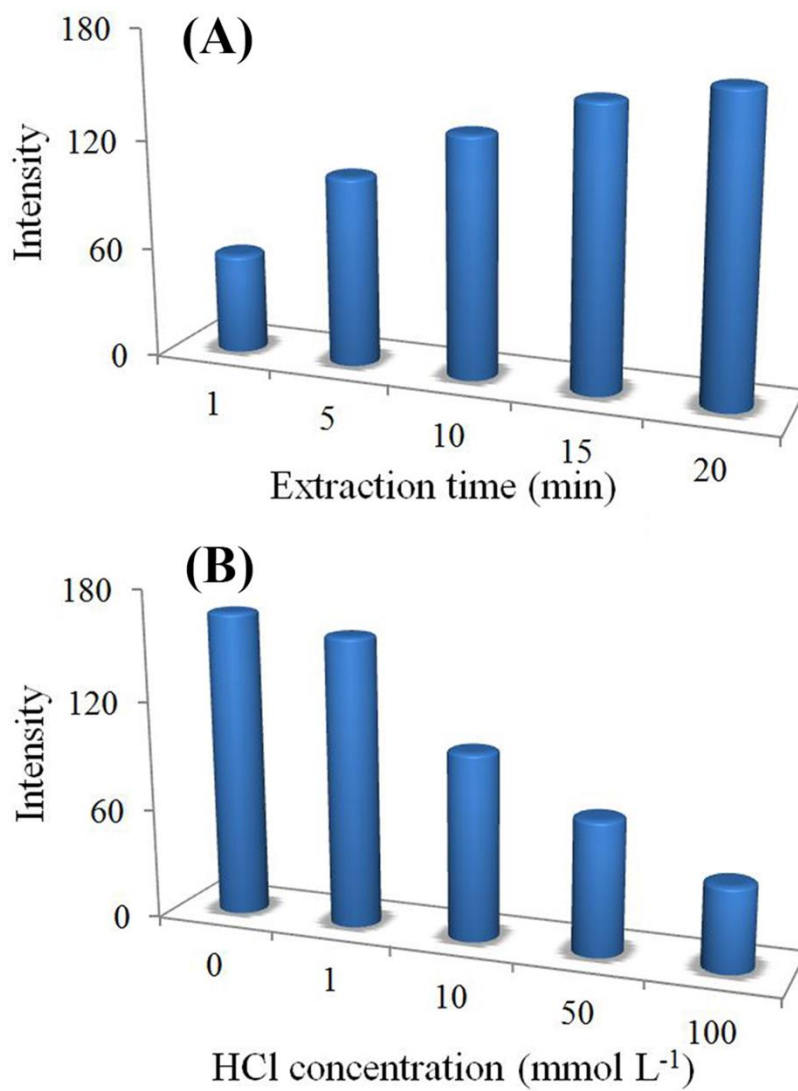




Fig. 3



1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Fig. 4



Analyst Accepted Manuscript

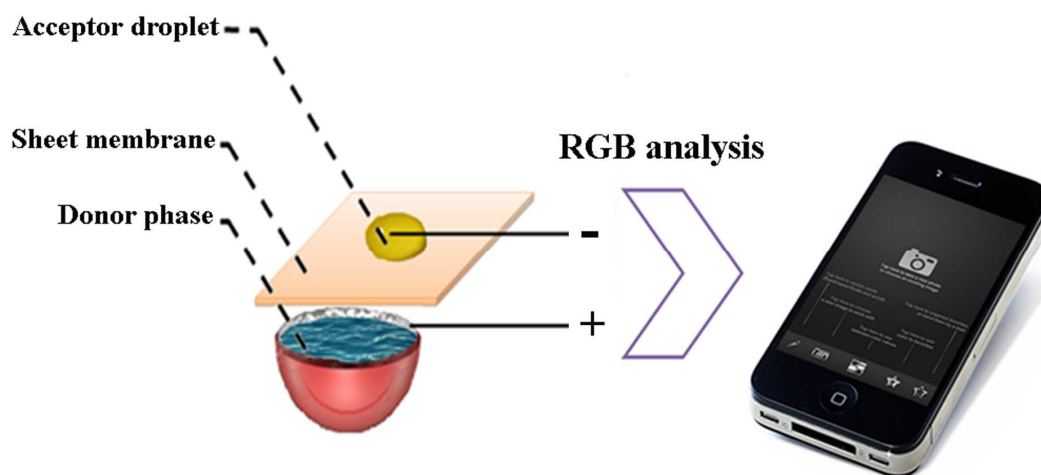
# Low voltage electrically stimulated lab-on-a-chip device followed by Red-Green-Blue analysis: A simple and efficient design for complicated matrices

Shahram Seidi<sup>a,\*</sup>, Maryam Rezazadeh<sup>b</sup>, Yadollah Yamini<sup>b</sup>, Niki Zamani<sup>c</sup>, Sara Esmaili<sup>c</sup>

<sup>a</sup>Department of Analytical Chemistry, Faculty of Chemistry, K.N. Toosi University of Technology, Tehran, Iran

<sup>b</sup>Department of Chemistry, Tarbiat Modares University, Tehran, Iran

<sup>c</sup>Farzanegan 1 Educational Center, National Organization for Development of Exceptional Talents, Tehran, Iran



An electrical field-induced lab-on-a-chip design followed by RGB analysis is introduced to develop a portable device for analysis of  $\text{Pb}^{2+}$ .

\* Corresponding author at: K.N. Toosi University of Technology, Faculty of Chemistry, Department of Analytical Chemistry, P.O. Box 16315-1618, Tehran, Iran. Tel.: +98(21)23064228; Fax: +98(21)22853650.  
E-mail address: [s.seidi@kntu.ac.ir](mailto:s.seidi@kntu.ac.ir) (S. Seidi).