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A multi-scale approach of the mechanisms underlying exopolysaccharides auto organization in the *Proteus mirabilis* extracellular matrix.

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For decades, the origin of the concentric ring pattern of bacterial swarming colonies has puzzled microbiologists. It was hypothesized that a periodic water activity variation originates a phase transition within the extracellular matrix water H bond network, which switches on and off the exopolysaccharides auto-organization. Both rheological and infrared spectroscopy measurements respectively performed at a molecular scale and on a currently migrating colony, have allowed to give a physical insight into the mechanisms which underlie the switch between swarming and consolidation phases. Thanks to *in situ* and real time infrared microspectroscopy, and thanks to the brilliance of the infrared beam at SOLEIL synchrotron, here we demonstrate that *Proteus mirabilis* swarming is triggered by a periodic variation of water activity at the colony's edge. A

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dynamic behavior emerges from the global properties of the multicellular entity which relies on the ability of the bacterial cells to tune exoproducts synthesis in order to undergo sharp transitions at a given water activity threshold.

## Introduction

Bacterial colonies are excellent candidates for studying fundamental problems of selforganization and pattern formation in complex biological systems. This is the reason why development biologists have paid attention to multicellularity<sup>1</sup> since such complex systems necessarily imply a high degree of signal integration leading to space and time correlations linking billions of individual cells. Hence, the interest of such studies significantly overpasses that of bacterial colonies and biofilms. In the case of Proteus mirabilis, it has been proposed that the periodicity of the swarming phenomenon<sup>2-5</sup> was linked to density-dependent thresholds in controlling the transitions between distinct phases<sup>3</sup>. Such correlations, often observed in bacterial multicellular entities, usually rely on chemical triggers, which mediate the quorum sensing (QS) cell-to-cell exchanges<sup>6</sup>. More precisely, QS allows every cell to sense the population extent and, beyond a given density threshold, to activate the genes responsible for differentiation or virulence expression. Regarding this point, the P. mirabilis swarming constitutes a unique complex biological oscillator. The population expanses through periodic and synchronous swarming phases which alternate with consolidation phases. If one excepts the very early stages of the colony formation, no chemical trigger like furanones or homoserine lactones are produced to warrant this unique synchronicity<sup>4</sup>. If the flhDC operon has been identified as the main genetic determinant for the vegetative to swarmer cell differentiation process<sup>7</sup>, genetics failed to reveal which factor was responsible for the sudden operon's switch at a given moment. Consequently,

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chemical triggers were actively, but in vain, looked for<sup>6</sup> since they are good candidates to convey information between cells, allowing thereby to locally promoting population cohesion<sup>4</sup>. This direction being a dead end, we turned to investigate the global properties of the colony and paid attention to the biochemical and functional properties of the extra cellular matrix (ECM) that constitutes the bacterial cells continuum<sup>8, 9</sup>. The *P. mirabilis* ECM biochemical analysis revealed a binary mixture of exopolisaccharides (EPS), a phenoglycolipid (PGL) and glycine betaine (GB), a well-known bacterial osmoprotectant. The presence of GB and of a PGL strongly suggested that the bacterial colony is submitted to a permanent osmotic stress, which in turn indicates that water content may constitute a constraint.

Here we present a multi-scale approach of the mechanisms which underlie the periodic and synchronous swarming of *P. mirabilis*. Earlier studies<sup>8, 9</sup> have suggested that ECM, due to its viscoelastic properties and its ability to form spherullites would behave as a semi-crystalline continuum. The present paper aims at bringing evidence that such properties are deeply involved in a periodic variation of water activity (a<sub>w</sub>), which in turn is responsible for the morphotype's switch between swarming and consolidation phases (see Fig. 10 in <sup>8</sup>). To demonstrate that this switch between consolidation and swarming phases is controlled by an EPS phase transition in relation to ECM water activity, experiments were performed to characterize more thoroughly the EPS viscoelastic properties and to monitor, *in situ* and in real-time, the EPS auto-organization along with the water H bonds network.

#### **Experimental Section**

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Strain, Media and Culture Conditions. The *P. mirabilis* wild-type strain WT19 corresponds to the clinical isolate U64507. WT19 was grown in LB medium at 37 °C. To obtain homogeneous populations of swarmer bacteria, 200  $\mu$ L of an overnight liquid culture was spread onto a LB agar (1.5%) plate, and the latter was incubated for 4 h at 37 °C. For studying the periodic swarming, a 3  $\mu$ L inoculum was deposited at the center of a Petri dish and was allowed to grow for up to 8 hours on the same agar solid medium.

# Experimental techniques.

Exopolysaccharides Extraction and Purification. EPS extraction and purification from actively swarming colonies have essentially been performed as described in <sup>8</sup>. As previously mentioned, the EPS fraction from a swarming colony is essentially characterized by two compounds with different molecular weights, one with a low molecular weight of 1 kDa and the other with a much higher molecular of 400 kDa, in a 20:1 ratio.

Polarized Microscopy. Purified EPS solutions at different concentrations (wt. %) have been prepared and were allowed to age between a sealed slide and coverslip. Observations are performed by using a BX-60 polarizing microscope and a quarter wave plate.

Rheology. All rheological measurements were carried out in oscillatory simple shear with a controlled strain rheometer (TA Instruments), equipped with a cone and plate geometry (cone angle: 2°, diameter: 5cm). The sample temperature was set at 20°C using a thermostatic bath. For each sample tested, strain sweep viscoelastic tests were first performed at a fixed 1 Hz frequency in order to determine the extent of the linear regime; then, frequency sweep experiments were

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carried out at a fixed strain in the linear regime in order to determine the linear viscoelastic moduli, that is the storage modulus G' and the loss modulus G", of the samples. G' characterizes the elastic energy stored in the sample structure, and G" the viscous dissipation within the sample. Purified EPS solutions at six different concentrations, 4.8 wt.%, 9 wt.%, 13 wt.%, 16.7 wt.%, 23 wt.%, and 28 wt.%, have been characterized.

Infrared Spectroscopy. The Mid Infrared (MIR) spectroscopy has for long been used to probe macromolecule structures and conformations since and, hence, is well suited to monitor EPS auto-organization <sup>12-14</sup> which shows up in the 1200-900 cm<sup>-1</sup> frequency domain<sup>12, 13</sup>. Water is also a strong absorbent in the MIR domain due to the O-H stretching (localized around 3400 cm<sup>-1</sup>) and the H-O-H bending (localized at 1640 cm<sup>-1</sup>) vibrations. Unfortunately, due to significant spectral overlap, essentially with proteins, the colony water content cannot be straightforwardly derived from absorbencies in these domains. This drawback may be bypassed by monitoring instead the water librational band located at around 615 cm<sup>-1 13-15</sup>. This band is rarely used since most of the optics and detectors of infrared spectrophotometers are nontransparent and not sensitive enough, respectively, to yield a signal with sufficient SNR to be interpreted. Another technical difficulty is that bacterial colonies exhibit high absorbency and diffusivity in the infrared domain which prevent from collecting a high quality specular reflected signal, the unique acquisition mode to get spectral information without any physical contact between the optics and a currently migrating colony. As a consequence, a very brilliant IR source is required, which is only available at a synchrotron radiation facility. Hence, to simultaneously monitor the EPS auto-organization phenomena and a<sub>w</sub> variations during a complete swarming phase, we used synchrotron infrared microspectroscopy (Continuum and NicPlan, Nicolet microscopes) allowing a satisfying (15 µm) 2D spatial resolution, and two types of IR detectors:

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a MCT detector (Continµm) to firstly assess *in situ* the occurrence of EPS auto-organization, and, secondly, a Si:B bolometer (NicPlan) which allows to record IR spectra in the 2000-400 cm<sup>-1</sup> wavenumber range which encompasses both EPS and water librational absorption bands. Spectra are the average of 126 scans and are collected at a 4 cm<sup>-1</sup> spectral resolution during a complete swarming cycle (consolidation-swarming-consolidation). It must be noted that no significant baseline drifts were observed nor artefacts generally arising from Mie scattering. Hence no Kramers-Kronig correction was applied to the collected spectra and intensities are expressed as log(1/R). This allows assessing that the observed spectral changes are not due to standing wave effects as described by Filik<sup>16</sup>.

#### Results and discussion

Semi-crystallinity of the EPS purified fraction. In the present study, ECM is assumed to play a major role in the switch between consolidation and swarming phases of *P. mirabilis* colonies. This hypothesis relies on the semi-crystalline properties of this continuum. Previous SEM experiments have shown that spherullite-like objects were observed in colonies<sup>9</sup>, whereas spherullites and needle-like crystalline objects were observed in purified ECM fractions<sup>8</sup>. Since a<sub>w</sub> variations within the colony are expected to drive reversible EPS auto-organization, we made attempts to study the behavior of EPS solutions at various concentrations to highlight the relation between a<sub>w</sub> and the formation of semi-crystalline objects. Hence, observation of EPS solutions at different concentrations (wt.%) were performed by using polarizing optical microscopy.

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**Figure 1**. Crystalline objects observed by polarized microscopy at different purified EPS weight fractions. The frame widths are 700  $\mu$ m.

Figure 1 displays different objects observed at weight concentrations ranging from 5 up to 28%. At low weight concentrations (<10%), the micrographs do not show any particular objects. At a concentration of about 13 wt.%, spherullite-like objects appear, with a strongly birefringent signature due to local order; they exhibit diameters ranging from 100 up to 200  $\mu$ m. If the water activity is further decreased, radially ordered crystalline needle-like objects are observed, which look very much like those observed by SEM in a swarming colony (see Fig. 6 in <sup>9</sup>). These needle-like objects exhibit an average length l of about 100  $\mu$ m and an average diameter D of about 17  $\mu$ m, leading to an average aspect ratio l/D of about 6 (about 50 needles were characterized). These observations clearly show that, in a *P. mirabilis* colony, such semicrystalline objects are derived from EPS auto-organization and are independent on any effect due to, or mediated by, the bacterial cells, since these objects are observed in EPS purified solutions. Above 16 wt.%, a new structure, governed by auto-organization phenomena, appears as suggested by the observation of crystalline needle-like objects organized in a branched network, which is expected to give an additional contribution to the ECM rigidity. It was hence necessary to further characterize the mechanical properties of the ECM, in the concentration range on both sides of the concentration corresponding to the above-described structural change.

Mechanical properties of the EPS purified fraction. For all samples tested, the limit of the linear viscoelastic regime, where G' and G" are independent of strain amplitude, lies between 0.1% and 1%. This result shows that a low strain is able to induce a non-linear viscoelastic behavior, suggesting that the structure of the EPS solutions, over the whole range of concentrations investigated, is rather fragile.

Figure 2 shows the storage modulus G' and the loss modulus G" as a function of frequency, at a strain chosen in the linear response regime, for a 4.8 wt% (Fig.2A) and 16.7 wt% (Fig.2B) EPS solution, respectively. At all EPS concentrations investigated in this work, the viscoelastic behavior of EPS solutions has the same features: both viscoelastic moduli have low values and are weakly dependent on frequency, at least in the frequency range investigated, and G'>>G". These results mean that all EPS solutions investigated in this work behave like soft viscoelastic solids, which suggests that their structure is that of a weak gel<sup>17</sup>, most likely due to the presence of numerous H bonds which connect EPS molecules, forming a fragile three-dimensional polymeric network, as discussed in a previous paper<sup>8</sup>.



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Figure 2 Storage modulus G' and loss modulus G'' as a function of frequency, at a fixed strain amplitude of 0.5%, for a 4.8 wt% (Left) and 16.7 wt% (Right) EPS solution.

The main difference between the low and high concentrated EPS solutions is the level of the moduli: the storage modulus of a 16.7 wt% EPS solution is about 2 decades higher than that of a 4.8 wt% EPS solution. In order to study the EPS concentration dependence of the storage G' at a fixed frequency, we plotted Figure 3.



Figure 3: Storage modulus G' as a function of EPS concentration, at a frequency of 1 Hz.

Figure 3 clearly shows that there are two concentration regimes: at EPS concentrations below about 13 wt%, G' has low values and exhibits a weak concentration dependence, whereas at EPS concentrations higher that 15 wt%, G' has much higher values and exhibits a much stronger concentration dependence. One may refer to Fig. 1 for the corresponding supramolecular organization present at each concentration.

The existence of a concentration threshold at about 13 wt%, characterized by a drastic G' increase, has to be linked with the concentration induced modification of the state of

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organization of the EPS macromolecules presented above. Indeed the presence of anisometric needle-like crystalline entities (the so-called spicules, see Fig. 1) in EPS solutions at concentrations higher than 10 wt% could be responsible for the drastic enhancement of elastic properties.

Knowing the characteristic dimensions of crystalline needles, and the percolation volume fraction of spheres,  $\Phi ps \sim 30\%$ <sup>18</sup>, the percolation volume fraction,  $\Phi p$ , of these objects can be inferred from the assumption that the volume occupied by cylinders is equivalent to that of spheres with a radius equal to half the average length of the cylinders. Indeed, if the needles are considered as cylinders, with diameter D and length L, then :

$$\frac{\Phi_p}{\Phi_{ps}} = \frac{3}{2} \left(\frac{D}{L}\right)^2$$

Knowing the average aspect ratio of the microcrystalline spicules  $L/D \sim 6$ , then  $\Phi p \sim 1.3\%$ ; thus only about 1 vol% spicules are needed to form a percolation network. We suggest that the EPS concentration threshold at about 13 wt% corresponds to the percolation threshold of the microcrystalline spicules, and that, above this concentration threshold, the significantly enhanced elastic response of EPS solutions is due to the superposition of the EPS transient polymeric network and the spicule percolation network.

Rheological experiments performed on EPS solutions have revealed a marked elastic behaviour with a sharp phase transition driven by water activity. Such a behaviour is the signature of EPS auto-organization, which in turn depends on the physico-chemical modulation of polysaccharides H bond networks. At low a<sub>w</sub>, a percolation network is formed which is expected to drastically hinder swarm cell motility. Since an alteration of the external medium

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mechanical properties drives the initial cell differentiation subsequent to the transfer of the strain from a liquid culture to an agar Petri dish<sup>10</sup>, we have hypothesized that the periodicity and synchronicity of the swarming could be ruled by a<sub>w</sub> variations at the edge of the colony. Both experimental observations and theoretical considerations<sup>9, 11</sup> converge to suggest that, due to the colony spreading on the agar as a thin (15 µm) film during a swarming phase, the resulting increased S/V ratio would increase the net water transfer from the agar to the colony. The resulting increased a<sub>w</sub> within the biofilm should consequently induce a disorganization of the percolation network which is responsible for the high storage modulus of the EPS solutions. It was then necessary to look for concomitant variations of both EPS supramolecular organization and water H bonds network. These attempts have been performed through infrared microspectroscopy on a currently swarming colony.

Real time study of EPS auto-organization. We firstly attempt to assess *in situ* the occurrence of EPS auto-organization by monitoring, in a non-invasive way, the EPS MIR spectral domain through an entire swarming cycle, *i.e.* between two consecutive consolidation phases. Every two minutes, a spectrum was collected at the colony's moving edge, which allows spatially mapping the spectral information. Figure 4 shows the resulting MIR spectra.

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**Figure 4**. Mean MIR spectrum (1800-900 cm<sup>-1</sup>) and difference spectra (inset) of a currently swarming colony. The figure displays the average (n=50) spectrum in the 1800-900 cm<sup>-1</sup> frequency domain and the inset shows the dynamic spectra (from which the mean spectrum has been subtracted). Proteins Amide I and II bands show up at 1650 cm<sup>-1</sup> (with a strong overlap of the water bending mode) and 1540-1580 cm<sup>-1</sup>, respectively whereas EPS show up in the 1200-900 cm<sup>-1</sup> frequency domain.



**Figure 5**. PCA of MIR spectra (1800-900 cm<sup>-1</sup>) collected during a swarming phase. A: The migration (dots) scale is zeroed at the beginning of the experiment and corresponds to an initial colony radius of approximately 3 mm, The PC1 score is also plotted (solid line); B: Three first principal components.

The data set was analysed by Principal Component Analysis (PCA), and the first principal component featuring 45% of the total variability was plotted as a function of the colony expansion (Fig. 5A). The resulting data show that most of the spectral variations can be safely assigned to EPS since the three first PCs, reflecting up to 90% of the total variability, are dominated by spectral features which correspond to the EPS absorption domain (Fig. 5B). It is

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noteworthy that the PC1 scores (Fig. 5A) undergo a reversible sharp transition just before the colony's edge switches between swarming and consolidation. Hence, it can be assessed that the EPS auto-organization which has been observed on purified EPS solutions<sup>8,19</sup>, also occurs *in situ* at the transition between consolidation and swarming phases.

In order to determine whether or not EPS auto-organization is linked to  $a_w$  variations within the colony, IR spectra were collected on a lower spectral window encompassing both the EPS and water librational mode frequency domains. This was made possible by the use of a Si:B bolometer, which is much more sensitive in this low frequency domain than the MCT detector.



**Figure 6**. Mean MIR spectrum (2000-400 cm<sup>-1</sup>) and difference spectra (inset) of a currently swarming colony. The figure displays the average (n=57) spectrum while the inset displays the corresponding dynamic spectra. The broad band located at 615 cm<sup>-1</sup> features the water librational mode; the dynamic spectra emphasize the variations of absorbance in this domain.



**Figure 7**. PCA of MIR spectra (2000-400 cm<sup>-1</sup>) collected during a swarming phase. Spectra were collected with the NicPlan microscope. A first PCA (A, B) was performed by using the whole spectral domain (2000-400 cm<sup>-1</sup>). A second PCA (C, D) was performed on the sole water absorption domain (900-400 cm<sup>-1</sup>). A and C: migration, dots; PC1 scores, solid line.

Fig. 6 shows the mean spectrum and the amplitude variations of the data set collected, as previously, during a full swarming cycle. Besides variations in the EPS spectral domain, significant variations are also observed in the spectral range that corresponds to the water librational mode, *i.e.* from 900 to 400 cm<sup>-1</sup>. To correlate EPS auto-organization observable between 2000 and 900 cm<sup>-1</sup> with a<sub>w</sub> variations observable from the 615 cm<sup>-1</sup> water librational band, a first PCA (Fig. 7A, B) was performed by using the whole spectral domain (2000-400 cm<sup>-1</sup>) which again reflects major changes in the EPS absorption domain as shown in Fig. 7B.

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To emphasize spectral variations of the water librational mode, a second PCA (Fig. 7C, D) was performed on the sole water absorption domain (900-400 cm<sup>-1</sup>) which demonstrates that during active swarming (here from 11h30 to 14h30), IR spectra exhibit positive PC1 loadings which correspond to low absorbencies at the low frequency band tail, and hence low a<sub>w</sub>. Indeed, the corresponding PC1 loadings (Fig. 7D) show that during swarming, the water band loses intensity over its lower energy side. This spectral evolution has been reported (see Fig. 2C in <sup>15</sup>) to reflect an increased crystallinity or, in other words, a larger H bonds connectivity. The data shown are from one representative experiment which has been reproduced on three distinct colonies. One must note that the PC1 score variation in the 900-400 cm<sup>-1</sup> domain is an order of magnitude less than one observed in the 2000-400 cm<sup>-1</sup> one.

To investigate the causality between the changes in EPS H bond networks and water activity, a 2D correlation analysis was performed<sup>20</sup>. The corresponding synchronous 2D correlation map (Fig. 8A) indicates that the spectral envelope between 950 and 1050 cm<sup>-1</sup> decreases as the 500 to  $800 \text{ cm}^{-1}$  domain increases. Indeed, in a 2D synchronous map, negative cross peaks reflect bands that evolve in an opposite manner during the process<sup>20</sup>. Information about the time sequence of these spectral variations can be derived from 2D asynchronous maps. The Fig. 8B shows a part of the upper left corner of the corresponding 2D asynchronous map. The negative cross peak centred at  $1000 / 700 \text{ cm}^{-1}$  allows to establish that the EPS vibrational bands are altered after the water ones<sup>20,21</sup>; hence, changes in the water H bond networks at the colony's edge precede any EPS auto-organization.



Figure 8. 2D IR synchronous (A) and asynchronous (B) maps.

Knowing that polysaccharides solutions allow forming a percolation network, the subsequent sharp increase of the elastic modulus must be linked to the periodic ability of the colony to swarm and consolidate. In this scheme, consolidation onset must be triggered by the connectivity transition, which appears in the abrupt increase of the elastic properties shown in Fig.3, which efficiently impairs swarming.

A wide infrared transparency window combined with the brilliance of the IR synchrotron beam at SOLEIL has allowed us, for the first time, to simultaneously monitor, in a non-invasive way and real time, the water H bond network and the EPS auto-organization within a currently swarming colony. Statistical data analysis (PCA and 2D correlation spectroscopy) have allowed

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establishing that  $a_w$  periodic variations and the resulting water H-bonding alterations do periodically occur and trigger EPS auto-organization. The progressive decrease of bulk water molecules during a swarming phase promotes additional elastic constrains due to the onset of a percolation network. Eventually, the inability of bacterial cells to keep swarming will trigger dedifferentiation to enter a new consolidation phase. Hence, the extensive correlation which links individual cells over large distances originates in a global property of the multicellular entity. The periodic water gradient at the colony's edge associated to volumetric variations of the colony which spreads from a "thick" (> 45 µm) consolidation terrace to a "thin" (16 µm) swarming film warrants for the synchrony of swarming. This exemplifies how biological properties and physical laws interplay: the bacterial cells synthesize a mixture of exoproducts (EPS, PGL, GB) whose structure is tuned to undergo marked transitions as passing over a given threshold. The *P. mirabilis* exoproducts differ whether the strain is grown in a liquid culture or on a solid medium<sup>8</sup>; this further supports that the ability of bacterial populations to cope with distinct environments tightly relies on the behaviour of such macromolecules.

In conclusion, the periodic and synchronous population behaviour is not controlled by a chemical trigger, but rather derives from the efficiency of supramolecular assemblies to respond to an external stimulus yielding thereby a high structural order over large dimensions which here has been characterized as a percolation network. These results concerning the *P. mirabilis* biologic oscillator are of broad significance since they demonstrate how the supramolecular behaviour of macromolecules promotes complex embedded dynamic cycles which root the evolution from unicellularity to multicellularity and, eventually, to pluricellularity : the permanent osmotic force allows polysaccharides self-organization to promote cyclic dissipative structures.

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ASSOCIATED CONTENT

**Supporting Information**. 2D synchronous and asynchronous full color maps are available on the journal web site

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# **Author Contributions**

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript. ‡These authors contributed equally.

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### **ABBREVIATIONS**

a<sub>W</sub>: water activity; ECM: extra cellular matrix; EPS: exopolysaccharides; ATR-FTIR: attenuated total reflexion-Fourier transform infrared; GB: glycinebetaine; LB: Luria Bertani; PGL: phenoglycolipid; SEM: Scanning Electron Microscopy; SVR: surface to volume ratio.

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A multi-scale approach of the mechanisms underlying exopolysaccharides auto-organization in the *Proteus mirabilis* extracellular matrix

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